

Article

Industrial Hemp Finola Variety Photosynthetic, Morphometric, Biomechanical, and Yield Responses to K Fertilization Across Different Growth Stages

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Abstract: The growing interest in *Cannabis sativa* as a highly used crop is present worldwide. There are limited data about the effect of potassium (K) fertilizer on industrial hemp yield for dual purposes (seed and stem production). The current study aimed to investigate the influence of adding two different K fertilizers, KCl and K₂SO₄, at two growth stages (flowering and ripening) on the productivity and chlorophyll *a* fluorescence (ChlF) of *Cannabis sativa*, variety Finola. Before sowing, different K treatments were applied: K₁—100 kg ha⁻¹ KCl (60% K) and K₂—100 kg ha⁻¹ K₂SO₄ (52% K, S 17%). The OJIP (O stands for “origin” (minimal fluorescence), P for “peak” (maximum fluorescence), and J and I for inflection points between the O and P levels) data were recorded and used for ChlF transients and individual ChlF parameters during vegetation. At harvest, the stem morphology parameters and yield (plant height, stem weight and diameter, and stem and seed yield), tensile strength, and the modulus of elasticity were determined. The results show the sensitivity of minimal (F₀) and maximal fluorescence (F_m), electron transport from Q_A to intersystem electron acceptors (ET₀/(TR₀ – ET₀)), and electron transport flux until PSI acceptors (RE₀/RC) to K fertilization. The parameters that described electron transport (ET₀/RC, ψE₀, and φE₀), performance index on absorption basis (PI_{ABS}, TR₀/DI₀, and φP₀), dissipation (DI₀/RC), and electron transport to photosystem I (φR₀ and δR₀/(1 – δR₀)) had a reaction only at the growth stage, indicating a change in their activity during the aging of the *Cannabis sativa* plants. The average stem height was 67.5 cm, and the stem diameter was 0.41 cm. The different K sources did not significantly influence the stem height and diameter, nor the dry stem (on average 12.2 t ha⁻¹) and seed yield (on average 1.85 t ha⁻¹). The tensile strength of individual hemp stems was the highest with K₂SO₄ (53.32 MPa) and the lowest with KCl (49.25 MPa). The stem stiffness by modulus of elasticity was about 5 GPa on average for all the treatments. In general, the photosynthetic parameters in this study varied more between the growth stages than between the different K fertilizer formulations. Moreover, based on the results of this study, it can be recommended to use both fertilizers, KCl and K₂SO₄, in dual-purpose industrial hemp production since no significant effect was found for the stem morphometric and biomechanical parameters as well as for the agronomic parameters.



Academic Editor: Tie Cai

Received: 14 January 2025

Revised: 15 February 2025

Accepted: 17 February 2025

Published: 19 February 2025

Citation: Varga, I.; Markulj Kulundžić, A.; Krolo, P.; Iljkić, D.; Tišma, M.; Kraus, I. Industrial Hemp Finola Variety Photosynthetic, Morphometric, Biomechanical, and Yield Responses to K Fertilization Across Different Growth Stages. *Agronomy* **2025**, *15*, 496. <https://doi.org/10.3390/agronomy15020496>

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Keywords: potassium fertilizer; *Cannabis sativa* L.; dual-purpose; chlorophyll *a* fluorescence; physiology; mechanobiology

1. Introduction

Industrial hemp is considered one of the oldest non-food crops used for fiber production [1–3]. The large hemp producers globally are Chile, France, the USA, and Canada. In Europe, in 2022, the areas with industrial hemp cultivation for seed production totaled around 8600 ha and for raw hemp stem, around 37,000 [4]. France is the largest producer, accounting for more than 60% of EU production, followed by Germany (17%), and the Netherlands (5%) [5]. Industrial hemp cultivation and processing were very popular during the 20th century, especially in Eastern Croatia, in the Slavonia and Baranja areas, and in Serbia, in the Vojvodina area [6,7]. Around 1000 ha is sown in Croatia, mainly for the purpose of seed production [8].

Unlike other crops, industrial hemp is somewhat controversial because some varieties of *Cannabis* genera are grown in a specific way and used to obtain psychoactive substances, which is prohibited by the law. The plant contains cannabinoids, of which Δ^9 -tetrahydrocannabinol (THC) is the most important. Thus, nowadays, renewed interest in industrial hemp cultivation is subject to certain regulations. The permitted THC content for industrial hemp varies from country to country. Due to the THC content, special regulations are in force, which mandate that the variety of industrial hemp being cultivated must have a THC content below 0.3% in Europe for hemp crops used for industrial purposes [9]. In the UK, the maximum content is up to 0.2% THC, and in New Zealand, it can vary from 0.35% to 0.50% [9]. According to Kaur et al. [10], a higher content is allowed in Australia and Mexico (1.0%), Malaysia (0.5%), and Canada (0.3%). In Croatia, the regulation [11] mandates that industrial hemp can be cultivated for food production if the THC content does not exceed 0.2%.

A major advantage of the increased interest in the re-cultivation of industrial hemp in Eastern Croatia is the change in the regulations, which now allow the use of whole plants, not just seeds [11,12]. These regulations offered an advantage that opened up many opportunities for the use and processing of stems and other plant parts not only for the food industry [13–15] but also in medicine, as an environmentally friendly material in numerous other processing branches, and as a raw material for bioethanol production [16–21]. Industrial hemp has a wide range of applications in sectors such as the automotive, paper, and textile industries, etc. Its seeds are rich in oil (25–38%), proteins (18–23%) [22], and essential fatty acids, with a high ratio of polyunsaturated to saturated fatty acids [23–25]. Industrial hemp has great potential as a renewable and environmentally friendly building material [26,27].

Macronutrients (N—nitrogen, P—phosphorus, K—potassium) are of great importance for field crops and are usually added to the soil [28–31]. A positive response of industrial hemp to nitrogen fertilization has been previously recognized, not only for biomass production but also for increased seed yield and plant height [32–34]. However, Struik et al. [35] reported that a relatively small amount of nitrogen is sufficient to meet the plants' nitrogen requirements. Research on P and K fertilization in industrial hemp is still limited. Ahmadi et al. [36] reported that P increases plant height, directly influencing plant biomass production and improving flowering. For industrial hemp cultivation, K is important for prolonging the flowering of male plants [37]. Zehler et al. [38] stated that K often depends on how it is combined in the fertilizer, which may influence crop yield and quality.

The most common formulation of K fertilizers is potassium chloride (KCl), followed by potassium sulfate (K_2SO_4). Even though chloride is an essential element, it can be used in small quantities, similarly to micronutrients. It is involved in osmosis and ionic balance within the cell [39]. Sulphate is a major plant nutrient since it is a component of proteins [40]. Both Cl^- and SO_4^{2-} regulate plant water uptake, but Cl is more hydrated and, therefore, has a greater swelling effect than SO_4^{2-} . Cl reduces transpiration, increases stomatal opening, and increases water uptake by the plants [38].

Very few studies have been published that have examined the yield of industrial hemp stem and seed in relation to photosynthetic activity parameters. As a short-day plant, hemp requires a photoperiod of 12–14 h of daylight [36]. Zehler et al. [38] reported that in higher doses of Cl, photosynthetic activity is lower due to lowered chlorophyll content, and on the contrary, SO_4^{2-} in plants is concerned with chlorophyll synthesis.

Recently, the importance of stem and leaf mechanobiology has accelerated, not only due to harvest and processing but also in breeding programs and crop loss due to lodging and stem deformation [41]. The tensile strength and stem stiffness are closely connected to plant species, stalk diameter, maturity, stem moisture, and cellulase content [42–44]. Due to their tensile strength and stiffness, industrial hemp fibers and stems can be used as reinforcements for composite materials. Liu et al. [45] investigated the influence of the industrial hemp plant's growth stage. However, research on the influence of K fertilizer treatment on the stiffness and tensile strength of industrial hemp stems is limited.

The cultivar Finola, which is used in this research, originates from Finland and is dioecious and auto-flowering. Lančaričová et al. [24] indicate that Finola needs fewer than 100 days to reach the maturity stage, and its buds contain low levels (<0.2%) of THC. The authors stated that it was the first cultivar registered as an oilseed crop in Europe. In Croatia, the Finola cultivar is the most common variety nowadays, mostly for seed production. Still, the potential of Finola utilization is not fully recognized. In Croatia, stem use was forbidden until 2019 [12] (OG 39/2019).

The aim of this study was to evaluate the dual-purpose production yield (stalk and seed yield) of industrial hemp of the Finola variety in eastern Croatia. In addition, since there are limited data on the influence of K fertilization on industrial hemp Finola cultivar yield production, the effect of the most common K fertilizers (KCl or K_2SO_4) on photosynthetic activity during growth and stem mechanobiology (stiffness and tensile strength), morphology parameters, and finally on the stem and seed yield was determined.

2. Materials and Methods

2.1. Field Trial

The field trial was conducted at the Tenja location in Eastern Croatia. The pre-crop was an opium poppy, and the basic cultivation was carried out to a depth of 25–30 cm during November 2020. In the spring of 2022, the winter furrow was closed, after which further supplementary soil cultivation was started to create an optimal seeding layer. Fertilizer was applied before sowing to avoid leaching of the Cl^- and SO_4^{2-} ions in the winter period. Presowing fertilization was conducted with different K fertilizers such as K_0 —without presowing fertilization (control treatment), K_1 —100 kg ha^{-1} KCl (60% K), and K_2 —100 kg ha^{-1} K_2SO_4 (52% K, S 17%). Other macronutrients were not added to exclude their influence on the obtained results.

For this study, the Finola variety was sown on 17 April 2022 in three replications in a randomized block design. A pneumatic seed drill was used for the industrial hemp sowing at an inter-row distance of 25 cm and a depth of 3 cm. The sowing rate was 30 kg ha^{-1} .

All weeds were manually removed from the field approximately every ten days. Even though the experiment was set up using non-organic practices, no agrochemicals were

used since there were no important pests or disease attacks, and there was no need for pesticide application.

2.2. Weather Data

At the time of sowing in April, according to the Croatian Meteorological and Hydrological Service [46], according to the distribution of percentiles, the temperature conditions in Osijek for April were described as cold (Figure 1), but the industrial hemp germination was satisfied. During May, there were no major deviations in temperature and precipitation. June 2022 was extremely warm throughout Croatia, and for the Osijek area, the average temperature in June was 3.2 °C lower than the long-term mean, while the amount of precipitation was average. The average air temperature in July 2022 was 1.8 °C higher as compared to the long term mean 1981–2010 for the Osijek area, with a lack of precipitation. Such weather conditions during the summer months have an unfavorable effect on the growth of industrial hemp, and the plant relatively quickly went into the flowering stage (generative stage).

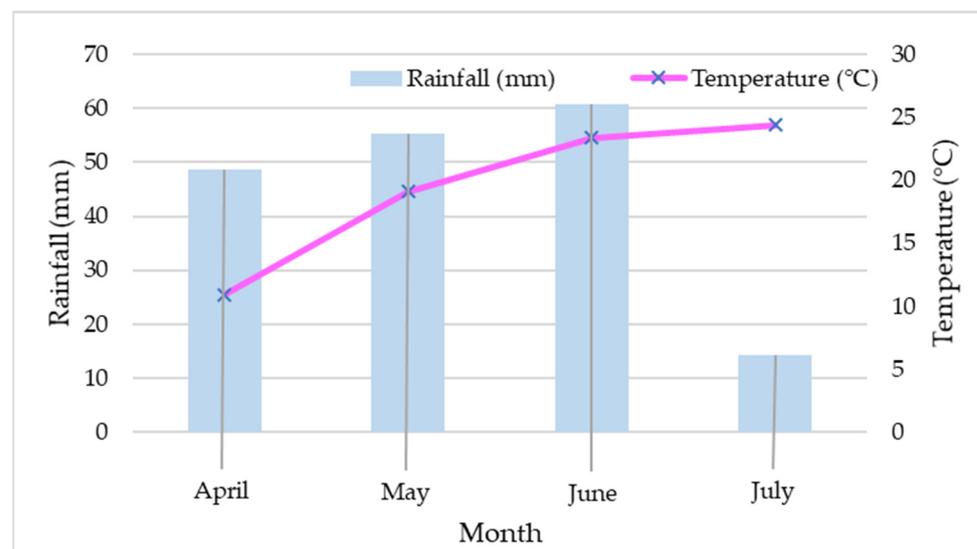


Figure 1. Weather data for industrial hemp during the 2022 vegetation period (Osijek location).

2.3. Measurements of Photosynthetic Activity and Chlorophyll Content

The fluorescence was analyzed using a portable chlorophyll fluorescence instrument, Handy PEA (Hansatech Instruments Ltd., King's Lynn, UK). Chlorophyll *a* fluorescence was assessed on the adaxial surface of the leaves at the base of the inflorescence of the main stem (fully grown leaf) of the *Cannabis* plant (Figure 2), subjecting the plants to a period of 30 min of darkness before measurement [47]. All measurements were conducted in the morning, between 7:00 and 8:00 a.m. Photosynthetic activity was determined in two plant growth stages, according to Mediavilla et al. [48]. The first measurement was on 5 July in the flowering stage, code 2202 (50% of bracts formed), and the second measurement was on 15 July in the seed maturity stage—ripening stage, code 2204 (50% of the seeds were hard). From each fertilization treatment, 10 plants were measured, and a total of 60 plants were measured in every growth stage. ChlF was measured after applying a saturating light pulse $> 3500 \mu\text{mol m}^{-2} \text{s}^{-1}$ for one second (s). The ChlF parameters (Table 1) were calculated following the methodology described by Strasser et al. [49] and Yusuf et al. [50]. Specific events of OJIP transient in the OK, OJ, JI, OI, JI, and IP phases were calculated and presented as different ΔV_{OP} , ΔV_{OK} (L band), ΔV_{OJ} (K band), ΔV_{OI} (J band), ΔV_{JI} (H and I bands), and ΔV_{IP} (G band) normalized to the controls (K0 treatment) per growth stage [50].



Figure 2. Photosynthetic activity measurements and leaf clip position for industrial hemp.

Table 1. List of chlorophyll *a* fluorescence parameter used in the analyses.

Parameter Label	Description of Chlorophyll <i>a</i> Fluorescence Parameters
F_0	Minimal fluorescence yield of the dark-adapted state
F_m	Maximal fluorescence yield of the dark-adapted state
ABS/RC	Absorption flux per active reaction center (RC)
DI_0/RC	Dissipation flux per active RC
TR_0/RC	Trapping flux per active RC
ET_0/RC	Electron transport flux per active RC
RE_0/RC	Electron flux reducing terminal electron acceptors at the photosystem I (PSI) acceptor side per RC
ϕP_0	Maximal photochemical quantum yield
ψE_0	The probability that a trapped exciton moves an electron into the electron transport chain beyond Q_A
ϕE_0	Quantum yield for electron transport
δR_0	The probability that an electron is transported from reduced PQ to the electron acceptor side of PSI
ϕR_0	The quantum yield of electron transport from Q_A^- to the PSI end electron acceptors
RC/ABS	Density of RC on chlorophyll <i>a</i> basis
TR_0/DI_0	Flux ratio trapping per dissipation
$ET_0/(TR_0 - ET_0)$	Electron transport from Q_A^- to intersystem electron acceptors
PI_{ABS}	Performance index on absorption basis
$\delta R_0/(1-\delta R_0)$	Electron transport from PQH ₂ to final PSI acceptors
PI_{TOTAL}	Performance index for energy conservation from exciton to the reduction in PSI terminal acceptors

Leaf chlorophyll content was determined with a chlorophyll content meter, model CL-01 (Hansatech Instruments, Pentney, UK).

2.4. Stem and Seed Harvest and Post-Harvest Measurements

The harvest was conducted on 26 July 2022. Whole plants were harvested by hand from each fertilization treatment and replicated from 2m² in three replications. At that time, the stem of the female plants was still green and inseeded. Female industrial hemp plants were light green to golden brown. Before harvesting the plants, the number of plants per unit area was determined, and then the proportion of male and female plants was determined. After the harvest, the yield (t ha⁻¹) of the industrial hemp stem (fresh

and air-dried stems) and seed yield (t ha^{-1}) was determined. The plants were dried, and after that, the stem diameter was determined.

From each treatment, ten individual plants were taken for measuring of the morphometric parameters. Stem height (cm) was measured with a ruler, representing the height of the whole plant, including the inflorescence. Stem diameter (cm) was determined by measuring 2 cm above and beyond each nodule measured with digital vernier calipers. The stem diameter represents the average of all measurements of the stem.

2.5. Macronutrient Stem Status

The stem for macronutrient status analysis was dried to a constant mass at $105\text{ }^{\circ}\text{C}$. After drying, the stems were milled in the laboratory mill (Retsch SM 100, Haan, Germany). This study determined three macronutrients: N, P, and K. A C/N analyzer was used to determine the N, P, and K macronutrient status after digestion with sulfuric acid and hydrogen peroxide and afterward was determined with AAS spectrometry for K. The P was determined using photometry at room temperature at wavelength 400 nm.

2.6. Tensile Testing of Industrial Hemp Stems

Stem tensile tests were conducted to determine the effects of the two types of K fertilizers, KCl and K_2SO_4 , on stem tensile strength and stiffness. The impact of fertilization on hemp stems has been compared to stems that have not been fertilized. Three sets of three industrial hemp samples were prepared for testing. The center part of the industrial hemp stem, which was bounded by the nodes, was chosen for the stem samples. The first group of samples was steamed without presowing fertilization (K_0); the second group of samples was fertilized with potassium chloride (KCl, K_1); and the third group of samples received potassium sulfate (K_2SO_4 , K_2). The samples are labeled x-y-z, with the first mark x indicating industrial hemp (IH) and the second mark y indicating K fertilization: K_0 , K_1 , and K_2 . The third mark, z, indicates the number of samples, which can range from one to three for each group of samples. The tests were conducted on stems with a complete cross-section. Before the test, the sample's length, external diameters at three points (ends and middle), and straw wall thickness (ends) were measured, and the average cross-sectional areas were calculated.

The sample was loaded under uniaxial tension using the Zwick/Roell Z600 material testing device (Ulm, Germany) to determine the maximum breaking force for the hemp stems. TestXpert II software was used for both the test management and data registration. Mechanical jaws with rubber inserts were used to fix the samples with capacities up to 10 kN and a load cell with a capacity of 50 kN. Figure 3 shows the test setup. The tension in the samples was achieved by controlling the displacement of a moving crosshead at a test speed of 0.5 mm/min. Testing is considered finished when the tensile strength drops to 80%. The moving crosshead's axial forces and displacements were measured and recorded during the test. The stresses and strains in the specimen were calculated using the measured forces and displacements.

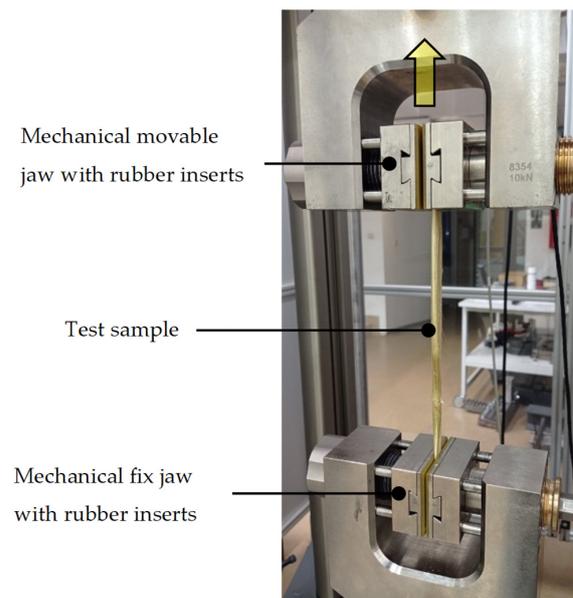


Figure 3. The tensile test setup of industrial hemp stem.

2.7. Statistical Analysis

The influence of different K treatments was tested with ANOVA in SAS Enterprise Guide 7.1. Differences between the means were shown at the $p < 0.05$ probability level. Mean values and standard deviations of ChlF parameters (relative units) are shown in Table 2. A factorial ANOVA for the ChlF parameters ($n = 60$) was used to determine the statistical differences between the fertilization treatments (K_0 , K_1 , and K_2) and the developmental stage (flowering and ripening) of the industrial hemp plants. Factorial ANOVA was followed by the Tukey post hoc honestly significant difference (HSD) test at $p < 0.05$. Correlation analysis was conducted for the stem morphometric parameters, seed and stem yield, stem stiffness and tensile strength, and macronutrient status in the stem dry matter. Pearson's coefficient of correlation was used to determine the correlations.

Table 2. Chlorophyll *a* fluorescence (ChlF) parameters for KCl and K_2SO_4 fertilization treatments in the flowering and ripening stage of *Cannabis* plants.

* ChlF Parameters	Flowering Stage			Ripening Stage			HSD _{GS}	HSD _{FT}	HSD _{GS × FT}
	Control (K_0)	KCl	K_2SO_4	Control	KCl	K_2SO_4			
F_0	598 ± 58	600 ± 51	624 ± 26	658 ± 110	501 ± 45	596 ± 77	ns	42.50	60.11
F_m	3958 ± 93	4021 ± 54	4036 ± 137	3811 ± 288	3081 ± 472	3554 ± 503	162.33	198.81	281.17
ABS/RC	2.103 ± 0.183	2.114 ± 0.269	2.136 ± 0.169	2.239 ± 0.228	1.926 ± 0.209	2.198 ± 0.369	ns	ns	ns
DI ₀ /RC	0.318 ± 0.045	0.318 ± 0.066	0.332 ± 0.044	0.389 ± 0.083	0.318 ± 0.054	0.381 ± 0.132	0.04	ns	ns
TR ₀ /RC	1.784 ± 0.147	1.796 ± 0.204	1.804 ± 0.127	1.850 ± 0.163	1.608 ± 0.163	1.817 ± 0.252	ns	ns	ns
ET ₀ /RC	0.964 ± 0.055	0.945 ± 0.049	0.928 ± 0.038	0.803 ± 0.081	0.780 ± 0.059	0.778 ± 0.033	0.03	ns	ns
RE ₀ /RC	0.395 ± 0.068	0.335 ± 0.083	0.321 ± 0.057	0.350 ± 0.064	0.341 ± 0.041	0.322 ± 0.049	ns	0.04	ns
ϕP_0	0.849 ± 0.012	0.851 ± 0.012	0.845 ± 0.009	0.828 ± 0.023	0.836 ± 0.015	0.830 ± 0.029	0.01	ns	ns
ψE_0	0.542 ± 0.039	0.531 ± 0.054	0.516 ± 0.030	0.436 ± 0.046	0.488 ± 0.045	0.434 ± 0.048	0.03	ns	ns
ϕE_0	0.461 ± 0.037	0.453 ± 0.050	0.436 ± 0.029	0.361 ± 0.045	0.408 ± 0.041	0.361 ± 0.047	0.02	ns	ns
δR_0	0.410 ± 0.069	0.353 ± 0.082	0.346 ± 0.058	0.435 ± 0.053	0.437 ± 0.039	0.414 ± 0.059	0.03	ns	ns
ϕR_0	0.191 ± 0.043	0.162 ± 0.049	0.152 ± 0.030	0.158 ± 0.033	0.178 ± 0.024	0.149 ± 0.024	ns	ns	ns
RC/ABS	0.479 ± 0.044	0.480 ± 0.057	0.471 ± 0.036	0.451 ± 0.046	0.525 ± 0.058	0.466 ± 0.072	ns	ns	ns
TR ₀ /DI ₀	5.657 ± 0.500	5.739 ± 0.517	5.480 ± 0.376	4.884 ± 0.721	5.130 ± 0.584	5.011 ± 0.858	0.32	ns	ns
ET ₀ /(TR ₀ − ET ₀)	1.199 ± 0.182	1.158 ± 0.236	1.072 ± 0.124	0.783 ± 0.143	0.967 ± 0.178	0.778 ± 0.147	0.09	0.11	ns
PI _{ABS}	3.316 ± 0.970	3.309 ± 1.166	2.807 ± 0.652	1.782 ± 0.626	2.670 ± 0.895	1.911 ± 0.737	0.45	ns	ns
$\delta R_0/(1 - \delta R_0)$	0.717 ± 0.207	0.566 ± 0.179	0.538 ± 0.124	0.783 ± 0.165	0.783 ± 0.129	0.721 ± 0.164	0.09	ns	ns
PI _{TOTAL}	2.499 ± 1.274	2.001 ± 1.114	1.537 ± 0.524	1.439 ± 0.708	2.082 ± 0.744	1.330 ± 0.551	ns	ns	ns

* ChlF parameters—chlorophyll *a* fluorescence parameters; all ChlF parameters abbreviations are described in Table 1. Tukey HSD test _{GS}—Tukey honestly significant difference test for growth stage, Tukey HSD test _{FT}—Tukey honestly significant difference test for fertilization treatment, Tukey HSD test _{GS × FT}—Tukey honestly significant difference test for interaction between growth stage and fertilization treatment; ns—not significant.

3. Results and Discussion

Plants need the availability of essential micro- and macro-elements for normal growth and development and for maintaining physiological processes, including photosynthesis [51]. In this study, the *in vivo* chlorophyll *a* fluorescence transient was analyzed, by which the change in the light stage of photosynthesis in *Cannabis* plants with the presowing application of KCl and K₂SO₄ fertilizers was analyzed. The plants were grown in a field to gain insight into the real effect of KCl and K₂SO₄ fertilizers on the photosystem II (PSII) and photosystem I (PSI) functions based on the ChlF transients and parameters.

3.1. Chlorophyll *a* Fluorescence Transients

The OJIP transients from the control industrial hemp leaves showed a typical polyphase rise with the basic steps of O-J-I-P. Both OJIP transients from the K fertilization *Cannabis* leaves showed deviations in steps I and J (Figure 4). Difference kinetics ΔV showed positive and negative trends depending on the K fertilization (Figure 4b–h). The L band and K band were positive in the flowering stage of the industrial hemp development. With the aging of the industrial hemp plants (ripening stage), the L and K bands became negative, with a greater pronounced influence when KCl (K₁) was applied. In the flowering stage of the industrial hemp, K fertilization reduces the energy connection of the PSII units (positive L band) [52].

On the other hand, at the ripening stage, the negativity of the L band indicates the opposite, which is why it can be said that the excitation energy was efficiently utilized, since the PSII units are more connected and better stability is created [50], which in this case was more pronounced when using KCl (K₁). Furthermore, K in flowering affected the deactivation of the oxygen-evolving complex and the increase in the size of the functional PSII antenna, as indicated by the positivity of the K band.

In this study, at the flowering stage, K affected the deactivation of the oxygen-evolving complex and the increase in the size of the functional PSII antenna, indicated by the positivity of the K band. However, the effect of K at the ripening stage with the occurrence of a negative K band is due to the faster transport of electrons on the donor side and/or the slower withdrawal of electrons from the acceptor side [53]. Also, in the I, H, and G bands, the curves behave the same way according to the stages of the industrial hemp development. The above bands provide insight into PSI's functioning and the reoxidation of the PQ pool from the PSI carrier. In the flowering stage, the I band is positive except for K₂SO₄ at the ripening stage. Changes from positive and negative bands and vice versa occurred at the ripening stage of the industrial hemp with a different reaction to KCl (K₁), and K₂SO₄ (K₂) fertilization. The influence of K fertilization can be seen most strongly in the H band. K₂SO₄ had a greater influence on the H band both at the flowering and ripening stages of the industrial hemp (greater deviations were created compared to the KCl treatment). The positivity of the H band indicates a decrease in PQ pool capacity and a higher reduction rate. On the other hand, the negative band represents an increase in the PQ pool [54].

In the case of the G band, in addition to the differences between the growth stages, a difference was found based on the applied fertilizer formulation, which caused a decrease in the PSI acceptor pool with reduced electron transport [53,55].

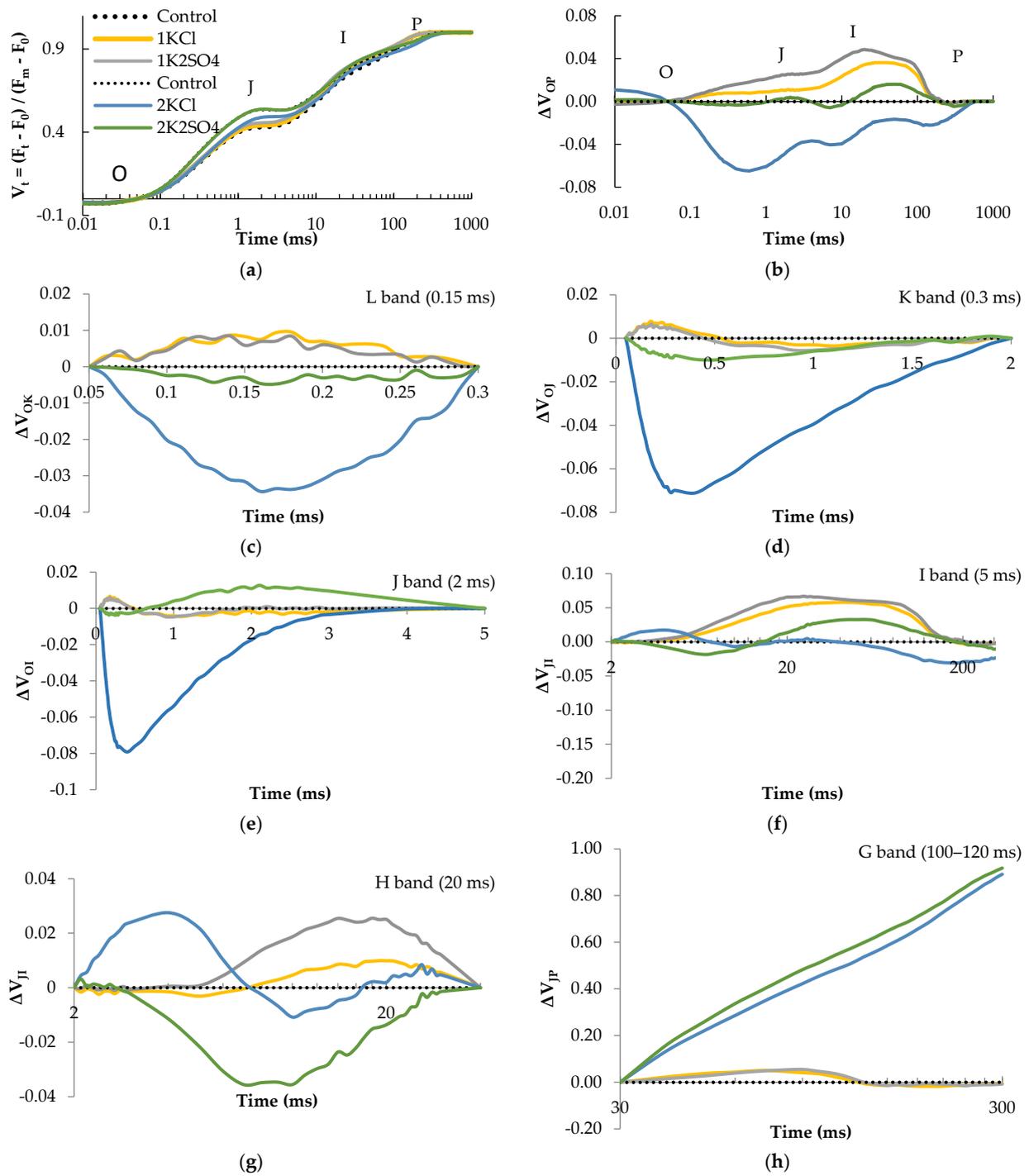


Figure 4. Expression of *Cannabis* plant response to KCl and K₂SO₄ soil fertilization in two growing stages (1—flowering; 2—ripening stage) are presented as kinetics of relative variable fluorescence V_t and as difference kinetics ΔV_{OP} (a,b). Difference kinetics ΔV_t , for individual bands, L (c), K (d), J (e), I (f), H (g), and G (h) are plotted at different time ranges. Each curve represents the average of ten measurements ($n = 10$) per treatment and growth stage. The control values (treatment without fertilization) per growth stage were used as referent values. The O, J, I, and P steps are indicated in V_t and ΔV_{OP} curves.

3.2. Chlorophyll a Fluorescence Parameters

Two-factorial ANOVA was used to determine the significance of the sources of variability: the growth stages (flowering and ripening), the fertilization treatments (K₀, K₁, and K₂), and the interactions between the growth stages and fertilization treatments. The

response of the chlorophyll *a* fluorescence parameters in the industrial hemp leaves based on the different K fertilizer doses at the two growth stages is presented in Table 2.

Significant minimum initial fluorescence (F_0) differences were confirmed only for the KCl (K_1) treatment. The control (K_0) and K_2SO_4 (K_2) groups were not statistically different, but KCl showed a trend of being 12.33% lower than the K_0 treatment. The interaction between the growth stages and fertilization treatment was found to be significant for the ripening stage when KCl fertilizer was applied and, compared to the K_0 and K_2SO_4 (K_2), the KCl (K_1) treatment affected F_0 by lowering the value, especially at the ripening stage. Then, it was 23.8% lower compared to the controls in the ripening stage. Generally, fertilization with K lowered the F_0 values in both growth stages compared to the K_0 treatment. This is also in accordance with the research of Kusaka et al. [56], which found that a lack of K causes an increase in the F_0 value in *Raphanus sativus* var. *sativus*.

Maximum fluorescence (F_m) varied significantly in response to the growth stage and fertilization treatment. The ripening stage and KCl (K_1) treatment were statistically different. The highest F_m value was determined in the flowering growth stage when applying K_2SO_4 (K_2), which was 4036 (relative units). The interaction revealed a significant difference in KCl (K_1) at the ripening stage, with the lowest F_m being 26.7% lower than in the K_0 treatment. In the flowering of the industrial hemp, the treatments with K increased the F_m , but the F_m value decreased at the ripening stage, which, according to Kalaji et al. [57], indicates the accumulation of inactive RC on PSII. Kusaka et al. [56] achieved lower F_m values in younger and older radish leaves due to K deficiency. The behavior of the F_0 and F_m values in this study confirmed that the industrial hemp plants, in both fertilization variants, KCl and K_2SO_4 , were sufficiently supplied with K. Hence, their values decreased under the influence of K.

The logical sequence of the behavior of F_0 and F_m in the fertilization treatments and growth stages also confirmed the significance of $ET_0/(TR_0 - ET_0)$ for the growth stage and treatment. These two parameters exactly determine $ET_0/(TR_0 - ET_0)$, i.e., electron transport from Q_A to the intersystem electron acceptors [49]. The difference between the KCl and K_2SO_4 treatments was 12.98%, with the highest values being seen in the KCl treatment. On the other hand, at the ripening stage, $ET_0/(TR_0 - ET_0)$ decreased by 26.25%.

Electron transport flux to the PSI acceptors (RE_0/RC) was confirmed to have a statistically significant difference in the fertilization treatment as a source of variability. The value of RE_0/RC decreased with the application of KCl by 9.4% and 13.7% with the application of K_2SO_4 . Also, reduced values of RE_0/RC were found in K deficiencies during the cultivation of radish [56]. The decrease in RE_0/RC that occurred due to the presence of K in the soil, which the plant adopted, describes the impaired movement of electron transport from Q_A^- to the end electron acceptors on the PSI acceptor side.

The absorption flux per active RC (ABS/RC), trapping flux per active RC (TR_0/RC), quantum yield of electron transport from Q_A^- to the PSI end electron acceptors (ϕR_0), density of RC on chlorophyll *a* basis (RC/ABS), and the performance index for energy conservation from the absorption all the way to the reduction in PSI end acceptors (PI_{TOTAL}) were not significant, regardless of the sources of variability. The mentioned parameters showed stability regardless of the testing conditions and performed their activities normally, i.e., were undisturbed. Although PI_{TOTAL} is known as the most sensitive ChlF parameter since it includes the most important functional steps of primary photochemistry and, hence, the vitality of photosynthetic units [50], this study did not show that PI_{total} was not affected by K fertilization or growth stage of industrial hemp. Researching nutrient deficiency in maize revealed that K deficiency did not significantly affect any of the tested photosynthetic ChlF parameters [58].

Furthermore, the treatments did not show significance, unlike the growth stage, which was significant for a maximum quantum yield of primary efficiency (ϕP_0), flux ratio trapping per dissipation (TR_0/DI_0), the performance index for energy conservation of the reduction in intersystem electron acceptors (PI_{ABS}), electron transport flux per active RC (ET_0/RC), the probability that a trapped exciton moves an electron into the electron transport chain beyond Q_A (ψE_0), quantum yield for electron transport (ϕE_0), dissipated energy flux per active RC (DI_0/RC), the probability that an electron is transported from the reduced PQ to the electron acceptor side of the PSI (δR_0), and electron transport from PQH_2 to the final PSI acceptors ($\delta R_0/(1 - \delta R_0)$). Kalaji et al. [58] also found no difference in ChlF parameters in K application to greenhouse-grown maize in nutrient solution. Also, a study of responses to nitrogen fertilization in *Cannabis sativa* suggests that although N affected the leaf N and chlorophyll content, the fluorescence parameters evaluated under field conditions remained unchanged [59].

The results of this study showed that the *Cannabis* plants had different values of ϕP_0 , TR_0/DI_0 , and PI_{ABS} according to the treatments, which did not show significant differences compared to the controls (K_0). However, the ET_0/RC , ψE_0 , ϕE_0 , ϕP_0 , TR_0/DI_0 , and PI_{ABS} parameters had higher values in the flowering stage, which was significant. ET_0/RC was lower by 16.84%, ψE_0 by 14.43%, ϕE_0 by 16.22%, ϕP_0 by 2.01%, TR_0/DI_0 by 10.97%, and PI_{ABS} by 32.54% compared to the flowering stage. This means that the activities of these parameters decrease with aging in industrial hemp plants. Also, DI_0/RC , δR_0 , and $\delta R_0/(1 - \delta R_0)$ showed statistical differences per growth stage, but their behavior was contrary to the abovementioned parameters. The ripening stage of the plant growth caused higher DI_0/RC , δR_0 , and $\delta R_0/(1 - \delta R_0)$ values by 1.42%, 15.68%, and 25.54%, respectively, i.e., their activities increase as the industrial hemp plants age.

PI_{ABS} shows the vitality of the photosynthetic units, which is why this is known as a very sensitive parameter [50]. Also, a very representative ChlF parameter in research is ϕP_0 , which proved less sensitive than PI_{ABS} . The reason for the same trend in the behavior of the mentioned parameters is that the behavior of ϕP_0 accompanies the changes in TR_0/DI_0 and PI_{ABS} because the values of ϕP_0 are used for the TR_0/DI_0 and PI_{ABS} calculation. Likewise, observing the parameters of electron transport (ET_0/RC , ET_0/TR_0 , and ET_0/ABS), the same conclusion was established because for the calculation of ET_0/ABS , the ϕP_0 and ET_0/TR_0 data are used, and for ET_0/RC the ET_0/TR_0 data are used. This is the same with the calculation $\delta R_0/(1 - \delta R_0)$, for which the values of δR_0 are used. This is the cause of the same behavior of the parameters according to the stages of growth and fertilization with K. Seliem et al. [60] found greater effectiveness in stimulating the vegetative growth of saffron when using different doses of K-silicate compared to K-sulfate. From the other point of view, precisely with this connection between the parameters, their changes according to the growth stages of the industrial hemp plants are visible. Earlier, a change in the tendency of the behavior of the parameters by growth stage was determined because the senescence of plants leads to changes in the behavior of the ChlF parameters, which also occurred in this research. The decrease in the value of the photosynthetic index and accompanying parameters with the aging of the industrial hemp plants, more precisely in the maturity stage, resulted in a loss of photosynthetic efficiency. The redox reaction after Q_A was inhibited, leading to additional electron transfer impairment between Q_A and Q_B . Likewise, the industrial hemp aging shows an increase in the parameters describing the flow of electrons in PSI, δR_0 , and $\delta R_0/(1 - \delta R_0)$. δR_0 depends on the electrons transferred to PSI from PQH_2 and the influx of electrons from the upper electron carrier. The aging-induced increase in δR_0 results from fewer donated electrons for PQH_2 reduction. During the senescence of *Alhagi sparsifolia* (Fabaceae) and wheat, photosynthesis efficiency and electron transfer changes were also detected [61,62].

Other photosynthetic parameters have been observed for industrial hemp. Struik et al. [35] found that industrial hemp radiation uses efficiency (PAR) changed throughout the season and reported that PAR decreased in earlier cultivars after flowering.

3.3. Chemical Composition of the Stem

The average content of macronutrients in the stem at the maturity stage was 1.16% N, 1.07% P, and 1.17% K (Figure 5). The K content was lower by 21.58% with KCl (K_1) and 26.62% lower with K_2SO_4 (K_2) compared to the control treatment (K_0). According to Iványi and Izsáki [63], in Hungary, which is close to this experimental station, the optimal time for collecting samples to determine micronutrient status in industrial hemp leaves is late May and early June, when satiation can be found if the leaves contain 5–6% N, 0.5–0.6% P, and 2.7–3.0% K.

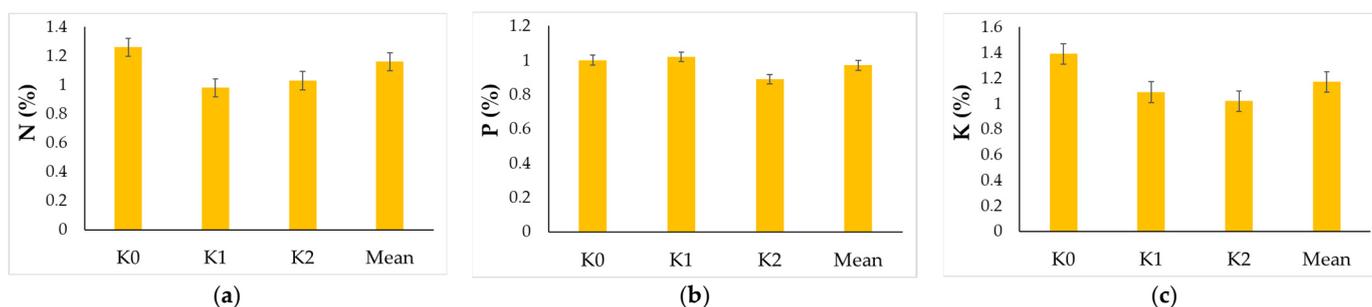


Figure 5. The macronutrient content (N, P, and K) in different industrial hemp stem samples (K_0 —control, K_1 —KCl, K_2 — K_2SO_4) (a) N (%), (b) P(%) and (c) K (%). Vertical error bars indicate a standard error of 5%.

In the present study, it was found that for both N and K, the content was highest in the control treatment (Figure 5a,c). Light et al. [64] reported that, even though the reason is not clear, there is a certain antagonism between N and Cl taken up by the crop, since it was found that N deficiency occurs when soil Cl is high. This may be the reason for the lowest N uptake by the industrial hemp plants in this study on the KCl treatment, even though the differences were not significant among the K sources (Figure 5a). In the present study, the highest concentrations of the other macronutrients, P and K, were determined for the industrial hemp in the K_0 treatment. Zehler et al. [37] stated that the uptake of the K^+ ions is 1.28 mEq with KCl, and is lower, at 0.74 mEq, with K_2SO_4 . This may be the reason for the lower K accumulation in the present study for the industrial hemp stem at harvest, due to the lower uptake of the K fertilizer source (Figure 5c).

According to Wilmer et al. [65], KCl in water dissociates into potassium (K^+) and chloride (Cl^-) ions, which contributes to a higher osmotic concentration compared to K_2SO_4 , which dissociates into K^+ and sulfate ions (SO_4^{2-}); thus, it may have an effect on water uptake and growth rates. Zehler et al. [37] stated in the case of nutrients such as K, the influence on the plant may depend on how it is chemically combined within the fertilizer with additional substances such as S and Cl. Naila Farooq et al. [66] argued that Cl content from KCl fertilizers may be a problem in arid and semiarid areas, due to salinity, so the authors recommend blending the KCl with composts, which significantly mitigates the salinity risks associated with the Cl^- ions and boosts nutrient availability, maize plant growth, and chlorophyll content. In sorghum plants, Amasiab et al. [67] found that with K_2SO_4 application (0, 10, 20, 30, 40, and 50 kg/ha), the content of the N in forage significantly ($p < 0.01$) increased from 1.4% (control) to 2.14% (50 kg/ha), which suggests that enhanced N content improves protein content, which is critical for forage quality. In the present study, the N content was slightly higher with the K_2SO_4 (K_2) application as compared to the KCl (K_1) treatment, but still, the content was decreased in comparison

with the control (K_0) treatment. Bakhsh et al. [68] found some differences in the source of K fertilizer for cereals. Thus, the authors stated that 37 kg K/ha of KCl was most effective in rice, wheat, and fallow–wheat rotations, whereas 37 kg K/ha of K_2SO_4 performed better in the maize–wheat rotation. Moreover, the authors stated that overall K_2SO_4 out-yielded KCl, likely due to the additional benefits of the sulfate (SO_4^{2-}) ion in improving nutrient availability and mitigating calcareous soil constraints. In calcareous and alkaline soils, Liu et al. [69] found that the potato tuber yield increased by 7.4% (200 kg/ha K_2SO_4) and even more, by 21.5% (300 kg/ha K_2SO_4), in comparison with the control treatment. Thus, the authors stated that K accumulation in critical growth stages such as tuber formation and starch accumulation was strongly correlated with increased potato yields, but also the application of K_2SO_4 significantly decreased soil pH, which enhanced phosphorus availability in calcareous soils.

3.4. Morphometric Parameters and Yield

In order to determine whether the formulation of K fertilizer affects the morphological properties of the stem and the chemical composition and fibers in the stem of industrial hemp, K fertilizer in two forms was applied as a presowing treatment. Generally, the effect of KCl and K_2SO_4 was not highly different for the morphometric parameters in this study. The different K fertilization sources did not significantly influence the number of plants per unit area. According to the results of this research, the average number of plants per unit area was 92,000 plants ha^{-1} (Table 3).

Table 3. The number of plants per unit area at harvest.

Treatment	No. Plants ha^{-1}	No. Male Plants ha^{-1}	No. Female Plants ha^{-1}
K_0	82.000	25.333	56.667
K_1	96.333	30.667	65.667
K_2	97.667	43.333	54.333
Mean	92.000	33.111	58.889
LSD _(0.05)	ns	ns	ns

Note: ns—non-significant.

The average share of male plants was 36% of the plants, while the number of female plants was 64%. Tang et al. [70] reported that for industrial hemp, the stem was higher at a lower density (45 plants $ha m^{-2}$) in comparison with the highest sowing density (67.5 plants m^{-2}). For industrial hemp for fiber production, Augustinović et al. [33] found that increasing plant density (from 100 to 300 seeds m^{-2}) resulted in a higher share of non-sprouted seeds, which confirms that fewer hemp plants survive denser sowing, i.e., a denser plant population after sprouting. According to Ranogajec et al. [71], in the period from 2014 to 2023, the average industrial hemp seed yield in Croatia varied from 0.4 t ha^{-1} (2014) to 1.5 t ha^{-1} (2023). Thus, based on the results of the present study, the seed yield was a little higher than the average yield in Croatia for 2023. The K treatment did not significantly influence the stem morphological parameters or stem and seed yield (Table 4). Even though the K_1 treatment did not significantly influence the morphometric parameters, the stem height, stem diameter, and stem and seed yield were on average the lowest as compared to the other K treatments (Table 4).

Table 4. Industrial hemp stems' morphometric and agronomic parameters.

Treatment	Stem Height (cm)	Stem Diameter (cm)	Stem Yield (t ha ⁻¹)	Dry Stem Yield (t ha ⁻¹) *	Seed Yield (t ha ⁻¹)
K ₀	79.7	0.45	16.0	11.9	1.96
K ₁	52.0	0.38	15.7	12.5	1.76
K ₂	79.7	0.39	17.0	12.2	1.82
Mean	67.5	0.41	16.2	12.2	1.85
LSD (0.05)	ns	ns	ns	ns	ns

Note: * air-dried mass per hectare; ns—non-significant

According to Vukadinović and Vukadinović [72], with a good supply of K for plants, the net assimilation is increased with a faster synthesis of reserve substances such as starch, sucrose, lipids, and proteins, but a K increase also strengthens of the plant and its turgor generation and helps it to resist drought and diseases, which is important for a high and stable yield of field crops. One of the most important roles of K is reducing stalk lodging for all field crops. Inadequate K fertilization leads to different stresses, such as water deficit. For industrial hemp, there is no newer literature about the influence of K formulation on stem yield and seed yield. According to Pasković [37], there was a recommendation for industrial hemp cultivation for fiber that it is better to avoid K from KCl fertilizer due to the harmful influence of Cl in K fertilizer, which leads to an increase in the volume of fiber groups and individual fiber cells. The volume expands, and the cell wall narrows so that the entire set of fibers becomes loose due to the deformed cells with a round or ovoid cross-section. Aubin et al. [34] found that K fertilization of industrial hemp has very limited or no influence on biomass and seed yield. Tsaliki et al. [73] stated that the productivity of industrial hemp is strongly affected by genotype and environmental conditions in Mediterranean conditions. Moreover, the authors reported that the industrial hemp stem of six genotypes (Santhica 27, Futura 75, Felina 32, Tygra, Bialobrzeshire, and Fedora 17) in a 3-year study was on average 5.7 mm and stem yield of 14.4 t ha⁻¹. Stack et al. [74] highlighted the importance of morphological and chemical compound traits among cultivars to gather data for plant breeders, especially on female inflorescences of *Cannabis sativa*. In Hungary, Iványi and Izsáki [64] reported that weather conditions have a great influence on industrial hemp stem yield, which was 12–17 t ha⁻¹ in a year with adequate precipitation, while in a year with lower precipitation, the yield was 6–8 t ha⁻¹. Kołodziej et al. [18] showed that different sowing rates significantly influence stem yield. Hence, the authors stated that between different sowing rates (from 5 to 60 kg ha⁻¹), the authors recommended 30 kg ha⁻¹ as an optimal sowing rate when hemp yielded 14.65 t ha⁻¹. Campiglia et al. [75], in a study of several varieties (Epsilon68, Fedora17, Felina32, Ferimon, Futura75, Santhica27, and Uso31) in Mediterranean conditions, found the dry stem yield was on average from 3.4 to 8.0 t ha⁻¹ of dry matter and further found that stem diameter was inversely correlated with plant density (6.7, 5.8 and 5.2 mm at 40, 80, and 120 plants m⁻², respectively). For the industrial hemp Finola variety, Varga et al. [76] found that stem diameter was on average 3.4 mm and the plant height was 71.2 cm, and that the seed mass per plant was significantly correlated ($p < 0.05$) with plant height with the application of K₂SO₄ fertilizer.

Saloner and Bernstein [77] examined secondary metabolites in different K supplies (15, 60, 100, 175, and 240 mg L⁻¹ K) of medicinal hemp and found that cannabinoid and terpenoid content decreased with the elevation of K supply. According to a study by Finnan and Burke [78], K supply did not significantly influence industrial hemp fiber yield. Ahmadi et al. [36] confirmed similar findings for K. According to the authors, K does not significantly influence plant biomass and seed yield, but this does not diminish its role as an essential element for plant growth. Light et al. [64] stated that Cl⁻ and SO₄²⁻ ions can

leach more rapidly in the soil system, and the adsorption of the SO_4^{2-} ions is more rapid in the plant tissue, occurring at a greater rate than for Cl^- ions. The formulation of K fertilizer proved to be important for several field crops such as potatoes [79], cotton [80], tobacco [81], tomato [82], and pepper [83] in terms of the way that chloride-free K fertilizers should be applied. K_2SO_4 instead of KCl for potatoes is preferred because Cl^- in the fertilizer can delay tuber development and decrease tuber yield [84]. Light et al. [64] mitigated the negative impact of KCl and found that KCl use did not influence potato quality. In a sugarcane pot experiment, Watanabe et al. [85] reported that sucrose concentration was reduced with KCl fertilizer but increased by K_2SO_4 fertilization. Hüvely and Vojnich [86] stated that chloride ions are generally harmful to vegetables such as pepper and that K sulfate is preferred (K_2SO_4). On the other hand, Cl^- is an essential element and, in some research, is found to influence plant development positively, but in higher doses, it can have a negative effect on plant growth, especially in early growth stages [87]. Fixen [88] reported that in wheat and barley, Cl^- is attributed to the suppression of the root and gives plants tolerance to disease. The amount of N has a greater influence on stem and seed yield formation than K. Wylie et al. [89] reported that up to 200 kg N ha^{-1} could increase plant height, stem diameter, biomass, and seed yields and that, on the contrary, 60 kg N ha^{-1} is sufficient for dual-purpose cultivars. Tang et al. [61] reported that fertilization rates over 300 kg ha^{-1} were not justified and that they even resulted in a higher level of stem bark content, concluding that the nutrient ratio should be a N:P:K ratio of 3:1:2, rather than 4:1:2.

3.5. Tensile Strength and Stiffness of Industrial Hemp Stem

After the two different K fertilization treatments (KCl and K_2SO_4), samples of the dry industrial hemp stems were subjected to tensile tests. The results were compared with samples that had not been fertilized (K_0). Each group consists of three test samples (Figure 6a). The relationship between stresses and strains are shown in Figure 6. Figure 6b shows samples of the industrial hemp after tensile failure.

In order to understand and observe the mechanical behavior of industrial hemp stem, tensile strength and the modulus of elasticity were determined. For this purpose, the middle part of the stem was taken for measuring. The tensile strength of the individual samples ranged from 38.23 to 63.59 MPa (Table 5). This is a result of the naturally heterogeneous and highly variable nature of industrial hemp stems. Unfertilized hemp has an average tensile strength of 51.78 MPa, KCl-fertilized hemp has a tensile strength of 49.25 MPa, and K_2SO_4 -fertilized hemp has a tensile strength of 53.32 MPa.

The modulus of elasticity was determined based on linear regression lines placed on the elastic part of the stress–strain curve (Figure 7). The average modulus of elasticity equals 4.99 GPa for IH- K_0 , 5.06 GPa for IH-KCl, and 5.07 GPa for IH- K_2SO_4 . The highest modulus of elasticity was obtained for sample IH-KCl-1 and was 6.51 GPa, while the lowest value of 3.37 GPa was obtained for sample IH-KCl-2.

Iványi and Izsáki [64] reported that N and K fertilization for industrial hemp leads to longer and thicker stems and higher fiber yield. Shah et al. [41] argue that the mechanical properties of plants are not only important for phenotyping but also for harvest and processing due to stem interaction with machines. Galedar et al. [43] highlighted that for alfalfa (*Medicago sativa* L.) tensile strength increased exponentially with a decrease in the moisture content and towards the lower regions, and that for the middle part of the stem, the tensile strength varied from 16.31 and 32.74 MPa.



Figure 6. Industrial hemp samples (a) before performing the tensile test and (b) after the tensile test.

Table 5. Tensile properties of industrial hemp stem.

Sample	Max F _c (N)	Average Cross-Section (mm ²)	Tensile Strength (MPa)	Average Tensile Strength (MPa)	Young's Modulus (GPa)	Average Young's Modulus (GPa)
IH-K ₀ (control treatment)						
IH-K ₀ -1	1055	25.61	41.18	51.78	4.77	4.99
IH-K ₀ -2	1095	17.72	63.59		5.19	
IH-K ₀ -3	1052	20.80	50.57		5.02	
IH-KCl treatment						
IH-KCl-1	921	18.01	51.14	49.25	6.51	5.06
IH-KCl-2	1337	22.91	58.37		3.37	
IH-KCl-3	945	24.76	38.23		5.29	
IH-K ₂ SO ₄ treatment						
IH-K ₂ SO ₄ -1	1066	20.79	51.27	53.32	5.15	5.07
IH-K ₂ SO ₄ -2	1272	21.00	60.57		6.00	
IH-K ₂ SO ₄ -3	836	17.38	48.12		4.05	

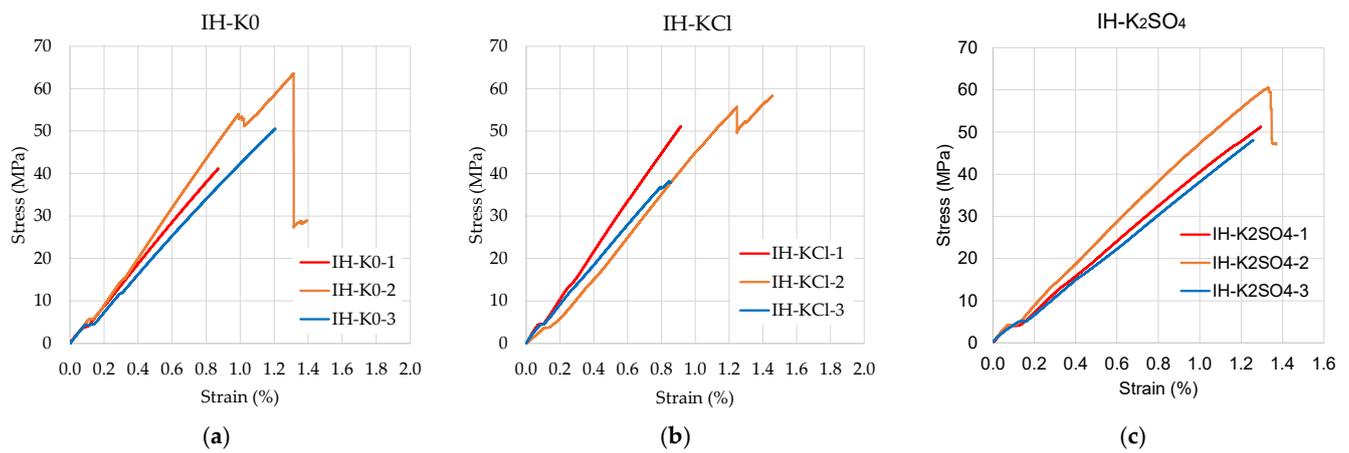


Figure 7. Stress–strain diagrams for industrial hemp stem: (a) IH-K0, (b) IH-KCl, and (c) IH-K₂SO₄.

3.6. Correlation Analysis of Industrial Hemp Stem

Correlation analysis showed a significant relationship between several parameters (Table 6). The highest very significant correlation coefficients were determined for industrial hemp macronutrient status (N, P, and K).

Table 6. Pearson’s correlation coefficients.

	No. of Plants Per Unit Area	Stem Height	Stem Diameter	Seed Yield	Stem Yield	N	P	K	Max Fc (N)	Average Cross-Section	Tensile Strength (MPa)	Young’s Modulus (GPa)
No. of plants per unit area	1											
Stem height	−0.395 ns	1										
Stem diameter	−0.506 ns	0.674 *	1									
Seed yield	0.003 ns	0.494 ns	0.242 ns	1								
Stem yield	0.688 *	−0.307 ns	−0.556 ns	−0.302 ns	1							
N	−0.029 ns	0.199 ns	0.504 ns	0.332 ns	−0.208 ns	1						
P	−0.204 ns	0.245 ns	0.721 *	0.354 ns	−0.372 ns	0.808 **	1					
K	−0.007 ns	0.114 ns	0.519 ns	0.430 ns	−0.163 ns	0.841 **	0.903 **	1				
Max Fc (N)	0.223 ns	−0.536 ns	−0.525 ns	−0.548 ns	0.486 ns	0.041 ns	−0.051 ns	0.051 ns	1			
Average cross-section	0.147 ns	−0.022 ns	0.081 ns	0.293 ns	−0.170 ns	0.264 ns	0.308 ns	0.431 ns	0.308 ns	1		
Tensile strength (MPa)	0.025 ns	−0.441 ns	−0.488 ns	−0.653 *	0.565 ns	−0.117 ns	0.613 *	0.305 ns	0.613 *	−0.545 ns	1	
Young’s modulus (GPa)	−0.158 ns	0.127 ns	−0.097 ns	0.413 ns	−0.306 ns	0.170 ns	0.218 ns	0.173 ns	−0.218 ns	−0.214 ns	0.021 ns	1

Note: ns—non-significant; * significance 0.05; ** significance 0.01.

Macronutrients play indispensable roles in plant growth and development, acting as structural components, energy carriers, and catalysts for biochemical processes. Nadeem et al. [90] stated that K is highly mobile within the plant and essential for younger plant parts, regulation of water uptake and transport, enzyme activation, and photosynthesis.

Nitrogen in the plants positively influences P uptake, but P deficiency negatively affects NO_3^- uptake from the soil [91–93]. Schleuss et al. [94] and Xie et al. [95] stated that plants often use N instead of P and vice versa, which can explain the synergistic plant growth responses to NP fertilization. Plant-available K in the soil (exchangeable K) has a considerable influence on N uptake [96]. The NH_4^+ and K^+ share similar valence and size properties in long-distance transport within the plant [97], which can explain the positive correlation of those elements in the industrial hemp stem determined in this study. For industrial hemp morphometric parameters and seed and stem yield, the significant ($p < 0.05$) correlation coefficients were determined for the number of plants per unit area and seed yield ($r = 0.688$), stem height, and diameter ($r = 0.674$). The stem diameter was positive, and there was a significant correlation with the amount of P in the stem dry matter ($r = 0.721$). Seed yield was in negative and significant ($p < 0.05$) correlation with tensile strength ($r = -0.653$).

4. Conclusions and Future Perspectives

Fertilization is a necessary agricultural practice that makes cultivating the desired crop more successful. This study aimed to analyze the influence of different K fertilizer formulations on the photosynthetic activity of industrial hemp leaves and stem morphology and biomechanics. Even though the forms of K fertilization that were used had no significant effect on the differences between the morphological parameters or stem and seed yield, further trials should be conducted on the application of macronutrients to Finola varieties. Although the tensile strength and module elasticity were not homogeneous, these results are valuable for the harvesting and further processing of the stems. These findings are very valuable considering the differences in cost between KCl (which is a little cheaper) and potassium fertilizers containing sulfate, like K_2SO_4 , which can certainly play a major role in agricultural decision-making. By using KCl as a fertilizer, farmers can reduce the costs of cultivation significantly, especially for crops that are not sensitive to chloride. In the present study, the industrial hemp variety Finola did not show any extreme influence regard the KCl application; thus, it can be recommended for future use in industrial hemp production.

In general, the applied KCl doses showed lower values than the control and K_2SO_4 for half of the chlorophyll *a* fluorescence parameters in the industrial hemp leaves. On the other hand, the values of the ChlF parameters measured at the maturity stage were mostly lower than those measured at the flowering stage. Although there were some changes in the efficiency of the photosystems I and II, our next step is to investigate the effect of higher doses of K fertilizers to gain insight into the photosynthetic activity at a higher level of availability of K from the soil to the cannabis plants.

Author Contributions: Conceptualization, I.K., I.V. and A.M.K.; methodology, A.M.K., D.I., M.T. and P.K.; software, A.M.K., P.K. and I.V.; formal analysis, A.M.K., P.K., M.T. and I.V.; investigation, I.K., A.M.K. and I.V.; writing—original draft preparation, I.V., A.M.K., P.K. and M.T.; writing—review and editing, D.I. and I.K.; visualization, A.M.K.; project administration, I.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research was a part of the project Rammed Earth for modeling and standardization in seismically active areas (RE-forMS project, UIP-2020-02-7363), funded by the Croatian Science Foundation (HRZZ).

Institutional Review Board Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Lash, R. Industrial hemp: The crop for the seventh generation. *Am. Indian Law Rev.* **2002**, *27*, 313. [CrossRef]
2. Fike, J. The history of hemp. In *Industrial Hemp as a Modern Commodity Crop*; ASA: Madison, WI, USA; CSSA: Madison, WI, USA; SSSA: Madison, WI, USA, 2019; pp. 1–25.
3. Pejić, B.; Vukčević, M.; Kostić, M. Hemp fibers in Serbia: Cultivation, processing and applications. In *Sustainable Agriculture Reviews 42: Hemp Production and Applications*; Springer: Cham, Switzerland, 2020; pp. 111–146.
4. FAOStat 2024. Available online: <https://www.fao.org/faostat/en/#data/QCL> (accessed on 25 October 2024).
5. Available online: https://agriculture.ec.europa.eu/farming/crop-productions-and-plant-based-products/hemp_en (accessed on 25 October 2024).
6. Varga, I.; Kraus, I.; Iljkić, D.; Jonjić, A.; Antunović, M. Tradicija proizvodnje industrijske konoplje u Hrvatskoj. *Sjemenarstvo* **2022**, *33*, 25–40. [CrossRef]
7. Visković, J.; Sikora, V.; Latković, D.; Zeremski, T.; Dunderski, D.; Astatkie, T.; Noller, J.S.; Zheljaskov, V.D. Optimization of hemp production technology for fiber and seed. *Ind. Crop. Prod.* **2024**, *219*, 119127. [CrossRef]
8. Croatian Bureau of Statistics. 2024. Available online: <https://dzs.gov.hr/> (accessed on 25 October 2024).
9. LL File No. 2022-021696, Regulation of Hemp, The Law Library of Congress, Global Legal Research Directorate. 2022. Available online: <https://www.loc.gov/item/2022666115> (accessed on 11 November 2024).
10. Kaur, N.; Brym, Z.; Oyola, L.A.M.; Sharma, L.K. Nitrogen fertilization impact on hemp (*Cannabis sativa* L.) crop production: A review. *Agron. J.* **2023**, *115*, 1557–1570. [CrossRef]
11. OG 18/2012 (Official gazette) Pravilnik o Uvjetima za Uzgoj Konoplje, Načinu Prijave Uzgoja Maka te Uvjetima za Posjedovanje Opojnih Droga u Veterinarstvu. Narodne Novine 18/2012. Available online: https://narodne-novine.nn.hr/clanci/sluzbeni/full/2012_02_18_505.html (accessed on 1 August 2024).
12. OG 39/2019 (Official gazette) Zakon o Izmjenama i Dopunama Zakona o Suzbijanju Zlouporebe Droga. Narodne Novine 39/2019. Available online: https://narodne-novine.nn.hr/clanci/sluzbeni/full/2019_04_39_799.html (accessed on 1 August 2024).
13. Klir, Ž.; Novoselec, J.; Antunović, Z. An overview on the use of hemp (*Cannabis sativa* L.) in animal nutrition. *Poljoprivreda* **2019**, *25*, 52–61. [CrossRef]
14. Yano, H.; Fu, W. Hemp: A Sustainable Plant with High Industrial Value in Food Processing. *Foods* **2023**, *12*, 651. [CrossRef] [PubMed]
15. Varga, I.; Kristić, M.; Lisjak, M.; Tkalec Kojić, M.; Iljkić, D.; Jović, J.; Kristek, S.; Markulj Kulundžić, A.; Antunović, M. Antioxidative Response and Phenolic Content of Young Industrial Hemp Leaves at Different Light and Mycorrhiza. *Plants* **2024**, *13*, 840. [CrossRef]
16. Idler, C.; Pecenka, R.; Fürll, C.; Gusovius, H.-J. Wet processing of hemp: An overview. *J. Nat. Fibers* **2011**, *8*, 59–80. [CrossRef]
17. Dunderski, D.; Jaćimović, G.; Crnobarac, J.; Visković, J.; Latković, D. Using Digital Image Analysis to Estimate Corn Ear Traits in Agrotechnical Field Trials: The Case with Harvest Residues and Fertilization Regimes. *Agriculture* **2023**, *13*, 732. [CrossRef]
18. Kołodziej, J.; Pudełko, K.; Mańkowski, J. Energy and Biomass Yield of Industrial Hemp (*Cannabis sativa* L.) as Influenced by Seeding Rate and Harvest Time in Polish Agro-Climatic Conditions. *J. Nat. Fibers* **2023**, *20*, 2159609. [CrossRef]
19. Visković, J.; Zheljaskov, V.D.; Sikora, V.; Noller, J.; Latković, D.; Ocamb, C.M.; Koren, A. Industrial Hemp (*Cannabis sativa* L.) Agronomy and Utilization: A Review. *Agronomy* **2023**, *13*, 931. [CrossRef]
20. Kovačić, Đ.; Radočaj, D.; Jurišić, M. Ensemble machine learning prediction of anaerobic co-digestion of manure and thermally pretreated harvest residues. *Bioresour. Technol.* **2024**, *402*, 130793. [CrossRef] [PubMed]
21. Visković, J.; Dunderski, D.; Adamović, B.; Jaćimović, G.; Latković, D.; Vojnović, Đ. Toward an Environmentally Friendly Future: An Overview of Biofuels from Corn and Potential Alternatives in Hemp and Cucurbits. *Agronomy* **2024**, *14*, 1195. [CrossRef]
22. Pospišil, M. *Ratarstvo II. dio—Industrijsko bilje*; Zrinski d.o.o.: Čakovec, Croatia, 2013.
23. Oseyko, M.; Sova, N.; Lutsenko, M.; Kalyna, V. Chemical aspects of the composition of industrial hemp seed products. *Ukr. Food J.* **2019**, *8*, 544–558. [CrossRef]
24. Lančaričová, A.; Kuzmiaková, B.; Porvaz, P.; Havrlentová, M.; Nemeček, P.; Kraic, J. Nutritional quality of hemp seeds (*Cannabis sativa* L.) in different environments. *J. Central Eur. Agric.* **2021**, *22*, 748–761. [CrossRef]
25. Gimeno-Martínez, D.; Igual, M.; García-Segovia, P.; Martínez-Monzó, J.; Navarro-Rocha, J. Characterisation of the Fat Profile of Different Varieties of Hemp Seeds (*Cannabis sativa* L.) for Food Use. *Biol. Life Sci. Forum* **2023**, *26*, 89. [CrossRef]
26. Cigasova, J.; Stevulova, N.; Junak, J. Innovative use of biomass based on technical hemp in building industry. *Chem. Eng. Trans.* **2014**, *37*, 685–690. [CrossRef]
27. Malabadi, R.B.; Kolkar, K.P.; Chalannavar, R.K. Industrial *Cannabis sativa* (Hemp fiber): Hempcrete-A Plant Based and Eco-friendly Building Construction Material. *Int. J. Res. Innov. Appl. Sci.* **2023**, *8*, 67–78. [CrossRef]
28. Kumar, S.; Kumar, S.; Mohapatra, T. Interaction between macro-and micro-nutrients in plants. *Front. Plant Sci.* **2021**, *12*, 665583. [CrossRef]

29. Neugschwandtner, R.W.; Száková, J.; Pachtrog, V.; Tlustoš, P.; Kulhánek, M.; Černý, J.; Kaul, H.-P.; Wagenristl, H.; Moitzi, G.; Euteneuer, P. Exchangeable and Plant-Available Macronutrients in a Long-Term Tillage and Crop Rotation Experiment after 15 Years. *Plants* **2022**, *11*, 565. [CrossRef]
30. Kristek, S.; Jović, J.; Martinović, M.; Jantoš, J.; Popović, B.; Lončarić, Z. The Application of Biopreparations as an Alternative to Chemical Fungicides in the Protection of Wheat. *Poljoprivreda* **2023**, *29*, 24–32. [CrossRef]
31. Iljkić, D.; Vuković, M.; Dvojković, K.; Horvat, D.; Szpunar-Krok, E.; Jańczak-Pieniążek, M.; Rastija, M. Variety, Chemical Protection and Biostimulator Effect on Winter Wheat Status. *Poljoprivreda* **2024**, *30*, 28–35. [CrossRef]
32. Venturi, G.; Amaducci, M.T. Effects of nitrogen fertilizer rate and sowing rate on yield and technological characteristics of *Cannabis sativa* L. *Riv. Di Agron.* **1997**, *31*, 616–623.
33. Augustinović, Z.; Pospišil, M.; Butorac, J.; Koren, M.A.; Ivanek-Martinčić, M.; Šumbera, N. Samoregulacija sklopa, odnos ženskih i muških biljaka i morfološka svojstva industrijske konoplje u ovisnosti o gustoći sjetve i gnojidbi dušikom. *Agron. Glas.* **2012**, *74*, 189–206.
34. Aubin, M.P.; Seguin, P.; Vanasse, A.; Tremblay, G.F.; Mustafa, A.F.; Charron, J.B. Industrial hemp responds to nitrogen, phosphorus, and potassium fertilization. *Crop Forage Turfgrass Manag.* **2015**, *1*, 1–10. [CrossRef]
35. Struik, P.C.; Amaducci, S.; Bullard, M.J.; Stutterheim, N.C.; Venturi, G.; Cromack, H.T.H. Agronomy of fibre hemp (*Cannabis sativa* L.) in Europe. *Ind. Crop. Prod.* **2000**, *11*, 107–118. [CrossRef]
36. Ahmadi, F.; Kallinger, D.; Starzinger, A.; Lackner, M. Hemp (*Cannabis sativa* L.) Cultivation: Chemical Fertilizers or Organic Technologies, a Comprehensive Review. *Nitrogen* **2024**, *5*, 624–654. [CrossRef]
37. Pasković, F. *Predivo Bilje: Dio. Konoplja, Lan i Pamuk*; Nakladni Zavod Znanje: Zagreb, Croatia, 1966; Volume 1.
38. Zehler, E.; Kreipe, H.; Gething, P.A. *Potassium Sulphate and Potassium Chloride. Their Influence on the Yield and Quality of Cultivated Plants*; International Potash Institute Bern: Bern, Switzerland, 1981.
39. Geilfus, C.-M. Chloride: From nutrient to toxicant. *Plant Cell Physiol.* **2018**, *59*, 877–886. [CrossRef]
40. Aarabi, F.; Naake, T.; Fernie, A.R.; Hoefgen, R. Coordinating sulfur pools under sulfate deprivation. *Trends Plant Sci.* **2020**, *25*, 1227–1239. [CrossRef]
41. Shah, D.U.; Reynolds, T.P.; Ramage, M.H. The strength of plants: Theory and experimental methods to measure the mechanical properties of stems. *J. Exp. Bot.* **2017**, *68*, 4497–4516. [CrossRef]
42. Genet, M.; Stokes, A.; Salin, F.; Mickovski, S.B.; Fourcaud, T.; Dumail, J.-F.; Van Beek, R. The influence of cellulose content on tensile strength in tree roots. *Plant Soil* **2005**, *278*, 1–9. [CrossRef]
43. Galedar, M.N.; Tabatabaefar, A.; Jafari, A.; Sharifi, A.; Rafiee, S.; Mohtasebi, S.S. Influence of moisture content, rate of loading and height regions on tensile strength of alfalfa stems. *Int. Agrophys.* **2009**, *23*, 27–30.
44. Durant, P.C.; Bhasin, A.; Juenger, T.E.; Heckman, R.W. Genetically correlated leaf tensile and morphological traits are driven by growing season length in a widespread perennial grass. *Am. J. Bot.* **2024**, *111*, e16349. [CrossRef]
45. Liu, M.; Fernando, D.; Daniel, G.; Madsen, B.; Meyer, A.S.; Ale, M.T.; Thygesen, A. Effect of harvest time and field retting duration on the chemical composition, morphology and mechanical properties of hemp fibers. *Ind. Crop. Prod.* **2015**, *69*, 29–39. [CrossRef]
46. Croatian Meteorological and Hydrological Service (2024). Available online: https://meteo.hr/index_en.php (accessed on 30 July 2023).
47. Markulj Kulundžić, A.; Sudarić, A.; Matoša Kočar, M.; Duvnjak, T.; Liović, I.; Mijić, A.; Varga, I.; Viljevac Vuletić, M. Detailed Insight into the Behaviour of Chlorophyll *a* Fluorescence Transient Curves and Parameters during Different Times of Dark Adaptation in Sunflower Leaves. *Agronomy* **2024**, *14*, 954. [CrossRef]
48. Mediavilla, V.; Jonquera, M.; Schmid-Slembrouck, I.; Soldati, A. Decimal code for growth stages of hemp (*Cannabis sativa* L.). *J. Int. Hemp Assoc.* **1998**, *65*, 68–74.
49. Strasser, R.J.; Tsimilli-Michael, M.; Srivastava, A. Analysis of the fluorescence transient. In *Chlorophyll Fluorescence: A Signature of Photosynthesis*; Advances in Photosynthesis and Respiration Series; George, C., Papageorgiou, C., Govindjee, Eds.; Springer: Dordrecht, The Netherlands, 2004; pp. 321–362.
50. Yusuf, M.A.; Kumar, D.; Rajwanshi, R.; Strasser, R.J.; Tsimilli-Michael, M.; Govindjee; Sarin, N.B. Overexpression of g-tocopherol methyl transferase gene in transgenic Brassica juncea plants alleviates abiotic stress: Physiological and chlorophyll *a* fluorescence measurements. *Biochim. Biophys. Acta* **2010**, *1797*, 1428–1438. [CrossRef] [PubMed]
51. Osman, K.T. Plant nutrients and soil fertility management. In *Soils*; Osman, K.T., Ed.; Springer: Berlin/Heidelberg, Germany, 2013; pp. 129–159.
52. Rastogi, A.; Zivcak, M.; Tripathi, D.K.; Yadav, S.; Kalaji, H.M.; Brestic, M. Phytotoxic effect of silver nanoparticles in *Triticum aestivum*: Improper regulation of photosystem I activity as the reason for oxidative damage in the chloroplast. *Photosynthetica* **2019**, *57*, 209–216. [CrossRef]
53. Zagorchev, L.; Atanasova, A.; Albanova, I.; Traianova, A.; Mladenov, P.; Kouzmanova, M.; Goltsev, V.; Kalaji, H.M.; Teofanova, D. Functional Characterization of the Photosynthetic Machinery in Smicronix Galls on the Parasitic Plant *Cuscuta campestris* by JIP-Test. *Cells* **2021**, *10*, 1399. [CrossRef] [PubMed]

54. Dimitrova, S.; Paunov, M.; Pavlova, B.; Dankov, K.; Kouzmanova, M.; Velikova, V.; Tsonev, T.; Kalaji, H.; Goltsev, V. Photosynthetic efficiency of two *Platanus orientalis* L. ecotypes exposed to moderately high temperature—JIP-test analysis. *Photosynthetica* **2020**, *58*, 657–670. [[CrossRef](#)]
55. Antunović Dunić, J.; Mlinarić, S.; Pavlović, I.; Lepeduš, H.; Salopek-Sondi, B. Comparative Analysis of Primary Photosynthetic Reactions Assessed by OJIP Kinetics in Three Brassica Crops after Drought and Recovery. *Appl. Sci.* **2023**, *13*, 3078. [[CrossRef](#)]
56. Kusaka, M.; Kalaji, H.; Mastalerczuk, G.; Dąbrowski, P.; Kowalczyk, K. Potassium deficiency impact on the photosynthetic apparatus efficiency of radish. *Photosynthetica* **2021**, *59*, 127–136. [[CrossRef](#)]
57. Kalaji, H.M.; Govindjee; Bosa, K.; Kościelniak, J.; Żuk-Golaszewska, K. Effects of salt stress on photosystem II efficiency and CO₂ assimilation of two Syrian barley landraces. *Environ. Exp. Bot.* **2011**, *73*, 64–72. [[CrossRef](#)]
58. Kalaji, H.M.; Schansker, G.; Ladle, R.J.; Goltsev, V.; Bosa, K.; Allakhverdiev, S.I.; Brestic, M.; Bussotti, F.; Calatayud, A.; Dąbrowski, P.; et al. Frequently asked questions about in vivo chlorophyll fluorescence: Practical issues. *Photosynth. Res.* **2014**, *122*, 121–158. [[CrossRef](#)]
59. Farnisa, M.M.; Miller, G.C.; Solomon, J.K.Q.; Barrios-Masias, F.H. Floral hemp (*Cannabis sativa* L.) responses to nitrogen fertilization under field conditions in the high desert. *PLoS ONE* **2023**, *18*, e0284537. [[CrossRef](#)] [[PubMed](#)]
60. Seliem, M.K.; Dewir, Y.H.; El-Mahrouk, M.E.; El-Ramady, H.; Fathy Elbehiry, F. The effects of potassium fertilizers on flowering, vegetative growth, and daughter corm yield of potted saffron. *Appl. Ecol. Environ. Res.* **2024**, *22*, 1829–1847. [[CrossRef](#)]
61. Tang, G.; Li, X.; Lin, L.; Guo, H. Combined effects of girdling and leaf removal on fluorescence characteristic of *Alhagi sparsifolia* leaf senescence. *Plant Biol.* **2015**, *17*, 980–989. [[CrossRef](#)] [[PubMed](#)]
62. Španić, V.; Šunić, K.; Duvnjak, J.; Hu, Y.; Katanić, Z. Chlorophyll *a* fluorescence during flag leaf senescence of field-grown winter wheat plants under drought conditions. *Ann. Appl. Biol.* **2023**, *183*, 80–92. [[CrossRef](#)]
63. Iványi, I.; Izsáki, Z. Effect of nitrogen, phosphorus, and potassium fertilization on nutritional status of fiber hemp. *Commun. Soil Sci. Plant Anal.* **2009**, *40*, 974–986. [[CrossRef](#)]
64. Light, S.E.; Sullivan, D.M.; Horneck, D.A. Timing of potassium chloride application effect on soil and potato uptake of chloride. *Agrosyst. Geosci. Environ.* **2022**, *5*, e20301. [[CrossRef](#)]
65. Wilmer, L.; Pawelzik, E.; Naumann, M. Comparison of the effects of potassium sulphate and potassium chloride fertilisation on quality parameters, including volatile compounds, of potato tubers after harvest and storage. *Front. Plant Sci.* **2022**, *13*, 920212. [[CrossRef](#)] [[PubMed](#)]
66. Naila Farooq, N.F.; Shamsa Kanwal, S.K.; Allah Ditta, A.D.; Azhar Hussain, A.H.; Muhammad Naveed, M.N.; Jamshaid, M.U.; Muhammad Iqbal, M.I. Comparative efficacy of KCl blended composts and sole application of KCl or K₂SO₄ in improving K nutrition, photosynthetic capacity and growth of maize. *Soil Environ.* **2018**, *37*, 68–74. [[CrossRef](#)]
67. Amasiab, E.O.; Eltaib, M.Z.; Abdalla, A.H. Effect of Potassium Sulfate Fertilizer on the Mineral Profile of Sorghum Forage and Their Uses as Ruminant Feed. *Int. J. Plant Soil Sci.* **2023**, *35*, 164–173. [[CrossRef](#)]
68. Bakhsh, A.; Khattak, J.K.; Bhatti, A.U. Comparative effect of potassium chloride and potassium sulfate on the yield and protein content of wheat in three different rotations. *Plant Soil* **1986**, *96*, 273–277. [[CrossRef](#)]
69. Liu, C.-A.; Liu, C.-C.; Zhang, R.-H.; Zhou, L.-M.; Jia, Y.; Gao, W.-J.; Li, J.-T.; Ma, Q.-F.; Siddique, K.H.; Li, F.-M. Yield-increase effects via improving soil phosphorus availability by applying K₂SO₄ fertilizer in calcareous–alkaline soils in a semi-arid agroecosystem. *Field Crop. Res.* **2013**, *144*, 69–76. [[CrossRef](#)]
70. Tang, K.; Wang, J.; Yang, Y.; Deng, G.; Yu, J.; Hu, W.; Guo, L.; Du, G.; Liu, F. Fiber hemp (*Cannabis sativa* L.) yield and its response to fertilization and planting density in China. *Ind. Crop. Prod.* **2022**, *177*, 114542. [[CrossRef](#)]
71. Ranogajec, L.; Antunović, M.; Stipešević, B.; Varga, I. A current status and production potential of industrial hemp in Croatia based on a SWOT analysis. *Poljoprivreda* **2024**, *30*, 56–63. [[CrossRef](#)]
72. Vukadinović, V.; Vukadinović, V. *Ishrana Bilja*; Poljoprivredni fakultet u Osijeku: Osijek, Croatia, 2011.
73. Tsaliki, E.; Kalivas, A.; Jankauskiene, Z.; Irakli, M.; Cook, C.; Grigoriadis, I.; Panoras, I.; Vasilakoglou, I.; Dhima, K. Fibre and Seed Productivity of Industrial Hemp (*Cannabis sativa* L.) Varieties under Mediterranean Conditions. *Agronomy* **2021**, *11*, 171. [[CrossRef](#)]
74. Stack, G.M.; Carlson, C.H.; Toth, J.A.; Philippe, G.; Crawford, J.L.; Hansen, J.L.; Viands, D.R.; Rose, J.K.C.; Smart, L.B. Correlations among morphological and biochemical traits in high-cannabidiol hemp (*Cannabis sativa* L.). *Plant Direct* **2023**, *7*, e503. [[CrossRef](#)] [[PubMed](#)]
75. Campiglia, E.; Radicetti, E.; Mancinelli, R. Plant density and nitrogen fertilization affect agronomic performance of industrial hemp (*Cannabis sativa* L.) in Mediterranean environment. *Ind. Crop. Prod.* **2017**, *100*, 246–254. [[CrossRef](#)]
76. Varga, I.; Iljkić, D.; Krolo, P.; Perić Fekete, A.; Kraus, I. The Source of K Fertilizer for Industrial Hemp (*Cannabis sativa* L.): Mechanical and Chemical Properties of Stem for Rammed Earth Walls. *Agriculture* **2024**, *14*, 2196. [[CrossRef](#)]
77. Saloner, A.; Bernstein, N. Effect of Potassium (K) Supply on Cannabinoids, Terpenoids and Plant Function in Medical Cannabis. *Agronomy* **2022**, *12*, 1242. [[CrossRef](#)]
78. Finnan, J.; Burke, B. Potassium fertilization of hemp (*Cannabis sativa*). *Ind. Crop. Prod.* **2013**, *41*, 419–422. [[CrossRef](#)]

79. Torabian, S.; Farhangi-Abriz, S.; Qin, R.; Noulas, C.; Sathuvalli, V.; Charlton, B.; Loka, D.A. Potassium: A Vital Macronutrient in Potato Production—A Review. *Agronomy* **2021**, *11*, 543. [[CrossRef](#)]
80. Tang, N.X.; Shen, J.X. A study on the effect of Cl on the growth and development of cotton. *Soils Fertil.* **1993**, *2*, 1–3.
81. Kafkafi, U.; Tarchitzky, J. *A Tool for Efficient Fertilizer and Water Management*; International Potash Institute (IPI): Paris, France, 2011; pp. 1–123.
82. Jarosz, Z. Effect of different types of potassium fertilization on the yielding of greenhouse tomatoes grown in various substrates. *Acta Sci. Pol. Hortorum Cultus* **2006**, *5*, 3–9.
83. Golcz, A.; Kujawski, P.; Markiewicz, B. Yielding of red pepper (*Capsicum annuum* L.) under the influence of varied potassium fertilization. *Acta scientiarum Polonorum. Hortorum Cultus* **2012**, *11*, 3–15.
84. Beringer, H.; Koch, K.; Lindhauer, M.G. Source: Sink relationships in potato (*Solanum tuberosum*) as influenced by potassium chloride or potassium sulphate nutrition. *Plant Soil* **1990**, *124*, 287–290. [[CrossRef](#)]
85. Watanabe, K.; Fukuzawa, Y.; Kawasaki, S.-I.; Ueno, M.; Kawamitsu, Y. Effects of potassium chloride and potassium sulfate on sucrose concentration in sugarcane juice under pot conditions. *Sugar Tech* **2016**, *18*, 258–265. [[CrossRef](#)]
86. Hüvely, A.; Vojnich, V.J. Effect of potassium chloride and sulphate nutrition of pepper plants yield. *Lucr. Științifice Manag. Agric.* **2016**, *18*, 71.
87. Varga, I.; Iljkić, D.; Tkalec Kojić, M.; Dobрева, T.; Markulj Kulundžić, A.; Antunović, M. Germination of industrial hemp (*Cannabis sativa* L.) at different level of sodium chloride and temperatures. *Agric. Conspec. Sci.* **2022**, *87*, 11–15.
88. Fixen, P.E. Crop Responses to Chloride. *Adv. Agron.* **1993**, *50*, 107–150. [[CrossRef](#)]
89. Wylie, S.E.; Ristvey, A.G.; Fiorellino, N.M. Fertility management for industrial hemp production: Current knowledge and future research needs. *GCB Bioenergy* **2021**, *13*, 517–524. [[CrossRef](#)]
90. Nadeem, F.; Hanif, M.A.; Majeed, M.I.; Mushtaq, Z. Role of macronutrients and micronutrients in the growth and development of plants and prevention of deleterious plant diseases—a comprehensive review. *Int. J. Chem. Biochem. Sci.* **2018**, *13*, 31–52.
91. Güsewell, S. N : P ratios in terrestrial plants: Variation and functional significance. *New Phytol.* **2004**, *164*, 243–266. [[CrossRef](#)]
92. Krouk, G.; Kiba, T. Nitrogen and Phosphorus interactions in plants: From agronomic to physiological and molecular insights. *Curr. Opin. Plant Biol.* **2020**, *57*, 104–109. [[CrossRef](#)] [[PubMed](#)]
93. Chen, X.; Wang, M.; Li, M.; Sun, J.; Lyu, M.; Zhong, Q.; Cheng, D. Convergent nitrogen–phosphorus scaling relationships in different plant organs along an elevational gradient. *AoB Plants* **2020**, *12*, plaa021. [[CrossRef](#)]
94. Schleuss, P.M.; Widdig, M.; Heintz-Buschart, A.; Kirkman, K.; Spohn, M. Interactions of nitrogen and phosphorus cycling promote P acquisition and explain synergistic plant-growth responses. *Ecology* **2020**, *101*, e03003. [[CrossRef](#)] [[PubMed](#)]
95. Xie, Y.; Li, L.; Wang, L.; Zhang, J.; Dang, Z.; Li, W.; Qi, Y.; Zhao, W.; Dong, K.; Wang, X.; et al. Relationship between Phosphorus and Nitrogen Concentrations of Flax. *Agronomy* **2023**, *13*, 856. [[CrossRef](#)]
96. Johnston, A.E.; Milford, G.F.J. *Potassium and Nitrogen Interactions in Crops*; Potash Development Association: York, UK, 2012.
97. Bar-Tal, A. The effects of nitrogen form on interactions with potassium. In *Nitrogen and Potassium Interactions*; International Potash Institute: Zug, Switzerland, 2011; Volume 9.

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