

Article

The Effect of Indigo (*Indigofera tinctoria* L.) Waste on Growth Performance, Digestibility, Rumen Fermentation, Hematology and Immune Response in Growing Beef Cattle

Nirawan Gunun¹, Chatchai Kaewpila², Waroon Khota², Sineenart Polyorach³, Thachawech Kimprasit², Wasana Phlaetita⁴, Anusorn Cherdthong⁵, Metha Wanapat⁵ and Pongsatorn Gunun^{2,*}

¹ Department of Animal Science, Faculty of Technology, Udon Thani Rajabhat University, Udon Thani 41000, Thailand

² Department of Animal Science, Faculty of Natural Resources, Rajamangala University of Technology Isan, Sakon Nakhon Campus, Phangkhon, Sakon Nakhon 47160, Thailand

³ Department of Animal Production Technology and Fisheries, Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang, Bangkok 10520, Thailand

⁴ Department of Plant Science, Faculty of Natural Resources, Rajamangala University of Technology Isan, Sakon Nakhon Campus, Phangkhon, Sakon Nakhon 47160, Thailand

⁵ Tropical Feed Resources Research and Development Center (TROFREC), Department of Animal Science, Faculty of Agriculture, Khon Kaen University, Khon Kaen 40002, Thailand

* Correspondence: pongsatorn.gu@rmuti.ac.th



Citation: Gunun, N.; Kaewpila, C.; Khota, W.; Polyorach, S.; Kimprasit, T.; Phlaetita, W.; Cherdthong, A.; Wanapat, M.; Gunun, P. The Effect of Indigo (*Indigofera tinctoria* L.) Waste on Growth Performance, Digestibility, Rumen Fermentation, Hematology and Immune Response in Growing Beef Cattle. *Animals* **2023**, *13*, 84. <https://doi.org/10.3390/ani13010084>

Academic Editors: Monica Isabella Cutrignelli, Maria N.T. Shipandeni and Bossima Ivan Koura

Received: 9 December 2022

Revised: 19 December 2022

Accepted: 22 December 2022

Published: 26 December 2022



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Simple Summary: Indigo waste is a by-product of the processing of natural indigo dye. Indigo waste could be utilized as a protein source in ruminant rations, which would reduce the cost of feed. We evaluated the effects of the inclusion of indigo waste in concentrate diets on the feed utilization, rumen fermentation, hematology, immune function and growth performance of growing beef cattle. The present findings suggest that the inclusion of indigo waste at low levels in concentrate diets maintains feed intake, digestibility, rumen fermentation or growth performance in growing beef cattle without affecting hematology or immune function.

Abstract: This experiment was conducted to assess the effect of indigo waste on the feed intake, digestibility, rumen fermentation, hematology, immune response and growth performance in growing beef cattle. Twenty crossbred beef cattle with an initial body weight (BW) of 145 ± 11 kg were fed four levels of indigo waste for 90 days in a trial. Additions of indigo waste at 0%, 10%, 20% and 30% in a concentrate diet using a completely randomized design (CRD). Cattle were fed concentrate at 1.8% BW, with rice straw fed ad libitum. The concentrate intake decreased linearly ($p = 0.01$) with the addition of indigo waste. The supplementation with indigo waste reduced dry matter (DM) and organic matter (OM) digestibility cubically ($p = 0.03$ and $p = 0.02$, respectively), while increasing neutral detergent fiber (NDF) digestibility cubically ($p = 0.02$). The final BW of beef cattle decreased linearly ($p = 0.03$) with the addition of indigo waste. The inclusion of indigo waste decreased the average daily gain (ADG) and gain-to-feed ratio (G:F) linearly ($p < 0.01$) from 0 to 90 days. The nutrient digestibility, ADG and G:F of beef cattle fed 10% indigo waste in the diet was similar when compared with the control (0% indigo waste). The ruminal pH, ammonia-nitrogen ($\text{NH}_3\text{-N}$) and total volatile fatty acid (VFA) concentrations were similar among treatments ($p > 0.05$). The proportion of acetate increased linearly ($p < 0.01$) but propionate decreased linearly ($p < 0.01$), resulting in an increase in the acetate to propionate ratio ($p < 0.01$) when cattle were fed with indigo waste supplementation. Increasing indigo waste levels did not influence blood urea nitrogen (BUN) levels, hematological parameters or immune responses (IgA, IgM and IgG) ($p > 0.05$). In conclusion, the inclusion of indigo waste at 10% in a concentrate diet did not have a negative effect on feed intake, nutrient digestibility, rumen fermentation, hematology, immune function or growth performance in growing beef cattle.

Keywords: indigo waste; feed intake; digestibility; average daily gain; volatile fatty acid; hematological indices; immune response; beef cattle

1. Introduction

In tropical areas, most ruminants are fed poor-quality roughages, especially rice straw [1,2]. The inclusion of concentrate, which is high in protein and other nutrients, can greatly enhance the efficiency of ruminant production [3]. However, the high cost and competitiveness of feedstuffs, especially soybean meal, has led to the search for alternative feeds, especially agricultural and agro-industrial by-products [4,5]. The sustainability of using by-products as feed for ruminants is based on their nutritive value, the availability of nutrients, rumen fermentation patterns, production responses and feed cost compared to conventional diets [6]. Consequently, they recirculate these wastes into the food supply chain [7]. Researchers have suggested that their waste products can be used as a source of protein in concentrate mixtures for ruminants at a rate of 10–30% [8–10]. In addition, the proper utilization of agro-industrial by-products in ruminant nutrition and the finding of new, inexpensive feed resources that promote ecological sustainability in feedstuffs are necessary [11,12].

Indigo (*Indigofera tinctoria* L.) is a legume plant classified in the family Fabaceae and distributed in Africa, South Asia and South East Asia, especially in Thailand [13]. Indigo is used in a variety of industries, including coloring food, cosmetics and pharmaceuticals but most commonly in textile products [14]. The indigo leaf contains 30.5% CP, 2.4% EE, 19.0% crude fiber and 36.6% carbohydrate [15]. Moreover, the indigo stem contains 5.1% CP, 2.0% EE, 54.5% crude fiber and 30.0% carbohydrate [15], as well as some phytonutrients, including condensed tannins, saponins and flavonoids [16,17]. Muda et al. [17] reported that supplementation with indigo leaf extract reduced fecal egg count but had no effect on growth performance or hematology in sheep.

A report from Thailand's Sakon Nakhon Province's Community Development Department showed that there were 120 groups of producers and enterprises producing indigo dye in 2015. Natural indigo dye making with indigo plant biomass is claimed to produce the best indigo dye purity, according to the traditional method [18]. After the dye extraction process, the remaining indigo waste consists of the stem and leaves. Through the recycling of by-products indigo waste can be used as animal feed and low-cost feed opportunities [19]. Indigo waste is believed to be rich in protein and has the potential to be used as a protein source to replace soybean meal in ruminant diets and reduce the cost of feed. In addition, the indigo plant has successfully served as a source for antibacterial, antioxidant, anti-inflammatory and immunomodulatory effects [20–23]. We hypothesize that the use of indigo waste will maintain feed utilization and growth performance while improving rumen fermentation, hematology and immune function in beef cattle. Therefore, the objective of this study is to assess the effects of the inclusion of indigo waste in concentrate on feed intake, nutrient digestibility, rumen fermentation, growth performance, hematological and immunological responses in beef cattle.

2. Materials and Methods

2.1. Ethical Procedure

The Animals Ethical Committee of the Rajamangala University of Technology Isan approved the animal care and experimental procedures (approval number 24/2564).

2.2. Animals, Treatments and Experimental Design

The study was conducted on the beef cattle farm of the Faculty of Natural Resources at Rajamangala University of Technology Isan, Sakon Nakhon Campus in Phangkhon, Sakon Nakhon, Thailand. After the dye extraction process, fresh indigo waste (leaf and stem) was collected from the indigo cloth community enterprise group, Baan Nohnrua, Pannanikom,

Sakon Nakhon, Thailand. They were sun-dried for four days, then ground before being added to the concentrate.

Twenty male crossbred (Brahman × Thai native) beef cattle with an average weight of 145 ± 11 kg were raised for a 90-day experiment. Each cattle was contained in a separate pen with access to fresh water. The cattle were fed concentrate at a rate of 1.8% of their body weight (BW), along with rice straw ad libitum, in two equal feedings at 08:00 h and 16:00 h. This study was conducted using a completely randomized design (CRD) to compare the indigo waste included in the concentrate at 0%, 10%, 20% and 30% on a DM basis (Table 1).

Table 1. Ingredients of the diet used in the experiment.

Item	Level of Indigo Waste (%DM)			
	0	10	20	30
Ingredient, kg dry matter (DM)				
Cassava chip	45.0	45.0	45.0	45.0
Rice bran	19.0	14.0	9.0	5.0
Soybean meal	14.0	11.0	8.5	7.0
Dried brewers' grains	17.5	15.5	13.0	8.5
Indigo waste	0.0	10.0	20.0	30.0
Molasses	2.0	2.0	2.0	2.0
Mineral and vitamin mixture	1.0	1.0	1.0	1.0
Urea	0.5	0.5	0.5	0.5
Salt	0.5	0.5	0.5	0.5
Sulfur	0.5	0.5	0.5	0.5

2.3. Feed Costs Analysis

The feed costs of the diets containing indigo waste were calculated using an input budgeting procedure according to Serrapica et al. [24]. However, the average costs of feed-stuffs at the local suppliers' gate were used in our calculation. The feed costs were adjusted based on the actual DM content and converted from baht to USD using 0.0286 currency. The feed costs (USD/kg DM) were cassava chip 0.32, rice bran 0.23, soybean meal 0.76, dried brewers' grains 0.41, indigo waste 0.06, molasses 0.29, mineral and vitamin mixture 1.51, urea 0.86, salt 0.29 and sulfur 0.91.

2.4. Data Collection and Sampling Procedures

Average daily gain (ADG) was estimated by weighing cattle at the beginning BW, 30 days, 60 days and the final BW at 90 days. Each morning, the offered and refused feed were recorded and taken for chemical analysis. Fecal samples were collected 56–60 days into the trial to conduct a digestibility test. Rectal sampling was used to obtain fresh feces (about 500 g). Each cattle's daily fresh fecal samples were pooled and chilled at 4 °C. Samples of feeds, refusals and feces were dried at 60 °C and ground (1-millimeter screen using the Cyclotech Mill; Tecator, Hoganas, Sweden). The contents of ash, ether extract (EE), crude protein (CP) [25], NDF and acid detergent fiber (ADF) [25,26] were determined. The modified vanillin-HCl procedure based on Burns [27] was used to measure the indigo waste's condensed tannin (CT) concentration. Methanol extraction was used to evaluate the crude saponins, as described by Kwon et al. [28] and modified by Pongchompu et al. [29]. The gross energy (GE) of the feeds was assessed by bomb calorimetry using an Oxygen bomb calorimeter (Parr Instrument Company, Moline, IL, USA), and acid-insoluble ash (AIA) was determined in the samples. AIA was created to assess the digestibility of nutrients [30].

On the 60th day of the experiment, 4 h after feeding, 200 mL of rumen fluid was taken with a stomach tube and a vacuum pump. The first 100 mL of the ruminal samples was thrown away to avoid contaminating them with saliva. The samples were then passed through four layers of cheesecloth and tested right away with a portable pH meter. Ruminal fluid samples were centrifuged at $16,000 \times g$ for 15 min at 4 °C, and the supernatant was kept at -20 °C. The ruminal samples were thawed and utilized to analyze $\text{NH}_3\text{-N}$ (Kjeltech Auto

1030 Analyzer, Tecator, Hoganas, Sweden) [31] and VFA (GC 8890; Agilent Technologies Ltd., Santa Clara County, CA, USA) [32].

Blood samples were taken at the same time as rumen fluid samples. Here, 10 mL of fresh blood was taken from the jugular vein of each animal. Blood urea nitrogen (BUN) was measured by Crocker's method [33]. A hematological analyzer (BCC-3000B; DIRUI, Gungoren/Istanbul, Turkey) was used to measure red blood cells (RBCs), hemoglobin, hematocrit, mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), white blood cells (WBCs), neutrophils, lymphocytes, monocytes, eosinophils and platelet count. The concentrations of immunoglobulins (IgA, IgG and IgM) were measured using the nephelometric method (Mispa-i3, Agappe Diagnostics Ltd., Ernakulam, Kerala, India).

2.5. Statistical Analysis

Using a completely randomized design, the general linear model (GLM) in SAS software was utilized to evaluate the data for variances [34]. Orthogonal polynomial contrasts (linear, quadratic and cubic) were used to compare the treatment trends statistically. To determine if an effect was significant, a $p < 0.05$ significance level was used.

3. Results

3.1. Feed Cost Analysis and Chemical Composition of Diets

The feed costs of the diets prepared without and with increasing indigo waste levels are presented in Table 2. Feed costs ranged from 29.25 to 39.59 USD/100 kg DM. The safe costs according to the replacements of indigo waste with soybean meal and dried brewer's grains were -3.63 , -7.08 and -10.35 USD/100 kg DM, respectively, for 10%, 20% and 30%. The indigo waste consists of 19.8% of CP, 46.6% of NDF, 32.4% of ADF, 5.4% of CT, 13.1% of saponins and 3487.5 kcal/kg DM of GE (Table 3). The local feed resources were used to create the concentrate diets, which had a 14.3–14.6% CP content. The NDF, ADF and GE content were increased according to the inclusion of indigo waste in the concentrate.

Table 2. Feed costs (USD/100 kg DM) of the experimental diets prepared with indigo waste.

Item	Level of Indigo Waste (%DM)			
	0	10	20	30
Cassava chip	14.29	14.29	14.29	14.29
Rice bran	4.41	3.25	2.09	1.16
Soybean meal	10.68	8.39	6.48	5.34
Dried brewers grains	7.10	6.29	5.28	3.45
Indigo waste	0.00	0.63	1.26	1.90
Urea	0.43	0.43	0.43	0.43
Molasses	0.57	0.57	0.57	0.57
Mineral and vitamin mixture	1.51	1.51	1.51	1.51
Salt	0.14	0.14	0.14	0.14
Sulfur	0.46	0.46	0.46	0.46
Total feeding costs	39.59	35.96	32.51	29.25
Safe costs (vs 0% indigo waste)	0.00	-3.63	-7.08	-10.35

Table 3. Chemical composition of concentrate, rice straw and indigo waste.

Item	Level of Indigo Waste (%DM)				Rice Straw	Indigo Waste
	0	10	20	30		
Chemical composition						
Dry matter, %	87.9	87.2	87.7	87.3	91.0	89.7
Organic matter, %DM	90.1	90.4	92.2	92.3	87.8	90.5
Crude protein, %DM	14.6	14.3	14.4	14.5	4.9	19.8
Neutral detergent fiber, %DM	43.1	46.0	52.1	59.5	73.7	46.6
Acid detergent fiber, %DM	27.8	28.1	29.8	30.5	51.9	32.4
Ash, %DM	9.9	9.6	7.8	7.7	12.2	9.5
Gross energy, kcal/kg DM	2861.2	3048.7	3214.8	3580.8	2797.9	3487.5
Condensed tannins, %DM	-	-	-	-	-	5.4
Crude saponins, %DM	-	-	-	-	-	13.1

3.2. Feed Intake and Digestibility

The increasing levels of indigo waste decreased linearly ($p = 0.01$) the concentrate intake from 0 to 90 days, but it did not affect the roughage intake ($p > 0.05$) (Table 4). The inclusion of indigo waste decreased total intake linearly ($p = 0.02$) from 61 to 90 days. The digestibility of CP and ADF was similar among the groups ($p > 0.05$), whereas DM and OM digestibility decreased cubically ($p = 0.03$ and $p = 0.02$, respectively) with increasing levels of indigo waste (Table 5). Furthermore, the digestibility of DM and OM were lowest in cattle fed with the inclusion of indigo waste at 30% in concentrate. The NDF digestibility increased cubically with the addition of indigo waste ($p = 0.02$).

Table 4. Effect of indigo waste on feed intake in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
Dry matter intake, kg/d								
Concentrate								
0 to 30 d	2.8	2.5	2.4	2.4	0.19	0.11	0.61	0.93
31 to 60 d	3.4	3.1	2.7	2.6	0.22	0.01	0.69	0.59
61 to 90 d	3.8	3.5	3.1	2.8	0.26	<0.01	0.94	0.98
0 to 90 d	3.3	3.0	2.7	2.6	0.21	0.01	0.79	0.87
Roughage								
0 to 30 d	1.9	1.7	1.8	1.8	0.21	0.69	0.52	0.73
31 to 60 d	2.2	1.9	1.9	1.9	0.20	0.38	0.32	0.75
61 to 90 d	2.2	2.0	2.0	2.0	0.21	0.42	0.65	0.74
0 to 90 d	2.1	1.9	1.9	1.9	0.19	0.47	0.47	0.73
Total intake								
0 to 30 d	4.7	4.2	4.2	4.2	0.36	0.27	0.52	0.81
31 to 60 d	5.6	5.0	4.6	4.6	0.40	0.07	0.47	0.88
61 to 90 d	6.1	5.5	5.2	4.7	0.39	0.02	0.84	0.85
0 to 90 d	5.5	4.9	4.7	4.5	0.37	0.07	0.60	0.93

Table 5. Effect of indigo waste on nutrient digestibility in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
Digestibility, %								
Dry matter	54.9	55.3	60.2	51.3	1.75	0.48	0.02	0.03
Organic matter	58.8	58.7	64.1	55.2	1.74	0.50	0.02	0.02
Crude protein	49.3	48.3	49.3	47.3	2.85	0.74	0.86	0.70
Neutral detergent fiber	49.1	50.8	60.8	56.1	1.90	<0.01	0.13	0.02
Acid detergent fiber	46.9	43.2	47.2	44.2	1.75	0.63	0.86	0.10

3.3. Performance

The BW in the cattle trial was similar among treatments ($p > 0.05$) on days 0, 30 and 60, whereas the BW on days 90 decreased linearly with the addition of indigo waste ($p = 0.03$) (Table 6). The ADG and G:F decreased linearly ($p < 0.01$) with increasing levels of indigo waste from 0 to 90 days. In addition, ADG and G:F were lower with the addition of indigo waste at 20–30% in concentrate diets.

Table 6. Effect of indigo waste on growth performance in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
Body weight, kg								
Initial	156.4	138.4	140.8	143.6	12.33	0.52	0.41	0.72
30 d	185.2	167.2	158.4	159.6	13.05	0.16	0.47	0.98
60 d	213.6	191.8	179.2	176.0	14.52	0.07	0.53	0.99
Final	238.4	214.2	197.6	190.4	15.55	0.03	0.59	0.98
ADG, kg/d								
0 to 30 d	0.96	0.96	0.58	0.52	0.09	<0.01	0.76	0.13
31 to 60 d	0.94	0.80	0.70	0.54	0.07	<0.01	0.89	0.76
61 to 90 d	0.82	0.76	0.62	0.48	0.09	0.01	0.67	0.85
0 to 90 d	0.91	0.84	0.63	0.51	0.08	<0.01	0.45	0.82
G:F								
0 to 30 d	0.20	0.22	0.15	0.13	0.03	0.02	0.46	0.14
31 to 60 d	0.18	0.16	0.15	0.12	0.01	0.02	0.64	0.93
61 to 90 d	0.14	0.14	0.12	0.10	0.02	0.04	0.51	0.86
0 to 90 d	0.17	0.18	0.14	0.12	0.06	<0.01	0.41	0.33

3.4. Rumen Fermentation

The use of indigo waste in concentrate feed for cattle also had no effect on ruminal pH, $\text{NH}_3\text{-N}$ or total VFA ($p > 0.05$) (Table 7). The proportions of propionate (C3), valerate (C5) and iso-valerate (i-C5) decreased linearly ($p < 0.01$, $p < 0.01$ and $p = 0.01$), while acetate (C2) and C2:C3 increased linearly ($p < 0.01$) by including levels of indigo waste.

Table 7. Effect of indigo waste on rumen fermentation in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
pH	6.8	6.9	6.9	6.9	0.07	0.38	0.23	0.78
$\text{NH}_3\text{-N}$, mg/dL	19.6	21.5	20.6	16.8	1.81	0.26	0.14	1.00
Total VFA, mmol/d	54.7	54.6	58.0	59.8	2.79	0.15	0.73	0.68
VFA, mol/100 mol								
Acetate (C2)	58.1	60.8	61.3	62.7	0.70	<0.01	0.37	0.32
Propionate (C3)	24.2	20.2	20.1	18.6	1.10	<0.01	0.26	0.29
Butyrate (C4)	13.8	15.4	15.1	15.5	0.77	0.19	0.45	0.42
Iso-butyrate (i-C4)	0.9	0.9	1.0	0.8	0.09	0.63	0.29	0.46
Valerate (C5)	1.5	1.4	1.3	1.3	0.05	<0.01	0.13	0.72
Iso-valerate (i-C5)	1.4	1.3	1.2	1.0	0.31	0.01	0.91	0.74
C2:C3	2.4	3.0	3.1	3.5	0.40	<0.01	0.58	0.27

3.5. Blood Urea Nitrogen and Hematological Parameters

The increasing levels of indigo waste did not affect the concentrations of BUN, RBCs, hemoglobin, hematocrit, MCV, MCH, WBCs, neutrophils, lymphocytes, monocytes, eosinophils and platelet count ($p > 0.05$) (Table 8).

Table 8. Effect of indigo waste on BUN and hematology in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
BUN, mg/dL	8.2	12.0	10.4	9.4	1.64	0.78	0.16	0.42
Red blood cells, 10 ¹² /L	4.9	4.8	4.9	5.1	0.40	0.73	0.71	0.99
Hemoglobin, g/dL	7.3	7.2	7.5	7.6	0.66	0.73	0.92	0.87
Hematocrit, %	22.8	21.8	22.6	22.8	1.97	0.92	0.76	0.78
MCV, 10 ⁶ /fL	46.2	45.8	46.2	45.4	0.43	0.32	0.65	0.32
MCH, pg	21.8	20.6	19.8	20.6	1.32	0.46	0.46	0.84
White blood cells, 10 ⁹ /L	13.9	14.2	10.7	12.9	1.79	0.42	0.59	0.25
Neutrophils, %	30.8	34.6	25.2	30.4	3.89	0.55	0.85	0.13
Lymphocytes, %	67.8	65.0	74.2	68.8	4.01	0.50	0.75	0.15
Monocytes, %	0	0	0	0	NA	NA	NA	NA
Eosinophils, %	1.4	0.4	0.6	0.8	0.57	0.54	0.31	0.64
Platelet count, 10 ⁹ /L	214.0	266.0	274.3	219.0	33.98	0.88	0.17	0.91

MCV—mean corpuscular volume; MCH—mean corpuscular hemoglobin.

3.6. Immune Response

The use of indigo waste in concentrate feed for beef cattle had no effect on IgA, IgM and IgG ($p > 0.05$) (Table 9).

Table 9. Effect of indigo waste on immune response in growing beef cattle.

Item	Level of Indigo Waste (%DM)				SEM	Contrast		
	0	10	20	30		Linear	Quadratic	Cubic
IgA, mg/dL	85.6	90.2	89.8	94.0	3.03	0.09	0.94	0.49
IgM, mg/dL	49.2	48.2	48.4	40.8	3.93	0.17	0.41	0.61
IgG, mg/dL	429.2	423.4	396.6	414.2	18.4	0.39	0.53	0.44

4. Discussion

The CP content of the indigo waste was 19.8%. However, Bhatta et al. [16] indicated that the CP of indigo leaf was 26.0% DM. Because indigo waste is composed of leaves and stems, it has a lower CP than indigo leaf. The inclusion of indigo waste increases the fiber and GE content in the concentrate. The indigo waste contained NDF, ADF and GE at 46.6%, 32.4% DM and 3487.5 kcal/kg DM, respectively. The treatment chemical composition indicates that adding indigo waste increased the fiber concentration and gross energy.

It is generally accepted that the NDF concentrations of the diet, which limit the DM digestibility of the diet, are a major factor limiting the voluntary DM intake of animals [35]. The inclusion of indigo waste in concentrate diets decreased concentrate intake, total intake and DM digestibility in the present study. Indigo waste (stem and leaf) has a high fiber content. Increased fiber content in concentrate diets with the addition of indigo waste results in reduced feed intake and the digestibility of DM and OM with increasing levels of indigo waste. Moreover, indigo waste has a high GE content, and GE increases when indigo waste levels are increased in concentrate diets. However, DM and OM digestibility declined. This could be due to the high-fiber energy of the by-product with reduced digestible energy when the addition of indigo waste, especially at 30% in concentrate, results in a decrease in beef cattle's DM and OM digestibility. The greater NDF digestibility of the diet was mainly due to enhanced hemicellulose digestion [36]. In the current study, increasing levels of indigo waste by 20% in concentrate diets improved digestibility. Similarly, Kongphitee et al. [37] reported that NDF digestibility increased with increasing levels of by-product in the diets of beef cattle. Lyu et al. [38] reported that the inclusion of by-products in diets enhanced the digestibility of NDF in dairy cows. However, ADF digestibility was not affected by indigo waste supplementation. This is plausible because a higher fraction of

the hemicellulose is present in indigo waste in concentrated diets, and therefore, NDF digestibility is increased but cellulose digestion is not affected.

ADG and feed efficiency are essential components of growing beef cattle production efficiency. ADG is an essential component of growing beef cattle production efficiency [39]. There was a linear decrease in ADG and G:F with an increasing level of indigo waste in growing beef cattle from 0 to 90 days. The ADG as well as the G:F ratio appear to be lower when cattle were fed diets containing 20–30% indigo waste from 0 to 90 days. Kanjanapruthipong et al. [40] reported that increasing NDF content in the diets decreased the digestibility of nutrients and ADG in dairy cattle. These results may be due to the high NDF content and also the lower DM intake, nutrient availability, and VFA, particularly propionate, when indigo waste was gradually increased in concentrate diets, resulting in decreased growth performance in growing beef cattle. The ADG required to obtain the target body weight is based on the body weight at the beginning of the trial and feed efficiency. The addition of indigo waste reduces the ADG and G:F, thereby decreasing the final BW (90 days of a trial). These results suggest that the use of indigo waste at 10% in concentrate diets is suitable for growth performance in growing beef cattle.

Ruminal pH is among the major fermentation factors that directly affect microbial ecology and, thereby, ruminal fermentation [41]. In our experiment, the rumen pH range for all diets was 6.8 to 6.9, and the optimum range for microbial activity in the rumen was 6.5–7.0 [4,42]. The ruminal pH was similar among treatments, which indicates that the inclusion of indigo waste did not change rumen ecology or fermentation in tropical beef cattle. The main nitrogen source for protein synthesis in the rumen is $\text{NH}_3\text{-N}$ [8]. In the present investigation, indigo waste levels had no effect on the ruminal $\text{NH}_3\text{-N}$ concentration, indicating that indigo waste seemed to have no effect on protein degradation by microorganisms in the rumen. The ruminal $\text{NH}_3\text{-N}$ concentrations ranged from 16.8 to 21.5 mg/dL, which is closer to the optimum range (15 to 30 mg/dL) [1,43].

The pattern of rumen fermentation was changed by the different diets fed to ruminants. As VFA production serves as an energy source for growth performance in ruminants, it is crucial to understand their metabolism [44]. Acetate, propionate and butyrate are the main VFAs produced in the rumen, and their concentrations vary depending on the feed ingredient, feed intake, digestibility, rumen ecology and the rate of passage [45]. When indigo waste was added to the diets, the rumen VFA profile changed, with an increase in acetate and a decline in propionate; this also caused the C2:C3 ratio to increase. The association between the C2:C3 ratio and feed has been explained by the metabolic properties of fiber- and starch-degrading bacteria [46]. Most structural carbohydrate fermentation, which leads to the production of acetate, is caused by cellulolytic bacteria. The major cellulolytic bacteria are thought to be *Ruminococcus albus*, *R. flavefaciens* and *Fibrobacter succinogenes*, which produce more acetate in the rumen [47]. Several different types of bacteria, including those in the family *Propionibacteriaceae*, produce propionate as an end product in the rumen [48]. A high concentration of starch in the diet is more likely to ferment into propionate production in the rumen, making it advantageous for the production of glucose, which helps the meat animal. [49]. The addition of a high-fiber by-product feed increased acetate levels while reducing propionate in the rumen [50]. Wanapat et al. [46] found that adding high amounts of structural carbohydrates to the diet increased the proportions of acetate in the rumen, which caused the C2:C3 ratio to be higher. This means that indigo waste diets had more fermentable structural carbohydrates, such as hemicellulose, which is thought to increase acetate production and decrease propionate production.

The BUN concentration is often used to assess protein supplies and metabolic concerns related to animal diseases [51]. The addition of indigo waste to concentrate had no effect on BUN, which ranged from 8.2 to 12.0 mg/dL, which was within the usual range of 8 to 14 mg/dL in tropical beef cattle [52,53]. Cattle health and nutrition and the cause of an abnormality or malfunction in cattle are frequently tested using hematological analysis [42,54]. The inclusion of indigo waste did not influence all hematological indicators. Similarly,

Muda et al. [17] reported that the hematology of sheep was not affected by indigo leaf extract supplementation. These results indicate that the addition of indigo waste as a feed had no negative effects on the health status of tropical beef cattle. Growing beef cattle had normal concentrations of RBCs, hemoglobin, hematocrit, WBCs, neutrophils, lymphocytes and eosinophils when compared to our previous study [42,51,55]. In addition, previous reports have demonstrated that concentrations of MCV, MCH [51,56], monocytes [57] and platelet count [57,58] in ruminant blood are within the accepted range.

Phytonutrients from tropical plants, which have been thought of as possible additions to animal feed, may affect immune and inflammatory responses. There was a high amount of phytonutrients such as total phenolics, total tannins, saponins and flavonoids in the indigo [22]. Indigo leaf extract has numerous pharmacological effects, including anti-inflammatory, antioxidant, antibacterial, antiviral and other activities [59]. In addition, this plant improved the immune response and demonstrated its immunostimulating efficacy *in vitro* and in rats [20,21]. In the current study, the addition of indigo waste had no influence on IgA, IgM or IgG immune responses. These results suggest that some phytonutrients may be water-soluble during the dye extraction procedure or that indigo waste contains crude leaf and stem, resulting in a weak effect on the immune response in cattle.

5. Conclusions

The inclusion of indigo waste in the concentrate did not affect the $\text{NH}_3\text{-N}$ concentrations, hematological parameters or immune response. However, the addition of indigo waste had an effect on the feed intake, digestibility of DM and OM and growth performance. Furthermore, incorporating indigo waste into the diets reduced propionate while increasing the acetate proportion. The addition of 10% indigo waste to the concentrate showed that it could be used as a source of protein and sustain growing beef cattle feed intake, nutrient digestibility, rumen fermentation and growth performance. In order to assess the effects of indigo waste on carcass characteristics and meat quality in beef cattle, more research needs to be conducted.

Author Contributions: Planning and design of the study, P.G. and N.G.; conducting and sampling, P.G., N.G., W.P. and S.P.; sample analysis, P.G., N.G., C.K. and W.K.; statistical analysis, P.G., N.G. and S.P.; manuscript drafting, P.G. and N.G.; manuscript editing and finalizing, P.G., N.G., T.K., S.P., A.C., M.W., C.K. and W.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research project is supported by Thailand Science Research and Innovation (TSRI) (contract No. FRB650059/SNK/08).

Institutional Review Board Statement: All animal procedures were approved by the Animal Ethical Committee of the Rajamangala University of Technology Isan (approval number 24/2564 on 29 January 2021).

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors are thankful the Department of Animal Science, Faculty of Natural Resources, Rajamangala University of Technology Isan, Sakon Nakhon Campus, and the Department of Animal Science, Faculty of Technology, Udon Thani Rajabhat University for the use of their research facilities.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Gunun, P.; Wanapat, M.; Anantasook, N. Effects of physical form and urea treatment of rice straw on rumen fermentation, microbial protein synthesis and nutrient digestibility in dairy steers. *Asian Australas. J. Anim. Sci.* **2013**, *26*, 1689–1697. [[CrossRef](#)] [[PubMed](#)]
2. Wanapat, M.; Foiklang, S.; Sukjai, S.; Tamkhonburi, P.; Gunun, N.; Gunun, P.; Phesatcha, K.; Norrapoke, T.; Kang, S. Feeding tropical dairy cattle with local protein and energy sources for sustainable production. *J. Appl. Anim. Res.* **2018**, *46*, 232–236. [[CrossRef](#)]
3. Makkar, H.P.; Beever, D. Optimization of feed use efficiency in ruminant production systems. In Proceedings of the FAO Symposium, Bangkok, Thailand, 27 November 2012; FAO Animal Production and Health Division: Rome, Italy, 2013.
4. Gunun, N.; Khejornsart, P.; Polyorach, S.; Kaewpila, C.; Kimprasit, T.; Sanjun, I.; Cherdthong, A.; Wanapat, M.; Gunun, P. Utilization of mao (*Antidesma thwaitesianum* Muell. Arg.) pomace meal to substitute rice bran on feed utilization and rumen fermentation in tropical beef cattle. *Vet. Sci.* **2022**, *9*, 585. [[CrossRef](#)] [[PubMed](#)]
5. Miranda, M.S.; Arcaro, J.R.P.; Saran Netto, A.; Silva, S.L.; Pinheiro, M.G.; Leme, P.R. Effects of partial replacement of soybean meal with other protein sources in diets of lactating cows. *Animal* **2019**, *13*, 1403–1411. [[CrossRef](#)] [[PubMed](#)]
6. Halmemies-Beauchet-Filleau, A.; Rinne, M.; Lamminen, M.; Mapato, C.; Ampapon, T.; Wanapat, M.; Vanhatalo, A. Review: Alternative and novel feeds for ruminants: Nutritive value, product quality and environmental aspects. *Animal* **2018**, *12*, s295–s309. [[CrossRef](#)] [[PubMed](#)]
7. Palangi, V.; Kaya, A.; Kaya, A.; Giannenas, I. Ecofriendly usability of mushroom cultivation substrate as a ruminant feed: Anaerobic digestion using gas production techniques. *Animals* **2022**, *12*, 1583. [[CrossRef](#)]
8. Gunun, N.; Ouppamong, T.; Khejornsart, P.; Cherdthong, A.; Wanapat, M.; Polyorach, S.; Kaewpila, C.; Kang, S.; Gunun, P. Effects of rubber seed kernel fermented with yeast on feed utilization, rumen fermentation and microbial protein synthesis in dairy heifers. *Fermentation* **2022**, *8*, 288. [[CrossRef](#)]
9. Rodrigues, T.C.G.C.; Santos, S.A.; Cirne, L.G.A.; Pina, D.S.; Alba, H.D.R.; de Araújo, M.L.G.M.L.; Silva, W.P.; Nascimento, C.O.; Rodrigues, C.S.; Tosto, M.S.L.; et al. Palm kernel cake in high-concentrate diets for feedlot goat kids: Nutrient intake, digestibility, feeding behavior, nitrogen balance, blood metabolites, and performance. *Trop. Anim. Health Prod.* **2021**, *53*, 454. [[CrossRef](#)]
10. Janicek, B.N.; Kononoff, P.J.; Gehman, A.M.; Doane, P.H. The effect of feeding dried distillers grains plus solubles on milk production and excretion of urinary purine derivatives. *J. Dairy Sci.* **2008**, *91*, 3544–3553. [[CrossRef](#)]
11. Yang, K.; Qing, Y.; Yu, Q.; Tang, X.; Chen, G.; Fang, R.; Liu, H. By-product feeds: Current understanding and future perspectives. *Agriculture* **2021**, *11*, 207. [[CrossRef](#)]
12. Besharati, M.; Palangi, V.; Moaddab, M.; Nemati, Z.; Pliego, A.B.; Salem, A.Z. Influence of cinnamon essential oil and monensin on ruminal biogas kinetics of waste pomegranate seeds as a biofriendly agriculture environment. *Waste Biomass Valorization* **2020**, *12*, 2333–2342. [[CrossRef](#)]
13. Senasri, N.; Sriyasad, P.; Suwanpakdee, S.; Chumnanka, N.; Tongkasee, P.; Sriputhorn, K. Toxicity of indigo dye-contaminated water on silver barb (*Barbonymus gonionotus*) and pathology in the gills. *EnvironmentAsia* **2022**, *15*, 106–115.
14. Tayade, P.B.; Adivarekar, R.V. Extraction of indigo dye from *Couroupita guianensis* and its application on cotton fabric. *Fash. Text.* **2014**, *1*, 16. [[CrossRef](#)]
15. Alagbe, J.O. Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and root of *Indigofera tinctoria*. *Int. J. Integr. Educ.* **2020**, *3*, 150–157. [[CrossRef](#)]
16. Bhatta, R.; Saravanan, M.; Baruah, L.; Sampath, K.T.; Prasad, C.S. Effect of plant secondary compounds on in vitro methane, ammonia production and ruminal protozoa population. *J. Appl. Microbiol.* **2013**, *115*, 455–465. [[CrossRef](#)]
17. Muda, I.; Prastowo, J.; Nurcahyo, W.; Sarmin, S. Anthelmintic effect of *Indigofera tinctoria* L on *Haemonchus contortus* obtained from sheep in Indonesia. *Vet. World* **2021**, *14*, 1272–1278. [[CrossRef](#)]
18. Pattanaik, L.; Naik, S.N.; Hariprasad, P. Valorization of waste *Indigofera tinctoria* L. biomass generated from indigo dye extraction process-potential towards biofuels and compost. *Biomass Convers. Biorefin.* **2019**, *9*, 445–457. [[CrossRef](#)]
19. Van Zanten, H.H.; Van Ittersum, M.K.; De Boer, I.J. The role of farm animals in a circular food system. *Glob. Food Secur.* **2019**, *21*, 18–22. [[CrossRef](#)]
20. Boothapandi, M.; Ramanibai, R. Immunomodulatory activity of *Indigofera tinctoria* leaf extract on in vitro macrophage responses and lymphocyte proliferation. *Int. J. Pharm. Pharm. Sci.* **2016**, *8*, 58–63.
21. Madakkannu, B.; Ravichandran, R. In vivo immunoprotective role of *Indigofera tinctoria* and *Scoparia dulcis* aqueous extracts against chronic noise stress induced immune abnormalities in Wistar albino rats. *Toxicol. Rep.* **2017**, *4*, 484–493. [[CrossRef](#)]
22. Sharma, V.; Agarwal, A. Physicochemical and antioxidant assays of methanol and hydromethanol extract of ariel parts of *Indigofera tinctoria* Linn. *Indian J. Pharm. Sci.* **2015**, *77*, 729–734. [[CrossRef](#)]
23. Vijayan, R.; Joseph, S.; Mathew, B. Indigofera tinctoria leaf extract mediated green synthesis of silver and gold nanoparticles and assessment of their anticancer, antimicrobial, antioxidant and catalytic properties. *Artif. Cells Nanomed. Biotechnol.* **2018**, *46*, 861–871. [[CrossRef](#)]
24. Serrapica, F.; Masucci, F.; De Rosa, G.; Calabrò, S.; Lambiase, C.; Di Francia, A. Chickpea can be a valuable local produced protein feed for organically reared, native bulls. *Animals* **2021**, *11*, 2353. [[CrossRef](#)] [[PubMed](#)]
25. AOAC. *Official Method of Analysis*, 20th ed.; Association of Official Analytical Chemists: Arlington, VA, USA, 2016.

26. Udén, P.; Robinson, P.H.; Wiseman, J. Use of detergent system terminology and criteria for submission of manuscripts on new, or revised, analytical methods as well as descriptive information on feed analysis and/or variability. *Anim. Feed Sci. Technol.* **2005**, *118*, 181–186. [[CrossRef](#)]
27. Burns, R.E. Method for estimation of tannin in the grain sorghum. *Agron. J.* **1971**, *163*, 511–512. [[CrossRef](#)]
28. Kwon, J.H.; Belanger, J.; Pare, M.R.; Yaylayan, V.A. Application of the microwave-assisted process (MAPTM) to the fast excretion of ginseng saponins. *Food Res. Int.* **2003**, *36*, 491–498. [[CrossRef](#)]
29. Pongchompu, O.; Wanapat, M.; Wachirapakorn, C.; Wanapat, S.; Cherdthong, A. Manipulation of ruminal fermentation and methane production by dietary saponins and tannins from mangosteen peel and soapberry fruit. *Arch. Anim. Nutr.* **2009**, *63*, 389–400. [[CrossRef](#)]
30. Van Keulen, J.; Young, B.A. Evaluation of acid insoluble ash as a neutral marker in ruminant digestibility studies. *J. Anim. Sci.* **1977**, *44*, 282–287. [[CrossRef](#)]
31. AOAC. *Official Methods of Analysis*, 16th ed.; Association of Official Analytical Chemists: Arlington, VA, USA, 1995.
32. Cai, Y. Analysis method for silage. In *Field and Laboratory Methods for Grassland Science*; Japanese Society of Grassland Science, Ed.; Tosho Printing Co., Ltd.: Tokyo, Japan, 2004; pp. 279–282.
33. Crocker, C.L. Rapid determination of urea nitrogen in serum or plasma without deproteinization. *Am. J. Med. Technol.* **1967**, *33*, 361–365.
34. Statistical Analysis Systems (SAS). SAS/STAT User's Guide. In *Statistical Analysis Systems Institute*, 5th ed.; SAS Institute Inc.: Cary, NC, USA, 1996.
35. Harper, K.J.; McNeill, D.M. The role iNDF in the regulation of feed intake and the importance of its assessment in subtropical ruminant systems (the role of iNDF in the regulation of forage intake). *Agriculture* **2015**, *5*, 778–790. [[CrossRef](#)]
36. Miron, J.; Adin, G.; Solomon, R.; Nikbachat, M.; Zenou, A.; Yosef, E.; Brosh, A.; Shabtay, A.; Asher, A.; Gacitua, H.; et al. Effects of feeding cows in early lactation with soy hulls as partial forage replacement on heat production, retained energy and performance. *Anim. Feed Sci. Technol.* **2010**, *155*, 9–17. [[CrossRef](#)]
37. Kongphitee, K.; Sommart, K.; Phonbumrung, T.; Gunha, T.; Suzuki, T. Feed intake, digestibility and energy partitioning in beef cattle fed diets with cassava pulp instead of rice straw. *Asian Australas. J. Anim. Sci.* **2018**, *31*, 1431–1441. [[CrossRef](#)] [[PubMed](#)]
38. Lyu, J.; Yang, Z.; Wang, E.; Liu, G.; Wang, Y.; Wang, W.; Li, S. Possibility of using by-products with high NDF content to alter the fecal short chain fatty acid profiles, bacterial community, and digestibility of lactating dairy cows. *Microorganisms* **2022**, *10*, 1731. [[CrossRef](#)] [[PubMed](#)]
39. Nielsen, M.K.; MacNeil, M.D.; Dekkers, J.C.M.; Crews, D.H., Jr.; Rathje, T.A.; Enns, R.M.; Weaber, R.L. Review: Life-cycle, total industry genetic improvement of feed efficiency in beef cattle: Blueprint for the Beef Improvement Federation. *Prof. Anim. Sci.* **2013**, *29*, 559–565. [[CrossRef](#)]
40. Kanjanapruthipong, J.; Buatong, N.; Buaphan, S. Effects of roughage neutral detergent fiber on dairy performance tropical conditions. *Asian Australas. J. Anim. Sci.* **2001**, *14*, 1400–1404. [[CrossRef](#)]
41. Li, M.; Penner, G.B.; Hernandez-Sanabria, E.; Oba, M.; Guan, L.L. Effects of sampling location and time, and host animal on assessment of bacterial diversity and fermentation parameters in the bovine rumen. *J. Appl. Microbiol.* **2009**, *107*, 1929–1934. [[CrossRef](#)]
42. Gunun, N.; Sanjun, I.; Kaewpila, C.; Foiklang, S.; Cherdthong, A.; Wanapat, M.; Polyorach, S.; Khota, W.; Kimprasit, T.; Kesorn, P.; et al. Effect of dietary supplementation of hydrolyzed yeast on growth performance, digestibility, rumen fermentation, and hematology in growing beef cattle. *Animals* **2022**, *12*, 2473. [[CrossRef](#)]
43. Cherdthong, A.; Khonkhaeng, B.; Seankamsorn, A.; Supapong, C.; Wanapat, M.; Gunun, N.; Gunun, P.; Chanjula, P.; Polyorach, S. Effects of feeding fresh cassava root with high-sulfur feed block on feed utilization, rumen fermentation, and blood metabolites in Thai native cattle. *Trop. Anim. Health Prod.* **2018**, *50*, 1365–1371. [[CrossRef](#)]
44. Wang, L.; Zhang, G.; Li, Y.; Zhang, Y. Effects of high forage/concentrate diet on volatile fatty acid production and the microorganisms involved in VFA production in cow rumen. *Animals* **2020**, *10*, 223. [[CrossRef](#)]
45. Bica, R.; Palarea-Albaladejo, J.; Lima, J.; Uhrin, D.; Miller, G.A.; Bowen, J.M.; Pacheco, D.; Macrae, A.; Dewhurst, R.J. Methane emissions and rumen metabolite concentrations in cattle fed two different silages. *Sci. Rep.* **2022**, *12*, 5441. [[CrossRef](#)]
46. Wanapat, M.; Gunun, P.; Anantasook, N.; Kang, S. Changes of rumen pH, fermentation and microbial population as influenced by different ratios of roughage (rice straw) to concentrate in dairy steers. *J. Agric. Sci.* **2014**, *152*, 675–685. [[CrossRef](#)]
47. Cherdthong, A.; Wanapat, M.; Saenkamsorn, A.; Supapong, C.; Anantasook, N.; Gunun, P. Improving rumen ecology and microbial population by dried rumen digesta in beef cattle. *Trop. Anim. Health Prod.* **2015**, *47*, 921–926. [[CrossRef](#)] [[PubMed](#)]
48. Chen, J.; Harstad, O.M.; Mcallister, T.; Drschi, P.; Holo, H. Propionic acid bacteria enhance ruminal feed degradation and reduce methane production in vitro. *Acta Agric. Scand. Sect. A—Anim. Sci.* **2020**, *69*, 169–175. [[CrossRef](#)]
49. Han, C.; Guo, Y.; Cai, X.; Yang, R. Starch properties, nutrients profiles, in vitro ruminal fermentation and molecular structure of corn processed in different ways. *Fermentation* **2022**, *8*, 315. [[CrossRef](#)]
50. Li, R.; Teng, Z.; Lang, C.; Zhou, H.; Zhong, W.; Ban, Z.; Yan, X.; Yang, H.; Farouk, M.H.; Lou, Y. Effect of different forage-to-concentrate ratios on ruminal bacterial structure and real-time methane production in sheep. *PLoS ONE* **2019**, *14*, e0214777. [[CrossRef](#)]
51. Oupppamong, T.; Gunun, N.; Tamkhonburee, C.; Khejornsart, P.; Kaewpila, C.; Kesorn, P.; Kimprasit, T.; Cherdthong, A.; Wanapat, M.; Polyorach, S.; et al. Fermented rubber seed kernel with yeast in the diets of tropical lactating dairy cows: Effects on feed intake, hematology, microbial protein synthesis, milk yield and milk composition. *Vet. Sci.* **2022**, *9*, 360. [[CrossRef](#)]

52. Supapong, C.; Cherdthong, A.; Wanapat, M.; Chanjula, P.; Uriyapongson, S. Effects of sulfur levels in fermented total mixed ration containing fresh cassava root on feed utilization, rumen characteristics, microbial protein synthesis, and blood metabolites in Thai native beef cattle. *Animals* **2019**, *9*, 261. [[CrossRef](#)]
53. Phesatcha, K.; Phesatcha, B.; Wanapat, M.; Cherdthong, A. The effect of yeast and roughage concentrate ratio on ruminal pH and protozoal population in Thai native beef cattle. *Animals* **2022**, *12*, 53. [[CrossRef](#)]
54. Saeed, O.A.; Sazili, A.Q.; Akit, H.; Alimon, A.R.; Samsudin, A.A. Effects of corn supplementation into PKC-urea treated rice straw basal diet on hematological indices and serum mineral level in lambs. *Animals* **2019**, *9*, 781. [[CrossRef](#)]
55. Gunun, P.; Gunun, N.; Khejornsart, P.; Ouppamong, T.; Cherdthong, A.; Wanapat, M.; Sililaophaisan, S.; Yuangklang, C.; Polyorach, S.; Kenchaiwong, W.; et al. Effects of *Antidesma thwaitesianum* Muell. Arg. pomace as a source of plant secondary compounds on digestibility, rumen environment, hematology, and milk production in dairy cows. *Anim. Sci. J.* **2019**, *90*, 372–381. [[CrossRef](#)]
56. Herman, N.; Trumel, C.; Geffré, A.; Braun, J.-P.; Thibault, M.; Schelcher, F.; Bourgès-Abella, N. Hematology reference intervals for adult cows in France using the Sysmex XT-2000iV analyzer. *J. Vet. Diagn. Investig.* **2018**, *30*, 678–687. [[CrossRef](#)] [[PubMed](#)]
57. George, J.W.; Snipes, J.; Lane, V.M. Comparison of bovine hematology reference intervals from 1957 to 2006. *Vet. Clin. Pathol.* **2010**, *39*, 138–148. [[CrossRef](#)] [[PubMed](#)]
58. Wood, D.; Quiroz-Rocha, G.F. Normal hematology of cattle. In *Schalm's Veterinary Hematology*, 6th ed.; Weiss, D.J., Wardrop, K.J., Eds.; Wiley: Ames, IA, USA, 2010; pp. 829–835.
59. Qi-Yue, Y.; Ting, Z.; Ya-Nan, H.; Sheng-Jie, H.; Xuan, D.; Li, H.; Chun-Guang, X. From natural dye to herbal medicine: A systematic review of chemical constituents, pharmacological effects and clinical applications of indigo naturalis. *Chin. Med.* **2020**, *15*, 127. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.