





Article

Phase IIa Clinical Biomarker Trial of Dietary Arginine Restriction and Aspirin in Colorectal Cancer Patients

Jason A. Zell ^{1,2,*} , Thomas H. Taylor ³, C. Gregory Albers ⁴ , Joseph C. Carmichael ⁵ , Christine E. McLaren ^{2,6}, Lari Wenzel ^{2,6} and Michael J. Stamos ⁵ 

¹ Division of Hematology/Oncology, Department of Medicine, University of California Irvine Medical Center, Orange, CA 92868, USA

² Chao Family Comprehensive Cancer Center, University of California Irvine Medical Center, Orange, CA 92868, USA

³ Department of Epidemiology & Biostatistics, University of California Irvine, Irvine, CA 92697, USA

⁴ Division of Gastroenterology, Department of Medicine, University of California Irvine Medical Center, Orange, CA 92868, USA

⁵ Division of Colorectal Surgery, Department of Surgery, University of California Irvine Medical Center, Orange, CA 92868, USA

⁶ Department of Medicine, University of California Irvine Medical Center, Orange, CA 92868, USA

* Correspondence: jzell@hs.uci.edu; Tel.: +1-714-456-5153; Fax: +1-714-456-2242

Simple Summary: The authors present a phase IIa clinical biomarker trial of colorectal cancer (CRC) survivors treated with a polyamine-inhibitory regimen (dietary arginine restriction plus daily aspirin 325 mg) as a potential future strategy for tertiary prevention of CRC (i.e., preventing colorectal adenomas and/or CRC among CRC survivors). In this study, 20 stage I-III CRC patients were treated for 12 weeks with the above-mentioned intervention, with assessments of calculated dietary arginine intake, plasma arginine levels, and rectal tissue mucosa samples assessed for polyamine levels at baseline and end-of-study. While dietary arginine intake and plasma arginine levels were significantly decreased by this intervention, rectal tissue polyamines (including tissue putrescine levels, which served as the primary study endpoint) were unaffected.



Citation: Zell, J.A.; Taylor, T.H.; Albers, C.G.; Carmichael, J.C.; McLaren, C.E.; Wenzel, L.; Stamos, M.J. Phase IIa Clinical Biomarker Trial of Dietary Arginine Restriction and Aspirin in Colorectal Cancer Patients. *Cancers* **2023**, *15*, 2103. <https://doi.org/10.3390/cancers15072103>

Academic Editors: Edoardo G. Giannini, Elisa Marabotto and Stefano Kayali

Received: 3 March 2023

Revised: 29 March 2023

Accepted: 29 March 2023

Published: 31 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: After potentially curative treatment, colorectal cancer (CRC) patients remain at high risk for recurrence, second primary CRC, and high-risk adenomas. In combination with existing data, our previous findings provide a rationale for reducing tissue polyamines as tertiary prevention in non-metastatic CRC patients. The goal of this study was to demonstrate rectal tissue polyamine reduction in optimally treated stage I-III CRC patients after intervention with daily oral aspirin + dietary arginine restriction. A single-institution phase IIa clinical trial was conducted. Patients were treated with aspirin 325 mg/day and an individualized dietary regimen designed to reduce arginine intake by $\geq 30\%$ over a 12-week study period. Dietary intake, endoscopy with rectal biopsies, and phlebotomy were performed pre- and post-intervention. The primary endpoint was to demonstrate $\geq 50\%$ decrease in rectal tissue putrescine levels from baseline as a measure of polyamine reduction in the target tissue. Twenty eligible patients completed the study. After study intervention, mean dietary arginine intake decreased from 3.7 g/day \pm 1.3 SD to 2.6 g/day \pm 1.2 SD (29.7% decrease, $p < 0.02$ by Sign test). Mean plasma arginine levels decreased from 46.0 ng/mL \pm 31.5 SD at baseline to 35 ng/mL \pm 21.7 SD ($p < 0.001$). Rectal tissue putrescine levels were 0.90 nMol/mg-protein pre-intervention and 0.99 nMol/mg-protein post-intervention ($p < 0.64$, NS). No significant differences were observed for the other tissue polyamines investigated: spermidine ($p < 0.13$), spermine ($p < 0.21$), spermidine:spermine ratio ($p < 0.71$). Among CRC survivors, treatment with daily oral aspirin and an individualized dietary arginine restriction intervention resulted in lower calculated dietary arginine intake and plasma arginine levels but did not affect rectal tissue polyamine levels.

Keywords: arginine restriction; aspirin; clinical trial; colorectal cancer; cancer prevention; dietary intervention; polyamines

1. Introduction

An estimated 106,970 new cases of colon cancer and 46,050 cases of rectal cancer will occur in the U.S. in 2023, making colorectal cancer (CRC) the third most common cancer among U.S. males and females [1]. Worldwide, CRC ranks as the 3rd most common form of cancer, with 1.9 million cases in 2020 (the most recent year for which we have global data); it ranks as the 2nd leading global cancer cause of death (916,000 deaths) [2]. Despite cure rates exceeding 90% for localized (stage I) disease, adherence to CRC screening is low (50–60%) and therefore 2/3rds of patients are diagnosed with regional (stage II, III) or advanced (stage IV) disease. As such, relative survival is estimated at just 65% [1]. Aside from risk of recurrence and CRC-specific mortality, CRC patients remain at high risk for multiple competing events. Such events include a 50% risk of recurrent adenoma and 27% risk of either a second primary colorectal cancer or high-risk adenoma (i.e., high-grade dysplasia, villous adenomas, large adenomas >1 cm, or multiple adenomas: three or more) [3–5].

The polyamine pathway has been implicated in carcinogenesis, including colorectal tumorigenesis. Germline mutations in the adenomatous polyposis coli (APC) tumor suppressor gene are found in Familial Adenomatous Polyposis (FAP); somatic APC mutations are found in the vast majority of sporadic colorectal cancers. APC regulates ornithine decarboxylase (ODC), which catalyzes polyamine synthesis from ornithine (an arginine-derived product) [6]. Without APC regulation of ODC, polyamine content increases in colorectal tissues, which promotes tumorigenesis [7]. Treatment with the chemopreventive agent L- α -difluoromethylornithine (DFMO, eflornithine), a suicide ODC inhibitor, decreases polyamine levels and suppresses tumorigenesis in the small intestine and colon of FAP-mouse models (*Apc*^{Min/+} mice) [7]. In a phase III clinical trial of colorectal adenoma patients, polyamine inhibition with combination of low-dose eflornithine (500 mg daily) and sulindac (150 mg daily) resulted in marked inhibition of colonic polyps, regardless of *Odc* genotype [8].

Recent studies have implicated polyamines in the regulation of immune response to immune checkpoint blockade in cancer treatment and demonstrated that polyamine levels are altered by interactions of the microbiome and the tumor microenvironment [9]. Intestinal bacteria can convert arginine directly to polyamines [10]. This route is not essential, however, as the hepatic and extra-hepatic arginases convert arginine to ornithine. Ornithine is itself converted by ODC to form polyamines. Thus, arginine represents an important substrate in the polyamine synthetic pathway. The major source of dietary arginine in humans is meat consumption—particularly beef consumption—but high quantities are found in pork, fish, and chicken [11]. Meat and meat products accounted for 38% of total daily arginine in a study of Finnish men who were estimated to have similar daily arginine intake to U.S. men (i.e., ~5000 mg/day) [10]. Red meat consumption has been associated with an increased risk of colon cancer in humans [12]; however, arginine's specific contribution to colorectal cancer risk has not been determined. Importantly, arginine is the key substrate for two competing metabolic pathways believed to be involved in carcinogenesis: polyamine synthesis and the nitric oxide (NO) synthase pathway (Figure 1). Nitric Oxide synthase 2 (NOS2) and other NO synthases catabolize arginine to form nitric oxide. Inducible NOS2 isoforms (in addition to cyclooxygenase, COX-2) are abundant in human colorectal adenomas [13]. NOS2 is also overexpressed in rat colon carcinoma tissues after azoxymethane treatment [14,15], an effect that aspirin inhibits [14]. *Apc*^{Min/+} *Nos2* knockout mice provided with dietary arginine showed fewer intestinal adenomas when compared to *Apc*^{Min/+} *Nos2*^{+/+} mice [16,17]. Prior experimental and epidemiologic analyses suggest that arginine-restriction [18] and NSAID use [19] confer benefits for colorectal carcinogenesis and survival, particularly among familial CRC patients.

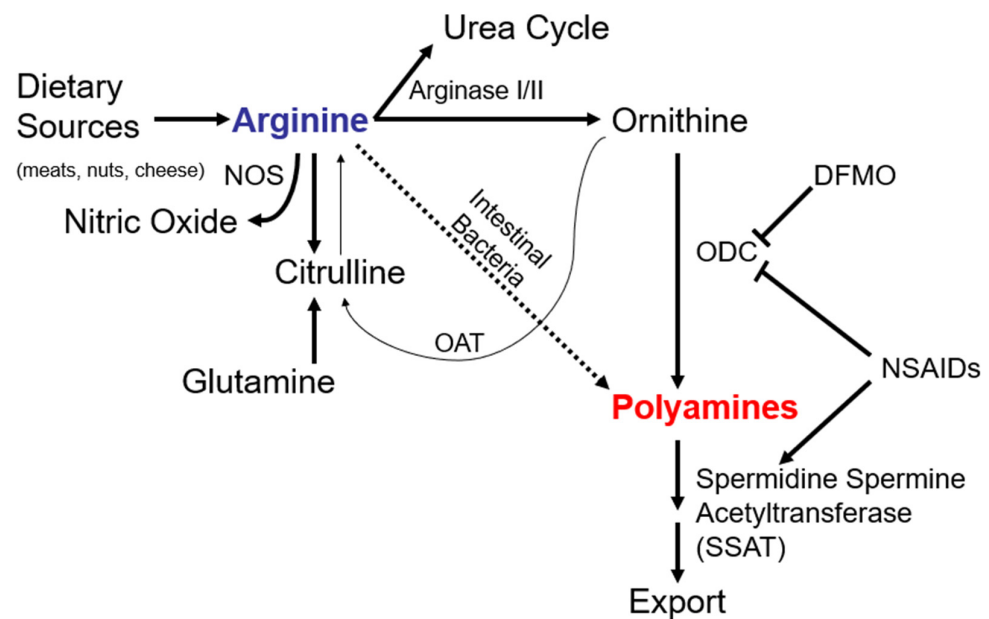


Figure 1. Arginine as the central substrate for polyamine and nitric oxide metabolism. Alternate pathways and inhibitors are indicated. NOS, nitric oxide synthase; DFMO, difluoromethylornithine (eflornithine); ODC, ornithine decarboxylase; NSAIDs, non-steroidal anti-inflammatory drugs; SSAT, spermidine/spermine N1-acetyltransferase; OAT, ornithine aminotransferase.

Despite potentially curative treatment, locally advanced colorectal cancer patients remain at considerable risk for multiple CRC-related events, and thus are the target for “tertiary cancer prevention” efforts (i.e., cancer prevention strategies aimed at reducing CRC-related events among patients with prior CRC history). Using our prior results in combination with the available literature, we designed a phase IIa clinical trial among CRC patients in order to detect favorable biomarker alterations in rectal tissue after a 12 week polyamine-inhibitory intervention comprised of dietary arginine restriction and daily aspirin use.

2. Methods

2.1. Study Design

This was a phase IIa clinical trial with pre- and post-intervention assessments of secondary endpoint biomarkers in colorectal cancer patients (ClinicalTrials.gov Identifier: NCT00578721). As such, all subjects received the intervention. The primary endpoint was to determine if a 12-week intervention consisting of daily enteric coated aspirin 325 mg and adherence to an arginine-restricted diet results in a significant $\geq 50\%$ decrease in rectal tissue putrescine levels in optimally treated stage I–III colorectal cancer patients. Here, rectal tissue putrescine represents a clinical biomarker. A total of 24 patients were enrolled to ensure that at least 20 completed the study. On-study procedures are shown in Figure 2.

Primary Telephone Counseling (PTC) Randomization Scheme: A subset of eligible participants were chosen at random to receive psychosocial telephone counseling (PTC). PTC was included as an exploratory aim to determine if it could positively reinforce behavioral changes and provide support during the trial. A randomization scheme was constructed for PTC assignment with block size of two (i.e., among 2 consecutive patients, one received PTC and the other did not). A set of consecutively numbered, sealed envelopes each contained a sheet giving the random assignment and spaces to record patient identity. Upon consent, each subsequent envelope was opened, revealing the assignment, and the form completed.

	Week						
	-2	0	4	6	8	12	13
Participant Identification	X						
Treatment							
Clinical Exam	X			X		X	
Dietician interview with 24-hr food recall assessments	XXX			XXX		XXX	
Compliance, and safety assessments				X		X	
Tests							
a. Routine CBC, CMP ¹ & UA	X			X		X	
b. 10ml Blood for SEB, <i>Odc</i> genotyping, plasma arginine and ornithine levels	X					X	
c. 10ml Blood for future testing.	X					X	
d. Urine Collection for UEB (*urinary polyamines, TBX2, PGEM)	X					X	
e. Rectal Biopsy (rectal mucosal tissue polyamines, * <i>Odc</i> and <i>Ssat</i> expression)	X ²						X ³

¹CMP includes sodium, potassium, chloride, CO₂, glucose, electrolyte balance, BUN, creatinine, total protein, albumin, calcium, alkaline phosphatase, AST, ALT, total bilirubin

²Endoscopy with biopsy

³Endoscopy with biopsy, within 2 weeks of planned end of study

"X" indicates timepoint for each assessment listed in table

*Not included in this analysis

Figure 2. Clinical procedures involved in the Phase IIa Clinical Trial.

2.2. Eligibility Criteria

Eligible patients included those age 18 years or older with a history of completely-resected AJCC stage I, II, or III colon or rectal cancer having no anticipated further treatment with radiation therapy or chemotherapy, and an ECOG Performance Status of 0–1. Patients with FAP or HNPCC were excluded. Patients having already received adjuvant (i.e., post-operative) radiation therapy to the colorectum were excluded. However, patients receiving neoadjuvant radiation therapy with or without chemotherapy prior to surgery for rectal cancer were eligible. Patients were excluded if they had a history of another invasive cancer within 5 years, excepting those with non-melanomatous skin cancer, melanoma in situ, Stage I cervical cancer, or Stage 0 CLL. Patients were required to have normal organ function. Patients already taking aspirin 325 mg/day at baseline were excluded, but those taking 81 mg/day aspirin were allowed on-study, substituting their daily 81 mg aspirin with protocol-defined 325 mg/day aspirin. The protocol did not allow enrolling patients taking calcium supplements (>520 mg/day), corticosteroids, NSAIDs, or anticoagulants on a regular or predictable intermittent basis. Patients with a history of abnormal wound healing or repair were excluded. Patients were excluded if they had personal history of colon resection >40 cm or inflammatory bowel disease. Pregnant or lactating women were not eligible. Premenopausal and perimenopausal women were required to be using adequate birth control methods. No history of allergies or adverse reactions to aspirin was allowed. No documented history of gastric/duodenal ulcer (or active treatment for such) within the last 12 months was allowed.

2.3. Intervention and On-Study Assessments

Patients were identified from the outpatient clinics at the University of California Irvine Medical Center (Orange, CA, USA). Upon obtaining written consent, the following clinical procedures were conducted: medical history & physical exam, baseline labs, *Odc* genotyping, three baseline 24 h Food Recall interviews (over a 2 week period) with estimation of total daily meat and arginine intake, plasma biomarker levels, and polyamine contents from the flat rectal mucosa at time of baseline flexible sigmoidoscopy. Participants were asked to complete a 24 h dietary recall interview by telephone with a dietitian approximately 3 separate times over a two week period at baseline, at mid-study (about 6 weeks), and again prior to the end of the study (12 weeks). Dietary intake data were collected and analyzed using Nutrition Data System for Research software version 6.0, developed

by the Nutrition Coordinating Center (NCC), University of Minnesota, Minneapolis, MN. The NDS-R software uses the multi-pass system of the interview methodology. Interview prompts guide interviewers and ensure standardized data entry, and dietary interviewers were certified by NCC for use of the NDS-R software. Upon review of dietary intake data, an individualized dietary plan was produced with the research dietitian to create a dietary prescription that would reduce estimated arginine intake by 30%. This was performed primarily through lowering meat intake by 50%, and lowering the intake of nuts, certain types of cheese, or other foods high in arginine content for the study duration. The dietitian discussed the diet with each subject to ensure that these dietary restrictions do not result in inadequate amounts of the recommended protein intake for the study period (i.e., 0.8 g protein per kg body weight per day). If the subject was not the person who regularly prepares food at home, the subject was asked to have the household food-preparer present when the dietary prescription was provided. In this manner all patients received an individualized $\geq 30\%$ arginine-restricted dietary prescription plus aspirin 325 mg PO daily x 12 weeks with nutritional education and compliance assessments. After 12 weeks on-study, patients underwent endoscopy with rectal biopsies for tissue polyamine content, and plasma biomarker levels, end of study H & P, labs, and 3 final 24 h food recall interviews for assessment of total arginine intake at end of study.

Dietary Assessment: Food consumption was assessed via three separate 24 h dietary recall interviews with a study dietitian during a 2 week period. These occurred at baseline, at mid-study (about 6 weeks) and again at end-of-study period (12 weeks), for a minimum of 9 dietary assessments over the course of study. Dietary factors that can influence the amount of intestinal polyamines and microbial activity, such as the amount and type of fiber and dietary protein, were also examined for a possible modifying effect on biomarker responsiveness [20].

Plasma Amino acid analysis: Arginine and ornithine concentrations were analyzed as described previously [10], as a measure of biochemical plasma responses to dietary arginine restriction. Plasma arginine correlates directly with arginine intake [10], and plasma ornithine levels can be monitored as a measure of adherence to an arginine-restricted diet [21,22]. Plasma samples were deproteinized with 5% (by volume) sulfosalicylic acid. One part 5% sulfosalicylic acid plus norleucine as an internal standard was added to 2 parts plasma, which was mixed thoroughly and kept at 4 °C for one hour. The samples were centrifuged at $12,000\times g$ for 15 min at 4 °C, and the supernatant was used for analysis. Arginine concentrations were analyzed via a Biochrom 20 amino acid analyzer (Pharmacia LKB Biochrom Ltd., Cambridge, UK), which is based on continuous flow ion exchange chromatography with ninhydrin detection of separate amino acids and lithium exchange ion.

Endoscopy with Rectal Mucosal Biopsy Procedure: At study entry (week 2) and at the end of study (week 13), endoscopy was performed and rectal mucosal biopsies were obtained for study evaluation. Eight biopsy specimens were obtained 8–12 cm above the anus and processed for tissue biomarker measurement. Biopsies were snap-frozen in liquid nitrogen and shipped to the University of Arizona for analysis.

Rectal Mucosa Tissue Polyamines: Polyamine content was determined as described previously [23], as this represents an important endpoint of treatment effect on the polyamine pathway. Eight small 4 mm² biopsy specimens were obtained; 3 were randomly selected for polyamine content. These methods measure putrescine, cadaverine, histamine, spermidine, spermine, and monoacetyl derivatives of putrescine, spermidine, and spermine [23]. Quality assurance procedures included regular measurements of standard polyamine preparations and use of internal standards in assessing polyamine amounts. Tumor-free portions of the rectum were collected, flushed with ice-cold saline, and stored frozen at −80 °C. Samples were processed and assayed for polyamine (putrescine, spermidine, spermine, and *N*¹-acetylspermine) content by reverse-phase high performance liquid chromatography with 1,7-diaminoheptane as an internal standard [24]. Protein content in each

sample was determined by the BCA protein assay kit (Pierce, Rockford, IL, USA). Data were expressed as nmol polyamine per mg protein.

2.4. Clinical Evaluation

At entry into the study and at 6 and 12 weeks or upon study discontinuation, patients underwent history and physical examination, and blood was drawn for CBC, with a metabolic panel to include liver function tests, BUN, creatinine, and urinalysis. All patients were evaluated for toxicity assessment from the time of first dose of aspirin and dietary arginine restriction (day 1), using Common Terminology for Adverse Events v4.0 (CTCAE, https://evs.nci.nih.gov/ftp1/CTCAE/CTCAE_4.03/Archive/CTCAE_4.02_2009-09-15_QuickReference_8.5x11.pdf) (accessed on 27 February 2023).

2.5. Statistical Analyses

The primary endpoint was percent reduction in rectal mucosa tissue putrescine levels from baseline to end of study. In prior research involving either colorectal adenoma patients [23] or experimental murine models [18], intestinal tissue putrescine reductions after polyamine-inhibitory treatments (DFMO, or celecoxib) ranged from 67–74%. Sample size calculations posited that percent reduction in putrescine would be 30% under the null and 60% under the alternative hypotheses, yielding a requirement for 20 subjects, using a 2-sided test ($\alpha = 0.05$, power = 80%); 24 patients were enrolled to allow for at least 15% attrition. The Sign Rank test was used to compare baseline to end-of-study rectal tissue putrescine levels, as well as levels of spermidine and spermine. All statistical analyses were conducted using SAS 9.1 statistical software (SAS Inc., Cary, NC, USA).

2.6. Ethical Considerations & Data Monitoring

This trial was approved by the Institutional Review Board of the University of California, Irvine. After discussing risks and benefits, all willing participants signed a written IRB-approved consent form. Participant safety was monitored by the UC Irvine Chao Family Comprehensive Cancer Center's Data Safety Monitoring Board (DSMB).

3. Results

3.1. Demographics

Among 24 eligible patients, 20 patients completed the study, including 10 rectal cancer patients and 10 colon cancer patients. By race/ethnicity, 60% of the patients were White, 40% were Asian, and 5% were Hispanic. 35% of participants were females. Mean age of all participants was 58 years. Patient clinical and pathological demographics are displayed in Table 1.

Table 1. Clinical and Demographic Characteristics. PTC, Psychosocial Telephone Counseling.

		Total (n = 20)
Gender	Male	13 (65%)
	Female	7 (35%)
Mean Age	Male	62.1 y (12.3)
	Female	51.7 y (6.9)
	Total	58.1 y (11.6)
Race	Asian	8 (40%)
	White	12 (60%)
Ethnicity	Hispanic	1 (5%)
	White	19 (95%)
	Cecum	1 (4.8%)
	Transverse	1 (4.8%)
Tumor Subsite Location	Sigmoid	5 (23.8%)
	Rectosigmoid	4 (19%)
	Rectum	6 (28.6%)
	Colorectum-NOS	3 (15%)

Table 1. *Cont.*

		Total (n = 20)
AJCC Stage	I	4 (19%)
	IIA	4 (19%)
	IIIA	3 (14.3%)
	IIIB	8 (38.1%)
	IIIC	1 (4.8%)
Tumor Grade	Well-Differentiated (Gr 1)	3 (15%)
	Moderately Differentiated (Gr 2)	10 (50%)
	Poorly Differentiated (Gr 3)	1 (5%)
	Missing	6 (30%)
Odc Genotype	AA	4 (20%)
	AG	7 (35%)
	GG	9 (45%)
Randomized to PTC Intervention?	Yes	8 (40%)
	No	7 (35%)
	N/A	5 (25%)

3.2. Dietary Analyses

Macronutrient intake from dietary intakes over the course of the study are presented in Table 2. Total reported kcal intake over the course of the study did not significantly differ pre- (median 1611 kcal/d) or post-intervention (1371 kcal/day, $p > 0.32$). When comparing relevant micronutrient data, the mean reported dietary arginine intake decreased after study intervention from 3.7 g/day \pm 1.3 SD to 2.6 g/day \pm 1.2 SD (29.7% decrease, $p < 0.02$ by Sign test (Figure 3). There was a significant correlation between the ranks of change in plasma arginine as a function of relative percent change in kCal ($Sr = 0.68$), with value of relative percent change in Kcal accounting for about 46 percent of the uncertainty in relative percent change of plasma arginine.

Table 2. Macronutrient data calculated from patient reported 24-h food recall surveys: minimum of 3 surveys done per patient at each timepoint: Baseline, Midpoint (6 weeks on-study), and End of study (12 weeks).

Variable	N	Mean	Median	Std Dev	Minimum	Maximum
Carbohydrate (g)						
Baseline	20	216.5	200.4	101.4	97.2	518.9
Midpoint	17	210.1	193.9	105.9	74.9	563.9
End of Study	17	189.7	182.5	58.0	75.0	298.1
Fiber (g)						
Baseline	20	18.0	18.4	7.3	6.6	31.8
Midpoint	17	18.5	16.5	10.5	5.6	48.2
End of Study	17	17.4	16.1	6.4	7.0	33.1
Fat (g)						
Baseline	20	56.5	49.8	29.6	16.7	160.4
Midpoint	17	51.4	52.4	32.9	17.6	154.6
End of Study	17	49.2	48.2	19.0	14.8	79.8
Protein (g)						
Baseline	20	65.4	62.9	20.9	27.8	118.7
Midpoint	17	55.9	54.3	20.5	27.8	112.4
End of Study	17	53.5	49.4	20.3	16.7	92.7
Energy (kcal)						
Baseline	20	1625.5	1595.1	682.2	661.4	3925.5
Midpoint	17	1515.7	1388.4	766.7	630.0	4090.4
End of Study	17	1409.8	1445.1	401.9	508.4	2255.3

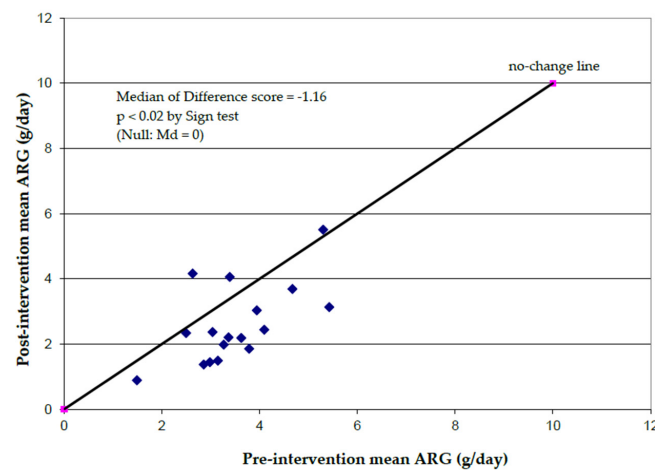


Figure 3. Dietary Arginine Intake, post- vs. pre-intervention (dietary arginine restriction and oral aspirin 325 mg daily). Measurements missing for $n = 3$ patients.

3.3. Plasma Arginine, Plasma Ornithine Levels

Mean plasma arginine levels decreased significantly from $46.0 \text{ ng/mL} \pm 31.53 \text{ SD}$ at baseline to $35 \text{ ng/mL} \pm 21.67 \text{ SD}$ at end of study (Mean difference -19.2 ng/mL , Sign Rank $p < 0.001$). Mean ornithine levels, however, did not significantly differ pre- (144.8 ng/mL) vs. post-intervention (133.5 ng/mL , mean difference -6.8 ng/mL , Sign Rank $p < 0.53$).

3.4. Rectal Tissue Polyamine Analysis

Rectal tissue polyamine levels were collected and analyzed for all patients; however, paired data were only available for 14 patients due to tissue processing issues affecting 6 patients (polyamine levels could not be calculated from submitted tissues). Rectal tissue putrescine levels (nMol/mg protein) did not differ statistically when comparing pre- ($0.90 \text{ nMol/mg protein}$) vs. post- ($0.99 \text{ nMol/mg protein}$) intervention biopsies (Sign-rank $p < 0.64$, $n = 14$, Figure 4). Similarly, no statistically significant differences were observed for rectal tissue changes at end of study vs. baseline for spermidine (Sign rank $p < 0.13$), spermine (Sign rank $p < 0.21$), or the spermidine to spermine ratio, SSR (Sign rank $p < 0.71$).

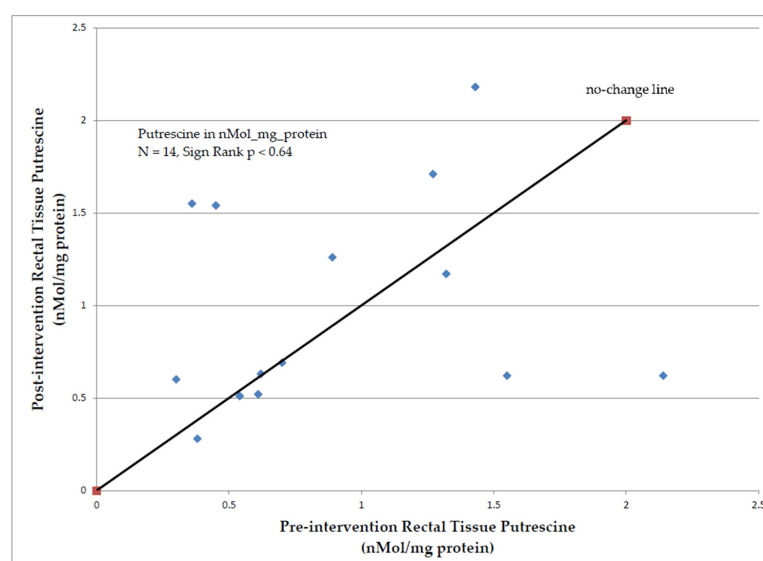


Figure 4. Rectal Tissue Putrescine Levels, post- vs. pre-intervention (dietary arginine restriction and oral aspirin 325 mg daily).

3.5. Adverse Events

Among the 20 patients completing the study, there were 7 therapy-related adverse events, all of which were grade 1 (diarrhea $n = 2$, fatigue $n = 3$, neuropathy $n = 1$, and myalgia $n = 1$). Of note, no clinically significant rectal bleeding occurred in any patient at baseline or end-of-study biopsies.

3.6. Quality of Life/Psychosocial Telephone Counseling (PTC) Intervention

Eight (8) patients were randomized to receive PTC. Of patients randomized to PTC, 71.4% achieved a lower dietary arginine consumption than the median decline, compared to 40% of patients randomized not to have PTC ($p < 0.34$, NS). Seven of eight rated each of the counseling sessions as “quite a bit” to “very” useful (highest score possible). In addition, seven of eight patients rated the counseling programs as “very helpful” (highest score possible). In response to the question, “Please indicate if any of the following changes occurred as a result of participating in the counseling sessions”, the majority checked a mean of five different changes (Range: 2–9 changes). Prominent changes included: improved diet, increased frequency of exercise, increased motivation for making healthy lifestyle improvements, and improved stress management.

4. Discussion

In this phase IIa tertiary prevention clinical trial of CRC survivors, an individualized dietary prescription to reduce dietary arginine intake by $\geq 30\%$ along with daily intake of 325 mg oral aspirin did not significantly reduce rectal tissue polyamine levels. As such, the polyamine-inhibitory intervention failed to demonstrate a biomarker effect in the target tissue of origin (i.e., rectal mucosa); this is one of the key criteria for further cancer prevention clinical trial development established in the landmark AACR consensus statement [25]. Of note, our trial demonstrated that this novel approach was safe and feasible, as we successfully achieved the 30% dietary arginine restriction goal among trial participants without substantial adverse effects, resulting in statistically significant decreased plasma arginine levels. As such, the lack of tissue biomarker response indicates that potential rectal tissue biomarker effects of 12 weeks of combination dietary arginine restriction and aspirin do not recapitulate the pharmacologic effects of polyamine inhibition through eflornithine and/or NSAIDs, as demonstrated in other studies.

Lowering rectal mucosal polyamines predicts pharmacologic treatment’s ability to prevent colorectal adenomas [26]. However, as mentioned, we did not observe a tissue polyamine-lowering effect of our dietary arginine restriction and aspirin intervention. It is possible that the 12 week intervention duration was too short for our intervention to result in 50% tissue putrescine reduction. This treatment duration was selected based on results from two separate trials involving tissue polyamine reductions after polyamine-inhibitory treatment with DFMO. In a prior colon cancer chemoprevention trial, treatment with oral DFMO at 0.4 g/m² resulted in a reduction of the rectal mucosa tissue spermidine:spermine ratio at 6 months that was similar to the reduction noted at 12 months [23]. In a small, phase IIa trial of nine prostate cancer patients, a short 4 week course of oral DFMO at 0.5 g/m² resulted in significant reductions of prostatic tissue polyamine content [27]. Thus, the selected duration here was intermediate between 1 and 6 months as noted in the above polyamine-inhibition trials using DFMO, and theoretically sufficient to observe the proposed changes in tissue polyamines pre- vs. post-intervention.

Aspirin and non-steroidal anti-inflammatory drugs (NSAIDs) inhibit intestinal cancer development in mouse models [18,28]. Clinical trials have implicated aspirin in premalignant colorectal adenoma regression and prevention [29–31], and cohort and case-control studies have shown that along with other NSAIDs, aspirin reduces CRC risk [32–35], particularly with prolonged use [34]. Lack of consensus, and unexpected cardiovascular toxicities associated with celecoxib use preclude recommending NSAIDs for primary prevention of CRC [36,37]. In a change from the 2016 US Preventive Services Task Force (USPSTF)

guidelines, new 2022 guidelines state there is currently insufficient evidence to support aspirin to reduce incidence or mortality of CRC [38].

A novel aspect of this phase IIa clinical trial is the Psychosocial Telephone Counseling (PTC) intervention. Psychosocial Telephone Counseling has shown benefit among cancer survivor cohorts in other studies, particularly in effects on overall quality of life, social support, and survivor-specific issues [39,40]. Interestingly, such PTC-based changes are marked by beneficial changes in patients' blood-based immune profile [39,40]. To our knowledge, these potentially efficacious PTC interventions have not been previously conducted among CRC survivors. While our numbers are small, seven of eight patients randomized to the PTC intervention reported that it was "very helpful" (highest score), and the majority indicated that PTC helped to improve diet, increase frequency of exercise, increase motivation for making healthy lifestyle improvements, and improve stress management. In addition, while not statistically significant, it is interesting to observe that patients randomized to PTC had numerically greater reductions in reported dietary arginine intake over the 12 week intervention period. Dietary interventions present unique challenges in the clinical trial setting. Given our encouraging but preliminary results, PTC may have a role in facilitating adherence to dietary treatments or other biobehavioral interventions in the clinical trial setting.

While adequately powered to address the study question, the study is limited by small sample size and was not powered to identify subtle polyamine tissue biomarker effects. In addition, the small sample size precludes one from making meaningful assessments of any potential differential effects by *Odc* genotype, which have been implicated as relevant in a larger clinical trial [41] and in two epidemiologic cohort studies—one of which demonstrated differential effects of meat consumption on outcomes based on *Odc* genotype [42,43]. By reducing dietary arginine, our study focused on reducing endogenous polyamine synthesis. Others have investigated effects of exogenous polyamine intake on CRC risk. In a study of individuals in China [44], differential effects based on specific dietary polyamine intake were reported (beneficial for putrescine and spermidine, adverse risk for spermine). In a separate analysis of data involving U.S. women enrolled in the Women's Health Initiative cohort study, no positive associations were observed for dietary polyamine intake and CRC risk, or CRC-specific mortality [45].

In our phase IIa clinical biomarker trial, patients were heterogeneous with respect to primary tumor site (colon, rectum), stage, and duration of remission since time of initial operation, which may have confounded the analysis. While the study (and intervention itself) relied on numerous dietary food recalls under the direction of a research dietitian, the nature of this methodology is dependent on patient recall and thus may be inaccurate.

Despite the negative rectal tissue biomarker results of this dietary and aspirin-based polyamine-inhibitory intervention, secondary and tertiary prevention of CRC through pharmacologic polyamine inhibition remain active areas of investigation. To this end, the randomized clinical trial of eflornithine, sulindac, and placebos in FAP patients (secondary prevention trial) provides some insights [46]. In the study, FAP patients randomized to one of three study arms (eflornithine + sulindac vs. eflornithine + sulindac-placebo vs. eflornithine-placebo + sulindac) revealed no significant differences on disease progression. However, among patients with an intact colorectum ("pre-colectomy group"), interesting yet non-significant differences were reported: time to disease progression was 39 months in the eflornithine-sulindac group, 25 months in the sulindac group, and 20 months in the eflornithine group. In a post-hoc analysis of this FAP trial, combination eflornithine-sulindac was shown to delay or prevent the need for lower gastrointestinal surgery in FAP patients [47]. Two additional U.S. National Cancer Institute (NCI)-supported phase III clinical trials are relevant to understanding pharmacologic polyamine-inhibitory agents as tertiary CRC prevention. Results of the large phase III clinical trial results of C80702 (ClinicalTrials.gov Identifier: NCT01150045), which randomized stage III colon cancer patients to a 3 year intervention of celecoxib or placebo after 1st randomization to 3 vs. 6 mo adjuvant FOLFOX chemotherapy, revealed no difference in disease-free survival

ascribed to celecoxib [48]. In a follow-up analysis of patients in this trial based on plasma inflammatory markers taken 3–8 weeks post-operatively (and prior to chemotherapy), higher inflammation postoperatively was associated with increased risk of recurrence or death [49]. Of note, analyses of the secondary clinical endpoints for C80702 (e.g., total adenoma recurrence, high-risk adenoma recurrence) have not yet been reported. Currently, accrual is nearing completion for the NCI-National Community Oncology Research Program (NCORP)-sponsored trial SWOG 0820: a double-blind randomized controlled clinical trial of eflornithine and sulindac vs. placebos in stage 0–III CRC patients (ClinicalTrials.gov Identifier: NCT01349881) [50]. The primary endpoint of S0820 is to determine if there is a $\geq 60\%$ reduction in high-risk adenomas and 2nd primary CRCs after 3 year intervention of oral eflornithine + sulindac vs. matching placebos in resected CRC patients.

In conclusion, our phase IIa clinical trial of a 12 week intervention of daily oral aspirin 325 mg plus an individualized dietary arginine restriction regimen did not result in rectal tissue polyamine changes in CRC survivors. Whether a more stringent dietary arginine restriction prescription, or a prolonged duration of intervention would have yielded different results is unknown and cannot be imputed from this study. Nevertheless, there is a disconnect between our results here and the tissue polyamine-lowering effects of various therapeutic prevention agents such as eflornithine, sulindac, aspirin, and various NSAIDs. As such, we await the results of the primary and secondary analyses of key NCI-sponsored CRC therapeutic prevention clinical trials, as well as ongoing basic and translational research that will shed light on potential future roles for polyamine inhibition as CRC prevention.

Author Contributions: Conceptualization, J.A.Z.; methodology, J.A.Z., T.H.T., C.G.A., J.C.C., C.E.M., L.W., M.J.S.; formal analysis, T.H.T.; investigation, J.A.Z., T.H.T., C.G.A., J.C.C., L.W., M.J.S.; data curation, J.A.Z., C.G.A., J.C.C., L.W.; writing—original draft preparation, J.A.Z.; writing—review and editing, J.A.Z., T.H.T., C.G.A., J.C.C., C.E.M., L.W., M.J.S.; project administration, J.A.Z.; funding acquisition, J.A.Z. All authors have read and agreed to the published version of the manuscript.

Funding: Research reported in this publication was supported by the University of California Irvine Department of Medicine, the National Cancer Institute of the National Institutes of Health under award numbers K23 CA133142, P30 CA062203 (support from the Chao Family Comprehensive Cancer Center’s Biobehavioral Shared Resource and Biostatistical Shared Resource), and the National Center for Research Resources Public Health Service grant M01 RR00827. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Institutional Review Board Statement: This trial was conducted in accordance with the Declaration of Helsinki, and approved by the Institutional Review Board of the University of California, Irvine. Protocol #20075892. Last approval date: 25 August 2022.

Informed Consent Statement: After discussing risks and benefits, all willing participants signed a written IRB-approved consent form.

Data Availability Statement: The data generated in this article will be shared upon reasonable request to the corresponding author.

Conflicts of Interest: The authors list the following disclosures: J.Z.: Tempus; C.A.: Nestle Health Sciences, AbbVie, Phathom Pharmaceuticals, DSI Pharmaceuticals, Salix Pharmaceuticals; J.C.: Johnson & Johnson; L.W.: Immunogen.

References

1. Siegel, R.L.; Miller, K.D.; Wagle, N.S.; Jemal, A. Cancer statistics, 2023. *CA Cancer J. Clin.* **2023**, *73*, 17–48. [[CrossRef](#)] [[PubMed](#)]
2. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* **2021**, *71*, 209–249. [[CrossRef](#)]
3. Chu, D.Z.; Chansky, K.; Alberts, D.S.; Meyskens, F.L., Jr.; Fenoglio-Preiser, C.M.; Rivkin, S.E.; Mills, G.M.; Giguere, J.K.; Goodman, G.E.; Abbruzzese, J.L.; et al. Adenoma recurrences after resection of colorectal carcinoma: Results from the Southwest Oncology Group 9041 calcium chemoprevention pilot study. *Ann. Surg. Oncol.* **2003**, *10*, 870–875. [[CrossRef](#)] [[PubMed](#)]

4. Martinez, M.E.; Baron, J.A.; Lieberman, D.A.; Schatzkin, A.; Lanza, E.; Winawer, S.J.; Zauber, A.G.; Jiang, R.; Ahnen, D.J.; Bond, J.H.; et al. A pooled analysis of advanced colorectal neoplasia diagnoses after colonoscopic polypectomy. *Gastroenterology* **2009**, *136*, 832–841. [\[CrossRef\]](#)
5. Sargent, D.J.; Patiyil, S.; Yothers, G.; Haller, D.G.; Gray, R.; Benedetti, J.; Buyse, M.; Labianca, R.; Seitz, J.F.; O’Callaghan, C.J.; et al. End points for colon cancer adjuvant trials: Observations and recommendations based on individual patient data from 20,898 patients enrolled onto 18 randomized trials from the ACCENT Group. *J. Clin. Oncol.* **2007**, *25*, 4569–4574. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Fultz, K.E.; Gerner, E.W. APC-dependent regulation of ornithine decarboxylase in human colon tumor cells. *Mol. Carcinog.* **2002**, *34*, 10–18. [\[CrossRef\]](#)
7. Erdman, S.H.; Ignatenko, N.A.; Powell, M.B.; Blohm-Mangone, K.A.; Holubec, H.; Guillen-Rodriguez, J.M.; Gerner, E.W. APC-dependent changes in expression of genes influencing polyamine metabolism, and consequences for gastrointestinal carcinogenesis, in the Min mouse. *Carcinogenesis* **1999**, *20*, 1709–1713. [\[CrossRef\]](#)
8. Meyskens, F.L., Jr.; McLaren, C.E.; Pelot, D.; Fujikawa-Brooks, S.; Carpenter, P.M.; Hawk, E.; Kelloff, G.; Lawson, M.J.; Kidao, J.; McCracken, J.; et al. Difluoromethylornithine plus sulindac for the prevention of sporadic colorectal adenomas: A randomized placebo-controlled, double-blind trial. *Cancer Prev. Res.* **2008**, *1*, 32–38. [\[CrossRef\]](#)
9. Holbert, C.E.; Cullen, M.T.; Casero, R.A., Jr.; Stewart, T.M. Polyamines in cancer: Integrating organismal metabolism and antitumour immunity. *Nat. Rev. Cancer* **2022**, *22*, 467–480. [\[CrossRef\]](#)
10. Venho, B.; Voutilainen, S.; Valkonen, V.P.; Virtanen, J.; Lakka, T.A.; Rissanen, T.H.; Ovaskainen, M.L.; Laitinen, M.; Salonen, J.T. Arginine intake, blood pressure, and the incidence of acute coronary events in men: The Kuopio Ischaemic Heart Disease Risk Factor Study. *Am. J. Clin. Nutr.* **2002**, *76*, 359–364. [\[CrossRef\]](#)
11. FoodData Central, U.S. Department of Agriculture, Agricultural Research Service. Available online: <https://fdc.nal.usda.gov/index.html> (accessed on 2 February 2023).
12. Giovannucci, E.; Willett, W.C. Dietary factors and risk of colon cancer. *Ann. Med.* **1994**, *26*, 443–452. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Ambs, S.; Merriam, W.G.; Bennett, W.P.; Felley-Bosco, E.; Ogunfusika, M.O.; Oser, S.M.; Klein, S.; Shields, P.G.; Billiar, T.R.; Harris, C.C. Frequent nitric oxide synthase-2 expression in human colon adenomas: Implication for tumor angiogenesis and colon cancer progression. *Cancer Res.* **1998**, *58*, 334–341. [\[PubMed\]](#)
14. Escribano, M.; Molero, L.; Lopez-Farre, A.; Abarrategui, C.; Carrasco, C.; Garcia-Mendez, A.; Manzarbeitia, F.; Martin, M.J.; Vazquez, M.; Sanchez-Fayos, P.; et al. Aspirin inhibits endothelial nitric oxide synthase (eNOS) and Flk-1 (vascular endothelial growth factor receptor-2) prior to rat colon tumour development. *Clin. Sci.* **2004**, *106*, 83–91. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Takahashi, M.; Fukuda, K.; Ohata, T.; Sugimura, T.; Wakabayashi, K. Increased expression of inducible and endothelial constitutive nitric oxide synthases in rat colon tumors induced by azoxymethane. *Cancer Res.* **1997**, *57*, 1233–1237.
16. Ahn, B.; Ohshima, H. Suppression of intestinal polyposis in Apc(Min/+) mice by inhibiting nitric oxide production. *Cancer Res.* **2001**, *61*, 8357–8360.
17. Yerushalmi, H.F.; Besselsen, D.G.; Ignatenko, N.A.; Blohm-Mangone, K.A.; Padilla-Torres, J.L.; Stringer, D.E.; Cui, H.; Holubec, H.; Payne, C.M.; Gerner, E.W. The role of NO synthases in arginine-dependent small intestinal and colonic carcinogenesis. *Mol. Carcinog.* **2006**, *45*, 93–105. [\[CrossRef\]](#)
18. Zell, J.A.; Ignatenko, N.A.; Yerushalmi, H.F.; Ziogas, A.; Besselsen, D.G.; Gerner, E.W.; Anton-Culver, H. Risk and risk reduction involving arginine intake and meat consumption in colorectal tumorigenesis and survival. *Int. J. Cancer* **2007**, *120*, 459–468. [\[CrossRef\]](#)
19. Zell, J.A.; Ziogas, A.; Bernstein, L.; Clarke, C.A.; Deapen, D.; Largent, J.A.; Neuhausen, S.L.; Stram, D.O.; Ursin, G.; Anton-Culver, H. Meat consumption, nonsteroidal anti-inflammatory drug use, and mortality among colorectal cancer patients in the California Teachers Study. *Cancer Prev. Res.* **2010**, *3*, 865–875. [\[CrossRef\]](#)
20. Bardocz, S.; Duguid, T.J.; Brown, D.S.; Grant, G.; Pusztai, A.; White, A.; Ralph, A. The importance of dietary polyamines in cell regeneration and growth. *Br. J. Nutr.* **1995**, *73*, 819–828. [\[CrossRef\]](#)
21. Kaiser-Kupfer, M.I.; Caruso, R.C.; Valle, D. Gyrate atrophy of the choroid and retina: Further experience with long-term reduction of ornithine levels in children. *Arch. Ophthalmol.* **2002**, *120*, 146–153. [\[CrossRef\]](#)
22. Kaiser-Kupfer, M.I.; Caruso, R.C.; Valle, D.; Reed, G.F. Use of an arginine-restricted diet to slow progression of visual loss in patients with gyrate atrophy. *Arch. Ophthalmol.* **2004**, *122*, 982–984. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Meyskens, F.L., Jr.; Gerner, E.W.; Emerson, S.; Pelot, D.; Durbin, T.; Doyle, K.; Lagerberg, W. Effect of alpha-difluoromethylornithine on rectal mucosal levels of polyamines in a randomized, double-blinded trial for colon cancer prevention. *J. Natl. Cancer Inst.* **1998**, *90*, 1212–1218. [\[CrossRef\]](#)
24. Seiler, N.; Knodgen, B. High-performance liquid chromatographic procedure for the simultaneous determination of the natural polyamines and their monoacetyl derivatives. *J. Chromatogr.* **1980**, *221*, 227–235. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Kelloff, G.J.; Lippman, S.M.; Dannenberg, A.J.; Sigman, C.C.; Pearce, H.L.; Reid, B.J.; Szabo, E.; Jordan, V.C.; Spitz, M.R.; Mills, G.B.; et al. Progress in chemoprevention drug development: The promise of molecular biomarkers for prevention of intraepithelial neoplasia and cancer—A plan to move forward. *Clin. Cancer Res.* **2006**, *12*, 3661–3697. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Thompson, P.A.; Wertheim, B.C.; Zell, J.A.; Chen, W.P.; McLaren, C.E.; LaFleur, B.J.; Meyskens, F.L.; Gerner, E.W. Levels of rectal mucosal polyamines and prostaglandin E2 predict ability of DFMO and sulindac to prevent colorectal adenoma. *Gastroenterology* **2010**, *139*, 797–805. [\[CrossRef\]](#)

27. Simoneau, A.R.; Gerner, E.W.; Phung, M.; McLaren, C.E.; Meyskens, F.L., Jr. Alpha-difluoromethylornithine and polyamine levels in the human prostate: Results of a phase IIa trial. *J. Natl. Cancer Inst.* **2001**, *93*, 57–59. [\[CrossRef\]](#)
28. Ignatenko, N.A.; Besselsen, D.G.; Stringer, D.E.; Blohm-Mangone, K.A.; Cui, H.; Gerner, E.W. Combination chemoprevention of intestinal carcinogenesis in a murine model of familial adenomatous polyposis. *Nutr. Cancer* **2008**, *60* (Suppl. S1), 30–35. [\[CrossRef\]](#)
29. Baron, J.A.; Cole, B.F.; Sandler, R.S.; Haile, R.W.; Ahnen, D.; Bresalier, R.; McKeown-Eyssen, G.; Summers, R.W.; Rothstein, R.; Burke, C.A.; et al. A randomized trial of aspirin to prevent colorectal adenomas. *N. Engl. J. Med.* **2003**, *348*, 891–899. [\[CrossRef\]](#)
30. Sandler, R.S.; Halabi, S.; Baron, J.A.; Budinger, S.; Paskett, E.; Keresztes, R.; Petrelli, N.; Pipas, J.M.; Karp, D.D.; Loprinzi, C.L.; et al. A randomized trial of aspirin to prevent colorectal adenomas in patients with previous colorectal cancer. *N. Engl. J. Med.* **2003**, *348*, 883–890. [\[CrossRef\]](#)
31. Steinbach, G.; Lynch, P.M.; Phillips, R.K.; Wallace, M.H.; Hawk, E.; Gordon, G.B.; Wakabayashi, N.; Saunders, B.; Shen, Y.; Fujimura, T.; et al. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N. Engl. J. Med.* **2000**, *342*, 1946–1952. [\[CrossRef\]](#)
32. Chan, A.T.; Giovannucci, E.L.; Schernhammer, E.S.; Colditz, G.A.; Hunter, D.J.; Willett, W.C.; Fuchs, C.S. A prospective study of aspirin use and the risk for colorectal adenoma. *Ann. Intern. Med.* **2004**, *140*, 157–166. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Asano, T.K.; McLeod, R.S. Nonsteroidal anti-inflammatory drugs and aspirin for the prevention of colorectal adenomas and cancer: A systematic review. *Dis. Colon Rectum* **2004**, *47*, 665–673. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Chan, A.T.; Giovannucci, E.L.; Meyerhardt, J.A.; Schernhammer, E.S.; Curhan, G.C.; Fuchs, C.S. Long-term use of aspirin and nonsteroidal anti-inflammatory drugs and risk of colorectal cancer. *JAMA* **2005**, *294*, 914–923. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Mahipal, A.; Anderson, K.E.; Limburg, P.J.; Folsom, A.R. Nonsteroidal anti-inflammatory drugs and subsite-specific colorectal cancer incidence in the Iowa women’s health study. *Cancer Epidemiol. Biomarkers Prev.* **2006**, *15*, 1785–1790. [\[CrossRef\]](#)
36. Arber, N.; Eagle, C.J.; Spicak, J.; Racz, I.; Dite, P.; Hajer, J.; Zavoral, M.; Lechuga, M.J.; Gerletti, P.; Tang, J.; et al. Celecoxib for the prevention of colorectal adenomatous polyps. *N. Engl. J. Med.* **2006**, *355*, 885–895. [\[CrossRef\]](#)
37. Bertagnolli, M.M.; Eagle, C.J.; Zauber, A.G.; Redston, M.; Solomon, S.D.; Kim, K.; Tang, J.; Rosenstein, R.B.; Wittes, J.; Corle, D.; et al. Celecoxib for the prevention of sporadic colorectal adenomas. *N. Engl. J. Med.* **2006**, *355*, 873–884. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Force, U.S.P.S.T.; Davidson, K.W.; Barry, M.J.; Mangione, C.M.; Cabana, M.; Chelmow, D.; Coker, T.R.; Davis, E.M.; Donahue, K.E.; Jaen, C.R.; et al. Aspirin Use to Prevent Cardiovascular Disease: US Preventive Services Task Force Recommendation Statement. *JAMA* **2022**, *327*, 1577–1584. [\[CrossRef\]](#)
39. Nelson, E.L.; Wenzel, L.B.; Osann, K.; Dogan-Ates, A.; Chantana, N.; Reina-Patton, A.; Laust, A.K.; Nishimoto, K.P.; Chicx-DeMet, A.; du Pont, N.; et al. Stress, immunity, and cervical cancer: Biobehavioral outcomes of a randomized clinical trial [corrected]. *Clin. Cancer Res.* **2008**, *14*, 2111–2118. [\[CrossRef\]](#)
40. Osann, K.; Wilford, J.; Wenzel, L.; Hsieh, S.; Tucker, J.A.; Wahi, A.; Monk, B.J.; Nelson, E.L. Relationship between social support, quality of life, and Th2 cytokines in a biobehavioral cancer survivorship trial. *Support. Care Cancer* **2019**, *27*, 3301–3310. [\[CrossRef\]](#)
41. Zell, J.A.; McLaren, C.E.; Chen, W.P.; Thompson, P.A.; Gerner, E.W.; Meyskens, F.L. Ornithine decarboxylase-1 polymorphism, chemoprevention with eflornithine and sulindac, and outcomes among colorectal adenoma patients. *J. Natl. Cancer Inst.* **2010**, *102*, 1513–1516. [\[CrossRef\]](#)
42. Zell, J.A.; Ziogas, A.; Ignatenko, N.; Honda, J.; Qu, N.; Bobbs, A.S.; Neuhausen, S.L.; Gerner, E.W.; Anton-Culver, H. Associations of a polymorphism in the ornithine decarboxylase gene with colorectal cancer survival. *Clin. Cancer Res.* **2009**, *15*, 6208–6216. [\[CrossRef\]](#)
43. Zell, J.A.; Lin, B.S.; Ziogas, A.; Anton-Culver, H. Meat consumption, ornithine decarboxylase gene polymorphism, and outcomes after colorectal cancer diagnosis. *J. Carcinog.* **2012**, *11*, 17. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Huang, C.Y.; Fang, Y.J.; Abulimiti, A.; Yang, X.; Li, L.; Liu, K.Y.; Zhang, X.; Feng, X.L.; Chen, Y.M.; Zhang, C.X. Dietary Polyamines Intake and Risk of Colorectal Cancer: A Case-Control Study. *Nutrients* **2020**, *12*, 3575. [\[CrossRef\]](#)
45. Vargas, A.J.; Ashbeck, E.L.; Wertheim, B.C.; Wallace, R.B.; Neuhaus, M.L.; Thomson, C.A.; Thompson, P.A. Dietary polyamine intake and colorectal cancer risk in postmenopausal women. *Am. J. Clin. Nutr.* **2015**, *102*, 411–419. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Burke, C.A.; Dekker, E.; Lynch, P.; Samadder, N.J.; Balaguer, F.; Huneburg, R.; Burn, J.; Castells, A.; Gallinger, S.; Lim, R.; et al. Eflornithine plus Sulindac for Prevention of Progression in Familial Adenomatous Polyposis. *N. Engl. J. Med.* **2020**, *383*, 1028–1039. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Balaguer, F.; Stoffel, E.M.; Burke, C.A.; Dekker, E.; Samadder, N.J.; Van Cutsem, E.; Lynch, P.M.; Wise, P.E.; Huneburg, R.; Lim, R.M.; et al. Combination of Sulindac and Eflornithine Delays the Need for Lower Gastrointestinal Surgery in Patients With Familial Adenomatous Polyposis: Post Hoc Analysis of a Randomized Clinical Trial. *Dis. Colon Rectum* **2022**, *65*, 536–545. [\[CrossRef\]](#)
48. Meyerhardt, J.A.; Shi, Q.; Fuchs, C.S.; Meyer, J.; Niedzwiecki, D.; Zemla, T.; Kumthekar, P.; Guthrie, K.A.; Couture, F.; Kuebler, P.; et al. Effect of Celecoxib vs Placebo Added to Standard Adjuvant Therapy on Disease-Free Survival Among Patients With Stage III Colon Cancer: The CALGB/SWOG 80702 (Alliance) Randomized Clinical Trial. *JAMA* **2021**, *325*, 1277–1286. [\[CrossRef\]](#)

49. Cheng, E.; Shi, Q.; Shields, A.F.; Nixon, A.B.; Shergill, A.P.; Ma, C.; Guthrie, K.A.; Couture, F.; Kuebler, P.; Kumar, P.; et al. Association of Inflammatory Biomarkers With Survival Among Patients With Stage III Colon Cancer. *JAMA Oncol.* **2023**, *9*, 404. [[CrossRef](#)]
50. Zell, J.; You, Y.N.; Boughey, J.C. PACES trial: Evaluating the effectiveness of eflornithine and sulindac in preventing colon adenomas. *Bull. Am. Coll. Surg.* **2015**, *100*, 70–71.

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.