



Review

Taxonomy and Translocations of African Mammals: A Plea for a Cautionary Approach

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Abstract: Ecotourism can fuel an important source of financial income for African countries and can therefore help biodiversity policies in the continent. Translocations can be a powerful tool to spread economic benefits among countries and communities; yet, to be positive for biodiversity conservation, they require a basic knowledge of conservation units through appropriate taxonomic research. This is not always the case, as taxonomy was considered an outdated discipline for almost a century, and some plurality in taxonomic approaches is incorrectly considered as a disadvantage for conservation work. As an example, diversity of the genus *Giraffa* and its recent taxonomic history illustrate the importance of such knowledge for a sound conservation policy that includes translocations. We argue that a fine-grained conservation perspective that prioritizes all remaining populations along the Nile Basin is needed. Translocations are important tools for giraffe diversity conservation, but more discussion is needed, especially for moving new giraffes to regions where the autochthonous taxa/populations are no longer existent. As the current discussion about the giraffe taxonomy is too focused on the number of giraffe species, we argue that the plurality of taxonomic and conservation approaches might be beneficial, i.e., for defining the number of units requiring separate management using a (majority) consensus across different concepts (e.g., MU—management unit, ESU—evolutionary significant unit, and ECU—elemental conservation unit). The taxonomically sensitive translocation policy/strategy would be important for the preservation of current diversity, while also supporting the ecological restoration of some regions within rewilding. A summary table of the main translocation operations of African mammals that have underlying problems is included. Therefore, we call for increased attention toward the taxonomy of African mammals not only as the basis for sound conservation but also as a further opportunity to enlarge the geographic scope of ecotourism in Africa.

Keywords: Africa; mammal subspecies; biotic homogenization; *Giraffa*; game tourism; taxonomy; *Panthera leo*; ECU



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1. Introduction

When dealing with Africa's biodiversity, we should not be tempted to fall into the belief of studying and conserving a piece of Eden that escaped the ruinous consequences of human impact. This is not certainly true, but see [1,2], yet wildlife diversity and abundance had—and still has—an obvious role in shaping Western perceptions of Africa as a unique place on Earth [3,4].

African biomes are also a unique laboratory for scientific research that explores the forces that governed evolutionary history. Dealing with such diverse issue as humans' evolutionary history or the relationship between tectonic activity and cichlids' speciation [5], Africa appears as a unique setting to reach a better knowledge of how biodiversity

developed. Moreover, African mammals have been a classic subject for scientific inquiries dealing with evolution for more than a century now, e.g., [6–17].

Mammal diversity, particularly the richness of relatively large-sized species, fueled at first a growing interest in Africa by hunters from all over the world. Later, a conservation movement developed (aside from a few pioneers such as Carl Akeley), mainly influenced by those researchers—such as George B. Schaller and Ian Douglas-Hamilton—studying charismatic species such as African elephants, lions, gorillas and chimpanzees. The same species and African wildlife biomass and diversity are now the target of a multi-millionaire tourism industry that makes wildlife a highly profitable business for private operators and African countries [18]. This has led to an increased role in active management of some mammal species, including a growing occurrence of translocations as a means to repopulate or restock protected areas that have lost their native stock of some of the most charismatic species [19–22], to realize the optimal management for threatened taxa (e.g., [23,24] in the case of the Cape mountain zebra) or for the ecological restoration of particular regions (e.g., [25,26]). Active management operations are often valuable from the conservation point of view, but some of them have been inappropriate or even damaging to the genetic integrity of autochthonous populations of particular species (cf. [27] for mitigation translocation cases), such as in the case of the wildebeest (see [28–30]), or whole communities (for evaluation of ungulate translocations, especially in Southern Africa, see [21,31,32]).

In the present contribution, we aimed to critically review translocations of some mammals in Africa as a conservation tool, partly using giraffes as a case-study because of the current progress in understanding their diversification across Africa and emphasizing some causes of concern relating the possible negative outcomes for the conservation of evolutionary history in a unique continent. Following an increasing emphasis on financial viability, many extralimital species—i.e., species that historically did not occur in an area—or stocks of atypical phenotypes (under “intentional genetic manipulation” [21]) were introduced into private and public reserves to increase public experiences with the intention of increasing ecotourism attractions [33]. Although translocations are not a totally new tool in African conservation, it seems that many current projects are being realized primarily for financial reasons rather than conservation considerations. Another factor that often determines and influences the translocations is the need to create private game reserves dedicated to trophy hunting. This phenomenon is especially widespread in South Africa [34], and, often, the purpose of increasing income is to the detriment of conservation because moving species and subspecies well outside their original ranges increases the risk of hybridization between closely related species or subspecies [35,36]. A similarly questionable type of operation is to create fenced private reserves with the aim of attracting tourists to observe animal species, especially large mammals, including those that had never been locally present there in historic times.

According to [37], there are at least seven types of translocation for which conservation is not the primary aim (note that species conservation may be an associated aim and protection of individual animals of threatened species may be a primary aim): non-lethal management of problem animals, commercial and recreational, biological control, aesthetic, religious, wildlife rehabilitation, and animal rights activism.

2. What Do We Know about Large Mammal Diversity?

Wildlife managers, like most tourists, are convinced that our knowledge of large African mammals is more than satisfactory, as demonstrated from the large number of books existing on the subject e.g., [38,39]. Field guides, in particular, are suspected to vehicle an assuring view concerning our taxonomic knowledge of mammals [40]. Most users are unaware that such tools as field guides are intended to help identify what people see in a given place (i.e., to distinguish a bushbuck, *Tragelaphus scriptus* (Pallas, 1766) from a sitatunga *T. spekii* Speke, 1863; taxonomy follows predominantly [41]) but say nothing about the number of taxa, their phyletic relationships, and the rank accorded inside the

sitatunga and the bushbuck concepts [42]. Diatribes regarding taxonomic subdivisions of even the most well-known African mammals are widespread (e.g., African elephants [43], ungulates [44], felids [45], and canids [46]), including many species that are commonly relocated through the African continent (see Table 1). Wildlife managers are often convinced that wildlife was ubiquitous before humans exerted a strong pressure on it, which also leads to local extirpations.

Table 1. Some selected cases of mammalian translocations in Africa, with uncertain outcomes, including goals and consequent problems. Taxonomy follows predominantly [41].

Species	Original Range	Translocated to	Purpose	Aftermath	References
<i>Damaliscus pygargus phillipsi</i> Harper, 1939	Northeast South Africa	Southwest South Africa, Botswana, Mozambique, Namibia, Swaziland, Zimbabwe, Angola	Hunting restocking, conservation, eco-tourism	Genetic integrity of <i>D. p. pygargus</i> (Pallas, 1767); invasive in Angola	[19,47,48]
<i>Connochaetes taurinus</i> (Burchell, 1824)	Northern South Africa	Southern South Africa	Hunting, restocking	Genetic integrity of <i>Connochaetes gnou</i> (Zimmermann, 1780)	[30]
<i>Equus zebra hartmannae</i> Matschie, 1898	Namibia	Western Cape, Eastern Cape (South Africa)	Introduction	Genetic integrity of <i>E. z. zebra</i> Linnaeus, 1758	[49]
<i>Equus grevyi</i> Oustalet, 1882	Kenya	Kenya (outside natural range)	Conservation (favor range expansion)	Genetic integrity due to hybridization with <i>E. quagga boehmi</i> Matschie, 1892	[50,51]
<i>Aepyceros melampus petersi</i> Bocage, 1879	Namibia, Angola	Angola, Namibia	Conservation	Possible hybridization with <i>A.m. melampus</i> (Lichtenstein, 1812)	[52]
<i>Hippotragus equinus koba</i> (Gray, 1872)	West Africa	South Africa	Hunting, restocking	genetic integrity of <i>H. e. equinus</i> (É. Geoffroy Saint-Hilaire, 1803)	[19,53]
<i>Redunca fulvorufula fulvorufula</i> (Afzelius, 1815)	South Africa	Namibia	Conservation	Introduction in a new area outside the historic range; possible competition with other species, sanitary problems, etc.	[54]
<i>Tragelaphus angasii</i> Angas, 1849	South Africa	Botswana, Namibia, Angola	Hunting	Competition and hybridization with <i>T. strepsiceros</i> (Pallas, 1766); competition with <i>T. scriptus</i> (Angola)	[47,55]
<i>Hippotragus niger</i> (Harris, 1838) ssp.	Tanzania, Zambia, Mozambique, Malawi	South Africa	Conservation	Genetic integrity of different subspecies; possible hybridization with <i>H. equinus</i>	[56,57]
<i>Hippotragus equinus</i> ssp.	Namibia, Botswana, South Africa	South Africa	Hunting, conservation	Genetic integrity of different subspecies	[58,59]
<i>Antidorcas marsupialis marsupialis</i> (Zimmermann, 1780)	South Africa	South Africa	Hunting, introduction	Possible hybridization with <i>A. m. hofmeyri</i> Thomas, 1926	[60]

Table 1. Cont.

Species	Original Range	Translocated to	Purpose	Aftermath	References
<i>Tragelaphus spekii gratus</i> P. L. Sclater, 1880 (?)	Unknown locality	The Gambia	Tourism, aesthetic reasons	Introduction? reintroduction? population probably extinct	[61]
<i>Diceros bicornis michaeli</i> Zukowski, 1965	Kenya	South Africa	Conservation restocking	Genetic integrity of <i>D. b. bicornis</i> (Linnaeus, 1758)	[62,63]
<i>Diceros bicornis michaeli</i>	South Africa (ranches), EAZA zoos	Tanzania	Conservation restocking, and reintroduction	Genetic integrity of <i>D. b. michaeli</i>	[64–66] (see reference [66] in case of ex situ stock)
<i>Diceros bicornis</i> ssp.	All areas	East Africa	Conservation	Possible genetic erosion of Masai Mara population with recognized traces of gene pool of <i>D. b. longipes</i> Zukowski, 1949	[66]
<i>Ceratotherium simum simum</i> (Burchell, 1817)	Kenya (ranch)	Uganda	Conservation?	Introduction of one allochthonous subspecies into previous range of extinct <i>C. s. cottoni</i> (Lydekker, 1908)	[67]
<i>Ceratotherium simum simum</i>	South Africa	Kenya, Zambia	Conservation	creation of new nuclei outside the historic range	[68]
<i>Beatragus hunteri</i> (Sclater, 1889)	Southeast Kenya	Kenya (Tsavo East NP)	Conservation	Creation of a new population outside the natural range	[69,70]
<i>Taurotragus oryx oryx</i> (Pallas, 1766)	South Africa	Senegal	Tourism, aesthetic reasons	Possible competition, and genetic integrity of <i>T. derbianus derbianus</i> (Gray, 1847)	[71–73]
<i>Kobus ellipsiprymnus ellipsiprymnus</i> (Ogilby, 1833)	South Africa	Senegal	Tourism, aesthetic reasons	Genetic integrity of <i>K. e. unctuosus</i> (Laurillard, 1842)	[71,72]
<i>Oryx gazella gazella</i> (Linnaeus, 1758)	South Africa	Senegal	Tourism, aesthetic reasons	Introduction in an area where it has never been present	[71,72]
<i>Panthera leo</i> cf. <i>melanochaita</i> (H. Smith, 1842) (ex <i>P. l. krugeri</i> (Roberts, 1929))	South Africa	Rwanda	“Reintroduction”	Introduction outside the natural range. In Rwanda, was formerly present as <i>P. l. azandica</i> (Allen, 1924)	[74]

Taxonomic research, on the contrary, evidences the existence of discrete morphological (and sometimes genetic) discontinuity inside globally perceived “species,” with geographic patterns that often follow well-known biogeographical subdivisions of Africa, and sometimes an abrupt taxa’s limits congruent with biomes and vegetation types changes are observed [75]. Although we may detect areas of apparent intergradations between clearly different “forms” (for the African buffalo, see [76,77]), there are few doubts about the existence of a long and complicated history of adaptation, separation, retraction, and

advancement of geographic ranges of different taxa following climate changes, which must be summarized by a not so simple taxonomy, as is often requested by some stakeholders and researchers [78,79]. Finally, complex taxonomies are also difficult to translate into national and international legislations; yet, this is vital for effective conservation policy to maintain current levels of biodiversity [80].

3. Translocations and Conservation

Table 1 shows some selected cases of translocation of African mammals that caused or may cause serious genetic-conservation, as well as sanitary problems or other undesirable conservation consequences.

Regrettably, in some cases, as with the black rhinoceros *Diceros bicornis*, the issue remains a highly academic one as extirpation of most rhinoceros populations preceded modern evidence-based conservation management [66,81]. In the latter decades, the increasing attention to tourism by several African countries led to growing attention to protected areas and, if needed, to the reintroductions of species that became historically extinct in these regions [25,82,83]. Hybridization between distinct taxa is a common result of translocations in South Africa, where several genera such as *Connochaetes* and *Aepyceros* are involved [21,84]. The same threat is now spreading elsewhere, a case in point being the managed populations of two *Taurotragus* species in the same area in Senegal; one being the critically endangered *Taurotragus derbianus derbianus* whose genetic integrity is potentially threatened by the imported *T. oryx oryx*—fortunately in this case, no hybrids have yet been detected based on microsatellite markers [73]. In the same private “reserves” in Senegal, several species, including *Ceratotherium simum simum*, *Kobus ellipsiprymnus ellipsiprymnus*, *Tragelaphus strepsiceros strepsiceros*, *Aepyceros melampus melampus*, *Oryx gazella gazella*, and *Giraffa camelopardalis giraffa* have been introduced from South Africa [71,72] (see also Table 1), which is a fact that raises some concern, especially in cases of animal escapes. Ironically, translocations may also be responsible for the extinction of pure genetic lineages via hybridization, thereby negatively impacting endangered, indigenous, and rare species. Owing to a general neglect of taxonomy—following the so-called “taxonomic inertia” period described by [40]—it is highly probable that the hybridization problem is greatly undervalued in conservation circles. However, a more subtle danger is the general distortions of the genetic landscape, especially of wild ungulates, as is often reported in North America and Europe [85] but also from Africa [86], which may preclude further research, and which may create management problems that cannot be properly anticipated.

Poor taxonomic knowledge coupled with translocations may also lead to conservation initiatives that have only an aesthetic value and, more seriously, may divert attention from real priorities. On the other hand, we know that funds and conservation interest is not homogeneous in Africa [87], and, therefore, we may accept that the extent of biodiversity we risk to lose is greater than we think, especially outside East and South Africa.

4. Giraffes as a Case Study

The last decade saw an unparalleled interest in giraffe taxonomy, starting with studies by [82,88,89]. We associate this unprecedented progress with the use of various data types, increased sampling of studies and availability of the data, various expert knowledge, and the tuning of arguments via numerous discussions [44,90–94]. Currently, we have increasing knowledge about giraffe morphological differentiation [44,77,88], phylogenetic structure and timing of differentiations of current or extinct populations [89,95–100], gene flow [44,100,101], unique genomic signatures [102], and potential ecological factors responsible for the restricted gene flow [75]. Giraffes are thus becoming the model case study for the testing of species delimitation in mammals, similarly as cetaceans became the extraordinary model for understanding the various aspects of their and general vertebrate evolution in unprecedented detail [103,104].

Departing from the classical monotypic taxonomic account dominating the 20th century, Colin Groves and Peter Grubb considered that an eight species arrangement better

interpreted available morphological data [44]; for the basic overview of giraffe taxonomy across the 20th and 21st centuries, see Table 2. This latter work, adopting a phylogenetic species concept, was not always accepted by several researchers, yet, in the *Giraffa* case, subsequent genetic works were unanimous in always accepting more than one species. Four species were accepted by [99] and more recently by [100]—*Giraffa camelopardalis* (Linnaeus, 1758); *G. reticulata* De Winton, 1899; *G. giraffa* (Boddaert, 1785); and *G. tippelskirchi* Matschie, 1898—while [97] recognized three species: *Giraffa camelopardalis*, *G. giraffa*, and *G. tippelskirchi*. Although a step forward, several gaps in available data and interpretation were evident, as too often these reviews were based on previous arrangements without a critical examination of geographic and taxonomic gaps. [94] rightly stressed the taxonomic history of the taxon *G. g. rothschildi* Lydekker, 1903 to evidence the lack of consensus and possible negative setbacks for conservation. A preponderance of scientists have followed the conservative IUCN protocol and continue to refer to the different kinds of giraffes using the one species/nine subspecies account [94], although this view can be quite dangerous, or at least controversial, for conservation. For example, the materials related to [99] specifically posters a “Giraffe conservation guide” and provided a clear and alarming conservation message to the public about the patchy distribution with declining trends in many northern populations. Recently, an important contribution came from [98], which also included genetic data from key museum specimens belonging to now extinct populations. Among the most important findings include the description of a new—historically extinct—taxon from Senegal, *G. camelopardalis senegalensis* Petzold, Magnant, and Hassanin, 2020, which went extinct around 1970; the revalidation of *G. g. wardi* Lydekker, 1904; and a stricter geographical delimitation of the nominal *G. c. camelopardalis*, which make this taxon, the Nubian giraffe, another example of a taxon “allowed to slip into extinction unnoticed” [40]. The better definition of conservation units or ECUs is a greater priority today than fixing the number of species, considering that taxonomy is particularly relevant when translocations are considered a key conservation component [105]. Petzold and collaborators [98] have contributed to better delineate conservation units, especially in the northern continental sector that has traditionally received scarcer conservation attention and where biogeographical barriers, such as the Nile Basin, have been scarcely considered in mammal taxonomic studies [106,107]. The western giraffe, *G. c. peralta* Thomas, 1898 emerges unanimously as a distinct taxonomic unit and hence as a conservation priority. Translocations to create more populations should be promoted, whereas the creation of breeding populations of extralimital taxa, such as the *G. c. giraffa* imported from South Africa to Senegal in the Bandia Private Reserve [108], pose more than a question regarding their conservation relevance and possible interference with long-term *G. c. peralta* conservation. Regrettably, it can be hypothesized that tourism development in West Africa may lead to further attempts to reconstruct a “true African wildlife experience” for naïve tourists through the importation of stocks from South African wildlife reserves, de facto enlarging a problem of genetic pollution that is already widespread in South Africa [19]. In another critical region, the Nile Basin, translocations have a critical role to play in re-establishing species to their former range [109]. It is vital however that each remaining nominal taxon/population is managed separately, avoiding premature lumping based on scattered evidence and a lack of awareness of biogeographical and ecological barriers. For example, [110] had already proposed a close phyletic relationship between the subspecies *rothschildi*, *cottoni* Lydekker, 1904 and *antiquorum* (Swainson, 1835), and although genetic data seems to confirm this, the adaptive significance of some morphological features of *rothschildi* (dark coloration and two additional posterior horns in the males) are still unknown. Considering the fact that some authors specify in detail some morphological differences between *camelopardalis* and *rothschildi*, and other taxa [110–112], it would be worthwhile to inspect their validity using modern statistical methods (e.g., multivariate methods, discriminant analysis). Still more controversial seems the inclusion of the taxon *congoensis* Lydekker, 1903 as a synonym of *antiquorum*, considering that his original descriptor considered it an intermediate between northern and southern giraffes because of completely spotted limbs (a southern character-

istic) and a well-developed frontal horn (a northern characteristic) [110,113]. The putative taxon *congoensis* is actually restricted to the Garamba National Park on the left side of the White Nile, whereas putative *cottoni* is found on the other side of the White Nile, which is apparently a non-trivial zoogeographical barrier.

In their recent genetic work [98], Petzold and collaborators evidenced how the nominal *G. c. camelopardalis* must now be considered extinct. This is further evidence of the relevance of sampling museum specimens from type locality [114] even if we have to correct these authors by circumscribing the type locality of *camelopardalis* to the Setit River (in present day Sudan and Eritrea), a well-known area for giraffe zoo collectors in the 19th and first half of the 20th centuries [115,116]. According to [98], the residual giraffe populations from the Gambella and Omo NPs in Southwest Ethiopia belong to *rothschildi*, but, as the only survivors in a complex biogeographical region east of the Nile that is still partially unexplored biologically [117], we recommend affording great priority to their conservation.

In summary, translocations of giraffes have a long tradition [105]. Some represent a helpful conservation management tool, as we mentioned above, but some others are questionable from a conservation perspective, at least until we have a clear understanding of separable conservation units in some key regions that have so far received little attention.

Table 2. Giraffe taxa recognized in the 20th and 21st centuries across several basic sources, which seemed to assess giraffes independently and/or using different data.

Source	[110,118]	[111]	[119]	[120]	[121]	[122]	[112]	[39]
Data Used	C+SSH	C+SSH	U(M)	U(M)	U(M)	U(M)	C	U(M)
<i>aethiopica</i>								
<i>africana</i>								
<i>angolensis</i>	x	x	x	x	x	x	x	x
<i>antiquorum</i>	x	x	x		x	x	x	
<i>australis</i>	<i>capensis</i>				<i>giraffa</i>	<i>giraffa</i>		
<i>biturigum</i>					<i>camelopardalis</i>	<i>camelopardalis</i>		
<i>camelopardalis</i>		x	x	x	x	x	x	x
<i>capensis</i>	x	x			<i>giraffa</i>	<i>giraffa</i>		
<i>congoensis</i>	x	x			<i>camelopardalis</i>	<i>camelopardalis</i>		x
<i>cottoni</i>	x	x			<i>rothschildi</i>	<i>rothschildi</i>		
<i>giraffa</i>			x	x	x	x	x	x
<i>hagenbecki</i>		<i>reticulata</i>			<i>reticulata</i>	<i>reticulata</i>		
<i>infumata</i>		x			<i>angolensis</i>	<i>angolensis</i>		
<i>maculata</i>		<i>capensis</i>			<i>giraffa</i>	<i>giraffa</i>		
<i>nigrescens</i>	x	<i>reticulata</i>			<i>reticulata</i>	<i>reticulata</i>		
<i>peralta</i>	x	x	x	x	x	x	x	x
<i>renatae</i>								
<i>reticulata</i>	x *	x	x	x	x	x	x	x
<i>rothschildi</i>	x	x	x	x	x	x	x	
<i>senegalensis</i>								
<i>schillingsi</i>	<i>tippelskirchi</i>	<i>tippelskirchi</i>			<i>tippelskirchi</i>	<i>tippelskirchi</i>		
<i>senariensis</i>		<i>camelopardalis</i>			<i>antiquorum</i>	<i>antiquorum</i>		
<i>thornicrofti</i>	x	x		x	x	x	x	x
<i>tippelskirchi</i>	x	x	x	x	x	x	x	x
<i>typica</i>	x	<i>camelopardalis</i>			<i>camelopardalis</i>			
<i>wardi</i>	x	x			<i>giraffa</i>			
valid taxa (sp.; ssp.)	2; 13	2; 13	1; 8	1; 8	1; 9	1; 9	1; 9	1; 8
Source	[123]	[88]	[41]	[82]	[124]	[125]		
Data used	U(M)	C+mtDNA+SSH+SSI	U(M)	mtDNA+STRs	U(M+G)	BSI+C+mtDNA+SSH+SSI		
<i>aethiopica</i>			<i>camelopardalis</i>					
<i>africana</i>			<i>camelopardalis</i>					
<i>angolensis</i>	<i>giraffa</i>	x or <i>giraffa</i> ?	<i>giraffa</i>	x *	x	x		
<i>antiquorum</i>	x	x or <i>camelopardalis</i> ?	<i>camelopardalis</i>	not sampled	x	x		
<i>australis</i>			<i>giraffa</i>					
<i>biturigum</i>			<i>camelopardalis</i>					
<i>camelopardalis</i>	x	x	x	not sampled	x	x		
<i>capensis</i>	<i>giraffa</i>		<i>giraffa</i>			<i>giraffa</i>		
<i>congoensis</i>	<i>antiquorum</i>		<i>camelopardalis</i>					

Table 2. Cont.

<i>cottoni</i>			<i>rothschildi</i>			<i>camelopardalis</i>
<i>giraffa</i>	x	x	x	x *	x	x
<i>hagenbecki</i>			<i>reticulata</i>			
<i>infumata</i>	<i>giraffa</i>		<i>giraffa</i>			<i>angolensis</i>
<i>maculata</i>			<i>giraffa</i>			
<i>nigrescens</i>			<i>reticulata</i>			
<i>peralta</i>	<i>antiquorum</i>	x or <i>camelopardalis</i> ?	<i>camelopardalis</i>	x *	x	x
<i>renatae</i>			<i>camelopardalis</i>			
<i>reticulata</i>	x	x	x	x *	x	x
<i>rothschildi</i>	<i>camelopardalis</i>	x	x	x *	x	<i>camelopardalis</i>
<i>senegalensis</i>						
<i>schillingsi</i>			<i>tippelskirchi</i>			
<i>senaariensis</i>			<i>camelopardalis</i>			
<i>thornicrofti</i>	x	x	x	not sampled	x	x
<i>tippelskirchi</i>	x	x	x	x *	x	x
<i>typica</i>			<i>camelopardalis</i>			
<i>wardi</i>	<i>giraffa</i>		<i>giraffa</i>			<i>giraffa</i>
valid taxa (sp.; ssp.)	1; 6	1; min. 6	1; 6	6; min. 6, up to 11	1; 9	1; 8
Source	[126]	[44,91]	[99–101]	[97,98]		
Data used	U	C+SSH+SSI+mtDNA	G+mtDNA+nDNA	C+mtDNA+nDNA+SSH+SSI		
<i>aethiopica</i>				G. c.		
<i>africana</i>				<i>camelopardalis</i>		
<i>angolensis</i>	x	x *	x (as subspecies of G. <i>giraffa</i>)	G. g. <i>giraffa</i>		
<i>antiquorum</i>	x	x *	x (as subsp. of G. <i>camelopardalis</i>)	x (as subsp. of G. <i>camelopardalis</i>)		
<i>australis</i>				G. g. <i>giraffa</i>		
<i>biturigum</i>				G. c.		
<i>camelopardalis</i>	x	x *	x *	<i>camelopardalis</i>		
<i>capensis</i>		<i>giraffa</i>		x *		
<i>congoensis</i>		<i>antiquorum</i> ?		G. g. <i>giraffa</i>		
<i>cottoni</i>		<i>camelopardalis</i>		G. c.		
<i>giraffa</i>	x	x *	x *	<i>antiquorum</i>		
<i>hagenbecki</i>				x *		
<i>infumata</i>		<i>giraffa</i>		G. c.		
<i>maculata</i>				<i>reticulata</i>		
<i>nigrescens</i>				G. g. <i>wardi</i>		
<i>peralta</i>	x	x *	x (as subsp. of G. <i>camelopardalis</i>)	G. g. <i>giraffa</i>		
<i>renatae</i>				G. c.		
<i>reticulata</i>	x	x *	x *	<i>reticulata</i>		
<i>rothschildi</i>	x	<i>camelopardalis</i>	G. c. <i>camelopardalis</i>	x (as subsp. of G. <i>camelopardalis</i>)		
<i>senegalensis</i>				x (as subsp. of G. <i>camelopardalis</i>)		
<i>schillingsi</i>				x (as subsp. of G. <i>camelopardalis</i>)		
<i>senaariensis</i>				G. t.		
<i>thornicrofti</i>	x	x *	x (as subsp. of G. <i>tippelskirchi</i>)	<i>tippelskirchi</i>		
<i>tippelskirchi</i>	x	x *	x *	G. c.		
				<i>antiquorum</i>		
				x (as subsp. of G. <i>tippelskirchi</i>)		
				<i>tippelskirchi</i>		
				x *		

Table 2. Cont.

<i>typica</i>				<i>G. c.</i>
				<i>camelopardalis</i>
<i>wardi</i>		<i>giraffa</i>		x (as subsp. of <i>G. giraffa</i>)
valid taxa (sp.; ssp.)	1; 9	8; not recognized	4; 7	3; 10

Taxa recognized as valid are labelled by “x.” Taxa recognized as species are labelled by “*.” Synonyms of specific taxa are specified when they were noted by particular authors. Abbreviations: BSI—body size, C—coloration, G—genomic data, mtDNA—mitochondrial DNA, nDNA—nuclear DNA, sp.—species, ssp.—subspecies, SSH—skull shapes, SSI—skull size, STRs—microsatellites, U—unspecified, U(M)—unspecified (presumably morphology), and U(M+G)—unspecified (presumably morphological and genetic data).

5. Discussion

As the current discussion about giraffe taxonomy is too focused on the number of species, we argue that the plurality of taxonomic and conservation approaches might be beneficial in order to unify conservation priorities, contra e.g., [127]; see also the comments on taxonomic instability in [128]. Taxonomists often use some particular species concept (for review see [129]), and population/evolutionary geneticists describe interesting results but often without presenting formal taxonomic actions [130,131] and/or alternatively using some standard conservation units—usually management unit (MU), evolutionary significant unit (ESU), cf. [132], or elemental conservation unit (ECU), cf. [133]. The reasons for diverse approaches are various, from the objective (e.g., lost or inaccessible type material, unsuccessful isolation of DNA from type series, difficulty to have samples from same regions) to the subjective (personal adherence to a particular approach based on various reasons or scarce knowledge of taxonomic practice); moreover, a consensus about the “species” label is often hardly obtainable, albeit units deserving conservation attention could be identical or very similar. This is also the case with giraffes (Table 2), because these populations—*angolensis*, *antiquorum*, *camelopardalis*, *giraffa*, *peralta*, *reticulata*, *rothschildi*, *thornicrofti*, and *tippelskirchi*—have been recognized as valid taxa in at least 13 of 18 reviewed sources since 1904 (Table 2). The concordance about the uniqueness of these populations and the latest assessments of giraffes, e.g., [97–101] are quite considerable. Therefore, we recommend using several criteria to define basic taxonomic and conservation units contemporarily (cf. [66] in the case of *Diceros bicornis* using MU, ESU, and higher level ESUs) in conservation management plans and programs in order to find the majority consensus on which “lineages” deserve conservation attention. This approach would meet integrative taxonomy standards [134]. Robuchon and collaborators [135] offered an excellent framework for evaluation of the impact of species splitting on species priority-setting that is more than recommended for accommodating new taxonomic knowledge in conservation strategies. Genomic data holds enormous potential to resolve species delimitation and recognize demographic history and adaptive potential in detail [136]; its usage should be recommended for future studies. The common practice (e.g., making sequences and other data available on GenBank and/or dryad data platforms as much as possible) should be continued, because it enables independent data testing and uses different approaches. Additionally, we highly recommend associating photographs and/or basic descriptions of the phenotype/size of the DNA voucher specimens, as proposed by [137].

As some current genomic assessments have recognized significant differentiation of populations, specifically in plains zebra *Equus quagga* Boddaert, 1785 [138] or tiger *Panthera tigris* (Linnaeus, 1758) [139], which have been recognized as undifferentiated based on small number of loci (zebras [130]) and some phenotype and ecological features (tigers [140]), we argue for a precautionary principle in conservation management [128,135,141].

Africa is a diverse continent with a growing ecotourism industry. Charismatic megavertebrates are particularly searched by tourists and “big game” hunters, but small-scale tourism may furnish valuable income to communities protecting valuable or appealing populations such as the Niger giraffe *G. (camelopardis) peralta*, the Bor maneless zebra *E. quagga borensis* Lönnberg, 1921, or the mountain nyala *Tragelaphus buxtoni* (Lydekker,

1910); income may also be generated by active promotion of a more sensitive mammal watching activity [142].

A further threat is represented by conservative taxonomies that are proposed by conservation groups without a real review of new and old data, as has been recently the case with the IUCN/SSC Cat Specialist Group [45]. Acceptance of a two-subspecies arrangement largely based on some genetic data seems to overly neglect the great phenotypic diversity still found, for instance, in lion *Panthera leo* (Linnaeus, 1758) [143,144] and ignores some studies that highlight genetic divergence promoted by ecological discontinuity [145,146] that seems to support a new paradigm to explain microevolutionary divergence in widespread, large-sized carnivores [147]. Therefore, without forgetting or underestimating the composite problems of lion conservation, and of coexistence/interaction with humans involving this highly charismatic species [148,149], taxonomic issues deserve to be included in the future conservation strategies of this charismatic species. Paradoxically, tourists (but also trophy hunters) may be greatly interested in phenotypic lion diversity, and works such as [143] may provide the input for further travels in different regions of Africa and the promotion of new conservation/tourism projects for little-known, overlooked populations such as those of Southwestern Ethiopia. Regrettably, genetic considerations alone have suggested merging quite distinct captive populations of two gazelle subspecies *Nanger dama dama* (Pallas, 1766) and *N. d. mhorh* (Bennett, 1833) [150]. Apart from pure scientific considerations [151], this line of action should preclude the possibility for Western Sahara communities to have a part in managing for their own welfare a unique ungulate taxon, which is an important incentive for local ecotourism; this is a view opposed to that recently presented in [152].

6. Conclusions

It should be emphasized that the taxonomically sensitive translocation policies [153] would be important for the preservation of current diversity but also for the ecological restoration of some regions within rewilding, which could be essential for the preservation of some unique (often refugee) species [154,155]. Considering the restricted distribution of some giraffe taxa [98,156] and its important role in communities as flower predators [157], giraffe translocations [105] have a great potential that should be utilized for future generations.

Furthermore, the creation in Africa of fenced private or state wildlife reserves filled with exotic stocks not only have a disputable educative effect on tourists and also on local wildlife managers, actually creating or perpetuating a homogenized idea of “African wildlife,” but even create the possibility of concrete dangers. In fact, some species can escape by crossing the fences, which has already happened, and the potential exists to create free allochthonous extra-range nuclei and also to undermine the genetic integrity of native subspecies possibly present in the areas surrounding the reserves.

It is time that conservation biologists recognize the immense threat that taxonomical oversight coupled with the great economic significance of tourism poses to the diversity and integrity of the “genetic landscape” (that is, the evolutionary landscape) of large mammals in Africa. If we wish to preserve such heritage, keeping it as unmodified as possible to future generations of African people and investigators, we need to act now with an urgent change of attitude toward these issues.

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References

1. Faith, J.T. Late Pleistocene and Holocene mammal extinctions on continental Africa. *Earth Sci. Rev.* **2014**, *128*, 105–121. [[CrossRef](#)]
2. Faith, J.T.; Rowan, J.; Du, A.; Paul, L.; Koch, P.L. Plio-Pleistocene decline of African megaherbivores: No evidence for ancient hominin impacts. *Science* **2018**, *362*, 938–941. [[CrossRef](#)]
3. Blixen, K.C. *Out of Africa*; Putnam: London, UK, 1937.
4. Akama, J.S. Western environmental values and nature-based tourism in Kenya. *Tour. Manag.* **1996**, *17*, 567–574. [[CrossRef](#)]
5. Schwarzer, J.; Swartz, E.R.; Vreven, E.; Snoeks, J.; Cotterill, F.P.D.; Misof, B.; Schliewen, U.K. Repeated trans-watershed hybridization among haplochromine cichlids (Cichlidae) was triggered by Neogene landscape evolution. *Proc. R. Soc. B* **2012**, *279*, 4389–4398. [[CrossRef](#)] [[PubMed](#)]
6. Haas, F.; Schwarz, E. Zur Entwicklung der afrikanischen Stromsysteme. *Geol. Rundsch.* **1913**, *4*, 603–607. [[CrossRef](#)]
7. Heller, E. The Geographical Barriers to the Distribution of Big Game Animals in Africa. *Geogr. Rev.* **1918**, *6*, 297–319. [[CrossRef](#)]
8. Schwarz, E. Huftiere aus West-und Zentralafrika. *Ergeb. Dtsch. Zent. Afr. Exped.* **1920**, *1*, 831–1044.
9. Lönnerberg, E. The development and distribution of the African fauna in connection with and depending upon climate changes. *Ark. Zool.* **1929**, *21*, 1–33.
10. Kingdon, J. *East African Mammals: An Atlas of Evolution in Africa*; Academic Press: London, UK, 1971; Volume 1.
11. Kingdon, J. *Island Africa*; Collins: London, UK, 1990.
12. Maglio, V.J.; Cooke, H.B.S. *Evolution of African Mammals*; Harvard University Press: Cambridge, MA, USA, 1978.
13. Vrba, E.S.; Denton, G.H.; Partridge, T.C.; Burckle, L.H. *Paleoclimate and Evolution, with Emphasis on Human Origins*; Yale University Press: New Haven, CT, USA; London, UK, 1996.
14. Bromage, T.; Schrenk, F. *African Biogeography, Climate Change, and Early Hominid Evolution*; Oxford University Press: Oxford, UK, 1999.
15. Schikora, T.F. Climate-Linked Temporal and Spatial Patterns in the Evolution of African Bovidae. Ph.D. Thesis, Johann Wolfgang Goethe-Universität, Frankfurt, Germany, 2000.
16. Cotterill, F.P.D. The Evolutionary History and Taxonomy of the Kobus Leche Species Complex of South-Central Africa in the Context of Palaeo-Drainage Dynamics. Ph.D. Thesis, University of Stellenbosch, Stellenbosch, South Africa, 2006.
17. Couvreur, T.L.; Dauby, G.; Blach-Overgaard, A.; Deblauwe, V.; Dessein, S.; Droissart, V.; Hardy, O.J.; Harris, D.J.; Janssens, S.B.; Ley, A.C.; et al. Tectonics, climate and the diversification of the tropical African terrestrial flora and fauna. *Biol. Rev.* **2021**, *96*, 16–51. [[CrossRef](#)]
18. Price, R.A. *The Contribution of Wildlife to the Economies of Sub Saharan Africa: K4D Helpdesk Report*; Institute of Development Studies: Brighton, UK, 2017.
19. Castley, J.G.; Boshoff, A.F.; Kerley, G.I.H. Compromising South Africa’s natural biodiversity—Inappropriate herbivore introductions. *S. Afr. J. Sci.* **2001**, *97*, 344–348.
20. Spear, D.; Chown, S.L. The extent and impacts of ungulate translocations: South Africa in a global context. *Biol. Conserv.* **2009**, *142*, 353–363. [[CrossRef](#)]
21. Russo, I.R.M.; Hoban, S.; Bloomer, P.; Kotzé, A.; Segelbacher, G.; Rushworth, I.; Birss, C.; Bruford, M.W. ‘Intentional Genetic Manipulation’ as a conservation threat. *Conserv. Genet. Resour.* **2019**, *11*, 237–247. [[CrossRef](#)]
22. Berger-Tal, O.; Blumstein, D.T.; Swaisgood, R.R. Conservation translocations: A review of common difficulties and promising directions. *Anim. Conserv.* **2020**, *23*, 121–131. [[CrossRef](#)]
23. Kotzé, A.; Smith, R.M.; Moodley, Y.; Luikart, G.; Birss, C.; Van Wyk, A.M.; Van Wyk, A.M.; Grobler, J.P.; Dalton, D.L. Lessons for conservation management: Monitoring temporal changes in genetic diversity of Cape mountain zebra (*Equus zebra zebra*). *PLoS ONE* **2019**, *14*, e0220331. [[CrossRef](#)] [[PubMed](#)]
24. Smith, R.M.; Bhoora, R.V.; Kotzé, A.; Grobler, J.P.; Dalton, D.L. Translocation a potential corridor for equine piroplasms in Cape mountain zebra (*Equus zebra zebra*). *Int. J. Parasitol. Parasites Wildl.* **2019**, *9*, 130–133. [[CrossRef](#)]
25. Correia, M.; Timéteo, S.; Rodríguez-Echeverría, S.; Mazars-Simon, A.; Heleno, R. Refaunation and the reinstatement of the seed-dispersal function in Gorongosa National Park. *Conserv. Biol.* **2017**, *31*, 76–85. [[CrossRef](#)]
26. Cromsigt, J.P.G.M.; te Beest, M.; Kerley, G.I.H.; Landman, M.; le Roux, E.; Smith, F.A. Trophic rewilding as a climate change mitigation strategy? *Philos. Trans. R. Soc. B* **2018**, *373*, 20170440. [[CrossRef](#)] [[PubMed](#)]
27. Germano, J.M.; Field, K.J.; Griffiths, R.A.; Clulow, S.; Foster, J.; Harding, G.; Swaisgood, R.R. Mitigation-driven translocations: Are we moving wildlife in the right direction? *Front. Ecol. Environ.* **2015**, *13*, 100–105. [[CrossRef](#)]
28. Ackermann, R.R.; Brink, J.S.; Vrahimis, S.; de Klerk, B. Hybrid wildebeest (*Artiodactyla: Bovidae*) provide further evidence for shared signatures of admixture in mammalian crania. *S. Afr. J. Sci.* **2010**, *106*, 1–4. [[CrossRef](#)]
29. Grobler, J.P.; Rushworth, I.; Brink, J.S.; Bloomer, P.; Kotze, A.; Reilly, B.; Vrahimis, S. Management of hybridization in an endemic species: Decision making in the face of imperfect information in the case of the black wildebeest—*Connochaetes gnou*. *Eur. J. Wildl. Res.* **2011**, *57*, 997–1006. [[CrossRef](#)]

30. Benjamin-Fink, N.; Reilly, B.K. Conservation implications of wildlife translocations; The state's ability to act as conservation units for wildebeest populations in South Africa. *Glob. Ecol. Conserv.* **2017**, *12*, 46–58. [CrossRef]
31. Spear, D.; Chown, S.L. Taxonomic homogenization in ungulates: Patterns and mechanisms at local and global scales. *J. Biogeogr.* **2008**, *35*, 1962–1975. [CrossRef]
32. Goss, J.R.; Cumming, G.S. Networks of wildlife translocations in developing countries: An emerging conservation issue. *Front. Ecol. Environ.* **2013**, *11*, 243–250. [CrossRef]
33. Maciejewski, K.; Kerley, G.I.H. Understanding tourists' preference for mammal species in private protected areas: Is there a case for extralimital species for ecotourism? *PLoS ONE* **2014**, *9*, e88192. [CrossRef]
34. van Hoven, W. Private game reserves in Southern Africa. In *Institutional Arrangements for Conservation, Development and Tourism in Eastern and Southern Africa: A Dynamic Perspective*; Springer: Dordrecht, The Netherlands, 2015; pp. 101–118. [CrossRef]
35. Lindsey, P.A.; Roulet, P.A.; Romañach, S.S. Economic and conservation significance of the trophy hunting industry in sub-Saharan Africa. *Biol. Conserv.* **2007**, *134*, 455–469. [CrossRef]
36. Muposhi, V.K.; Gandiwa, E.; Makuza, S.M.; Bartels, P. Ecological, physiological, genetic trade-offs and socio-economic implications of trophy hunting as a conservation tool: A narrative review. *J. Anim. Plant Sci.* **2017**, *27*, 1–14.
37. Seddon, P.J.; Strauss, W.M.; Innes, J. Animal translocations: What are they and why do we do them? In *A Reintroduction Biology: Integrating Science and Management*, 1st ed.; Ewen, J.G., Armstrong, D.P., Parker, K.A., Seddon, P.J., Eds.; Wiley-Blackwell: Oxford, UK, 2012; pp. 1–32. [CrossRef]
38. Dorst, J.; Dandelot, P. *A Field Guide to the Larger Mammals of Africa*; Collins: London, UK, 1970.
39. Kingdon, J. *The Kingdon Field Guide to African Mammals*; Academic Press: San Diego, CA, USA, 1997.
40. Gippoliti, S.; Cotterill, F.P.D.; Zinner, D.; Groves, C.P. Impacts of taxonomic inertia for the conservation of African ungulate diversity: An overview. *Biol. Rev.* **2018**, *93*, 115–130. [CrossRef]
41. Grubb, P. Artiodactyla. In *Mammal Species of the World: A Taxonomic and Geographic Reference*; Wilson, D.E., Reeder, D.M., Eds.; The Johns Hopkins University Press: Baltimore, MD, USA, 2005; pp. 637–722.
42. Gippoliti, S.; Groves, C.P. Cryptic problematic species and troublesome taxonomists: A tale of the Apennine bear and the Nile white rhinoceros. In *Problematic Wildlife II*; Angelici, F.M., Rossi, L., Eds.; Springer: Cham, Switzerland, 2020; pp. 509–527. [CrossRef]
43. Grubb, P.; Groves, C.P.; Dudley, J.P.; Shoshani, J. Living African elephants belong to two species: *Loxodonta africana* (Blumenbach, 1797) and *Loxodonta cyclotis* (Matschie, 1900). *Elephant* **2000**, *2*, 1–4. [CrossRef]
44. Groves, C.P.; Grubb, P. *Ungulate Taxonomy*; The Johns Hopkins University Press: Baltimore, MD, USA, 2011.
45. Kitchener, A.C.; Breitenmoser-Würsten, C.; Eizirik, E.; Gentry, A.; Werdelin, L.; Wilting, A.; Yamaguchi, N.; Abramov, A.V.; Christiansen, P.; Driscoll, C.; et al. A revised taxonomy of the Felidae: The final report of the Cat Classification Task Force of the IUCN/SSC Cat Specialist Group. *Cat News* **2017**, *Special Issue 11*, 1–80.
46. Gippoliti, S.; Lupi, L. A note on the wild canids (Carnivora: Canidae) of the Horn of Africa, with the first evidence of a new-forgotten-species for Ethiopia *Canis mensesi* Noack, 1897. *Bonn Zool. Bull.* **2020**, *69*, 111–115. [CrossRef]
47. Walker, J.F. Will Secret Wildlife Imports Doom Ultra-Rare Giant Sable? Available online: <https://www.nationalgeographic.com/animals/article/150521-angola-giant-sable-antelope-operation-noahs-ark-south-africa> (accessed on 22 May 2021).
48. van Wyk, A.M.; Dalton, D.L.; Hoban, S.; Bruford, M.W.; Russo, I.M.; Birss, C.; Grobler, P.; van Vuuren, B.J.; Kotzé, A. Quantitative evaluation of hybridization and the impact on biodiversity conservation. *Ecol. Evol.* **2017**, *7*, 320–330. [CrossRef]
49. Moodley, Y.; Harley, E.H. Population structuring in mountain zebras (*Equus zebra*): The molecular consequences of divergent demographic histories. *Conserv. Genet.* **2005**, *6*, 953–968. [CrossRef]
50. Cordingley, J.E.; Sundaresan, S.R.; Fischhoff, I.R.; Shapiro, B.; Ruskey, J.; Rubenstein, D.I. Is the endangered Grevy's zebra threatened by hybridization? *Anim. Conserv.* **2009**, *12*, 505–513. [CrossRef]
51. Schieltz, J.M.; Rubenstein, D.I. Caught between two worlds: Genes and environment influence behaviour of plains × Grevy's zebra hybrids in central Kenya. *Anim. Behav.* **2015**, *106*, 17–26. [CrossRef]
52. Green, W.C.H.; Rothstein, A. Translocation, hybridisation and the endangered black-faced impala. *Conserv. Biol.* **1998**, *12*, 475–480. [CrossRef]
53. Measey, J.; Hui, C.; Somers, M.J. Terrestrial Vertebrate Invasions in South Africa. In *Biological Invasions in South Africa*; van Wilgen, B., Measey, J., Richardson, D., Wilson, J., Zengeya, T., Eds.; Springer: Cham, Switzerland, 2020; pp. 115–151. [CrossRef]
54. Taylor, A.; Avenant, N.; Schulze, E.; Viljoen, P.; Child, M.F. A conservation assessment of *Redunca fulvorufula fulvorufula*. In *The Red List of Mammals of South Africa, Swaziland and Lesotho*; Child, M.F., Roxburgh, L., Do Linh San, E., Raimondo, D., Davies-Mostert, H.T., Eds.; South African National Biodiversity Institute and Endangered Wildlife Trust: Pretoria and Gauteng, South Africa, 2016; pp. 1–7.
55. Furstenburg, D. Nyala *Tragelaphus angasii*. In *The New Game Rancher*; Oberem, P., Oberem, P.T., Eds.; Briza Publications: Queenswood, South Africa, 2016; pp. 1–14.
56. Matthee, C.A.; Robinson, T.J. Mitochondrial DNA population structure of roan and sable antelope: Implications for the translocation and conservation of the species. *Mol. Ecol.* **1999**, *8*, 227–238. [CrossRef]
57. Furstenburg, D. Sable Antelope *Hippotragus niger*. In *The New Game Rancher*; Oberem, P., Oberem, P., Eds.; Briza Publications: Queenswood, South Africa, 2016; pp. 1–13.

58. Alpers, D.L.; Van Vuuren, B.J.; Arctander, P.; Robinson, T.J. Population genetics of the roan antelope (*Hippotragus equinus*) with suggestions for conservation. *Mol. Ecol.* **2004**, *13*, 1771–1784. [CrossRef] [PubMed]
59. Barrie, A. Translocation of Roan Antelope in South Africa and the Effect This Has Had on the Genetic Diversity of the Species. Mini Dissertation, University of Johannesburg, Johannesburg, South Africa, 2015. Available online: <http://hdl.handle.net/10210/2100> (accessed on 22 May 2021).
60. Furstenburg, D. Springbok *Antidorcas marsupialis*. In *The New Game Rancher*; Oberem, P., Oberem, P.T., Eds.; Briza Publications: Queenswood, South Africa, 2016; pp. 1–18.
61. Starin, E.D. Notes on sitatunga in The Gambia. *Afr. J. Ecol.* **2000**, *38*, 339–342. [CrossRef]
62. Carter, N. *Arm'd Rhinoceros*; Andre Deutsch: London, UK, 1965.
63. Penzhorn, B.L. A summary of the re-introduction of ungulates into South African National Parks (to December 1970). *Koedoe* **1971**, *14*, 145–159. [CrossRef]
64. Knight, M.H.; Kerley, G.I.H. Black rhino translocations within Africa. *Afr. Insight* **2009**, *39*, 70–83. [CrossRef]
65. Fyumagwa, R.D.; Nyahongo, J.W. Black rhino conservation in Tanzania: Translocation efforts and further challenges. *Pachyderm* **2010**, *47*, 59–65.
66. Moodley, Y.; Russo, I.-R.M.; Dalton, D.L.; Kotzé, A.; Muya, S.; Haubensak, P.; Bálint, B.; Munimanda, G.K.; Deimel, C.; Setzer, A.; et al. Extinctions, genetic erosion and conservation options for the black rhinoceros (*Diceros bicornis*). *Sci. Rep.* **2017**, *7*, 41417. [CrossRef] [PubMed]
67. Sheil, D.; Kirkby, A.E. Observations on Southern white rhinoceros *Ceratotherium simum simum* translocated to Uganda. *Trop. Conserv. Sci.* **2018**, *11*, 1–7. [CrossRef]
68. Emslie, R.; Brooks, M. *African Rhino: Status Survey and Conservation Action Plan*; IUCN/SSC African Rhino Specialist Group: Gland, Switzerland; Cambridge, UK, 1999.
69. Grimwood, I.R. Airlift for Hunter's antelope: Rescue operation in Kenya. *Oryx* **1964**, *7*, 164–167. [CrossRef]
70. Hofmann, R.R. Hirola: Translocation to Tsavo NP and new scientific information. *Gnusletter* **1996**, *15*, 2–5.
71. Vincke, X.; Hornick, J.-L.; Njikam, N.I.; Leroy, P. Gestion de la faune sauvage au Sénégal: Comparaison du Parc National du Niokolo Koba et de la Réserve privée de Bandia. *Ann. Med. Vet.* **2005**, *149*, 232–237.
72. Vermeulen, C. La réserve faunique de Bandia (Sénégal): Modèle ou contre-modèle pour l'Afrique de l'Ouest? *Parcs et Réserves* **2010**, *65*, 23–27.
73. Kubátová, A.; Štochlová, K.; Brandlová, K.; Jůnková Vymyslická, P.; Bolfíková, B.C. Comparison of divergent breeding management strategies in two species of semi-captive eland in Senegal. *Sci. Rep.* **2020**, *10*, 8841. [CrossRef]
74. Hall, S. The king of the jungle is back. Rwanda welcomes lions at Akagera National Park after 21 years' absence. *Eye Mag.* **2015**, 32–33.
75. Thomassen, H.A.; Freedman, A.H.; Brown, D.M.; Buermann, W.; Jacobs, D.K. Regional differences in seasonal timing of rainfall discriminate between genetically distinct East African giraffe taxa. *PLoS ONE* **2013**, *8*, e77191. [CrossRef]
76. Grubb, P. Variation and incipient speciation in the African buffalo. *Z. Säugetierkd.* **1972**, *37*, 121–144.
77. Grubb, P. Morphocline evolution in ungulates. In *Antelopes, Deer, and Relatives*; Vrba, E.S., Schaller, G.B., Eds.; Yale University Press: New Haven, CT, USA; London, UK, 2000; pp. 156–170.
78. Taylor, P.J.; Denys, C.; Cotterill, F.P.D. Taxonomic anarchy or an inconvenient truth for conservation? Accelerated species discovery reveals evolutionary patterns and heightened extinction threat in Afro-Malagasy small mammals. *Mammalia* **2019**, *38*, 313–329. [CrossRef]
79. Gippoliti, S. Species delimitation in mammals: A comment on Zachos (2018). *Mamm. Biol.* **2019**, *94*, 127–131. [CrossRef]
80. Gippoliti, S.; Capula, M.; Ficetola, G.F.; Salvi, D.; Andreone, F. Threatened by legislative conservationism? The case of the critically endangered Aeolian lizard. *Front. Ecol. Evol.* **2017**, *5*, 130. [CrossRef]
81. Rookmaaker, K. The black rhino needs a taxonomic revision for sound conservation. *Int. Zoo News* **2005**, *52*, 280–282.
82. Brown, D.M.; Brenneman, R.A.; Koepfli, K.-P.; Pollinger, J.P.; Milá, B.; Georgiadis, N.J.; Louis, E.E., Jr.; Grether, G.F.; Jacobs, D.K.; Wayne, R.K. Extensive population genetic structure in the giraffe. *BMC Biol.* **2007**, *5*, 57. [CrossRef]
83. Huntley, B.J.; Russo, V.; Lages, F.; Ferrand, N. *Biodiversity of Angola Science & Conservation: A Modern Synthesis*; Springer International Publishing: Cham, Switzerland, 2019. [CrossRef]
84. Miller, S.M.; Moeller, C.-H.; Harper, C.K.; Bloomer, P. Anthropogenic movement results in hybridisation in impala in southern Africa. *Conserv. Genet.* **2020**, *21*, 653–663. [CrossRef]
85. de Jong, J.F.; van Hooft, P.; Megens, H.-J.; Crooijmans, R.P.M.A.; de Groot, G.A.; Pemberton, J.M.; Huisman, J.; Bartoš, L.; Iacolina, L.; van Wieren, S.E.; et al. Fragmentation and Translocation Distort the Genetic Landscape of Ungulates: Red Deer in the Netherlands. *Front. Ecol. Evol.* **2020**, *8*, 535715. [CrossRef]
86. Heller, R.; Okello, J.B.A.; Siegismund, H. Can small wildlife conservancies maintain genetically stable populations of large mammals? Evidence for increased genetic drift in geographically restricted populations of Cape buffalo in East Africa. *Mol. Ecol.* **2010**, *19*, 1324–1334. [CrossRef] [PubMed]
87. Brockington, D.; Scholfield, K. Expenditure by conservation nongovernmental organizations in sub-Saharan Africa. *Conserv. Lett.* **2010**, *3*, 106–113. [CrossRef]
88. Seymour, R. Patterns of Subspecies Diversity in the Giraffe, *Giraffa camelopardalis* (L. 1758): Comparison of Systematic Methods and Their Implications for Conservation Policy. Ph.D. Thesis, University of Kent, Canterbury, UK, 2001.

89. Hassanin, A.; Ropiquet, A.; Gourmand, A.-L.; Chardonnet, B.; Rigoulet, J. Mitochondrial DNA variability in *Giraffa camelopardalis*: Consequences for taxonomy, phylogeography and conservation of giraffes in West and central Africa. *C. R. Biol.* **2007**, *330*, 265–274. [[CrossRef](#)] [[PubMed](#)]
90. Mitchell, G.; Skinner, J.D. On the origin, evolution and phylogeny of giraffes *Giraffa camelopardalis*. *Trans. R. Soc. S. Afr.* **2003**, *58*, 51–73. [[CrossRef](#)]
91. Groves, C. Giraffe taxonomy: Where are we now? *Giraffid* **2015**, *9*, 8–9.
92. Bercovitch, F.B.; Berry, P.S.M.; Dagg, A.; Deacon, F.; Doherty, J.B.; Lee, D.E.; Mineur, F.; Muller, Z.; Ogden, R.; Seymour, R.; et al. How many species of giraffe are there? *Curr. Biol.* **2017**, *27*, R123–R138. [[CrossRef](#)] [[PubMed](#)]
93. Fennessy, J.; Winter, S.; Reuss, F.; Kumar, V.; Nilsson, M.A.; Vamberger, M.; Fritz, U.; Janke, A. Response to “How many species of giraffe are there?”. *Curr. Biol.* **2017**, *27*, R123–R138. [[CrossRef](#)] [[PubMed](#)]
94. Bercovitch, F.B. Giraffe taxonomy, geographic distribution and conservation. *Afr. J. Ecol.* **2020**, *58*, 150–158. [[CrossRef](#)]
95. Bock, F.; Fennessy, J.; Bidon, T.; Tutching, A.; Marais, A.; Deacon, F.; Janke, A. Mitochondrial sequences reveal a clear separation between Angolan and South African giraffe along a cryptic rift valley. *BMC Evol. Biol.* **2014**, *14*, 219. [[CrossRef](#)]
96. Stanton, D.W.G.; Hart, J.; Galbusera, P.; Helsen, P.; Shephard, J.; Kümpel, N.F.; Wang, J.; Ewen, J.G.; Bruford, M.W. Distinct and diverse: Range-wide phylogeography reveals ancient lineages and high genetic variation in the endangered Okapi (*Okapia johnstoni*). *PLoS ONE* **2014**, *9*, e101081. [[CrossRef](#)]
97. Petzold, A.; Hassanin, A. A comparative approach for species delimitation based on multiple methods of multi-locus DNA sequence analysis: A case study of the genus *Giraffa* (Mammalia, Cetartiodactyla). *PLoS ONE* **2020**, *15*, e0217956. [[CrossRef](#)]
98. Petzold, A.; Magnant, A.-S.; Edderai, D.; Chardonnet, B.; Rigoulet, J.; Saint-Jalme, M.; Hassanin, A. First insights into past biodiversity of giraffes based on mitochondrial sequences from museum specimens. *Eur. J. Taxon.* **2020**, *703*, 1–33. [[CrossRef](#)]
99. Fennessy, J.; Bidon, T.; Reuss, F.; Kumar, V.; Elkan, P.; Nilsson, M.A.; Vamberger, M.; Fritz, U.; Janke, A. Multi-locus analyses reveal four giraffe species instead of one. *Curr. Biol.* **2016**, *26*, 2543–2549. [[CrossRef](#)] [[PubMed](#)]
100. Coimbra, R.T.F.; Winter, S.; Kumar, V.; Koepfli, K.-P.; Gooley, R.M.; Dobrynin, P.; Fennessy, J.; Janke, A. Whole-genome analysis of giraffe supports four distinct species. *Curr. Biol.* **2021**, *31*, 1–10. [[CrossRef](#)]
101. Winter, S.; Fennessy, J.; Janke, A. Limited introgression supports division of giraffe into four species. *Ecol. Evol.* **2018**, *8*, 10156–10165. [[CrossRef](#)]
102. Agaba, M.; Ishengoma, E.; Miller, W.; McGrath, B.C.; Hudson, C.N.; Bedoya Reina, O.C.; Ratan, A.; Burhans, R.; Chikhi, R.; Medvedev, P.; et al. Giraffe genome sequence reveals clues to its unique morphology and physiology. *Nat. Commun.* **2016**, *7*, 11519. [[CrossRef](#)]
103. Gatesy, J.; Geisler, J.H.; Chang, J.; Buell, C.; Berta, A.; Meredith, R.W.; Springer, M.S.; McGowen, M.R. A phylogenetic blueprint for a modern whale. *Mol. Phylogenet. Evol.* **2013**, *66*, 479–506. [[CrossRef](#)] [[PubMed](#)]
104. McGowen, M.R.; Gatesy, J.; Wildman, D.E. Molecular evolution tracks macroevolutionary transitions in Cetacea. *Trends Ecol. Evol.* **2014**, *29*, 336–346. [[CrossRef](#)] [[PubMed](#)]
105. Muller, Z.; Lee, D.E.; Scheijen, C.P.J.; Megan, K.L.; Strauss, M.K.L.; Kerry, D.; Carter, K.D.; Deacon, F. Giraffe translocations: A review and discussion of considerations. *Afr. J. Ecol.* **2020**, *58*, 159–171. [[CrossRef](#)]
106. Kostin, D.S.; Martynov, A.A.; Komarova, V.A.; Alexandrov, D.Y.; Yihune, M.; Kasso, M.; Bryja, J.; Lavrenchenko, L.A. Rodents of Choke Mountain and surrounding areas (Ethiopia): The Blue Nile gorge as a strong biogeographic barrier. *J. Vertebr. Biol.* **2020**, *69*, 20016. [[CrossRef](#)]
107. Angelici, F.M.; Colangelo, P.; Gippoliti, S. Out of Europe: Investigating *Hystrix cristata* (Rodentia: Hystricidae) skull morphometric geographic variability in Africa. *Biogeographia* **2021**, *36*, a001. [[CrossRef](#)]
108. Malyjurkova, L.; Hejzlarova, M.; Vymyslicka, P.J.; Brandlova, K. Social Preferences of Translocated Giraffes (*Giraffa camelopardalis giraffa*) in Senegal: Evidence for Friendship among Females? *Agric. Trop. Subtrop.* **2014**, *47*, 5–13. [[CrossRef](#)]
109. Brown, M.B.; Bolger, D.T.; Fennessy, J. All the eggs in one basket: A countrywide assessment of current and historical giraffe population distribution in Uganda. *Glob. Ecol. Conserv.* **2019**, *19*, e00612. [[CrossRef](#)]
110. Lydekker, R. On the Subspecies of *Giraffa camelopardalis*. *Proc. Zool. Soc. Lond.* **1904**, *74*, 202–227. [[CrossRef](#)]
111. Krumbiegel, I. *Die Giraffe: Unter Besonderer Berücksichtigung der Rassen; Monographien der Wildsäugetiere*; Verlag Dr. Paul Schöps: Leipzig, Germany, 1939; Volume VIII, pp. 1–98.
112. MacClintock, D.; Mochi, U. *A Natural History of Giraffes*; Charles Scribner’s Sons: New York, NY, USA, 1973.
113. Schouteden, H. Note sur la giraffe du Congo. *Rev. Zool. Afr.* **1913**, *2*, 134–137.
114. Gippoliti, S. Everything mammal conservation biologists always wanted to know about taxonomy (but were afraid to ask). *J. Nat. Conserv.* **2020**, *54*, 125793. [[CrossRef](#)]
115. Tedesco Zammarrano, V. *Fauna e Caccia*; Ministero delle Colonie: Rome, Italy, 1930.
116. Gippoliti, S.; Hagos, F.; Angelici, F.M. Eritrean ungulates in Italian museums as benchmark for taxonomy and conservation planning. *Hystrix* **2018**, *29* (XI ATIt Congress Suppl.), 30.
117. Gippoliti, S. On the Taxonomy of *Erythrocebus* with a re-evaluation of *Erythrocebus poliophaeus* (Reichenbach, 1862) from the Blue Nile Region of Sudan and Ethiopia. *Primate Conserv.* **2017**, *31*, 53–59.
118. Lydekker, R. Two undescribed giraffe. *Nature* **1911**, *87*, 484. [[CrossRef](#)]
119. Haltenorth, T. Klassifikation der Säugetiere: Artiodactyla I (18). In *Handbuch der Zoologie*; Walter de Gruyter & Co.: Berlin, Germany, 1963; Volume 8, pp. 1–167.

120. Spinage, C.A. *The Book of the Giraffe*; Collins: London, UK, 1968.
121. Ansell, W.F.H. Artiodactyla (Excluding the Genus *Gazella*). In *The Mammals of Africa—An Identification Manual for African Mammals*; Meester, J.A., Setzer, H.W., Eds.; Smithsonian Institution Press: Washington, DC, USA, 1971; pp. 1–84.
122. Dagg, A.I. *Giraffa camelopardalis*. *Mamm. Species* **1971**, *5*, 1–8. [[CrossRef](#)]
123. East, R. *African Antelope Database 1998*; IUCN/SSC Antelope Specialist Group IUCN: Gland, Switzerland; Cambridge, UK, 1999.
124. Skinner, J.D.; Mitchell, G. Family Giraffidae (Giraffe and Okapi). In *Handbook of the Mammals of the World*; Wilson, D.E., Mittermeier, R.A., Eds.; Lynx Edicions: Barcelona, Spain, 2011; Volume 2, pp. 788–802.
125. Ciofolo, C.; Le Pendu, Y. *Giraffa camelopardalis* Giraffe. In *Mammals of Africa*; Kingdon, J., Hoffmann, M., Eds.; Bloomsbury Publishing: London, UK, 2013; Volume VI, pp. 98–109.
126. Shorrocks, B.; Bates, W. *The Biology of African Savannas*, 2nd ed.; Oxford University Press: Oxford, UK, 2015.
127. Garnett, S.T.; Christidis, L. Taxonomy anarchy hampers conservation. *Nature* **2017**, *546*, 25–27. [[CrossRef](#)]
128. Robovský, J.; Melichar, L.; Gippoliti, S. Zoos and conservation in the Anthropocene: Opportunities and problems. In *Problematic Wildlife II*; Angelici, F.M., Rossi, L., Eds.; Springer: Cham, Switzerland, 2020; pp. 451–484. [[CrossRef](#)]
129. Groves, C.P.; Cotterill, F.P.D.; Gippoliti, S.; Robovský, J.; Roos, C.; Taylor, P.J.; Zinners, D. Species definitions and conservation: A review and case studies from African mammals. *Conserv. Genet.* **2017**, *18*, 1247–1256. [[CrossRef](#)]
130. Lorenzen, E.D.; Arctander, P.; Siegismund, H.R. High variation and very low differentiation in wide ranging plains zebra (*Equus quagga*): Insights from mtDNA and microsatellites. *Mol. Ecol.* **2008**, *17*, 2812–2824. [[CrossRef](#)] [[PubMed](#)]
131. Moritz, C. Defining ‘Evolutionarily Significant Units’ for Conservation. *Trends Ecol. Evol.* **1994**, *9*, 373–375. [[CrossRef](#)]
132. Minelli, A. The galaxy of the non-Linnaean nomenclature. *Hist. Philos. Life Sci.* **2019**, *41*, 31. [[CrossRef](#)]
133. Wood, C.C.; Gross, M.R. Elemental Conservation Units: Communicating extinction risk without dictating targets for protection. *Conserv. Biol.* **2008**, *22*, 36–47. [[CrossRef](#)]
134. Schlick-Steiner, B.C.; Steiner, F.M.; Seifert, B.; Stauffer, C.; Christian, E.; Crozier, R.H. Integrative taxonomy: A multisource approach to exploring biodiversity. *Annu. Rev. Entomol.* **2010**, *55*, 421–438. [[CrossRef](#)] [[PubMed](#)]
135. Robuchon, M.; Faith, D.P.; Julliard, R.; Leroy, B.; Pellens, R.; Robert, A.; Thévenin, C.; Véron, S.; Pavoine, S. Species splitting increases estimates of evolutionary history at risk. *Biol. Conserv.* **2019**, *235*, 27–35. [[CrossRef](#)]
136. Stanton, D.W.G.; Frandsen, P.; Waples, R.K.; Heller, R.; Russo, I.-R.; Orozco-terWengel, P.A.; Pedersen, C.E.T.; Siegismund, H.R.; Bruford, M.W. More grist for the mill? Species delimitation in the genomic era and its implications for conservation. *Conserv. Genet.* **2019**, *20*, 101–113. [[CrossRef](#)]
137. Groves, C. The genus *Cervus* in eastern Eurasia. *Eur. J. Wildl. Res.* **2006**, *52*, 14–22. [[CrossRef](#)]
138. Pedersen, C.E.T.; Albrechtsen, A.; Etter, P.D.; Johnson, E.A.; Orland, L.; Chikhi, L.; Siegismund, H.R.; Heller, R. A southern African origin and cryptic structure in the highly mobile plains zebra. *Nat. Ecol. Evol.* **2018**, *2*, 491–498. [[CrossRef](#)]
139. Liu, Y.-C.; Sun, X.; Driscoll, C.; Miquelle, D.G.; Xu, X.; Martelli, P.; Uphyrkina, O.; Smith, J.L.D.; O’Brien, S.J.O.; Luo, S.-J. Genome-Wide Evolutionary Analysis of Natural History and Adaptation in the World’s Tigers. *Curr. Biol.* **2018**, *28*, 1–10. [[CrossRef](#)]
140. Wilting, A.; Courtiol, A.; Christiansen, P.; Niedballa, J.; Scharf, A.K.; Orlando, L.; Balkenhol, N.; Hofer, H.; Kramer-Schadt, S.; Fickel, J.; et al. Planning tiger recovery: Understanding intraspecific variation for effective conservation. *Sci. Adv.* **2015**, *1*, e1400175. [[CrossRef](#)] [[PubMed](#)]
141. Gippoliti, S.; Cotterill, F.P.D.; Groves, C.P.; Zinner, D. Poor taxonomy and genetic rescue are possible co-agents of silent extinction and biogeographic homogenization among ungulate mammals. *Biogeographia* **2018**, *33*, 41–54. [[CrossRef](#)]
142. Dinets, V.; Hall, J. Mammalwatching: A new source of support for science and conservation. *Int. J. Biodivers. Conserv.* **2018**, *10*, 154–160. [[CrossRef](#)]
143. Lupták, P.; Csurma, L. The external variability and taxonomy of recent and extinct subspecies of lion (*Panthera leo*). *Gazella* **2009**, *36*, 33–150.
144. Zamudio, K.R.; Bell, R.C.; Mason, N.A. Phenotypes in phylogeography: Species’ traits, environmental variation, and vertebrate diversification. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 8041–8048. [[CrossRef](#)]
145. Moore, A.E.; Cotterill, F.P.D.; Winterbach, C.W.; Winterbach, H.E.K.; Antunes, A.; O’Brien, S.J. Genetic Evidence for Contrasting Wetland and Savannah Habitat Specializations in Different Populations of Lions (*Panthera leo*). *J. Hered.* **2016**, *107*, 101–103. [[CrossRef](#)]
146. Dures, S.G.; Carbone, C.; Savolainen, V.; Maude, G.; Gotelli, D. Ecology rather than people restrict gene flow in Okavango-Kalahari lions. *Anim. Conserv.* **2020**, *23*, 505–515. [[CrossRef](#)]
147. Shafer, A.B.A.; Wolf, J.B.W. Widespread evidence for incipient ecological speciation: A meta-analysis of isolation-by-ecology. *Ecol. Lett.* **2013**, *16*, 940–950. [[CrossRef](#)] [[PubMed](#)]
148. Bauer, H.; Chapron, G.; Nowell, K.; Henschel, P.; Funston, P.; Hunter, L.T.B.; Macdonald, D.W.; Packer, C. Lion (*Panthera leo*) populations are declining rapidly across Africa, except in intensively managed areas. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 14894–14899. [[CrossRef](#)]
149. Trinkel, M.; Angelici, F.M. The Decline in the lion population in Africa and possible mitigation measures. In *Problematic Wildlife: A Cross-Disciplinary Approach*; Angelici, F.M., Ed.; Springer: Cham, Switzerland, 2016; pp. 45–68.
150. Senn, H.; Banfield, L.; Wachter, T.; Newby, J.; Rabeil, T.; Kaden, J.; Kitchener, A.; Abaigar, T.; Luisa Silva, T.; Maunder, M.; et al. Splitting or lumping? A conservation dilemma exemplified by the critically endangered Dama Gazelle (*Nanger dama*). *PLoS ONE* **2014**, *9*, e98693. [[CrossRef](#)]

151. Schreiber, A.; Moreno, E.; Groves, C.; Robovský, J. Systematics and management units of Dama Gazelle *Nanger dama*. *Gnusletter* **2018**, *35*, 8–12.
152. Garnett, S.T.; Thomson, S.A. Are the implications for conservation of a major taxonomic revision of the world's birds' simply serendipity? *Anim. Conserv.* **2020**, *23*, 355–356. [[CrossRef](#)]
153. IUCN/SSC. *Guidelines for Reintroductions and Other Conservation Translocations: Version 1.0*; IUCN Species Survival Commission: Gland, Switzerland, 2013.
154. Kerley, G.I.H.; Kowalczyk, R.; Cromsigt, J.P.G.M. Conservation implications of the refuge species concept and the European bison: King of the forest or refuge in a marginal habitat? *Ecography* **2012**, *35*, 519–529. [[CrossRef](#)]
155. Ali, A.H.; Amin, R.; Evans, J.S.; Fischer, M.; Ford, A.T.; Kibara, A.; Goheen, J.R. Evaluating support for rangeland-restoration practices by rural Somalis: An unlikely win-win for local livelihoods and hirola antelope? *Anim. Conserv.* **2018**, *22*, 144–156. [[CrossRef](#)]
156. O'Connor, D.; Stacy-Dawes, J.; Muneza, A.; Fennessy, J.; Gobush, K.; Chase, M.J.; Brown, M.B.; Bracis, C.; Elkan, P.; Rabeil, T.; et al. Updated geographic range maps for giraffe, *Giraffa* spp., throughout sub-Saharan Africa, and implications of changing distributions for conservation. *Mamm. Rev.* **2019**, *49*, 285–299. [[CrossRef](#)]
157. Fleming, P.; Hofmeyr, S.; Nicolson, S.; Du Toit, J. Are giraffes pollinators or flower predators of *Acacia nigrescens* in Kruger National Park, South Africa? *J. Trop. Ecol.* **2006**, *22*, 247–253. [[CrossRef](#)]