End-to-End Continuous Small-Scale Drug Substance Manufacturing: From a Continuous In Situ Nucleator to Free-Flowing Crystalline Particles

Anne Cathrine Kufner †, Marc Meier † and Kerstin Wohlgemuth *

1. Introduction

Crystallization is of particular importance in the pharmaceutical industry, being involved in about 90% of the manufacturing processes of drug substances or active pharmaceutical ingredients (APIs) [1]. APIs are commonly obtained as crystalline solids at the end of the downstream process. Therefore, APIs are mainly highly specific and, thus, chemically complex, resulting in low bioavailability [2]. Accordingly, the specific adjustment of particle specifications, such as particle size, is crucial concerning the dissolution behavior of APIs. Crystallization as a separation method highly influences the critical quality attributes (CQAs) of the product particles, such as the particle size distribution (PSD), product purity, and particle morphology [3–5]. The crystallization process starts from a prepurified solution during workup after drug substance synthesis. In the crystallization step, the target component is isolated as a solid phase from a liquid phase. The subsequent solid–liquid separation, washing, and drying of the particles complete the crystal process chain (CPC) and aim to preserve the PSD, achieving a high product purity and low residual moisture [6].
In particular, the processes within the CPC during drug substance manufacturing are still predominantly operated in batch mode. However, a paradigm shift towards continuous processes is currently taking place, as these provide numerous benefits. These include the elimination of batch-to-batch variability, increased space–time yield, and simplified scale-up by, for example, numbering-up due to reduced plant volumes. This enables the use of research and development equipment for industrial production just by increasing the runtime, resulting in a reduced time to market [7,8].

Developing an end-to-end continuous manufacturing process for a small-scale API production of 250–1000 kg a⁻¹, from the raw material treatment to the final dosage form [9], requires, especially for pharmaceutical production, the realization of continuous upstream, crystallization, product isolation, and final formulation processes, ensuring the continuous production of on-spec products [4]. This study focuses on the continuous CPC from continuous crystallization to continuous product isolation. A major challenge is the connection of the respective unit operations for continuous crystallization and product isolation [9,10]. In our previous study, the successful coupling of continuous crystallization with a slug flow crystallizer (SFC) and continuous product isolation with a continuous vacuum screw filter (CVSF) was demonstrated [11]. However, continuous seeding of the SFC was required, which is a bottleneck of the continuous CPC. Accordingly, the decoupling of the nucleation and crystal growth processes into separate units eliminates the bottleneck by providing independent control of both processes, leading to improved product quality control, as well as an increased process efficiency [10]. With regard to continuous crystal growth, various concepts have already been elaborated, which are classified into two classes: mixed-suspension mixed-product removal (MSMPR) and tubular plug flow crystallizers (PFCs) [4,5,8,9]. In this work, the SFC was used as a continuous crystal growth zone. The SFC belongs to the class of tubular PFCs. The concept of the SFC is based on a segmented flow of two immiscible fluids, resulting in a narrow and equal residence time distribution (RTD) of the solid and liquid phases despite laminar flow conditions. Therefore, backmixing effects can be avoided, and a reproducible narrow PSD at the end of the process is achieved [12]. In this work, synthetic air is used to segment the product stream, enabling an effective separation of the segmenting fluid and a high product purity in the subsequent downstream process. The wall friction is responsible for the generation of internal vortices in the slugs, resulting in an effective and gentle suspension of the particles without the use of external equipment. In terms of the specific adjustment of the desired particle size, the SFC offers various degrees of freedom. Based on the kinematically limited crystal growth, the residence time (RT) can be regulated by the tubing length and the volume flow rate. In addition, the driving force for crystal growth in the SFC is realized by cooling via a tube-in-tube system along the SFC, whereby the crystal growth can specifically be influenced by the cooling rate. More detailed information concerning the specific setup of the used SFC can be found in the literature [9,10,13–15].

Commonly, the apparatuses for continuous crystal growth processes require the supply of seed crystals [16]. However, the addition of seed crystals in a crystallization process could lead to process contamination, which must be avoided, particularly in the pharmaceutical industry [15]. Moreover, the particle properties of the seed crystals are crucial for the final product quality, resulting in high requirements for the seed crystal preparation process and, thus, a high time investment [15,17]. Furthermore, long-term operation cannot be realized due to the continuous supply of seed crystals. Accordingly, the demand for a continuous in situ nucleator in the context of an end-to-end continuous CPC is high. In this study, the design of a continuous in situ nucleator for coupling to various types of continuous crystallizers is presented. There are various concepts available for the realization of continuous nucleation. These include gassing-induced nucleation [18], sonocrystallization [19], an impinging jet [20], and antisolvent nucleation. The selected nucleation method for the designed nucleator in this work is antisolvent nucleation, which is based on lowering the solubility
of a solute via the addition of an antisolvent [17,21]. This method requires a low energy input and is suitable for thermally sensitive substances [21]. Additionally, the precise adjustment of the degree of supersaturation is possible via the mass of the added antisolvent independent of the temperature. Various technical concepts ensuring a continuous, effective, and rapid mixing process of two liquid streams, as fundamental requirements of an in situ nucleator based on antisolvent nucleation or precipitation, are presented in the literature. A microporous tube-in-tube microchannel reactor [22], a spinning disc reactor [23], and high-gravity antisolvent precipitation [24] were investigated for the antisolvent precipitation of nanoparticles of poorly water-soluble substances. Membrane-assisted antisolvent crystallization as antisolvent nucleation technology was tested for the substance system erythritol/water with ethanol as an antisolvent by Li et al. [25]. Engler et al. [26] investigated T-shaped convective passive micromixers with respect to mixing quality and mixing time. These T-mixers were considered in the study of Engler et al. [26] for application as microreactors for chemical reactions. The aim was to realize an efficient mixing process with short mixing times to ensure that the mixing process was completed before the reaction started. This objective can also be applied to the process of antisolvent nucleation, where the mixing time should be shorter than the induction time. A T-shaped convective passive mixer offers rapid and effective diffusive micromixing with the support of convective mixing under laminar conditions [26]. Moreover, a T-shaped micromixer provides a higher mixing efficiency and smaller dead zones than conventional Y-shaped micromixers [27]. Since the concepts of antisolvent precipitation or nucleation presented above are very challenging due to the complex relationships between process and design parameters, a passive T-shaped micromixer was selected as the most promising concept for the design of a continuous in situ nucleator in this study. Accordingly, a T-mixer represents a simple, robust, compact, and reliable unit without moving components and an external energy input, keeping the investment, operating, and maintenance costs comparatively low. In the context of an end-to-end continuous small-scale drug substance manufacturing process, starting from continuous nucleation in the nucleator to crystal growth in the SFC, continuous product isolation is required to obtain free-flowing particles at the end of the process.

Continuous product isolation, including solid–liquid separation, washing, and drying, is essential in terms of maintaining CQAs. The patented continuous vacuum screw filter (CVSF) combines the process steps of filtration, washing, and drying in a modular design [28]. Due to the modular setup of the CVSF, different configurations of the filtration, washing, and drying modules can be realized, resulting in a high flexibility of the apparatus regarding various applications and operating points [6,29]. The respective modules are tubular glass bodies, which are connected via flanges. The suspension from the preceding crystallization process is fed into the first module, where the filtration step takes place using an applied vacuum and a filter frit incorporated at the bottom of the glass body. The formed filter cake is transported axially through the apparatus by a rotating screw into subsequent modules for further process steps like washing and drying. Fundamental requirements for continuous product isolation are the preservation of the CQAs designed in the preceding crystallization process and the production of free-flowing particles at the end of the process [6]. In previous studies, for bipyramidal L-alanine particles in an aqueous solution and a CVSF configuration consisting of filtration and two-stage washing for suspension volume flow rates of up to 84 mL min⁻¹ and a solid loading of 6 wt.-%, we demonstrated that the targeted residual moisture of less than 1%, the preservation of PSD, and a narrow RTD of the solid phase are ensured [6,29]. Furthermore, the coupling of the SFC as a continuous crystallizer with the CVSF for continuous product isolation was successfully proven [11].

The coupling of various continuous unit operations requires an extensive characterization of the individual apparatuses in order to meet the high requirements of a reproducible high product quality. The characterization of an apparatus includes the understanding of the relationships between the input and output parameters and is
essential to ensure the compatibility of different apparatuses (nucleator, crystal growth zone, and product isolation) in the context of a continuous integrated process [5,8]. The definition of CQAs is decisive for the quality-by-design approach in order to evaluate the influence of the critical process parameters (CPPs) on the integrated process performance [8].

The design of the nucleator needs to meet the requirements for compatibility with the respective crystallizer used. Accordingly, the nucleator must generate a sufficiently high number of nuclei to prevent fouling in the growth zone and to achieve the highest possible yield without the occurrence of secondary nucleation. We postulate the following hypotheses:

1. In the passive T-mixer, a sufficiently rapid micromixing process of the feed and antisolvent streams is ensured for equal inlet stream velocities.
2. As an antisolvent for continuous nucleation, the same solvent used for particle washing within the crystal process chain is suitable.
3. The continuously generated nuclei or the solid loading in the continuous nucleator can be estimated from the mixing point in a ternary solubility diagram.

Our work is divided into the following three parts: (1) the design and characterization of a continuous in situ nucleator for coupling with various continuous crystallizers, answering the three above mentioned hypotheses; (2) continuous particle design considering nucleation and crystal growth by coupling the nucleator with a slug flow crystallizer (SFC) as the crystal growth zone for particles with different aspect ratios, ranging from nearly spherical to needle-shaped particles; and (3) the demonstration of a fully continuous end-to-end small-scale manufacturing process for crystalline particles consisting of the nucleator, the SFC as the crystal growth zone, and the CVSF for product isolation.

2. Materials and Analytics

2.1. Substances

The model substance used as a target molecule in this study was L-alanine, since it provides similar properties to high-value APIs but is easier to handle in the laboratory and much cheaper. L-alanine was sourced with a purity of $\geq 99.7\%$ from Evonik Industries AG (Essen, Germany). Ultrapure, deionized, and bacteria-free water acted as the main solvent and was provided by the purification system Milli-Q® from Merck KGaA (Darmstadt, Germany). The binary saturation concentration of L-alanine $c_{\text{als}}$ in water depending on the respective temperature $\vartheta^*$ was determined and regressed by Wohlgemuth et al. [30] (Equation (1)):

$$c_{\text{als}}(\vartheta^*) [g \, g_{\text{solution}}^{-1}] = 0.11238 \cdot \exp (9.0849 \cdot 10^{-3} \cdot \vartheta^*[\text{°C}])$$ (1)

Ethanol absolute was used as an antisolvent for nucleation and as a wash liquid during product isolation. It was purchased from VWR Chemicals (Darmstadt, Germany) with a purity of $\geq 99.5\%$.

L-glutamic acid was used as a habit modifier during L-alanine crystallization. With the aid of the addition of L-glutamic acid to a clear solution, various particle shapes of L-alanine, depending on the concentration of L-glutamic acid, can be realized. L-glutamic acid was purchased from Alfa Aesar (Kandel, Germany) with a purity of $\geq 99\%$. All substances were used within this study without any pretreatment.

2.2. Analytics

The characterization of the nucleator requires qualitative and quantitative investigations of the nuclei generated in situ. Therefore, the output of the nucleator was analyzed using light microscopy images, gravimetric concentration measurements, and a dynamic image analysis (DIA) at specific RTs.
The light microscopy images were used for the qualitative evaluation of particles in terms of size and shape. In this work, suspension samples were placed on a Petri dish and examined directly by utilizing a light microscope (DM2700 M from Leica, Wetzlar, Germany) with an equipped camera (FLEXACAM C1 from Leica, Wetzlar, Germany).

The gravimetric concentration measurements represent a loss-on-drying method and were used for the measurement of the liquid l-alanine concentration to determine the solid loading, \( w_{\text{solid}} \), of the suspension after the use of the continuous nucleator. The suspension samples were filtered using a syringe top filter (CHROMAFIL® Xtra PTFE-45/25, 0.45 µm from Macherey-Nagel GmbH & Co. KG, Düren, Germany) and collected in polypropylene (PP) vials to ensure a particle-free solution. After an initial pre-drying step for 24 h in a drying oven at 50 °C, a subsequent intensive drying process in a vacuum dryer for 48 h at 50 °C was carried out. Corresponding to the procedure described in our previous work [6], the solute concentration was calculated. Furthermore, \( w_{\text{solid}} \) was determined by the difference between the l-alanine concentration of a suspension sample without and with filtration, according to the procedure described above. Moreover, the relative yield (\( Y_{\text{rel}} \)) of the crystallization process was determined based on gravimetric concentration measurements of l-alanine in the liquid phase (\( c_{\text{Ala}} \)) and the theoretical saturation concentration at a specific temperature (\( c^{\ast}_{\text{Ala}} \)) according to Equation (2):

\[
Y_{\text{rel}} [\%] = \left(1 - \frac{c_{\text{Ala}} - c^{\ast}_{\text{Ala}}}{c^{\ast}_{\text{Ala}}} \right) \cdot 100\%
\]  

The DIA was conducted with a QICPIC image analysis sensor from Sympatec GmbH (Clausthal-Zellerfeld, Germany) and a LIXELL dispersion module to evaluate the particle size, distribution, and shape quantitatively. For a reliable assessment of the developed nucleator, the analysis of small particle sizes had to be ensured. For this reason, the M4 module was used, enabling a reliable detection of particles in a size range from 9.9 µm to 750 µm [31]. The measured projection areas were analyzed using an in-house Matlab script [32,33] with respect to the PSD. Isotropic particles, such as nearly spherical bipyramidal particles, were evaluated based on the equivalent diameter. However, anisotropic particles, such as needles, were characterized by the minor axis length (MIL) and major axis length (MAL) of an ellipse fitted to the respective particle projection area, enabling the calculation of an axis length distribution (ALD), which is strongly coupled to the PSD [34–36]. A density plot illustrates the probability distribution of anisotropic particles with a certain MIL and MAL. The focus point of the density plot represents the most probable MIL (MIL\( _{\text{ch}} \)) and MAL (MAL\( _{\text{ch}} \)), serving as characteristic parameters of the PSD of anisotropic particles. In addition, the aspect ratio (AR) was calculated via the quotient of MAL\( _{\text{ch}} \) to MIL\( _{\text{ch}} \) for further particle characterization. Detailed information concerning the DIA of anisotropic particles can be found in our previous study [37].

3. Design and Characterization of the Continuous Nucleator

In this section, the design of the continuous in situ nucleator based on antisolvent nucleation for various types of crystallizers is presented. The model substance system used in this study was l-alanine/water. Since ethanol was identified as a suitable wash liquid for particle washing, ethanol was applied as an antisolvent [38].

3.1. Design of the Continuous Nucleator

A convective passive T-mixer as a continuous in situ nucleator must ensure that mixing times (micromixing) are shorter than induction times to prevent local supersaturation and guarantee consistent high product quality for long-term continuous operation [15,17]. Engler et al. [26] describe in their study that, in a T-mixer with a 180° inlet configuration, the impingement process of the inlet streams that flow into each other can be classified into three different laminar flow regimes based on the Reynolds number in
the mixing channel \( (Re_M, \text{Equation (3)}) \): stratified flow \((0 < Re_M < 50)\), vortex flow \((50 < Re_M < 150)\), and engulfment flow \((150 < Re_M < 300)\).

\[
Re_M = \frac{\rho_{\text{mix}} \cdot v_M \cdot L}{\eta_{\text{mix}}}
\]  \hspace{1cm} (3)

\[
\rho_{\text{mix}} = \frac{1}{(w_{i,\text{mix}} \cdot \rho_i) + (w_{j,\text{mix}} \cdot \rho_j) + (w_{k,\text{mix}} \cdot \rho_k)}
\]  \hspace{1cm} (4)

\[
\eta_{\text{mix}} = \frac{1}{(w_{i,\text{mix}} \cdot \eta_i) + (w_{j,\text{mix}} \cdot \eta_j) + (w_{k,\text{mix}} \cdot \eta_k)}
\]  \hspace{1cm} (5)

\( Re_M \) is composed of the product of the density of the mixture \( \rho_{\text{mix}} \) (Equation (4)), the velocity in the mixing channel \( v_M \), and the characteristic length \( L \) divided by the dynamic viscosity of the mixture \( \eta_{\text{mix}} \) (Equation (5)). The characteristic length \( L \) of a circular cross-section is equal to the inner diameter (ID).

Flow patterns have a massive influence on the mixing quality and mixing time. Accordingly, the impingement of the inlet streams that flow into each other is increased with increasing \( Re_M \), generating intensive mixing in the impingement zone to achieve shorter mixing times. For an efficient mixing process supported by convection, it is essential to generate engulfment flow in a T-mixer. Depending on the design and process parameters, the transition from vortex flow to engulfment flow is achieved at different \( Re_M \) values, indicated by the corresponding turnover Reynolds number in the mixing channel \( (Re_{M,T0}) \). Based on this, the highest possible velocities of the inlet streams must be realized in order to maximize \( Re_M \). An increase in the velocities of the inlet streams can be accomplished by increasing the volume flow rate or reducing the cross-sectional area. Since in practical application, the feed volume flow rate is set through a defined product capacity and cannot be varied freely, an increase in the velocity of the inlet streams was achieved by minimizing the cross-sectional area, here, the inner diameter of the T-mixer. For a worst-case estimation, the minimum volume flow rate was set to the often applied value of 10 mL min\(^{-1}\) [11,12,39,40].

The study of ENGLER ET AL. [26] shows that, for a cross-sectional area ratio \( (\Phi, \text{Equation (6)}) \) between the mixing channel \( (A_M) \) and the inlet channels \( (A_{\text{Inlet}}) \) of \( \Phi = 2 \), \( Re_{M,T0} \) is approximately 150. This represents the minimum \( Re_M \) in the mixing channel for the design of a T-mixer with a 180° inlet configuration in order to create engulfment flow and, thus, realize short mixing times.

\[
\Phi = \frac{A_M}{A_{\text{Inlet}}} = \frac{d_{i,M}^2}{d_{i,\text{feed}}^2}
\]  \hspace{1cm} (6)

Moreover, design parameters for a convective passive T-mixer (180° inlets) were defined. A fundamental condition is the equality of feed and antisolvent inlet velocities to avoid backmixing. Additionally, the inlet volume flow rates were equalized to provide identical inlet diameters and, thus, equal front faces of the impinging inlet streams in the T-mixer configured with a 180° inlet. This is a crucial factor for the mixing process and ensured the validity of the correlations from Engler et al. [26]. Consequently, the antisolvent volume flow rate was aligned to the set feed volume flow rate. Considering \( \Phi = 2 \), an inlet ID \( (d_{i,\text{feed}}) \) of 0.75 mm (lower manufacturing limit) resulted in an ID of the mixing channel \( (d_{i,M}) \) of 1.06 mm (Figure 1).
Batch cooling crystallization commonly focuses on seed loading in a range of 0.1–1.5 wt.-%, with seed crystals in a size range from 40 to 330 µm at relatively low cooling rates (e.g., $w_{solid}$ of 1.5 wt.-% for a cooling rate of 0.15 K min$^{-1}$) [38–40]. Since continuous crystallization aims to realize high cooling rates (e.g., 1.5 K min$^{-1}$) in order to achieve high space–time yields, a higher surface area provided by nuclei is required for efficient crystal growth. Therefore, as a further restriction in the context of continuous cooling crystallization, a $w_{solid}$ of at least 1 wt.-% was targeted in the design of the T-mixer as a continuous nucleator. In this study, the composition of the antisolvent mixture used in the T-mixer as a nucleator was determined graphically in corresponding ternary solubility diagrams. Considering that the volume flow rates of the feed ($Q_{Feed}$) and the antisolvent mixture ($Q_{AS}$) were set to the same value so that both streams met at the same velocity, the distances for the feed ($\dot{m}_{Feed}$) and antisolvent mixture ($\dot{m}_{AS}$) mass flow rates were calculated using Equations (7–9) to determine the mixing point (MP) graphically.

$$\frac{Q_{Feed}}{Q_{AS}} = 1$$  \hspace{1cm} (7)

$$\frac{\dot{m}_{Feed}}{\dot{m}_{AS}} = \frac{\rho_{Feed}}{\rho_{AS}}$$  \hspace{1cm} (8)

$$\dot{m}_{Feed} + \dot{m}_{AS} = \dot{m}_{tot}$$  \hspace{1cm} (9)

![Figure 1](image1.png)

**Figure 1.** Design of the T-mixer with 180° inlets as continuous nucleator: (left) illustration of a cross-section created using the software Fusion 360 (Version 2.0.17457) with ID dimensions; (right) photo of the 3D printed T-mixer.

![Figure 2a](image2a.png)

**Figure 2a.** Ternary solubility diagrams of L-alanine/water/ethanol at 20 °C and 1 bar. The solid blue line represents the solubility curve based on the experimental data from AN ET AL. [41]. The solid green line describes the underlying conode for nucleation, the crystallization process until thermodynamic equilibrium is reached. Additionally, the orange dot ($w_{EGH,AS}$) indicates the
composition of the liquid phase in the equilibrium state. Furthermore, the dashed red line connects the feed solution (F, blue triangle) and antisolvent mixture (AS, red triangle, \( w_{\text{ETHOHmix}} \)) to be mixed, and the intersection between the dashed red line and the green line defines the mixing point (MP, black dot). Panel (a) shows the calculation of the ratio of the feed and antisolvent mass flow rate based on the location of the MP on the conode by applying the lever rule (Equations (7)–(9)). Panel (b) displays the calculation of \( w_{\text{solid}} \) based on the location of the MP on the conode by using the lever rule (Equation (10)). Different operating points for the T-mixer with 180° inlets based on graphical construction are shown in Table S1.

The total mass flow rate \( \dot{m}_{\text{tot}} \) in Equation (9) represents the total length of the dashed red line. The densities of the feed \( (\rho_{\text{Feed}}) \) and antisolvent mixture \( (\rho_{\text{AS}}) \) solutions are calculated using Equation (4) according to the corresponding compositions. Generally, based on Equations (7)–(9), different MPs are obtained for different antisolvent mixture compositions, which simultaneously results in different \( w_{\text{solid}} \) values. The latter can be calculated using the ratio of the solid mass flow rate (\( \dot{m}_{\text{solid}} \)) to the mass flow rate of the solution (\( \dot{m}_{\text{solution}} \)) according to Equation (10), which can be graphically determined by applying the lever rule illustrated in Figure 2b. In the example shown, this results in a \( w_{\text{solid}} \) of approximately 5 wt.-%.

\[
\dot{m}_{\text{solution}} = \frac{\dot{m}_{\text{solid}}}{\dot{m}_{\text{solution}}} \quad (10)
\]

According to the lever rule, the ternary solubility diagram indicates that the dilution of ethanol as an antisolvent with water is required so that an MP results, where \( w_{\text{solid}} \) is reduced to 1 wt.-%, as given in Table S2 in the Supporting Information.

For the characterization experiments, a \( w_{\text{ETHOH,AS}} \) of 50 wt.-% was chosen as an example, resulting in a \( w_{\text{ETHOH,AS}} \) of 23 wt.-%, a \( w_{\text{solid}} \) of 2.26 wt.-%, and an \( R_M \) of 362 according to the graphical analysis. This operating point was promising concerning the generation of a sufficient quantity of nuclei and an effective micromixing process.

### 3.2. Manufacturing of a Continuous Nucleator

In this study, 3D printing was used as a manufacturing method for the nucleator. Considering the low flexibility of the T-mixer with regard to the changing of process parameters, in-house 3D printing enabled the fast, precise, and robust manufacturing of differently configured T-mixers. In particular, masked stereolithography printing provided relatively fast and precise small-scale manufacturing using a temperature-resistant resin (High Temp V2) purchased from Formlabs (Somerville, MA, USA). With an adjusted layer height of 50 µm, the printer (Type Saturn from Elegoo, Shenzhen, China) offers a high accuracy and high surface quality with a low material consumption and a relatively high printing speed. The lowest manufacturing limit for circular channels is a diameter of 0.75 mm.

### 3.3. Experimental Setups and Procedures

Characterization experiments were conducted at ambient temperature \( (\theta_{\text{ambient}} \approx 20 \, ^\circ\text{C}) \) to exclude the influence of potential heat losses on the performance of the T-mixer as a continuous nucleator and obtain reliable data. In order to evaluate the degradation of the induced supersaturation via the addition of the antisolvent in the T-mixer, the formed nuclei were quantified at different RTs (6 s, 5 min, 10 min, and 15 min) after the mixing process. The experimental setups are depicted in Figure 3.
Figure 3. Schematic of experimental setup for the characterization of the T-mixer with 180° inlet configuration as continuous nucleator. (a) Experimental setup for the analysis of generated nuclei after a short RT of 6 s provided by the periphery. (b) Extended setup by a batch vessel, enabling the adjustment of specific RTs (5 min, 10 min, and 15 min).

The feed solution and antisolvent mixture were provided in Schott glass bottles ($V = 0.5$ L), whereby the saturated feed solution was prepared for a saturation temperature $\theta^* = 20^\circ$C in accordance with Equation (1). Each solution was conveyed using a peristaltic pump (REGLO Digital MS-4/12, 12 rollers from ISMATEC Industry Solutions GmbH, Grevenbroich, Germany). The used pump tubes (TYGON LMT-55 from IDEX Health & Science GmbH Wertheim-Mondfeld, Germany) had an ID of 3.17 mm or 1.75 mm with a wall thickness of 0.9 mm or 0.75 mm, respectively. Considering the scale of the volume flow rates, tubes with an ID of 1.75 mm were used for precise pumping performance at low volume flow rates. As depicted in Figure 3a, the outlet of the T-mixer was directly connected to further tubing for analytics. The volume flow rate of the outlet stream of the T-mixer and the tubing length (47.5 cm) caused a $n_{RT}$ of approximately 6 s until samples were withdrawn and analytics applied. Since the nucleation and growth kinetics were unknown, experiments were performed to investigate the supersaturation degradation as a function of RT qualitatively. Therefore, approximately 30 mL of the product stream was collected in a stirred batch vessel after the mixing process in the T-mixer (Figure 3b). Subsequently, analyses were performed after each specific RT (5 min, 10 min, and 15 min).

3.4. Results

The results of the in-line DIA showed that a very low number of projection areas of particles were detected. Since the lower detection limit of the DIA is 9.9 µm and the critical nuclei size is unknown, the in-line DIA is not valid for the analysis of instantly formed nuclei smaller than 9.9 µm.

The microscopic analysis of the outlet stream of the T-mixer 20 s after sampling (Figure 4b) indicated that considerably more nuclei were visible than in the in-line DIA (Figure 4a). Furthermore, the microscope image of the same sample 60 s after sampling (Figure 4c) revealed a significantly increased number of nuclei qualitatively.
Figure 4. Panel (a) displays a screenshot from the in-line DIA. Additionally, light microscopic images of the same sample after an RT of approximately 6 s provided by the periphery taken 20 s (b) and 60 s (c) after sampling are shown.

It was assumed that the induced supersaturation was not completely degraded instantly via nucleation after an RT of approximately 6 s provided by the periphery due to the low number of nuclei detected by the in-line DIA (Figure 4a), whereas the microscopic images depict a comparatively high particle density. Moreover, the microscopic images (Figure 4b,c) indicate anisotropic nuclei, which could be confirmed after 5 min, 10 min, and 15 min RT. The experiments indicated increasing $w_{\text{solid}}$ of needles with increasing RT due to nucleation and subsequent crystal growth, as shown for 15 min RT in Figure 5a,b. Thus, a $w_{\text{solid}}$ of 0.17 wt.-% after 5 min RT, a $w_{\text{solid}}$ of 0.26 wt.-% after 10 min RT, and a $w_{\text{solid}}$ of 0.60 wt.-% after 15 min RT were determined. Consequently, it could not be assumed that the induced supersaturation was completely degraded after 15 min since, based on thermodynamics, a $w_{\text{solid}}$ of 2.26 wt.-% should result. Since the particles were anisotropic, the PSD was determined using the ALD approach, as described in Section 2.2. Therefore, MIL$_{ch}$, MAL$_{ch}$, and the resulting AR served as characterization parameters for the PSD of the anisotropic particles obtained in this experiment (Figure 5c).

Figure 5. Light microscopic images of the same sample after an RT of 15 min in the batch vessel 20 s (a) and 60 s (b) after sampling. (c) ALD of anisotropic particles after 15 min inside the batch vessel. The data points are plotted as functions of MIL, MAL, AR, and measurement probability. The red star indicates the centroid of the data points and, thus, the most probable projection area to be measured, characterized by MIL$_{ch}$, MAL$_{ch}$, and AR.

Furthermore, the T-mixer was operated for 90 min. Within this time, no encrustations occurred, and the operating performance was not restricted. Thus, operating times beyond 90 min could be realized, proving that the T-mixer is suitable for long-term operation.
In conclusion, the T-mixer with 180° inlets was considered a potentially suitable long-term operating continuous in situ nucleator based on antisolvent nucleation for coupling with different types of crystallizers. Since thermodynamic equilibrium was not instantly reached in the continuous in situ nucleator, a \( w_{\text{solid}} \) of approximately 1 wt.-% was not instantly generated for subsequent crystal growth. Accordingly, a higher theoretical \( w_{\text{solid}} \) could be targeted in the graphical design of the nucleation process to realize an instantaneously real \( w_{\text{solid}} \) of about 1 wt.-% (Table S1). Nevertheless, the nuclei formed after an RT of 6 s, being below the lower detection limit of the DIA, could provide a sufficient surface area for efficient crystal growth in the subsequent crystallizer due to the high surface-to-volume ratio. Therefore, a higher \( w_{\text{solid}} \) would be required compared to the smaller nuclei generated in situ to provide the same surface area for the subsequent crystal growth. Moreover, the compatibility of the in situ nucleator with a crystallizer could be ensured despite the residual supersaturation by appropriately adjusting the process parameters of the crystallizer. In order to prove the hypothesis that small nuclei at a \( w_{\text{solid}} \) below 1 wt.-% provide a sufficient surface area for crystal growth in the subsequent continuous crystallizer and that the compatibility of the continuous in situ nucleator with a continuous crystallizer can be ensured despite the residual supersaturation of the nucleator, the T-mixer as a continuous in situ nucleator was tested via coupling to the SFC as a growth zone.

4. Continuous Particle Design Considering Nucleation and Crystal Growth

In this section, the adaption of the nucleator for coupling with an SFC is shown and validated following the procedure described in Section 3. Furthermore, continuous cooling crystallization experiments with the nucleator coupled with the SFC as a growth zone were carried out for differently shaped particles to show its broad applicability.

4.1. Adaption of the Nucleator for Coupling with a Slug Flow Crystallizer

The coupling of the continuous nucleator to the SFC requires the consideration of the mass fraction of ethanol as an antisolvent in the mixture since it affects the slug flow stability and, thus, the performance of the SFC. High slug stability is aimed at avoiding the migration of particles to neighboring slugs and, thus generating a narrow RTD of the liquid and solid phases due to the characteristic pattern of alternating slug flow [12,42]. Furthermore, a high slug stability provides a narrow slug length distribution (SLD), ensuring uniform growth conditions in the slugs and, thus, a reproducible high product quality. A high slug stability is characterized by convex slugs, requiring the absence of a wall film, the so-called dry pattern. The wall film formation depends on the dynamic three-phase contact angle (\( \theta_{\text{dyn}} \)) between the tubing material and the gas and the liquid phases. In this context, the interfacial tension between the gas and liquid phases (\( \sigma_{\text{G/L}} \)) is crucial. For the generation of nuclei in the continuous nucleator, ethanol (antisolvent) is added to the aqueous L-alanine solution, which increases the ethanol concentration in the mixture and decreases \( \sigma_{\text{G/L}} \) and \( \theta_{\text{dyn}} \). Using the experimental strategy from our previous publication, it is possible to identify a well-suited operating range of the SFC for the respective substance system to apply continuous cooling crystallization [43]. From this, the ethanol concentration in the mixture should not exceed a \( w_{\text{ETHOH, mix}} \) of 10 wt.-% for a total volume flow rate of 40 mL min\(^{-1}\) at 20 °C for the applied SFC setup.

In the graphical construction of the nucleation process in the T-mixer with a 180° inlet configuration, it is evident that, considering the restrictions of the equal inlet volume flow rates of the feed and antisolvent, for a maximum \( w_{\text{ETHOH, mix}} \) of 10 wt.-%, as well as a generated \( w_{\text{solid}} \) of at least 1 wt.-%, a high dilution results. Due to the high dilution, the MP is located outside the miscibility gap, and no supersaturation is generated for nucleation (Figure S1). For this reason, it is not possible to operate the T-mixer with 180° inlets with equal inlet volume flow rates and guarantee a maximum \( w_{\text{ETHOH, mix}} \) of 10 wt.-%, as well as a \( w_{\text{solid}} \) of at least 1 wt.-%. Therefore, the nucleator concept is adapted for a
T-mixer with 90° inlets in order to realize equal inlet velocities for the feed solution and antisolvent mixture. Accordingly, the ID dimensions of the feed and antisolvent inlets are modified, enabling equal velocities at the MP and avoiding backmixing during the mixing process.

Furthermore, regarding the crystallization experiments and the coupling of the T-mixer with the SFC, the design of the nucleator should be constructed for use at \( \vartheta = 50 \, ^\circ\text{C} \). Therefore, as a further restriction, it must be taken into account that the classification of a stable operation zone for ensuring the favored slug formation mechanism and a stable slug flow at a total volume flow rate of 40 mL min\(^{-1}\) and a \( w_{\text{ETH, mix}} \) of 10 wt.-% is only valid for \( \vartheta = 20 \, ^\circ\text{C} \) [43]. Nevertheless, the influence of the temperature increase to \( \vartheta = 50 \, ^\circ\text{C} \) for coupling with the SFC can be qualitatively evaluated from the capillary number, \( Ca \). Since \( \sigma_{G/L} \) is reduced with increasing temperature, \( Ca \) is increased, leading to the conclusion that an enhanced wall film is present. Consequently, it is expected that forcing the favored slug formation mechanism at \( \vartheta = 50 \, ^\circ\text{C} \) requires a decrease in \( Ca \). This is realized by designing T-mixers with 90° inlets, providing a \( w_{\text{ETH, mix}} \) of 5 wt.-%. The graphical construction can be seen in Figure 6. The constructed and 3D printed T-mixer is shown in Figure 7.

![Figure 6](image_url)

**Figure 6.** Ternary solubility diagram of l-alanine/water/ethanol at 50 °C and 1 bar. The solid blue line represents the solubility curve based on the experimental data from \textsc{An et al.} [41]. The solid green line describes the underlying conode of the nucleation process until thermodynamic equilibrium is reached. The dashed red line connects the feed solution (F, blue triangle) and antisolvent mixture (AS, red triangle, \( w_{\text{ETH, AS}} \)) to be mixed, and the intersection between the dashed red line and the green line defines the MP (black dot). By using the lever rule, \( w_{\text{solid}} \) as well as the ratio of the feed and antisolvent mass flow rate can be calculated based on the location of the MP on the conode. Graphical construction of nucleation process for the 90° T-mixer for \( w_{\text{ETH, mix}} \) of 5 wt.-% and \( w_{\text{solid}} \) of 1 wt.-%.
Figure 7. Design of the T-mixer with 90° inlets as continuous nucleator for $w_{\text{EtOH mix}}$ of 5 wt.-% at $\vartheta = 50$ °C: Left—illustration of a cross-section created using the software Fusion 360 (Version 2.0.17457) with ID dimensions. The IDs of the feed and the mixing channel depend on the process parameters. Right—photo of the 3D printed T-mixer.

Before coupling the nucleator to the SFC, the T-mixer and its performance were first examined. Characterization experiments examining the T-mixer as a continuous in situ nucleator were conducted at $\vartheta = 20$ °C to evaluate the performance of the nucleator without heat losses. The design and process parameters of the T-mixer with 90° inlets for the characterization (Design 1) and coupling experiments with the SFC as a growth zone (Design 2) are summarized in Table 1.

### Table 1. Graphically determined process parameters and resulting design parameters of T-mixers with 90° inlets and equal velocities at the mixing point, providing $w_{\text{EtOH mix}}$ of 5 wt.-%.

<table>
<thead>
<tr>
<th>Design</th>
<th>1</th>
<th>2</th>
</tr>
</thead>
<tbody>
<tr>
<td>T-Mixer Configuration</td>
<td>90°</td>
<td>90°</td>
</tr>
<tr>
<td>$\vartheta$/°C</td>
<td>20</td>
<td>50</td>
</tr>
<tr>
<td>$d_{i,\text{AS}}$/mm</td>
<td>0.75</td>
<td>0.75</td>
</tr>
<tr>
<td>$d_{i,\text{Feed}}$/mm</td>
<td>2.61</td>
<td>2.27</td>
</tr>
<tr>
<td>$d_{i,M}$/mm</td>
<td>2.61</td>
<td>2.27</td>
</tr>
<tr>
<td>$w_{\text{EtOH mix}}$/wt.-%</td>
<td>5</td>
<td>5</td>
</tr>
<tr>
<td>$w_{\text{solid}}$/wt.-%</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>$q_{\text{feed}}$/mL min$^{-1}$</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>$q_{\text{AS}}$/mL min$^{-1}$</td>
<td>0.83</td>
<td>1.10</td>
</tr>
<tr>
<td>$w_{\text{EtOH,AS}}$/wt.-%</td>
<td>72</td>
<td>61</td>
</tr>
<tr>
<td>$R e_{\text{AS}}$</td>
<td>88</td>
<td>190</td>
</tr>
</tbody>
</table>

4.2. Validation Experiments for SFC Nucleator

The characterization experiments for the mixer with 90° inlets were carried out according to the process parameters listed in Table 1 (Design 1) to validate the formation of nuclei and to verify the suitability as a continuous nucleator for the SFC. The results are shown in Figure 8.
The resulting shape of the isotropic particles was characterized as mainly bipyramidal (Figure 8a). In-line DIA measurements were conducted twice. Despite the low particle number, a narrow PSD with a defined mean particle size was obtained (Figure 8b, blue data points).

The low number of measured particles suggests a relatively high residual supersaturation at the time of measurement in the QICPIC flow cell after an RT of approximately 10 s provided by the periphery. Therefore, analogous to the characterization experiments of the T-mixer with 180° inlets (Section 3.3), at-line gravimetric concentration measurements were conducted after 5, 10, and 15 min of RT to evaluate whether a steady-state value is established after a certain time period. For the configuration used in this section, the mass fraction \( w_{\text{solid}} \) slightly increased during the observed RT of 15 min (Table S1), indicating slow nucleation and growth kinetics. The DIA after an RT of 15 min (Figure 8b, orange data points) showed a high number of particles detected with a \( d_{50,5} \) of 52.54 ± 4.20 µm and a PSD width of \( d_{90-10,3} = 64.82 ± 1.85 \mu \text{m} \). The low standard deviations show that the nucleator design with 90° provides highly reproducible results.

The characterization experiments were conducted for 90 min each without the occurrence of encrustations or restricted operating performance. Hence, this nucleator is assumed to be suitable for long-term operation and coupling to the SFC, serving as a growth zone.

### 4.3. Experimental Setup and Procedure

The setup used for the continuous crystallization experiments containing the nucleator in front of the SFC as a growth zone is shown in Figure 9. The feed solution \( V = 5 \text{ L} \) was prepared for a saturation temperature of \( T^\circ = 50 \text{ °C} \) (Equation (1)) and stirred for 1 h at 55 °C at a rotational stirrer speed of 250 rpm. This ensured that the solute L-alanine was completely dissolved and that a crystal-free solution could be generated. After one hour, the antisolvent mixture \( V = 0.3 \text{ L} \) was prepared and stirred at 150 rpm. Afterwards, the feed solution and antisolvent mixture were pumped into the T-mixer, serving as a continuous nucleator. In order to compensate heat losses in the periphery between the vessels \( L_{\text{feed line}} = 103.4 \text{ cm}; L_{\text{AS line}} = 49.4 \text{ cm} \) and the nucleator, as well as between the nucleator and the SFC \( L_{\text{connection line}} = 10 \text{ cm} \), the feed vessel was tempered to 52 °C and the antisolvent vessel to 53 °C during the operation. Furthermore, as illustrated in Figure 9, the feed line, the antisolvent line, and the connection line were isolated in order to improve temperature control.
The pumps and tubing used for these experiments are identical to those used in the experimental setup described previously for the independent investigation of the nucleator (Section 3.3.). The subsequent slug formation zone is realized with a T-junction (PP, θ_{PP/\text{water/air}} = 102°), which is fed with the suspension from the nucleator from the top and with synthetic air from the bottom. The volume flow rate of the synthetic air is regulated with a high-resolution needle valve (NV-001-HR from Bronkhorst) and a flow meter (El-Flow-Select from Bronkhorst). In addition, the synthetic air is heated up to the solution’s temperature via a tube-in-tube construction to avoid wall crystallization in the T-junction due to the temperature difference between the suspension and the synthetic air. The constructive connection from the T-junction to the SFC tubing determines the slug stability. Since the IDs of the T-junction (d_t = 3 mm) and the SFC tubing (d_f = 3.18 mm) are almost equal, a flush connection can be realized. The SFC tubing is manufactured of fluorinated ethylene–propylene (FEP) and has a total length of 26.54 m. FEP is characterized by a strong hydrophobicity and a high wetting angle of θ_{FEP/\text{water/air}} = 108.5° so that the risk of wall film formation in the SFC can be minimized.

The driving force for crystallization within the growth zone is realized via cooling. For this purpose, a co-current tube-in-tube construction is used around the FEP tubing, which allows for the gentle cooling of the suspension and the flexible adjustment of different cooling rates via the final temperature and the volume flow rate of the tempering medium (water) \cite{12}.

At the outlet of the crystallizer, analyses for qualitative and quantitative evaluations of product particles are conducted, as described in Section 2.2. In addition, the stability and consistency of slug flow are observed with a non-invasive video analysis. Therefore, the FEP tubing passes through a transparent glass box (14 cm × 6 cm × 6 cm) filled with degassed water, illuminated by a LED lamp. This minimizes reflections and the light effects of the environment and creates a high contrast. Depending on the adjusted temperature at the end of the SFC, the glass box is filled with appropriately tempered water to prevent wall crystallization due to temperature differences. The videos are recorded with a Canon EOS M6 camera and processed with an in-house Matlab script to determine the slug length and its distribution \cite{44,45}.

The overall crystallization process, including nucleation and the growth zone, is evaluated with respect to the relative yield, long-term operability, reproducibility, and product quality in terms of PSD. Therefore, one operating point (OP1) in triplicate is tested for the validation of the continuous nucleator and its suitability for coupling with the SFC for the proof of concept. The selection of the experimental parameters is based on
preliminary experiments so that the occurrence of wall crystallization can be avoided and stable and long-term operation of the connected apparatus can be guaranteed. Nevertheless, this operating point is known to be not optimal regarding the suspension of particles, so agglomeration events may frequently occur.

In order to enhance the suspension, the suspension correlation developed in our previous publication [46] is used to determine the velocity necessary for homogeneous horizontal and vertical particle suspension inside the slugs. Therefore, two experiments (OP2) are conducted to qualitatively show the suitability of the coupled nucleation and growth zone, producing high-quality products. The total liquid volume flow rate is increased, as this has been shown to be the CPP for suspension [44]. However, this leads to a reduction in $\tau_{SFC}$ at constant $L_{tubing}$, and, thus, less time remains for crystal growth inside the SFC. The design and process parameters of both operating points are displayed in Table 2.

Table 2. Experimental and resulting parameters for a T-mixer with 90° inlets ($w_{ETH, mix} = 5$ wt.-%, Design 2) as continuous nucleator coupled with the SFC ($L_{tubing} = 26.54$ m) for two different operating points (OP1 and OP2). The experiments for OP1 and OP2 were performed three and two times, respectively.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>OP1</th>
<th>OP2</th>
</tr>
</thead>
<tbody>
<tr>
<td>$Q_L$/mL min$^{-1}$</td>
<td>5.55</td>
<td>11.09</td>
</tr>
<tr>
<td>$\varepsilon_{L0}/\text{m}^{-3}$</td>
<td>$0.43 \pm 0.03$</td>
<td>$0.41 \pm 0.01$</td>
</tr>
<tr>
<td>$\tau_{SFC}$/min</td>
<td>$16.44 \pm 1.21$</td>
<td>$7.74 \pm 0.13$</td>
</tr>
<tr>
<td>$\vartheta_{SFC, start}$/°C</td>
<td>$50.35 \pm 0.11$</td>
<td>$49.46 \pm 0.01$</td>
</tr>
<tr>
<td>$\vartheta_{SFC, end}$/°C</td>
<td>$25.64 \pm 0.41$</td>
<td>$34.75 \pm 0.08$</td>
</tr>
<tr>
<td>$K$/K min$^{-1}$</td>
<td>$1.51 \pm 0.09$</td>
<td>$1.90 \pm 0.02$</td>
</tr>
</tbody>
</table>

Regarding the production of APIs in the pharmaceutical industry, the feasibility of a continuous crystallization process for non-bipyramidal, anisotropic particle shapes is of major importance since most APIs exhibit a high AR [47]. Accordingly, the T-mixer with 90° inlets as a continuous nucleator based on antisolvent nucleation was qualitatively examined with respect to its potential for operation with various crystal shapes of L-alanine. The T-mixer characterization experiments in Section 3.4. indicate that, with regard to antisolvent nucleation, the particle shape is influenced by the ethanol concentration. Thus, the experimental results revealed that bipyramidal particles were generated at $w_{ETH, mix} = 5$ wt.-% and that needle-shaped particles were obtained at $w_{ETH, mix} = 23$ wt.-%. Since explicitly for the coupling with the SFC, the formation of a specific particle shape by adjusting $w_{ETH, mix}$ is not possible due to the operational limitation regarding the ethanol concentration for stable slug flow, the influence of the addition of L-glutamic acid on the particle shape in the context of a quaternary substance system, L-alanine/L-glutamic acid/water/ethanol, was evaluated. L-glutamic acid was added at a concentration of 5.8 g kg$^{-1}$ regarding the mass of water to the feed solution (L-alanine/water) at $\vartheta^* = 50$ °C. The experiments were performed using the process parameters of OP2 (Table 2) for enhanced particle suspension in the slugs in duplicate. The relatively high $K$ of 1.90 K min$^{-1}$ favored the formation of the largest possible optically detectable particles to ensure a reliable microscopic examination of the particle shape.

4.4. Results

The results of the crystallization experiments with the 90° T-mixer as a continuous nucleator coupled with the SFC according to the operating points for a ternary substance system in Table 2 are given in Table 3 and Figure 10. During the entire processing time of the OP1 experiments of about 45 min each, the desired squeezing mechanism inside the slug formation zone was observed, confirming the assumptions made according to the findings of KUFNER ET AL. [43]. Furthermore, the low standard deviations with respect to
the process parameters $\varepsilon_{LB}$, $\vartheta_{SFC,start}$, and $\vartheta_{SFC,end}$ ensured a reliable evaluation of the experiments and their reproducibility. In terms of $Y_{rel}$, $81.52 \pm 3.61\%$ was obtained within an RT of $16.44 \pm 1.21$ min using a moderate mean cooling rate ($\bar{\kappa}$) of $1.51 \pm 0.09$ K min$^{-1}$.

![Image](image_url)

**Figure 10.** Microscopic images (top) and pictures of slug flow with suspended particles taken at the end of the SFC (bottom) for inhomogeneous ((a), black, OP1) and homogeneous ((b), green, OP2) suspension states during the crystallization process inside the SFC.

Regarding the particle size, a $d_{50.3}$ of $484.14 \pm 54.60$ µm and a broad particle size width of $d_{90-10} = 530.47 \pm 50.21$ µm were measured via the at-line DIA procedure. This confirmed that the suspension of particles was not sufficient to distribute the particles equally in the horizontal and vertical directions within the slugs (Figure 10a, bottom), leading to a high contact time of the particles. This enabled the formation of solid bridges between them, resulting in agglomerates, which was qualitatively confirmed via microscopic images (see examples in Figure 10a, top). The particle shapes were bipyramidal and corresponded to the shapes obtained in the characterization experiments of the 90° T-mixer shown in Section 4.2.

In the case of OP2, no single particles or particle clusters were visible to the naked eye at the end of the SFC tubing, but a white shimmer was visible within the entire liquid slug (Figure 10b, bottom), proving the assumption that the suspension state was sufficient. The microscopic image shows that small, bipyramidal single particles formed (Figure 10b, top), so the operating point was capable of producing particles with a low agglomeration degree. Unfortunately, this could not be conclusively confirmed via at-line DIA, as the particles were too small for measurement.

### Table 3. Experimental results of the crystallization processes using the process parameters given in Table 2.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>OP1</th>
<th>OP2</th>
</tr>
</thead>
<tbody>
<tr>
<td>$Y_{rel}/%$</td>
<td>$81.52 \pm 3.61$</td>
<td>$65.65 \pm 2.07$</td>
</tr>
<tr>
<td>$n_{particle}/$</td>
<td>$494 \pm 248$</td>
<td>n.a. *</td>
</tr>
<tr>
<td>$d_{50.3}/\mu m$</td>
<td>$484.14 \pm 54.60$</td>
<td>n.a. *</td>
</tr>
<tr>
<td>$d_{90-10}/\mu m$</td>
<td>$530.47 \pm 50.21$</td>
<td>n.a. *</td>
</tr>
</tbody>
</table>

* PSD could not be measured due to the high end temperature at the SFC, so influences during the collection of the suspension, as well as during the measurement, cannot be excluded.
Figure 11 shows the results of using the quaternary substance system L-alanine/L-glutamic acid/water/ethanol to demonstrate the potential application of the T-mixer as a continuous nucleator for producing particles with an increasing aspect ratio. During the experiments, optically, no particles were detected in the slugs via the naked eye (see Figure 11, bottom). This qualitatively suggests that the quaternary substance system had a broader metastable zone width (MZW) than the ternary substance system used before. But, Figure 11 shows the corresponding microscopic image of the suspension at the outlet of the SFC, showing small needle-shaped particles. Consequently, the influence of L-glutamic acid on the particle shape was verified. Furthermore, the microscopic image qualitatively shows a narrow PSD with a relatively small mean particle size. This aligns with the observation that no particles were optically detected in the SFC due to the relatively small particle sizes. Moreover, the small particle sizes indicate an effective suspension of the particles in the slugs reflected qualitatively by the formation of single or low agglomerated particles.

The experimental results presented in this section qualitatively demonstrate that the T-mixer with 90° inlets as a continuous nucleator coupled with the SFC as a growth zone in the context of continuous crystallization is also feasible for the production of non-bipyramidal anisotropic particles.

5. Fully Continuous End-to-End Small-Scale Manufacturing

In this section, the fully continuous end-to-end small-scale manufacturing process of crystalline particles, from continuous nucleation to free-flowing particles, was realized by adding the continuous particle isolation unit resulting in free-flowing particles.

5.1. Experimental Setup and Procedure

The experimental setup is depicted in Figure 12. The integrated process is divided into three stages, and the experimental conditions of all stages are summarized in Table 4.
Figure 12. Schematic of experimental setup for end-to-end continuous small-scale manufacturing of free-flowing particles containing a continuous nucleator (T-mixer with 90° inlet configuration) in stage 1, continuous crystallization (SFC) in stage 2, and continuous particle isolation consisting of filtration and two-stage washing (CVSF), as well as a heater between SFC and CVSF, in stage 3.

Table 4. Experimental process and resulting parameters for the integrated process consisting of coupled nucleator, SFC, heater, and CVSF based on triple experiments.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>T-mixer (90° inlets)</td>
<td></td>
</tr>
<tr>
<td>$Q_L$/mL min$^{-1}$</td>
<td>5.55</td>
</tr>
<tr>
<td>$Q_{Feed}$/mL min$^{-1}$</td>
<td>5</td>
</tr>
<tr>
<td>$Q_{AS}$/mL min$^{-1}$</td>
<td>0.55</td>
</tr>
<tr>
<td>$w_{ELOH,AS}$/wt.-%</td>
<td>61</td>
</tr>
<tr>
<td>$w_{ELOH,mix}$/wt.-%</td>
<td>5</td>
</tr>
<tr>
<td>SFC</td>
<td></td>
</tr>
<tr>
<td>$Q_L$/mL min$^{-1}$</td>
<td>5.55</td>
</tr>
<tr>
<td>$\varepsilon_{LO}$</td>
<td>0.44 ± 0.00</td>
</tr>
<tr>
<td>$\tau_{SFC}$/min</td>
<td>16.72 ± 0.04</td>
</tr>
<tr>
<td>$\vartheta_{SFC,start}$°C</td>
<td>48.89 ± 0.24</td>
</tr>
<tr>
<td>$\vartheta_{SFC,end}$°C</td>
<td>23.63 ± 0.19</td>
</tr>
<tr>
<td>$\kappa$/K min$^{-1}$</td>
<td>1.51 ± 0.03</td>
</tr>
<tr>
<td>$w_{solid}$/wt.-%</td>
<td>2.73 ± 0.56</td>
</tr>
<tr>
<td>Heater</td>
<td></td>
</tr>
<tr>
<td>$\vartheta_{heater,TM,IN}$°C</td>
<td>51</td>
</tr>
<tr>
<td>$\vartheta_{heater,TM,OUT}$°C</td>
<td>31</td>
</tr>
<tr>
<td>CVSF</td>
<td></td>
</tr>
<tr>
<td>$n_{screw}$/rpm</td>
<td>1</td>
</tr>
<tr>
<td>$\Delta p_{set}$/mbar</td>
<td>400</td>
</tr>
<tr>
<td>$Q_{wash}$/mL min$^{-1}$</td>
<td>15</td>
</tr>
<tr>
<td>$\alpha$/%</td>
<td>5</td>
</tr>
<tr>
<td>$\tau_{CVSF, id}$/min</td>
<td>32</td>
</tr>
</tbody>
</table>
Stage 1 comprises the continuous nucleator realized by the T-mixer with the 90° inlet configuration of design 2 according to Table 1. Subsequently, continuous crystallization takes place in the SFC in stage 2, with the process parameters of OP1 (see Table 2). OP1 provides a higher $Y_{rel}$ and a lower SFC outlet temperature than OP2, ensuring a higher $w_{solid}$ after crystallization, which is needed for higher downstream performance inside the CVSF (described below) and a reliable analysis at the SFC outlet with minimal temperature influence. Finally, the continuous particle isolation steps, namely, solid–liquid separation and particle washing, are performed using the patented CVSF in stage 3 to achieve free-flowing particles at the end of the CPC [28,37].

The coupling of the SFC to the CVSF was investigated by us in a previous study at two different operating points [11]. For continuous crystallizers, residual supersaturation at the outlet is a major challenge with respect to coupling with subsequent unit operations, especially for solid–liquid separation based on filtration. The residual supersaturation of the mother liquor can potentially lead to the clogging of the filter medium, decreasing the performance of continuous solid–liquid separation. As described in our previous work [11], the residual supersaturation of the mother liquor at the continuous crystallizer outlet can be degraded by rapidly heating to a slightly undersaturated state without dissolving the product particles due to a short RT in this state before filtration. Consequently, a tube-in-tube heater was implemented between the SFC and CVSF. The outlet temperature of the SFC from the coupling experiments of the T-mixer with the 90° inlet configuration and the SFC (Section 4.3.) was used for the operating temperature of the heater, as well as for the adjustment of the operating parameters of the CVSF. Based on a total volume flow rate of the SFC outlet stream of approximately 12.8 mL min$^{-1}$ and an ID of 3.18 mm of the inner FEP tubing of the heater, a tubing length for the heater of 19 mm was chosen to provide an RT of merely 7 s in the heater ($t_{heater}$) and, thus, rapid heating while avoiding particle dissolution. The preservation of the PSD before and after the heater for $t_{heater} < 10$ s was proven in preliminary experiments in our previous work [11]. The outer tubing of the heater was made of polyvinyl chloride with an ID of 15 mm. The tube-in-tube heater was operated in co-current mode by using a thermostat (CC-K6 from Peter Huber Kältemaschinenbau SE, Offenburg, Germany). Regarding the outlet temperature of the SFC of 24 °C, the mother liquor was rapidly heated by 7 K to 31 °C in the heater to ensure a slightly undersaturated state before entering the CVSF.

The modular CVSF in stage 3 consists of tubular, double-walled cylindrical modules for the process steps of filtration and washing. As investigated in previous studies for the substance system l-alanine/water with bipyramidal l-alanine particles, the CVSF setup providing filtration and two-stage washing guarantees dry, free-flowing particles without additional drying [37]. More detailed information regarding the CVSF setup can be found in previous publications [6,29]. The suspension from the SFC was continuously heated in the tube-in-tube heater and supplied into the first module of the CVSF, where the initial solid–liquid separation was performed via vacuum filtration. The vacuum in the modules was applied by corresponding vacuum pumps (PC 2004 Vario from Vacuubrand GmbH & Co. KG, Mönchengladbach, Germany), and the filtrates were collected in respective vessels. The formed filter cake was axially transported simultaneously to the first washing step in the subsequent module by a slowly rotating screw. The wash liquid in the first washing module was a mixture of ethanol and water at a volumetric ratio of 4:1. The composition was chosen to prevent nucleation due to an antisolvent effect of the remaining mother liquor in the filter cake after the filtration process. In the second washing module, pure ethanol was applied as a wash liquid. Hollow cone nozzles (220.054 from Lechler GmbH, Metzingen, Germany) were used for a homogenous distribution of the wash liquids on the filter cake. Peristaltic pumps (07528-30 from Masterflex SE, Gelsenkirchen, Germany) were utilized to pump the wash liquids to the nozzles of the corresponding washing module. In the washing modules, the displaced liquid from the pores of the filter cake and the applied wash liquid were simultaneously removed via vacuum filtration analogous to the sole filtration process in the first module.
The filling degree, \( \alpha \), is an essential parameter for CVSF operation and performance. Therefore, \( \alpha \) should be adjusted to a minimum of 50% to ensure an effective filtration and washing process [6,29]. Since the volume flow rates of the suspensions \( Q_{\text{susp}} \) and \( w_{\text{solid}} \) are predefined by the preceded crystallization process in the SFC, \( \alpha \) was set according to the rotational screw speed \( n_{\text{screw}} \). \( n_{\text{screw}} \) directly influences \( \alpha \) in the CVSF. Accordingly, a maximum, non-optimal \( \alpha \) of 5% was realized at a minimum \( n_{\text{screw}} \) of 1 rpm. The volume flow rates of the wash liquids \( Q_{\text{wash}} \) were each set to the minimum volume flow rate of 15 mL min\(^{-1}\) to provide the lowest possible wash ratio in both washing modules.

Gravimetric concentration measurements at the outlet of the SFC were conducted, enabling the calculation of the achieved \( Y_{\text{rel}} \) of the continuous crystallization process in the SFC. Moreover, the PSDs were measured using at-line DIA at the inlet and outlet of the CVSF in order to demonstrate that the CVSF operation ensures the preservation of the particle properties designed in the preceding continuous crystallization process in the SFC. Finally, the residual moisture of the particles at the outlet of the CVSF was determined via gravimetric concentration measurements in order to guarantee free-flowing particles at the end of the fully continuous CPC.

5.2. Results

From the concentration measurements at the outlet of the SFC, a high \( Y_{\text{rel}} \) of 87.5 ± 2.5% was achieved in the continuous crystallization step based on three experiments. Furthermore, confirming the experimental results in Section 4.4., the continuous nucleation in the T-mixer with 90° inlets coupled with the SFC provided reproducible results in terms of the achieved \( Y_{\text{rel}} \). The PSDs determined at the inlet and outlet of the CVSF of one experiment are depicted in Figure 13 together with representative images from DIA.

![Figure 13](image-url)  
Figure 13. Results at the inlet and outlet of the CVSF. Left—the PSDs of the product particles at the inlet (CVSF\text{IN}, red) and outlet (CVSF\text{OUT}, blue) of the CVSF. Right—exemplary DIA binary images of particles at the inlet (top, red) and outlet (bottom, blue).

Based on the PSD in Figure 13, a \( d_{50,3,\text{IN}} \) of 377.25 \( \mu \text{m} \) was determined. This value is in the same magnitude as the \( d_{50,3} \) from the coupling experiments in Section 4.4., indicating a reproducible continuous nucleation and growth process. Furthermore, a
of \( d_{90-10.3, IN} \) of 258.92 \( \mu m \) was derived. Compared to our previous study regarding the coupling of the SFC with the CVSF based on seed supply (350–560 \( \mu m \)) [11], a narrower PSD was achieved in this work by using the T-mixer with 90° inlets as a continuous in situ nucleator. The PSD at the CVSF outlet was characterized by a \( d_{50.3, OUT} \) of 353.29 \( \mu m \) and a \( d_{90-10.3, OUT} \) of 267.60 \( \mu m \). Consequently, the mean particle size (\( d_{50.3} \)) and the particle size width (\( d_{90-10.3} \)) at the inlet and outlet of the CVSF showed a high level of agreement, demonstrating the preservation of the CQAs during CVSF operation.

The reproducibly measured residual moisture of 3 ± 0.16% in three experiments indicates almost dry, free-flowing particles, despite the low \( \alpha \) of 5%, leading to a restricted deliquoring process and, thus, a higher residual moisture.

Regarding the long-term operation of the entire integrated process, the SFC was operated for 45 min until increased wall crystallization inside the SFC and the heater occurred. The CVSF exhibited a constant performance over the total 60 min operation time. Accordingly, no filter clogging occurred, as evidenced by a constant pressure drop in the filtration module (Figure 14a) and a constant filtration performance (Figure 14b). This suggests that the supersaturated mother liquor at the outlet of the SFC was sufficiently undersaturated in the heater in order to prevent filter clogging in the filtration module of the CVSF.

![Figure 14. Results of the filtration module during CVSF operation. (a) Pressure drop of the vacuum pump over the process time (blue line). The vertical green line indicates the end of the suspension supply from the SFC. The horizontal dotted red line represents the set pressure drop \( \Delta P_{set} \). (b) Filtrate mass over the process time until the end of the suspension supply into the CVSF.](image)

The experimental results presented in this section demonstrate for OP1 (Table 2) that the fully continuous end-to-end small-scale manufacturing process, consisting of the T-mixer with a 90° inlet configuration, the SFC, and the CVSF, provided a high \( Y_{rel} \) of approximately 80% in the continuous crystal growth process in the SFC. Additionally, the preservation of the CQAs during CVSF operation could be inferred from the high agreement of PSDs in terms of the mean particle size (\( d_{50.3} \)) and particle size width (\( d_{90-10.3} \)) at the inlet and outlet of the CVSF. Furthermore, a low residual moisture of approximately 3% was reproducibly determined at the CVSF outlet, indicating free-flowing particles at the CVSF outlet, despite a low, non-optimal \( \alpha \) of 5% in the CVSF.

6. Conclusions

Small-scale production in the pharmaceutical industry (250–100 kg a\(^{-1}\)) is undergoing a turnaround in terms of processing methods. Thus, more and more research is being performed in the direction of continuous production, which, however, brings challenges at a small scale like this. There are already some concepts for continuous cooling crystallization and a few concepts for downstream crystallization, but there is still a lack
of suitable methods that provide continuous nucleation and, thus, continuous in situ seeds for crystallization. This leads to the fact that long-term operation for the continuous production of crystalline products is not easily possible.

Our study follows this line of thought, introducing a novel continuous nucleator designed based on phase diagrams and making use of antisolvent nucleation. The choice of antisolvent used for nucleation should ideally match the solvent used in the washing step. For the rapid and precise creation of the nucleator at a small scale, meeting diverse requirements for nucleation and subsequent growth processes, fast 3D printing was used. The suitability of the nucleator for continuous, long-term operation in combination with various crystallizers was assessed to ensure an adequate number of nuclei for the subsequent growth process and substantial consumption of supersaturation, minimizing the risk of fouling in the crystallizer. The developed nucleator in the form of a T-mixer could be implemented in front of various crystallizers. Here, its implementation was shown for a slug flow crystallizer (SFC) with the aim of producing high-quality product particles. These results obtained using the continuous nucleator coupled with the SFC as a growth zone are qualitatively comparable to the results of batch cooling crystallization [11] and SFC crystallization in the literature [11,12], which, however, were carried out with the binary substance system l-alanine/water. In this context, the concept of continuous in situ nucleation compared to seed crystal addition provides both a reduced time investment, since the production of seed crystals with high-quality specifications is not required, and the prevention of potential contamination due to the in situ production of nuclei.

Furthermore, with a connection to a continuous vacuum screw filter (CVSF), the fully continuous crystal process chain (CPC) up to the achievement of free-flowing particles was completed, and the experimental results demonstrate that the fully continuous end-to-end small-scale manufacturing of free-flowing particles is possible, achieving reproducibly high relative yields in the crystal growth process in the SFC of approximately 80%, preserving the critical quality attributes (CQAs) during CVSF operation and resulting in residual moistures of around 3% with the configuration used, despite a low, non-optimal filling degree of 5% in the CVSF. The modular setup of the CVSF provides the option to extend the CVSF via a drying module in order to further reduce the residual moisture below 1% and obtain dry, free-flowing particles at the end of the integrated process. Additionally, the CVSF offers the potential to realize higher filling degrees independent from the suspension volume flow rate and solid loading by increasing the shaft diameter of the screw. This will further improve the flexibility of the CVSF with regard to compatibility with various types of continuous crystallizers and operation points. A summary of all the results of the nucleator characterization and coupling experiments is provided in Table S3.

With respect to long-term operation, the processing time could be extended by implementing a cleaning-in-place (CIP) system when fouling is detected in the SFC. Moreover, proof-of-concept experiments were performed with the model substance system l-alanine/water/ethanol, resulting in limited achievable yields and solid loadings in the SFC based on the solubility data. Consequently, the integrated process consisting of the T-mixer with a 90° inlet configuration as a continuous nucleator, the SFC for continuous crystallization, and the CVSF for continuous product isolation must comprehensively be investigated with respect to different substance systems and operating windows.

The end-to-end continuous small-scale manufacturing process for free-flowing crystalline particles investigated in this study shows that, even under non-optimized conditions, the concepts used enable the fast production of high-quality crystalline particles, which is of high importance during the drug development phase, especially for new drug substances.
**Supplementary Materials**: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/cryst13121675/s1. Table S1: Results of gravimetric concentration measurements at 5, 10 and 15 min RT for the 90° T-mixer designed for the SFC coupling according to the procedure described in Section 3.3; Table S2: Different operating points for the T-mixer with 180° inlets based on the graphical construction in the ternary solubility diagram for L-alanine/water/ethanol at 20 °C and 1 bar; Table S3: Design and process parameters as well as the experimental results of this study. The results of the characterization ex-periments of the T-mixer with 180° and 90° inlets as continuous in-situ nucleator and coupling experiments of the T-mixer with 90° inlet configuration with the SFC as well as with the SFC and the CVSF are listed; Figure S1: Ternary solubility diagram of L-alanine/water/ethanol at 20 °C and 1 bar. The solid blue line represents the solubility curve based on the experimental data from AN ET AL. [41]. The solid green line describes the underlying cone of the nucleation process until thermodynamic equilibrium is reached. The dashed red line connects the feed (F, blue triangle) and antisolvent mixture (AS, red triangle, \( w\text{_{EtlOH,AS}} \)) to be mixed and the intersection between the dashed red line and the green line defines the MP (black dot). By using the lever rule, \( w\text{_{solid}} \) as well as the ratio of the feed and antisolvent mass flow rate can be calculated based on the location of the MP on the cone. Graphical construction of nucleation process using the 180° inlet T-mixer.

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**Abbreviations**

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tbody>
<tr>
<td>ALD</td>
<td>Axis length distribution</td>
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<tr>
<td>API</td>
<td>Active pharmaceutical ingredient</td>
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<td>AR</td>
<td>Aspect ratio</td>
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<tr>
<td>AS</td>
<td>Antisolvent</td>
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<tr>
<td>CPC</td>
<td>Crystal process chain</td>
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<td>CPP</td>
<td>Critical process parameter</td>
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<td>CQA</td>
<td>Critical quality attribute</td>
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<tr>
<td>CVSF</td>
<td>Continuous vacuum screw filter</td>
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<td>DIA</td>
<td>Dynamic image analysis</td>
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<tr>
<td>F</td>
<td>Feed</td>
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<tr>
<td>FEP</td>
<td>Fluorinated ethylene–propylene</td>
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<tr>
<td>ID</td>
<td>Inner diameter</td>
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<tr>
<td>MAL</td>
<td>Major axis length</td>
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<tr>
<td>MAL(_{ch})</td>
<td>Characteristic major axis length</td>
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<tr>
<td>MIL</td>
<td>Minor axis length</td>
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<tr>
<td>MIL(_{ch})</td>
<td>Characteristic minor axis length</td>
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<tr>
<td>MP</td>
<td>Mixing point</td>
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<tr>
<td>MSMPR</td>
<td>Mixed-suspension mixed-product removal</td>
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<tr>
<td>MZW</td>
<td>Metastable zone width</td>
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<tr>
<td>NZ</td>
<td>Nucleation zone</td>
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<tr>
<td>OP</td>
<td>Operating point</td>
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<tr>
<td>PFC</td>
<td>Plug flow crystallizer</td>
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<tr>
<td>PP</td>
<td>Polypropylene</td>
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<td>PSD</td>
<td>Particle size distribution</td>
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<td>Abbreviation</td>
<td>Description</td>
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<tr>
<td>RT</td>
<td>Residence time</td>
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<tr>
<td>RTD</td>
<td>Residence time distribution</td>
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<tr>
<td>SFC</td>
<td>Slug flow crystallizer</td>
</tr>
<tr>
<td>SLD</td>
<td>Slug length distribution</td>
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**Latin Symbols**

- \( A \): Area/\( m^2 \)
- \( c \): Concentration/\( g \cdot g^{-1} \)
- \( c^* \): Saturation concentration/\( g \cdot g^{-1} \)
- \( Ca \): Capillary number/-
- \( d \): Diameter/\( m \)
- \( h \): Height/\( m \)
- \( L \): Characteristic length/\( m \)
- \( m \): Mass/\( kg \)
- \( \dot{m} \): Mass flow rate/\( kg \cdot s^{-1} \)
- \( n \): Particle number density of class \( i/- \)
- \( Q \): Volume flow rate/\( mL \cdot min^{-1} \)
- \( r \): Radius/\( m \)
- \( Re \): Reynolds number/-
- \( S \): Supersaturation/-
- \( V \): Volume/\( m^3 \)
- \( v \): Velocity/\( m \cdot s^{-1} \)
- \( w \): Mass fraction/-
- \( Y \): Yield/\( % \)

**Greek Symbols**

- \( \alpha \): Filling degree/-
- \( \Delta c \): Concentration difference/\( g \cdot g^{-1} \)
- \( \varepsilon \): Liquid hold-up/-
- \( \eta \): Dynamic viscosity/\( Pa \cdot s \)
- \( \theta \): Three-phase contact angle/\( \circ \)
- \( \theta \): Temperature/\( ^\circ C \)
- \( \kappa \): Cooling rate/\( K \cdot min^{-1} \)
- \( \rho \): Density/\( kg \cdot m^{-3} \)
- \( \sigma \): Surface tension/\( N \cdot m^{-1} \)
- \( \tau \): Residence time/\( s \)
- \( \Phi \): Cross-sectional area/-

**Indices**

- \( 0 \): Initial state
- \( 10,3 \): 10% of the distribution by volume
- \( 50,3 \): Median by volume
- \( 90,3 \): 90% of the distribution by volume
- \( * \): Saturation
- \( Ala \): \( L \)-alanine
- \( ambient \): Ambient temperature
- \( AS \): Antisolvent
- \( AS \) line: Tubing of the antisolvent line
- \( BV \): Batch vessel
- Connection line: Tubing between nucleator outlet and SFC inlet
- \( dry \): Dry solid
- \( dyn \): Dynamic
- \( empty \): Empty vessel
- \( EtOH \): Ethanol
- \( Feed \): Feed stream
- \( Feed \) line: Tubing of the feed line
- \( G/L \): Between gas and liquid phases
- \( i \): Inner
- \( L \): Liquid
- \( M \): Mixing channel
References


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