

Article

Basidiomycetes Associated with *Alnus glutinosa* Habitats in Andros Island (Cyclades, Greece)

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Abstract: Alluvial forests dominated by black alder (*Alnus glutinosa*) are widespread in Europe along river banks and watercourses forming a habitat of renowned ecological/conservation importance. Despite the considerable interest this habitat has attracted in terms of the associated fungal diversity, very few pertinent data are available from the eastern Mediterranean. Andros island (Aegean Sea, Greece) hosts the southernmost population of *A. glutinosa* in the Balkan Peninsula; such stands have been systematically inventoried for several years in respect to macrofungi. In total, 187 specimens were collected and studied by examining morphoanatomic features and by evaluating (when necessary) the outcome of sequencing the internal transcribed spacer (ITS) region of nuclear ribosomal DNA (nrDNA) to elucidate their identity and obtain an insight into phylogenetic relationships. As a result, 106 species were recorded, 92 are saprotrophic and 14 form ectomycorrhizae (ECM) with alders. Twenty-one species are first national records, while 68 other species are reported for the first time from this habitat in Greece. Several findings of particular interest due to their rarity, ecological preferences and/or taxonomic status are presented in detail and discussed, e.g., six *Alnicola* taxa, *Cortinarius americanus*, *Lactarius obscuratus*, *Paxillus olivellus* and *Russula pumila* (among the ECMs), and the saprotrophs *Entoloma uranochroum*, *Gymnopilus arenophilus*, *Hyphoderma nemorale*, *Lepiota ochraceofulva*, *Phanerochaete livescens* and *Psathyrella hellebosensis*.

Keywords: macrofungi; Basidiomycota; mushroom diversity; ectomycorrhiza; saprotroph; alder; Aegean Sea; Mediterranean; *Alnicola*

1. Introduction

Alluvial forests with *Alnus glutinosa* Gaertn. and *Fraxinus excelsior* L. (priority habitat 91E0*; Annex I, Directive 92/43/EEC) are distributed throughout Europe, but they are generally rare and threatened since only remnants exist, mainly in central and northern Europe [1]. Alder stands are considerably less frequent in the Mediterranean region, where the repercussions of changes in the hydrological cycle caused by global warming and climate destabilization are much more evident [2]. The southernmost limit of the priority habitat 91E0* in the Balkan Peninsula is located in Andros island (Figure 1), i.e., the northernmost in the Cyclades and situated at a transition zone between continental Greece and other islands of the Aegean Archipelago. From the geomorphological point of view, it is characterized by a remarkably intense relief and by many rivulets and streams of constant flow, which are unique among most of Central and South Aegean islands. *A. glutinosa* trees demonstrate a patchy distribution in Andros, predominantly occurring along the main streams within the Site of Community Importance (SCI) GR4220001 and in altitudes ranging from sea level to as high as 850 m above sea level (a.s.l.), very close to the highest peaks of the island. In many cases black alders are mixed with *Platanus orientalis* L., *Fraxinus ornus* L. and/or *Nerium oleander* L. (in lower altitudes), while they also form pure stands, as it is the case at the estuaries of the Vori stream in NE Andros.

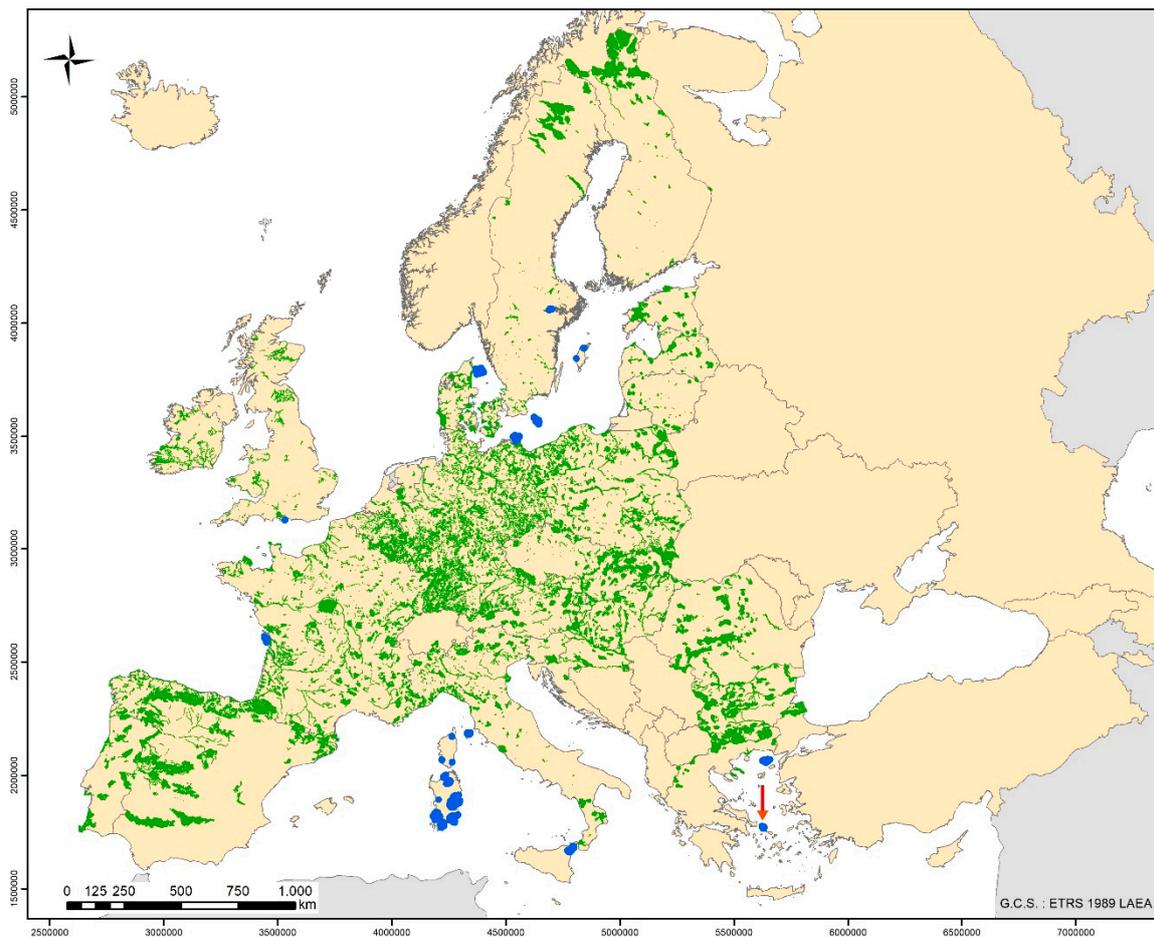


Figure 1. Map presenting Natura 2000 sites, which include the priority habitat 91E0* in continental Europe (in green) and in islands (in blue); Andros island is indicated by the red arrow. Data from <https://www.eea.europa.eu/data-and-maps/data/natura-6>.

Alder trees are known to form symbiotic relationships with nitrogen-fixing actinomycetes of the genus *Frankia* Brunchorst [3,4], with arbuscular mycorrhizal fungi (AM) of Glomeromycota [5,6] and with various ectomycorrhizal (ECM) fungi of Ascomycota and Basidiomycota [7–9]. European alder stands have been relatively well-studied in terms of both macro- and microfungal communities, and approx. 1000 species of saprotrophic and ECM macrofungi were reported [10–15]. In addition, mycocoenological studies from Europe and North America suggested that ECM fungi of *Alnus* spp. exhibit a remarkably high degree of host specificity compared to other tree species [8,16], while the analysis of both sporophores and ectomycorrhizae evidenced that alders have a low number (<50) of ECM symbionts worldwide [17–19].

Limited knowledge is available on the diversity of fungi associated with alders in Greece, and only preliminary data are reported in the few pertinent publications [20,21]. On the other hand, Andros is the only island of the Aegean Archipelago where a systematic inventory of macrofungi is in progress for more than 20 years. Biotopes characterized by river banks, springs and alluvial forests, where *A. glutinosa* is often the dominant tree species, were forayed in the past and 37 mushroom species were reported from this particular habitat in Andros, including ECM symbionts as well as xylotrophic, litter and/or humus saprotrophs [22–24]. Among the latter, *Entoloma alnicola* Noordel. & Polemis was described as new species for science and it is still known from the type locality only [25].

Since 2017, mycodiversity studies in alder stands of Andros were intensified in the frame of a LIFE Nature project (LIFE16-NAT_GR_000606), which -among others- aims at the conservation and restoration of the priority habitat 91E0* in the island. Hence, during the last few years, new

sites with alder stands were repeatedly forayed (in addition to those previously investigated), and a large number of new collections were made. These, together with previously sampled—but still unidentified—specimens, were subjected to detailed morphoanatomical examination in conjunction with sequencing and phylogenetic analyses (where judged necessary) in order to assess their identity. Moreover, in several occasions, past relevant reports on recorded taxa were revised/re-evaluated according to the latest respective taxonomic and phylogenetic concepts. Hence, this work presents an updated compilation of available data on the diversity of macrofungi in a habitat of significant interest occurring at the limits of its distribution in Europe.

2. Materials and Methods

2.1. Sampling of Biological Material

Data presented in this inventory are based on specimens collected from 10 sampling sites covering almost the entire area of *A. glutinosa* distribution in Andros island, which appears mainly within (or marginally out) the SCI GR4220001, extending from sea-level to an altitude of ca. 850 m a.s.l. (Figure 2; Table S1). The biological material examined for the purpose of this work was sampled in 38 forays performed during the last 25 years from late October to April; more than half of those (#23) were conducted in the period from 2017 to 2020. In total, 187 specimens found exclusively under alder trees or directly on their wood, woody residues or leaf-litter were collected, and voucher specimens are deposited in the Fungarium of the Laboratory of General and Agricultural University of Athens (ACAM).

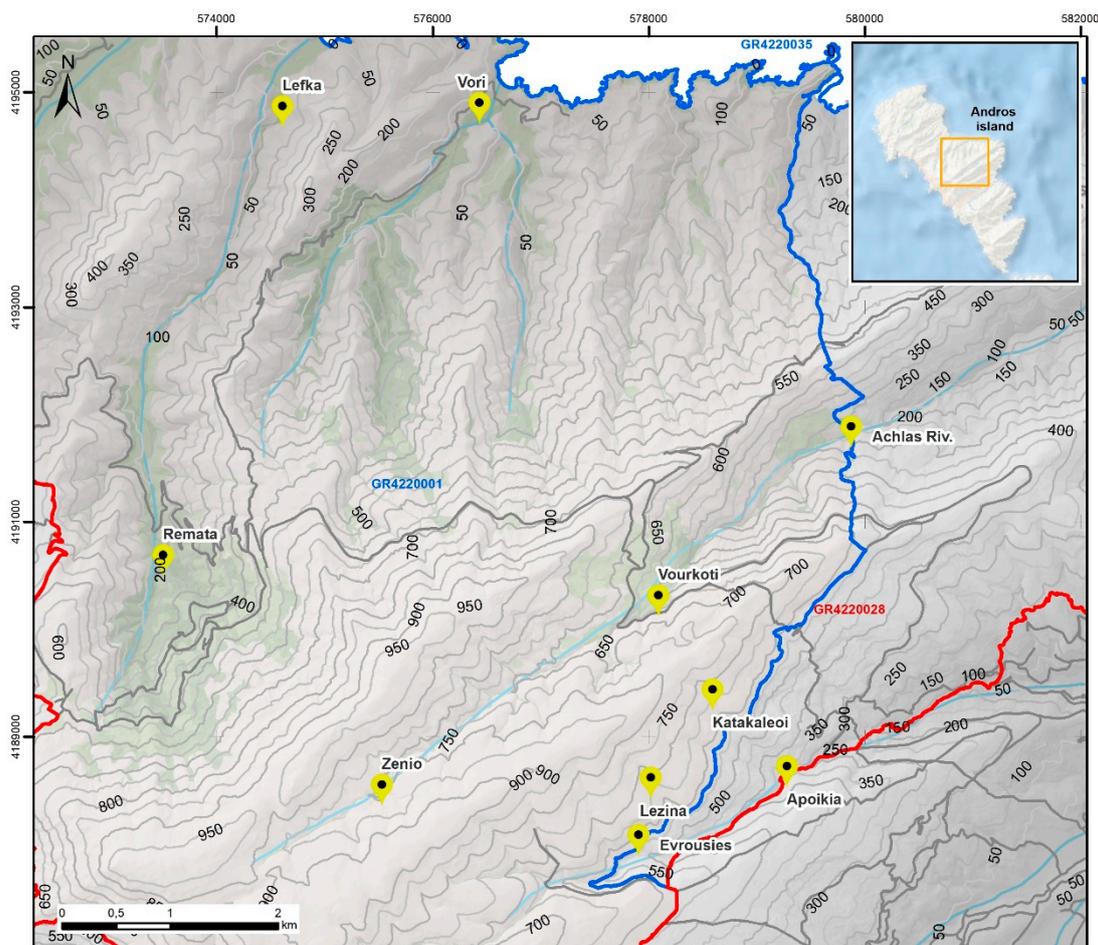


Figure 2. Sampling sites (in yellow marking) in the *Alnus glutinosa* habitat and relative position/size of the area under investigation within Andros island (map in upper right corner).

2.2. Morpho-Anatomical Features in Basidiomes

The morphological study included in situ recording of macroscopic features of taxonomic interest, while ex-situ examination involved observations of morphoanatomical characters in dried specimens. Sections were mounted and observed in KOH 3–5% (*w/v*), in Melzer's reagent, in cotton-blue, in cresyl-blue and in sulfovaniline solution. Observations were performed with the use of a Zeiss AxioImager A2 microscope under bright field and differential interference contrast (DIC); microphotographs were taken with the aid of a mounted digital camera (Axiocam). For all examined specimens a minimum of 30 mature basidiospores were measured and the resulting measurements as well as additional observations of other essential microscopical features (hymenial cystidia, pileipellis etc.) were used for determination of the species examined in accordance to pertinent identification keys and monographs (e.g., [26–36]).

2.3. DNA Extraction, Amplification and Sequencing

When deemed necessary, DNA sequencing and phylogenetic analyses were performed. Total genomic DNA was obtained from dried basidiomes and DNA extraction was performed through the use of the Nucleospin Plant II DNA kit (Macherey and Nagel, Düren, Germany) by following the manufacturer's protocol. The internal transcribed spacer (ITS; ITS1, 5.8S, ITS2) region within the nuclear ribosomal RNA gene cluster was examined by using the primers ITS1/ITS4 [37]. Polymerase chain reactions (PCR) were performed in 50 µL containing 50 ng DNA template, 0.25 µM of each primer, 0.2 mM of each dNTP, 1 × HiFi Buffer (Takara BIO INC., Shiga, Japan) and 1 U HiFi Taq DNA polymerase (Takara BIO INC., Shiga, Japan). PCR reactions were performed as follows: 94 °C for 5 min, followed by 35 cycles of 94 °C for 30 s, 50 °C for 30 s and 72 °C for 1 min, and a final extension at 72 °C for 10 min. PCR products were run in 1% agarose gels and purified using Invitrogen PureLink kit (Thermo Fisher Scientific, Seoul, S. Korea), and were submitted for sequencing to CeMIA SA (Larissa, Greece). The same PCR primers were used for sequencing. Chromatograms were checked with the aid of BioEdit v. 7.2.5 software [38]. Then sequences were examined against GenBank built-in search tools for obtaining information which could confer at identifying the material under study. A total of 61 validated sequences generated in this work were deposited in GenBank and the accession numbers MT458502 to MT458562 were obtained.

2.4. Phylogenetic Analysis of Sequence Data

A total of 42, 29 and 22 ITS sequences corresponding to selected species of the genera *Alnicola* Kühner (and *Naucoria* (Fr.) P. Kumm.), *Lactarius* Pers. and *Paxillus* Fr. (including 12, 5 and 4 sequences generated in this work), respectively, were subjected to phylogenetic analysis. In addition, species of the same or other genera were used as outgroups in each case. Multiple sequence alignment of each ITS rDNA dataset was conducted using the Q-INS-I algorithm as implemented in the online version of MAFFT v. 7 [39]. Alignments were reviewed, manually adjusted at misaligned sites and trimmed at the same position through MEGA X [40] before being used for further analysis.

Phylogenetic relationships of taxa for each alignment were inferred by using maximum likelihood (ML) and Bayesian inference (BI) through the CIPRES web portal (www.phylo.org; Miller et al. 2010). ML analyses were conducted by RAxML BlackBox online server (<http://phylobench.vital-it.ch/raxml-bb/>) [41] using default parameters and calculating bootstrap statistics according to the program recommendations for the best-scoring ML tree. BI analyses were performed by MrBayes v. 3.2.1 [42]. The best-fit substitution model for each dataset was selected according to the corrected Akaike information criterion (cAIC), as implemented in jModeltest v.2 [43]. The TPM2uf+G, TPM1uf+G and SYM+G models were selected for the *Alnicola*, *Lactarius* and *Paxillus* datasets, respectively. To estimate posterior probabilities, Markov chain Monte Carlo (MCMC) simulation was implemented in two parallel independent runs of four chains, one cold and three heated, with trees sampled every 1000 generations until the standard deviation of split frequencies is below 0.05; the first 25% of trees were omitted as

burn-in. A 50% majority rule consensus tree was built and visualized with iTOL [44]. Clades with ML bootstrap support (MLB) $\geq 65\%$ and Bayesian posterior probability (BPP) $\geq 95\%$ were considered as significantly supported.

3. Results and Discussion

The study of 187 specimens of macrofungi associated with the *A. glutinosa* priority habitat in Andros led to the identification of 106 species (74 genera) of basidiomycetes. Among them, 14 (13%) are ECM species (Table 1) strictly associated with alders [18,19]. The other 92 (87%) are saprotrophic; 70 (66%) saproxylic and 22 (21%) saprotrophic on soil, humus or leaf-litter (Table 2). Interestingly, 10 ECM and 11 saprotrophic species are first national records, while other 68 are reported for the first time from this habitat in Greece. Identification of specimens to species was performed by examining their morphoanatomic features and by evaluating (when necessary) the outcome of ITS sequencing and phylogenetic analysis; in the latter case, the respective GenBank accession numbers are provided (Tables 1 and 2). Selected findings of particular interest are presented (and discussed) by providing brief descriptions and comments on characters of potentially diagnostic value.

Table 1. Ectomycorrhizal (ECM) fungi identified during the study: species name, specimen code/collection date, locality and GenBank accession numbers for ITS sequences generated. First national records for Greece are indicated by an asterisk (*) before the species name.

a/a	Species Name	Specimen Code/ Collection Date	Locality	GenBank Accession No.
1	<i>*Alnicola escharoides</i> (Fr.) Romagn.	EP.17-A1344/11-Nov-2017	Zenio	
		EP.17-A1420/24-Nov-2017	Vori	
		EP.18-A1548/22-Feb-2018	Vori	MT458538
		EP.18-A1561/1-Nov-2018	Zenio	MT458539
		EP.18-A1571/2-Nov-2018	Vourkoti	MT458540
		EP.19-A1636/16 Nov 2019	Katakalaioi	
2	<i>*Alnicola inculata</i> (Peck) Singer	EP.17-A1346/11 Nov 2017	Zenio	MT458541
3	<i>*Alnicola luteolofibrillosa</i> Kühner	EP.17-A1430/24-Nov-2017	Vori	MT458542
4	<i>*Alnicola subconspersa</i> (Kühner ex P.D. Orton) Bon	EP.17-A1421/24-Nov-2017	Vori	MT458543
		EP.19-A1637/16-Nov-2019	Katakalaioi	MT458544
5	<i>Alnicola striatula</i> (P.D. Orton) Romagn.	EP.04-A679/15-Nov-2004	Evrousies	
		EP.19-A1614/14-Nov-2019	Evrousies	MT458545
6	<i>*Alnicola umbrina</i> (R. Maire) Kühner	EP.04-A678/15-Nov-2004	Evrousies	
		EP.17-A1377/2-Nov-2017	Lezina	MT458546
		EP.18-A1572/2-Nov-2018	Vourkoti	MT458547
		EP.19-A1607/12 Nov 2019	Zenio	MT458548
		EP.19-A1638/16-Nov-2019	Katakalaioi	
		EP.19-A1646/17-Nov-2019	Achlas riv.	
		EP.19-A1666/2-Dec-2019	Remata	MT458549
7	<i>*Cortinarius americanus</i> A.H. Sm.	EP.19-A1622/15-Nov-2019	Vourkoti	
8	<i>Gyrodon lividus</i> (Bull.) Sacc.	EP.14-A1263/1-Nov-2014	Vori	
		EP.17-A1428/24-Nov-2017	Vori	
9	<i>*Inocybe calospora</i> Quél.	EP.18-A1570/2-Nov-2018	Vourkoti	MT458550
10	<i>*Lactarius obscuratus</i> (Lasch) Fr.	EP.17-A1347/11-Oct-2017	Zenio	MT458551
		EP.17-A1566/1-Nov-2018	Zenio	MT458552
		EP.17-A1576/2-Nov-2018	Vourkoti	MT458553
		EP.19-A1645/17-Nov-2019	Achlas riv.	MT458554
		EP.19-A1664/30-Nov-2019	Remata	MT458555

Table 1. Cont.

a/a	Species Name	Specimen Code/ Collection Date	Locality	GenBank Accession No.
11	<i>*Paxillus olivellus</i> P.-A. Moreau, J.-P. Chaumeton, Gryta & Jarge	EP.95-A028/13-Nov-1995	Achlas riv.	MT458556
		EP.02-A353/22-Sep-2002	Evrousies	
		EP.04-A670/23-Oct-2004	Remata	
		EP.04-A673/24-Oct-2004	Achlas riv.	
		EP.14-A1266/1-Nov-2014	Vori	
		EP.17-A1348/11-Nov-2017	Zenio	
		EP.17-A1396/23-Nov-2017	Evrousies	
		EP.17-A1426/24-Nov-2017	Vori	
		EP.18-A1552/22-Feb-2018	Vori	
12	<i>*Russula pumila</i> Rouzeau & F. Massart	EP.18-A1575/2-Nov-2018	Vourkoti	MT458560
13	<i>Tomentella stuposa</i> (Link) Stalpers	EP.02-A327/29-Apr-2002	Vori	MT458561
14	<i>Tomentella subvilacina</i> (Ellis & Holw.) Wakef.	EP.02-A452/11-Oct-2002	Achlas riv.	MT458562
		EP.17-A1437/24-Nov-2017	Vori	

Table 2. Saprotrophic basidiomycetes identified during the study: species name, specimen code/collection date, locality, type of substrate and GenBank accession numbers for ITS sequences generated. First national records for Greece are indicated by an asterisk (*) before the species name.

a/a	Species Name	Specimen Code/Collection Date	Locality	Substrate Type	GenBank Accession No.
1	<i>Abortiporus biennis</i> (Bull.) Singer	EP.18-A1582/02-Nov-2018	Vourkoti	fallen trunk	
2	<i>Agaricus moelleri</i> Wasser	EP.19-A1613/14-Nov-2019	Katakalaioi	leaf-litter	MT458502
3	<i>Amaropostia stiptica</i> (Pers.) B.K. Cui, L.L. Shen & Y.C. Dai	EP.17-A1423/24-Nov-2017	Vori	dead stump	MT458503
4	<i>Armillaria gallica</i> Marxm. & Romagn.	EP.17-A1443/24-Nov-2017	Vori	dead stump	
5	<i>Armillaria mellea</i> (Vahl) P. Kumm.	EP.95-A021/12-Nov-1995	Remata	dead stump	
		EP.18-A1584/02-Nov-2018	Vourkoti	standing trunk	
		EP.19-A1651/29-Nov-2019	Vori	trunk base	
6	<i>Auricularia auricula-judae</i> (Bull.) Quéf.	EP.18-A1539/22-Feb-2018	Vori	standing trunk	
		EP.19-A1672/02-Dec-2019	Remata	standing trunk	
7	<i>Bjerkandera adusta</i> (Willd.) P. Karst.	EP.19-A1670/02-Dec-2019	Remata	fallen trunk	
8	<i>Botryobasidium candicans</i> J. Erikss.	EP.01-A275/26-Dec-2001	Vori	fallen trunk	
		EP.11-A1023/05-Jan-2011	Vori	fallen trunk	
		EP.17-A1434/24-Nov-2017	Vori	fallen trunk	
9	<i>Brevicellicium olivascens</i> (Bres.) K.H. Larss. & Hjortstam	EP.17-A1352/11-Nov-2017	Zenio	fallen trunk	MT458504
10	<i>Calocera cornea</i> (Batsch) Fr.	EP.17-A1440/25-Nov-2017	Vori	dead stump	
		EP.19-A1616/14-Nov-2019	Evrousies	dead stump	
11	<i>Ceriporia purpurea</i> (Fr.) Donk	EP.17-A1350/11-Nov-2017	Zenio	fallen trunk	MT458505
		EP.17-A1363/11-Nov-2017	Zenio	fallen trunk	
12	<i>Chondrostereum purpureum</i> (Pers.) Pouzar	EP.17-A1467/28-Nov-2017	Achlas riv.	standing trunk	MT458506
		EP.19-A1678/02-Dec-2019	Remata	standing trunk	
13	<i>Clavaria fragilis</i> Holmsk.	EP.19-A1639/16-Nov-2019	Katakalaioi	soil	
14	<i>Clitocybe nebularis</i> (Batsch) P. Kumm.	EP.18-A1580/02-Nov-2018	Vourkoti	leaf litter	

Table 2. Cont.

a/a	Species Name	Specimen Code/Collection Date	Locality	Substrate Type	GenBank Accession No.
15	<i>Clitocybe phyllophila</i> (Pers.) P. Kumm.	EP.18-A1579/02-Nov-2018	Vourkoti	leaf litter	
16	<i>Clitopilus hobsonii</i> (Berk.) P.D.Orton	EP.02-A331/29-Apr-2002	Vori	fallen trunk	
17	<i>Coniophora puteana</i> (Schumach.) P. Karst.	EP.11-A1017/5-Jan-2011 EP.17-A1411/24-Nov-2017 EP.19-A1679/02-Dec-2019	Vori Lefka Remata	fallen trunk fallen trunk fallen trunk	MT458507 MT458508 MT458509
18	<i>Coprinellus disseminates</i> (Pers.) J.E. Lange	EP.01-A260/26-Dec-2001 EP.19-A1654/29-Nov-2019	Vori Vori	rotten wood around stump	
19	<i>Coprinellus radians</i> (Fr.) Vilgalys, Hopple & Jacq. Johnson	EP.19-A1655/29-Nov-2019 EP.19-A1668/02-Dec-2019	Vori Remata	woody residues woody residues	MT458510
20	* <i>Coprinopsis melanthinia</i> (Fr.) Örstadius & E. Larss.	EP.17-A1412/24-Nov-2017 EP.19-A1659/29-Nov-2019	Lefka Vori	woody residues woody residues	MT458511
21	<i>Corioloopsis gallica</i> (Fr.) Ryvarden	EP.11-A1016/05-Jan-2011	Vori	standing trunk	
22	<i>Crepidotus luteolus</i> (Lamotte) Sacc.	EP.01-A268/26-Dec-2001	Vori	woody residues	
23	<i>Delicatula integrella</i> (Pers.) Fayod	EP.19-A1634/16-Nov-2019 EP.19-A1640/17-Nov-2019	Katakalaioi Achlas riv.	trunk base bark living tree	
24	<i>Entoloma alnicola</i> Noordel. & Polemis	EP.02-A364/22-Sep-2002	Evrousies	soil	
25	<i>Entoloma incanum</i> (Fr.) Hesler	EP.02-A362/22-Sep-2002 EP.04-A674/24-Oct-2004 EP.19-A1633/16-Nov-2019	Evrousies Achlas riv. Katakalaioi	soil soil soil	
26	<i>Entoloma juncinum</i> (Kühner & Romagn.) Noordel.	EP.02-A362/22-Sep-2002	Evrousies	soil	
27	<i>Entoloma mougeotii</i> (Fr.) Hesler	EP.02-A446/11-Oct-2002	Achlas riv.	soil	
28	* <i>Entoloma uranochroum</i> Hauskn. & Noordel.	EP.19-A1615/14-Nov-2019	Evrousies	soil/leaf-litter	
29	<i>Exidiopsis galzinii</i> (L.S. Olive) K. Wells	EP.02-A448/11-Oct-2002	Achlas riv.	fallen trunk	
30	<i>Fibroporia citrine</i> (Bernicchia & Ryvarden) Bernicchia & Ryvarden	EP.17-A1356/11-Nov-2017	Zenio	fallen trunk	
31	<i>Fomes fomentarius</i> (L.) Fr.	EP.20-A1681/05-Jan-2020	Vori	standing trunk	
32	<i>Fuscoporia torulosa</i> (Pers.) T. Wagner	EP.04-A668/15-Oct-2004 EP.04-A672/24-Oct-2004 EP.17-A1447/26-Nov-2017 EP.19-A1677/02-Dec-2019	Evrousies Achlas riv. Achlas riv. Remata	standing trunk standing trunk standing trunk standing trunk	
33	<i>Ganoderma adspersum</i> (Schulzer) Donk	EP.14-A1264/01-Nov-2014 EP.17-A1422/24-Nov-2017	Vori Vori	standing trunk standing trunk	
34	<i>Ganoderma resinaceum</i> Boud.	EP.19-A1662/30-Nov-2019	Remata	standing trunk	
35	* <i>Gymnopilus arenophilus</i> A. Ortega & Esteve-Rav.	EP.03-A659/04-Nov-2003 EP.17-A1408/24-Nov-2017 EP.19-A1674/02-Dec-2019	Apoikia Lefka Remata	rotten stump rotten stump rotten stump	MT458512 MT458513
36	<i>Gymnopilus junonius</i> (Fr.) P.D. Orton	EP.04-A667/15-Oct-2004 EP.04-A825/02-Dec-2004 EP.17-A1385/14-Nov-2017 EP.18-A1587/05-Nov-2018	Apoikia Apoikia Evrousies Vori	standing trunk standing trunk standing trunk standing trunk	
37	<i>Gymnopus brassicolens</i> (Romagn.) Antonín & Noordel.	EP.17-A1425/24-Nov-2017 EP.19-A1673/02-Dec-2019	Vori Remata	woody residues woody residues	
38	* <i>Hydropus floccipes</i> (Fr.) Singer	EP.19-A1658/29-Nov-2019	Vori	standing trunk	
39	<i>Hyphoderma medioburiense</i> (Burt) Donk	EP.17-A1361/11-Nov-2017 EP.17-A1381/12-Nov-2017	Zenio Katakalaioi	fallen trunk twigs	MT458514

Table 2. Cont.

a/a	Species Name	Specimen Code/Collection Date	Locality	Substrate Type	GenBank Accession No.
40	<i>*Hyphoderma nemorale</i> K.H. Larss.	EP.02-A326/29-Apr-2002	Vori	fallen trunk	MT458515
41	<i>Hyphoderma setigerum</i> (Fr.) Donk	EP.02-A332/29-Apr-2002 EP.17-A1357/11-Nov-2017	Vori Zenio	fallen trunk fallen trunk	MT458516
42	<i>*Hyphodermella corrugate</i> (Fr.) J. Erikss. & Ryvarden	EP.17-A1398/23-Nov-2017	Evrousies	fallen branch	MT458517
43	<i>*Lepiota ochraceofulva</i> P.D. Orton	EP.19-A1627/15-Nov-2019	Vourkoti	leaf-litter	MT458518
44	<i>*Lepista ovispora</i> (J.E. Lange) Gulden	EP.19-A1608/12-Nov-2019	Zenio	leaf-litter	
45	<i>Lepista nuda</i> (Bull.) Cooke	EP.18-A1564/01-Nov-2018	Zenio	leaf-litter	
46	<i>Leucoagaricus melanotrichus</i> (Malençon & Bertault) Trimbach	EP.19-A1604/12-Nov-2019	Zenio	leaf-litter	
47	<i>Leucopaxillus gentianeus</i> (Qué.) Kotl.	EP.18-A1577/01-Nov-2018	Zenio	leaf-litter	
48	<i>Marasmius rotula</i> (Scop.) Fr.	EP.02-A360/22-Sep-2002	Evrousies	twigs	
49	<i>Melanoleuca excissa</i> (Fr.) Singer	EP.17-A1415/24-Nov-2017	Vori	soil	MT458519
50	<i>Merulius tremellosus</i> Schrud.	EP.04-A680/15-Nov-2004 EP.02-A323/29-Apr-2002	Evrousies Vori	fallen trunk fallen trunk	
51	<i>Mycena galericulata</i> (Scop.) Gray	EP.17-A1349/11-Nov-2017 EP.19-A1625/15-Nov-2019 EP.19-A1629/16-Nov-2019 EP.19-A1668/02-Dec-2019	Zenio Vourkoti Katakalaioi Remata	around trunk dead stump around trunk dead stump	MT458520 MT458521
52	<i>Mycena haematopus</i> (Pers.) P. Kumm.	EP.01-A262/26-Dec-2001	Vori	rotten trunk	
53	<i>Mycena pseudocorticola</i> Kühner	EP.17-A1449/26-Nov-2017 EP.19-A1667/02-Dec-2019	Achlas riv. Remata	bark living tree bark living tree	
54	<i>Mycena sanguinolenta</i> (Alb. & Schwein.) P. Kumm	EP.19-A1642/17-Nov-2019	Achlas riv.	twigs	
55	<i>Mycetinis scorodoni</i> (Fr.) A.W. Wilson & Desjardin	EP.19-A1644/17-Nov-2019	Achlas riv.	twigs	
56	<i>Mycocacia aurea</i> (Fr.) J. Erikss. & Ryvarden	EP.02-A325/29-Apr-2002 EP.11-A1026/05-Jan-2011	Vori Vori	fallen trunk fallen trunk	
57	<i>Mycocacia uda</i> (Fr.) Donk	EP.17-A1351/11-Nov-2017	Zenio	fallen trunk	
58	<i>Mycocaciella bispora</i> (Stalpers) Erikss. & Ryvarden	EP.01-A272/26-Dec-2001 EP.02-A330/29-Apr-2002	Vori Vori	fallen trunk fallen trunk	
59	<i>Paralepista flaccida</i> (Sowerby) Vizzini	EP.18-A1563/01-Nov-2018 EP.18-A1581/02-Nov-2018	Zenio Vourkoti	leaf-litter leaf-litter	
60	<i>Parasola kuehneri</i> (Uljé & Bas) Redhead, Vilgalys & Hopple	EP.18-A1546/22-Feb-2018	Vori	soil	
61	<i>Peniophora tamaricicola</i> Boidin & Malençon	EP.02-A329/29-Apr-2002	Vori	fallen trunk	
62	<i>Peniophorella praetermissa</i> (P. Karst.) K.H. Larss.	EP.01-A276/26-Dec-2001 EP.17-A1355/11-Nov-2017	Vori Zenio	fallen trunk fallen trunk	MT458522
63	<i>Perenniporia ochroleuca</i> (Berk.) Ryvarden	EP.02-A461/11-Oct-2002	Achlas riv.	dead wood	
64	<i>*Phanerochaete livescens</i> (P. Karst.) Volobuev & Spirin	EP.01-A273/26-Dec-2001 EP.18-A1541/22-Feb-2018	Vori Vori	fallen trunk fallen trunk	MT458523
65	<i>Phellinus lundellii</i> Niemelä	EP.17-A1342/13-Apr-2017	Zenio	standing trunk	
66	<i>Phlebia rufa</i> (Pers.) M.P. Christ.	EP.17-A1354/11-Nov-2017 EP.20-A1682/05-Jan-2020	Zenio Vori	fallen trunk fallen trunk	MT458524 MT458525

Table 2. Cont.

a/a	Species Name	Specimen Code/Collection Date	Locality	Substrate Type	GenBank Accession No.
67	<i>Phlebiopsis ravenelii</i> (Cooke) Hjortstam	EP.11-A1018/05-Jan-2011	Vori	fallen trunk	
68	<i>Phloeomana alba</i> (Bres.) Redhead	EP.19-A1641/17-Nov-2019	Achlas riv.	bark living tree	MT458526
69	<i>Phloeomana speirea</i> (Fr.) Redhead	EP.17-A1378/12-Nov-2017	Lezina	twigs	MT458527
70	<i>Physisporinus vitreus</i> (Pers.) P. Karst.	EP.20-A1684/05-Jan-2020	Vori	fallen trunk	
71	<i>Pilatotrampa ljubarskyi</i> (Pilát) Zmitrovich	EP.18-A1551/22-Feb-2018	Vori	fallen trunk	MT458528
72	<i>Pleurotus ostreatus</i> (Jacq.) P. Kumm.	EP.01-A261/26-Dec-2001 EP.18-A1586/05-Nov-2018	Vori Vori	standing trunk standing trunk	
73	<i>Pluteus cervinus</i> (Schaeff.) P. Kumm.	EP.01-A321/29-Apr-2002 EP.19-A1623/15-Nov-2019 EP.19-A1676/02-Dec-2019	Vori Vourkoti Remata	dead wood dead wood fallen trunk	MT458529 MT458530
74	<i>Pluteus nanus</i> (Pers.) P. Kumm.	EP.19-A1653/29-Nov-2019	Vori	woody residues	
75	<i>Pluteus salicinus</i> (Pers.) P. Kumm.	EP.19-A1657/29-Nov-2019	Vori	fallen branch	
76	* <i>Pluteus podospileus</i> Sacc. & Cub.	EP.19-A1643/17-Nov-2019	Achlas riv.	fallen twigs	
77	<i>Postia balsamea</i> (Peck) Jülich	EP.19-A1663/30-Nov-2019	Remata	fallen trunk	
78	<i>Psathyrella candolleana</i> (Fr.) Maire	EP.02-A336/04-Jun-2002 EP.19-A1624/15-Nov-2019	Achlas riv. Vourkoti	woody residues woody residues	MT458531
79	<i>Psathyrella corrugis</i> (Pers.) Konrad & Maubl.	EP.19-A1601/16-Oct-2019	Vourkoti	soil/buried wood	MT458532
80	* <i>Psathyrella hellebosensis</i> Deschuyteneer & A. Melzer	EP.17-A1409/24-Nov-2017	Lefka	woody residues	MT458533
81	<i>Psathyrella microrhiza</i> (Lasch) Konrad & Maubl.	EP.19-A1603/12-Nov-2019	Vourkoti	woody residues	MT458534
82	<i>Psathyrella prona</i> (Fr.) Gill.	EP.02-A363/22-Sep-2002	Evrousies	soil	
83	<i>Psathyrella vinosofulva</i> P.D. Orton	EP.17-A1431/24-Nov-2017	Vori	soil	MT458535
84	<i>Radulomyces confluens</i> (Fr.) M.P. Christ.	EP.11-A1022/05-Jan-2011	Vori	fallen trunk	
85	<i>Steccherinum ochraceum</i> (Pers. ex J.F. Gmel.) Gray	EP.18-A1540/22-Feb-2018	Vori	fallen trunk	
86	<i>Stereum hirsutum</i> (Willd.) Pers.	EP.17-A1397/23-Nov-2017 EP.17-A1463/28-Nov-2017 EP.18-A1544/22-Feb-2018 EP.20-A1691/25-Jan-2020	Evrousies Achlas riv. Vori Lefka	trunk/branch trunk/branch trunk branch	
87	<i>Trechispora nivea</i> (Pers.) K.H. Larss.	EP.02-A328/29-Apr-2002 EP.20-A1683/05-Jan-2020	Vori Vori	fallen trunk fallen trunk	MT458536
88	<i>Trametes versicolor</i> (L.) Lloyd	EP.11-A1019/05-Jan-2011 EP.18-A1542/22-Feb-2018	Vori Vori	fallen trunk standing trunk	
89	<i>Tubaria furfuracea</i> (Pers.) Gillet	EP.19-A1652/29-Nov-2019	Vori	woody residues	
90	<i>Tulostoma fimbriatum</i> Fr.	EP.17-A1431/24-Nov-2017	Vori	soil	
91	<i>Vitreoporus dichrous</i> (Fr.) Zmitr.	EP.02-A445/11-Oct-2002 EP.17-A1439/25-Nov-2017 EP.18-A1538/22-Feb-2017	Achlas riv. Vori Vori	fallen trunk fallen branch fallen branch	
92	<i>Xyloдон raduloides</i> Riebesehl & Langer	EP.11-A1025/05-Jan-2011 EP.18-A1543/22-Feb-2018 EP.19-A1671/02-Dec-2019	Vori Vori Remata	fallen trunk fallen trunk fallen trunk	MT458538

3.1. The ECM Element

Among the ECM macrofungi recorded (Table 1), the genus *Alnicola* is represented by six species (Figure 3); five of them form part of the sect. *Alnicola* sensu Moreau [45], which is characterized by urticoid cheilocystidia, and one of the sect. *Submelinoideae* Singer with clavate or capitate cheilocystidia [46].



Figure 3. Species of the genus *Alnicola* recorded in Andros alder stands: *A. escharoides* basidiomes, basidiospores and cheilocystidia (a–c); *A. umbrina* basidiomes, basidiospores and cheilocystidia (d–f); *A. striatula* basidiomes, basidiospores and cheilocystidia (g–i); *A. subconspersa* basidiomes, basidiospores and cheilocystidia (j–l); *A. luteolofibrillosa* basidiomes, basidiospores and cheilocystidia (m–o); *A. inculta* basidiomes, basidiospores, basidia and cheilocystidia (p–s). Bars: basidiomes, 1 mm; basidiospores and basidia, 10 μ m; cheilocystidia, 20 μ m.

Different opinions exist regarding the genus name in pertinent literature since some European authors as well as the Index Fungorum prefer to conserve the name *Naucoria* (Fr.) P. Kumm., whereas Moreau, in his nomenclatural revision, rejected this name in favour of *Alnicola* Kühner [45]; the latter approach is accepted by other European mycologists, the Mycobank, and is also adopted in this work. Moreover, the taxonomy of species of the sect. *Alnicola* remains problematic and, consequently, a phylogenetic analysis was performed to deal with this issue.

The most often found *Alnicola* species in our study were *A. umbrina* (R. Maire) Kühner and *A. escharoides* (Fr.) Romagn., i.e., two of the most common taxa associated with alders in Europe; both constitute new national records for Greece. Particularly *A. escharoides* (syn. *A. citrinella* Moreau & A. de Haan [47]) is distinguished from all other (more or less brownish) species found in Andros by its pale yellowish-buff non striate pileus, the amygdaliform to navicular spores, with prominent ornamentation, measuring $9.9\text{--}11.8 \times 5.3\text{--}5.9 \mu\text{m}$, $Q = 1.9\text{--}2.1$ (Figure 3a–c). Following phylogenetic analysis, our specimens are positioned in a distinct group (albeit not adequately supported) together with other sequences from material identified as *A. escharoides* and *A. citrinella* (Figure 4).

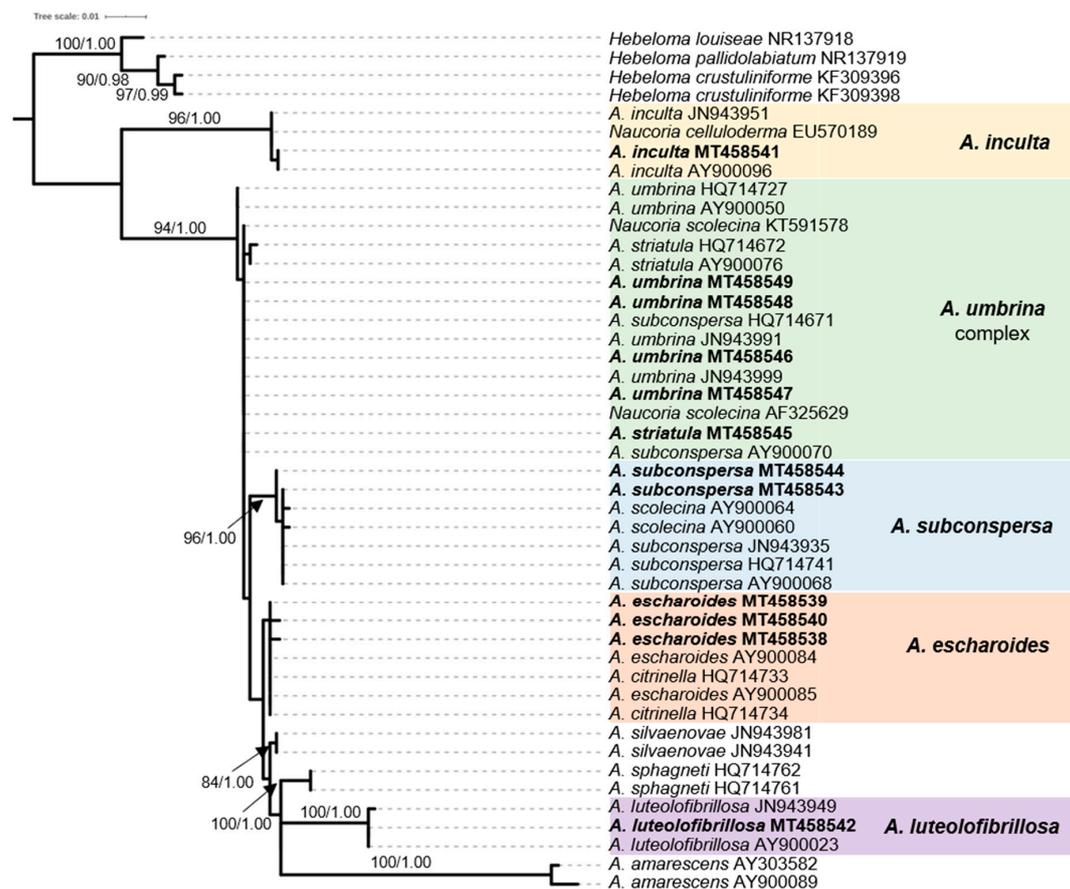


Figure 4. Phylogeny of *Alnicola* species derived from rDNA ITS sequences through ML analysis. Branches are labelled when MLB > 65% and BPP > 0.95. *Hebeloma* species (*H. louiseae*, *H. pallidolabiatum*, *H. crustuliniforme*) were used as outgroups. Boxes include sequences from specimens recorded in the *Alnus glutinosa* habitat.

On the other hand, *A. umbrina* (Figure 3d–f) is hereby considered as a species complex following the nomenclatural concept of Moreau [45] and the outcome of the phylogenetic study by Rochet et al. [19]. According to our observations, *A. umbrina* shows a rather large morphological variability with dark brown hygrophanous pilei bearing prominent striations up to their centre when wet, becoming much lighter and indistinctly striate only at margin when dry. Basidiospores are variable in size and shape,

often somewhat elongated fusiform, weakly to moderately verrucose, measuring $10.7\text{--}13.6 \times 5.2\text{--}6.1 \mu\text{m}$, $Q = 1.9\text{--}2.4$. Sequences generated in this work clustered together with material identified as *A. umbrina*, *N. scolecina* (Fr.) Quél., *A. striatula* (P.D. Orton) Romagn. and *A. subconspersa* (Kühner ex P.D. Orton) Bon into a group that was not adequately supported (Figure 4). However, the morphological features of specimens identified as *N. scolecina* in Europe are very similar to descriptions of *A. umbrina* [22,33,48,49]. Therefore, *N. scolecina* and *A. umbrina* form part of the same complex and the question whether they constitute different entities or not remains open and in need of further research.

One collection representing another closely related taxon, previously reported as *N. striatula* P.D. Orton (Figure 3g–i) from alder stands in Andros [22], derived from the same site during our recent forays. According to Moreau (2005), *A. striatula* might merely correspond to a pale form of *A. umbrina*, but our morphological studies revealed some noteworthy differences when compared to specimens hereby named *A. umbrina*, i.e., pileus always very prominently striate, smooth and shiny, and (most importantly) significantly smaller basidiospores measuring $8.2\text{--}10.0 \times 4.5\text{--}5.6 \mu\text{m}$, $Q = 1.7\text{--}1.9$; these features are in accordance to previous descriptions of *N. striatula* [33,48,50]. As evidenced from our phylogenetic analysis (Figure 4), this particular collection forms part of the *A. umbrina* complex (together with the other two *A. striatula* sequences included in the tree) by using ITS alone; however, since it is morphologically distinct and fits to the widely accepted taxonomic concept of *A. striatula*, we provisionally retain it in this inventory as a separate taxon, until a future multigene approach shows otherwise.

A similar looking species to *A. umbrina*—but less common in Andros—is *A. subconspersa* (Figure 3j–l). The most prominent distinguishing features versus our *A. umbrina* specimens are the non (or very faintly) striate pileus as well as the size and shape of spores, being wider, amygdaliform to navicular and more prominently ornamented, measuring $10.9\text{--}12.9 \times 6.0\text{--}6.8 \mu\text{m}$, $Q = 1.7\text{--}2.0$. It is noteworthy that *A. subconspersa* forms a well-supported phylogenetic group including sequences labelled as *A. scolecina* (Fr.) Romagn. (Figure 4), which is indicative of the morphological affinity of these taxa that had apparently led to the development of ambiguous species concepts.

Another collection representing a member of the sect. *Alnicola* was recorded in the alluvial littoral forest of Vori; it corresponds to *A. luteolofibrillosa* Kühner and constitutes the first report of this species in Greece (Figure 3m–o). It is morphologically characterized by non-striate, pale buff, fibrillose to tomentose pilei, with abundant whitish veil remnants on stipe and pileal margin; the respective sequence falls within a highly-supported terminal subgroup corresponding to this species (Figure 4). Lastly, *A. inculta* (Peck) Singer (Figure 3p–s) was recorded only at Zenio (i.e., the site with the highest altitude among those of this study, 850 m) and is reported for the first time in Greece. It forms part of the sect. *Submelinoideae*, and, according to Moreau [45] is conspecific to the taxon widely referred as *N. celluloderma* P.D. Orton, as it is also evidenced by our phylogenetic analysis (Figure 4). Morphologically, this species is easily distinguished from all aforementioned taxa thanks to the clavate to capitate cheilocystidia characterizing members of sect. *Submelinoideae* and the 2-spored basidia.

The most common ECM mushroom in alder stands of Andros belongs to the genus *Paxillus*; it was the first recorded *Alnus*-specific symbiont in the island 25 years ago, and was later repeatedly found in this particular habitat (Figure 5a–d). It was initially identified as *P. rubicundulus* P.D. Orton [22]; however, sequencing of recent collections revealed that it forms part of the newly described taxon *P. olivellus* Moreau P-A, Chaumeton J-P, Gryta H, Jargeat P [51]. Although clearly separated by molecular approaches, *P. olivellus* can be hardly distinguished from *P. rubicundulus* and *P. adelphus* Chaumeton JP, Gryta H, Jargeat P, Moreau P-A on the basis of morphology alone, i.e., only by the olivaceous tinges of the young basidiomes and the basidiospores shape, which are ovoid to ellipsoid in *P. olivellus*, cylindrical in *P. rubicundulus* and short cylindrical in *P. adelphus* [51]. Such features were observed in our specimens since olivaceous tints were always evident in young basidiomes, and spores were ovoid to ellipsoid measuring $6.7\text{--}8.1 \times 4.5\text{--}5.2 \mu\text{m}$, $Q = 1.39\text{--}1.66$. In addition, ITS sequences from our material originating from various sites in the habitat under study were very similar or identical to those corresponding to *P. olivellus* (including the type), and formed a terminal subgroup with high

support (Figure 6). Therefore, this particular species seems to be the only representative of the genus *Paxillus* in the black alder stands of Andros island.

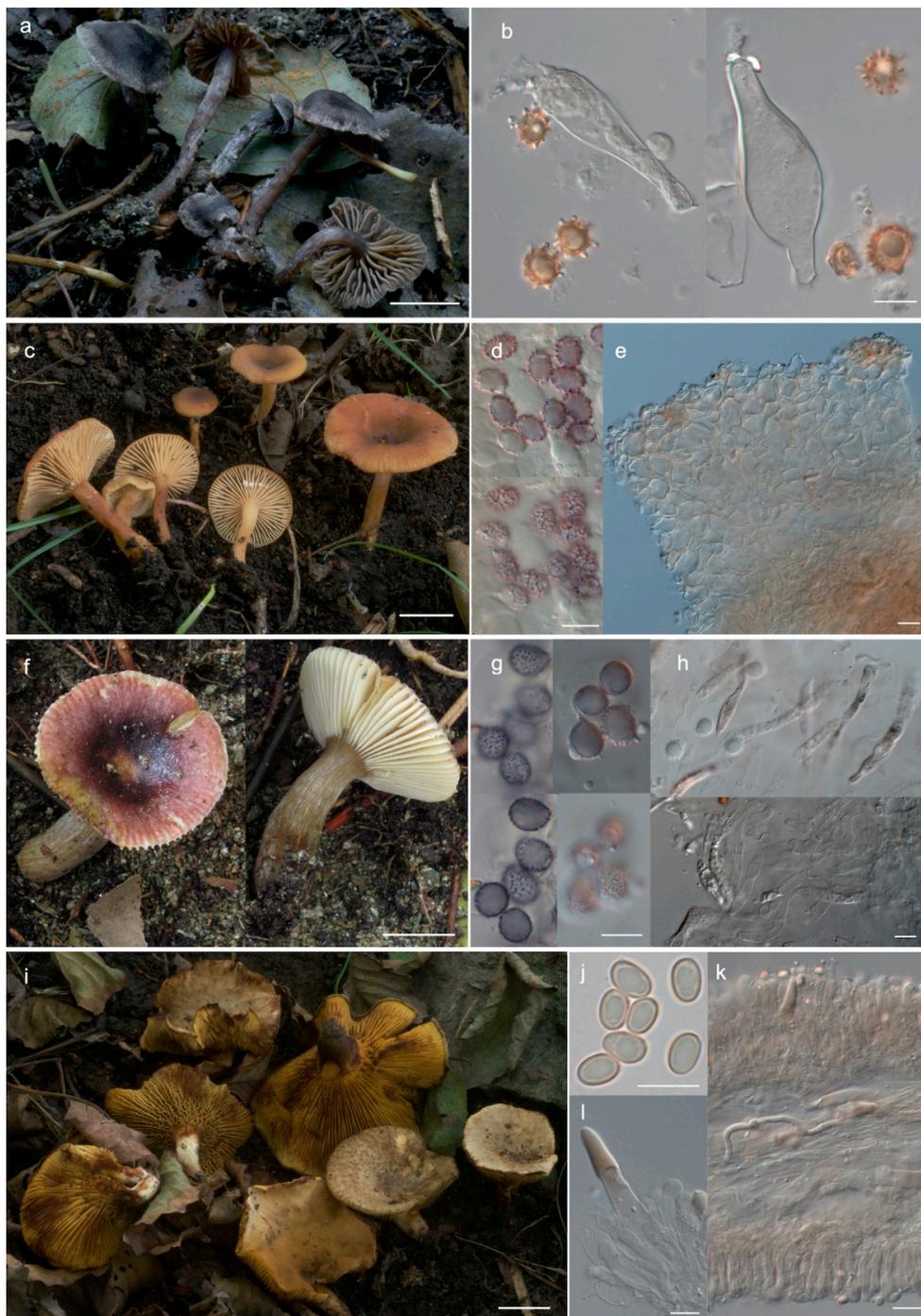


Figure 5. Alder-associated ECM fungi recorded in Andros: *Paxillus olivellus* basidiomes (a; bar 1 mm), basidiospores (b; bar 10 μ m), section of lamella (c; bar 20 μ m), hymental cystidium and basidia (d; bar 20 μ m); *Lactarius obscuratus* basidiomes (e; bar 1 mm), basidiospores (f; bar 10 μ m), pileipellis (g; bar 20 μ m); *Russula pumila* basidiomes (h; bar 1 mm), basidiospores (i; bar 10 μ m), pileipellis (j; bar 20 μ m); *Cortinarius americanus* basidiomes (k; bar 1 mm); *Inocybe calospora* basidiospores, basidium and pleurocystidium (l; bar 10 μ m).

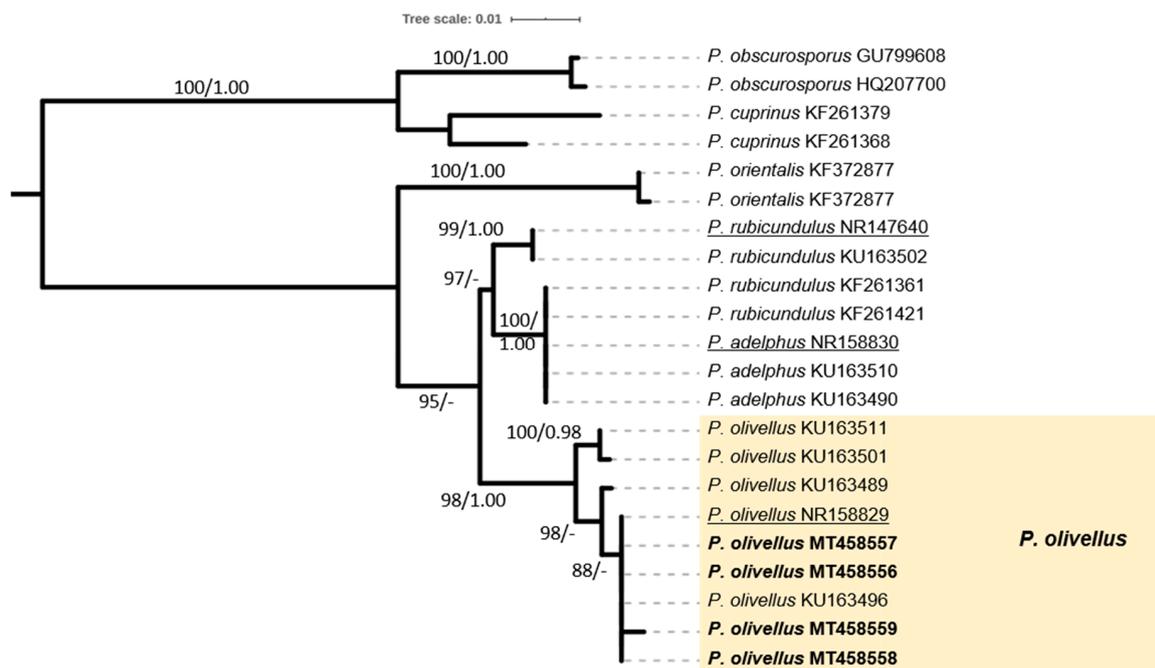


Figure 6. Phylogeny of *Paxillus* species derived from rDNA ITS sequences through ML analysis. Branches are labelled when MLB > 65%, and BPP > 0.95. *P. cuprinus* and *P. obscuroporus* were used as outgroups. The coloured box includes sequences from specimens recorded in the *Alnus glutinosa* habitat.

Lactarius obscuratus (Lasch) Fr. is one of the few *Alnus*-specific ECM symbionts of this particular genus; it was found in several inland collection sites dominated by *A. glutinosa*, but not in the alluvial (littoral) forest of Vori (Figure 5e–g). Phylogenetic analysis of our sequences derived from several collections confirmed that they belong to this particular species (Figure 7). However, the respective terminal subgroup in our phylogenetic tree is composed of sequences named either *L. obscuratus* or *L. cyathuliformis* Bon, which is due to the different interpretations existing about this taxon (J. Nuytinck, pers. comm.). In the study of Rochet et al. [19], it is referred to as *L. cyathuliformis*, whereas the correct name for the same group is *L. obscuratus* according to Wisitrassameewong et al. [52]. Since the basidiospores average size in our collections (measuring $7.6\text{--}8.5 \times 6.1\text{--}6.3 \mu\text{m}$) is in agreement with the concept of *L. obscuratus* (sensu Heilmann-Clausen et al. [31]; according to the same authors, spores of *L. cyathuliformis* have an average size of $8.3\text{--}9.9 \times 7.0\text{--}7.7 \mu\text{m}$), we adopt this name for the specimens included in this work. The genus *Russula* Pers. is represented by *R. pumila* Rouzeau & F. Massart (Figure 5h–j) detected in one site only (Vourkoti). Although *R. pumila* is synonymous to *R. alnetorum* Romagn. according to both Index Fungorum and Mycobank, there are different opinions about the synonymy of these two taxa and to the best of our knowledge this issue has not been resolved yet (S. Adamcik, pers. comm.). *R. pumila* is reported to occur mainly in lowlands with *A. glutinosa*, as opposed to *R. alnetorum*, which is mostly recorded in subalpine habitats with *A. viridis* [53,54]; therefore, we adopt the use of the former name.

Tomentella sublilacina (Ellis & Holw.) Wakef. and *T. stuposa* (Link) Stalpers were previously recorded in Andros (in the alluvial alder forest of Vori; [20]) and identified on the basis of their morphology. These names are provisionally retained here due to the absence of precise taxonomic information concerning alder-specific *Tomentella* species. It should be noted that the ITS sequences generated in the frame of this work represent phylogenetically distinct taxa corresponding to entities named “*aff. sublilacina*” and “*aff. stuposa*” in previous studies referring to material originating from alder hosts only [18,55].

It is noteworthy that the first ever report of an alniphilous *Cortinari* species in Greece derives from a single collection of *C. americanus* A.H. Sm. (Figure 5k), which forms part of a small group of

species within the subgenus *Telamonia* (Fr.) Trog known to be associated with *A. glutinosa* and *A. incana* in Europe [33,56]. *C. americanus* is characterized by the minute size, pileus not exceeding 2 cm in diameter, with dark violet to blackish colour, and spores smaller than $10 \times 6 \mu\text{m}$ [33,56]; our collection has spores measuring $7.6\text{--}8.4 \times 4.9\text{--}5.6 \mu\text{m}$.

Last, *Inocybe calospora* Quél., recorded for the first time in Greece, was collected in Vourkoti only; it is an easily identified species thanks to its unique star-shaped spiny spores (Figure 5). Among all ECM species included in this inventory it is the only one which is not considered to be exclusively associated with alders, and reported from diverse damp deciduous forests of Europe [33,56].

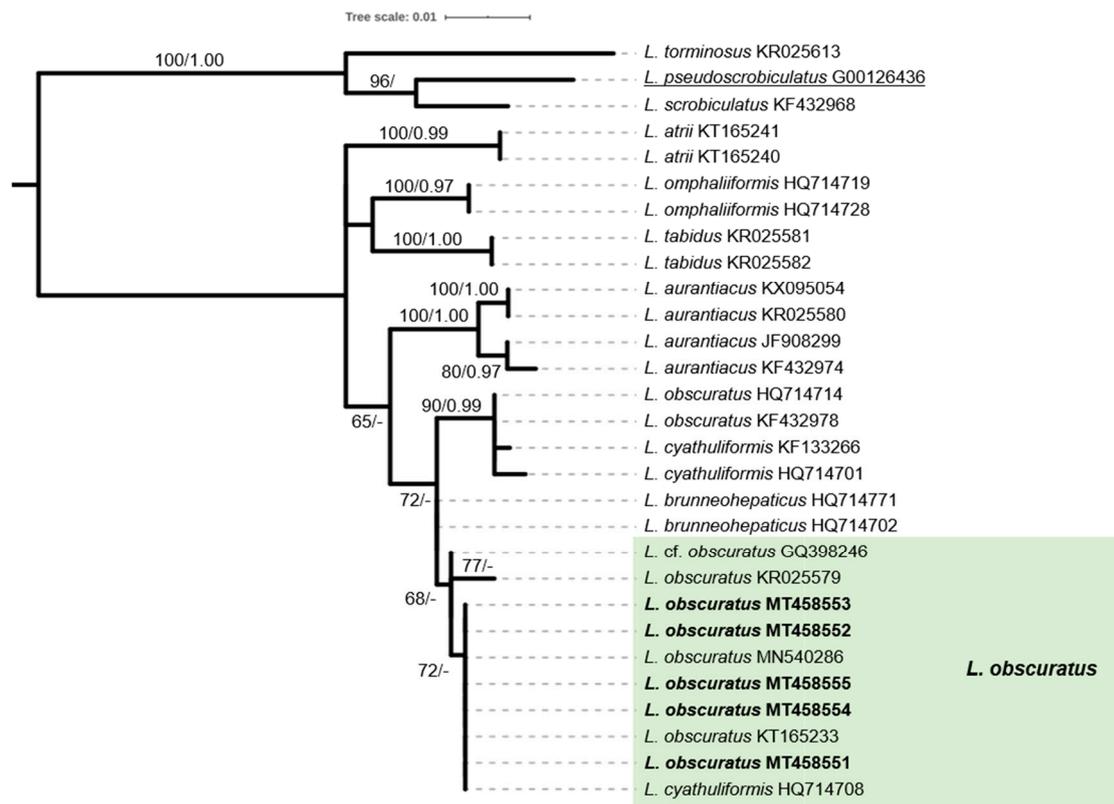


Figure 7. Phylogeny of *Lactarius* species derived from rDNA ITS sequences through ML analysis. Branches are labelled when MLB > 65%, and BPP > 0.95. *L. torminosus*, *L. scrobiculatus* and *L. pseudoscrobiculatus* were used as outgroup. The colored box includes sequences from specimens recorded in the *Alnus glutinosa* habitat.

3.2. The Saproxylic Element

By far the highest number of species recorded in this inventory correspond to white-rot and brown-rot basidiomycetes found on various wood parts of *A. glutinosa* (Table 2). This is quite anticipated since alder trees have a life-span which rarely exceeds 100 years; therefore, they produce large amounts of dead wood. Moreover, in contrast to ECM species, wood-rotting fungi do not show any specificity to alders, with only few exceptions including the common in northern Europe plant-pathogenic polypore *Inonotus radiatus* (Sowerby) P. Karst. [15,57–60], which, however, was not among our findings. Most of the recorded species are wood rotting basidiomycetes that are quite common throughout Europe on deciduous tree species including alders [13,61,62]. The best represented genera of saproxylic fungi were *Hyphoderma* Wallr. (three spp.), *Mycena* (Pers.) Roussel (four spp.), *Pluteus* Fr. (four spp.) and *Psathyrella* (Fr.) Quél. (four spp. on woody residues or buried wood). In total, seven species recorded on dead wood or bark of living alder trees are recorded for the first time in Greece and are presented in more detail below.

Among corticioid basidiomycetes, *Hyphoderma nemorale* K.H. Larss. is a distinct, widely distributed but rare species in Europe [35], which was identified by re-examining an old collection from Vori alluvial forest and further confirmed by ITS sequencing. The presence of thick-walled subicular hyphae and of two types of hymenial cystidia (i.e., short ventricose and subcapitate, and more seldom long tubular with characteristic constrictions, sometimes moniliform, which was the case in our specimen) are the main diagnostic features of this species [63]. *Hyphodermella corrugata* (Fr.) J. Erikss. & Ryvarden (Figure 8a) is fairly common and widespread in Europe [35]. It is easily identified thanks to its characteristic cystidioid hyphal ends appearing in bundles that are heavily incrustated [24]. *Phanerochaete livescens* (P. Karst.) Volobuev & Spirin (Figure 8b) is a species closely related to *Ph. sordida* (P. Karst.) J. Erikss. & Ryvarden which was recently described by using both morphological and phylogenetic criteria [64]. In accordance to the pertinent description, our specimens possessed cystidia with thickened walls to the acute apex, densely covered by crystals, as opposed to the accidentally encrusted, obtuse and thin-walled towards the apex cystidia of *Ph. sordida*. Moreover, the identity of our specimen was confirmed by the respective ITS sequence which was identical to those of *Ph. livescens* as determined by Volobuev et al. [64].



Figure 8. Saproxylic species recorded on *Alnus glutinosa* wood and litter in Andros: *Hyphodermella corrugata* (a) *Phanerochaete livescens* (b) *Pluteus podospileus* (c) *Delicatula integrella* (d) *Coprinopsis melanthina* (e) *Gymnopilus arenophilus* (f) *Hydropus floccipes* (g) *Lepiota ochraceofulva* (h) *Lepista ovispora* (i) *Psathyrella hellebosensis* (j) *Entoloma uranochroum* (k) Bar: 1 mm.

Among the agaricoid wood-inhabiting fungi, the genus *Pluteus* Fr. is hereby represented by four species, of which *P. podospileus* Sacc. & Cub. (Figure 8c) is reported for the first time in Greece. It belongs to the sect. *Celluloderma* Fay. subsection *Mixtini* Sing. ex Sing, and possesses a pileipellis made up of both fusiform and broadly clavate elements. *P. thomsonii* (Berk. & Broome) Dennis is very similar morphologically but it differs in the absence of pleurocystidia and the shape of cheilocystidia, which are characteristically rostrate [65]. The record of *Delicatula integrella* (Pers.) Rat. (Figure 8d) is worth mentioning since it was reported only once before in Greece, in the content of a regional field-guide [66]. *D. integrella* forms whitish-mycenoid mushrooms of minute size with pileus diameter measuring (in our specimens) 0.4–0.6 cm, reduced, almost vein-like lamellae and non-amyloid, amygdaliform-fusoid spores. It is considered widespread and common in Europe and N. America, and grows on decaying wood and wood debris of deciduous trees [34]. *Coprinopsis melanthina* (Fr.) Örstadius & E. Larss. (Figure 8e) is a striking-looking psathyrelloid species, easily identified due to the relatively large basidiomes with woolly to squamulose pileus (measuring 2–6 cm in diam.) and stipe (up to 6.0 × 0.8 cm), absence of pleurocystidia, almost colourless basidiospores, devoid of germ-pore, measuring 9.8–11.8(13.5) × 5.6–6.5 µm in our collections. This is a rather uncommon European species growing on and around rotten stumps in humid deciduous forests [33]; the only other record of this species in Greece derives from Crete (G. Konstandinidis, pers. comm.).

Gymnopilus arenophilus A. Ortega & Esteve-Rav. (Figure 8f) was described from continental areas of Spain [67] and from maritime dunes in France, under or near Mediterranean pines, on sandy soil by being attached to wood debris or wood, often burnt or buried in the sand [68]. Two of our collections from the alluvial *A. glutinosa* habitat at Lefka were sequenced and found to correspond to this species. Apparently, no native pines exist in Andros while both specimens were growing on rotten alder stumps, a fact that largely expands the so far known ecological and geographical range of this Mediterranean species. Morphologically, our specimens possessed features that fit well to the taxonomic concept of *G. arenophilus* i.e., smooth to fibrillose pileal surface, bitter taste, ellipsoid to subamygdaliform, moderately verrucose spores, measuring 8–10 × 5.5–6.5 µm, lageniform cheilocystidia often with subcapitate apex, 25–45 × 5–8 µm and absence of pleurocystidia. On the other hand, the size of basidiomes was significantly larger, with pilei up to 10 cm in diam. and a sturdy stipe often thicker than 1 cm. One previous collection of ours also found on rotten alder stump, and originally identified as *G. picreus* (Pers.) P. Karst. [22], is now re-assessed as *G. arenophilus*.

Hydropus floccipes (Fr.) Singer (Figure 8g) is a rare mycenoid species generally found to grow on decayed trunks of deciduous trees in damp forests, and is characterized by non-amyloid, subglobose spores, not blackening basidiomes, and typically scabrous stipe with grey-brown spots [33,69]. In addition, our specimens possessed yellowish stipe, previously reported for *H. floccipes* var. *luteipes* Ortega & Zea described from Spain [70]; the latter is otherwise microscopically identical and of unknown phylogenetic status. Two unpublished reports of *H. floccipes* exist from Greece (D. Sofronis and G. Konstandinidis, pers. comm.).

3.3. Litter and Other Terrestrial Decomposers

Apart ECM and saproxylic fungi, several other mushroom species were recorded under black alder trees, and therefore constitute a part of the fungal diversity of the *A. glutinosa* priority habitat in Andros. Needless to say, none of these species is specifically linked to alders; instead they are considered ‘generalists’ to be found in both deciduous and coniferous forests. Among them, the following four species are recorded for the first time in Greece. *Lepiota ochraceofulva* P.D. Orton (Figure 8h) is a rather rare (but widespread) species in Europe, reported from *Fagus* and other deciduous trees on humus-rich, loamy soil [71,72]; it forms highly toxic mushrooms containing amanitins. Our single collection consisted of few basidiomes growing on a thick layer of leaf-litter under *A. glutinosa*. They are characterized by pilei of up to 7 cm in diam., with orange-brown scales; lamellae forming a distinct collarium and reddish-orange in maturity; spores ellipsoid to oblong, dextrinoid, not metachromatic in cresyl-blue, measuring 5.4–7.5(8.1) × 3.5–4(4.5) µm; basidia (2)4-spored, clamped; cheilocystidia short

clavate to cylindrical, rarely papillate, often in chains; pileipellis, a hymeniderm, composed of more or less clavate elements up to 50 μm long.

Lepista ovispora (J.E. Lange) Gulden (Figure 8i) is an uncommon (albeit widespread) European species recorded only once during this study on leaf litter under alders. Typical diagnostic features are the densely caespitose habit, the relatively fleshy basidiomes, the brown hygrophanous pileus with pruinose surface [73]. In addition, our specimens possessed spores ovoid to broadly ellipsoid, finely punctate, $4.7\text{--}6.8 \times 3.8\text{--}4.4 \mu\text{m}$, clamped basidia and no cystidia. *Psathyrella hellebosensis* D. Deschuyteneer, A. Melzer (Figure 8j) was recently described from Belgium [74] and was later reported from riparian alder habitats in Italy [75]. The morphological features of our collection are in agreement with the morphology of Belgian and Italian basidiomes, but since it corresponds to a rarely reported species, a detailed description of our material is hereby provided: pileus up to 3 cm in diam., hygrophanous from dark reddish-brown to greyish-beige, with scanty remains of veil; lamellae subdistant with whitish edge; stipe $2\text{--}4 \times 0.2\text{--}0.3$ cm, not rooting; spores $7.3\text{--}8.7 \times 4.5\text{--}5.7 \mu\text{m}$, $Q = 1.43\text{--}1.73$, ovoid to angular in face-view and not or weakly phaseoliform in side-view, not opaque; lamellae edge sterile composed exclusively of sphaeropendunculate paracystidia (no pleurocystidioid paracystidia were observed); pleurocystidia $34\text{--}48 \times 11\text{--}17 \mu\text{m}$, utriform. The material was collected from wet soil by the alluvial stream banks. This species shows high phylogenetic affinity to *P. thujina* A. H. Sm. by using ITS sequences only; however, it is clearly separated when the *tef-1 α* marker is added in the phylogenetic analysis, while it is also distinguished by its distinctly larger and prominently phaseoliform spores [75].

Previous studies on the mycodiversity of Andros island reported the occurrence of four *Entoloma* species, one of them was new to science, i.e., *E. alnicola* Noordel. & Polemis [22,25]. Our recent field work resulted in other interesting collections of *Entoloma* spp. for which the identity and phylogenetic relationships to closely allied taxa are still under investigation. However, by using morphology alone, the presence of a rare European species was confirmed, namely *E. uranochroum* Hauskn. & Noordel. (Figure 8k) recorded for the first time in an alder habitat. This beautiful dark blue-violet mushroom was so far reported from subalpine meadows on calcareous soil in Austria (type locality) and the French Alps. Moreover, its striking microscopical features, e.g., the large fusiform cheilocystidia with granular yellowish-brown content, place it in the distinct section *Ramphocystotae* (Largent) Noordel., together with only one other European representative, namely *E. rhynchocystidium* Noordel. & Liiv [76].

4. Conclusions

A long-term study of the diversity of macrofungi in alder stands of Andros resulted in an inventory consisting of 106 species of basidiomycetes, including 21 taxa recorded for the first time in Greece. The majority of findings corresponded to saprotrophs (#92, mainly wood-rotting fungi) and the rest were ECM species. Considering the limited size of the area under study in a small Aegean island, the outcome of this work in terms of the number of taxa and variability is indicative of the wealth of the *A. glutinosa* priority habitat. However, the black alder stands in Andros have suffered considerably from floods in the past (as a consequence of fires that destroyed vegetation in the surrounding mountains which acted as a physical barrier protecting from downhill water runoffs) and their regeneration is hindered due to grazing by feral goats. The importance of fungi in the conservation/restoration of such natural habitats was demonstrated in the past [77,78], and recent activities focus at improving the status of the degenerated alder stands by exploiting indigenous ECM fungi as inoculants to young alder seedlings prior to their transplantation on site. Moreover, new knowledge about mushroom diversity and the ecological role of this group of organisms seems to enhance considerably people's perception and awareness, and hence facilitates implementation of conservations actions which are currently under way in selected alder stands of Andros.

Supplementary Materials: The following is available online at <http://www.mdpi.com/1424-2818/12/6/232/s1>, Table S1: Details of the 10 sampling sites in Andros island from where basidiomes were collected: locality name, coordinates, altitude (m a.s.l.) and surface of the study area (m²).

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