What’s Inside the Hole? A Review of European Dendrolimnetic Moth Flies (Diptera: Psychodidae: Psychodinae)

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Abstract: We conducted an extensive literature review in search of records of dendrolimnetic Psychodinae, with additional field sampling of the European species of Psychodinae associated with water-filled tree holes. After checking more than 100 publications, only 11 specific published records involving dendrolimnetic Psychodinae were found. Our results show that six genera, represented by 13 species of Psychodinae, are associated with 13 species of plant trees. As a result of our field sampling, we report Lepiseodina latipennis (Sará, 1953) and Telmatoscopus bartai (Ježek, 2004) comb. nov. for the first time in Germany. Furthermore, we redescribe L. latipennis based on freshly collected material with a closer examination of the holotype. Derived from our findings, we review the genera Lepiseodina Enderlein, 1937 and Telmatoscopus Eaton, 1904, providing an identification key for the males of both genera. In addition, we synonymize Krivosheinoscopus Ježek, 2001 syn. nov. under Telmatoscopus, changing combination of Telmatoscopus ussuricus (Ježek, 2001) comb. nov. and Telmatoscopus bartai (Ježek, 1994), additionally, we change combination and a sononymy of Tematoscopus wagneri (Salmanna, 1982) comb. et syn. nov. under Telmatoscopus advena (Eaton, 1893). Furthermore, we describe for the first time the female and eggs of Telmatoscopus advena. Moreover, we provide the first published DNA barcodes (COI) for Telmatoscopus bartai, Lepiseodina latipennis (Sará, 1953), Lepiseodina rothschildi (Eaton, 1912), and Lepiseodina tristis (Meigen, 1830). Finally, we also discuss the taxonomy and ecology of the European dendrolimnetic species of the subfamily Psychodinae.

Keywords: water-filled tree holes; DNA barcoding; moth flies; new taxa; integrative taxonomy

1. Introduction

Phytotelmata (singular phytotelma) derives from the Greek words “phyto”, which means plant, and “telma”, meaning pond, and was first proposed by Varga [1], based on his observations, to define the microhabitats product of water-filled bodies in plant surfaces [2]. Within the phytotelmata, we can find the dendrotelmata (singular dendrotelma), deriving from the Greek words “dendron”, meaning tree, and “telma”, which circumscribe the body waters only to water-filled tree holes. These dendrotelmata are considered an integral microhabitat inside forest ecosystems, as they provide a suitable space for development, prey, and water sources for many organisms ranging from invertebrates to vertebrates [3,4]. Species that relate to or inhabit these water-filled tree holes are known as dendrolimnetic, from the Greek words “dendron”, and “limnētês”, which means relating to or inhabiting the open water of a body of fresh water.

Dendrotelmata are commonly found in old trees, dead or alive, and are a considered crucial component of ecosystems, especially with the modern forestry management where the number of mature and over-mature trees (commonly referred as old trees) has decreased [5]. The main sources of nutrients flowing in the tree holes consist of leaf litter and arthropod cadavers, while the quality and composition of these nutrients vary across species.
and habitats [6,7]. The fact that these microhabitats have a small size, discrete boundaries, and are naturally replicated in nature, makes them attractive for studies of community structure and functionalities [7]. Researchers have reported the usage of dendrotelmata by amphibians during development, as a water source for reptiles and small mammals, and, as bathing sites for birds and bats [4]. Nonetheless, mainly invertebrates with aquatic development have been reported developing inside these microhabitats, with some reports of other invertebrates that use them as a water source [4,8]. Among them, the most common organisms found inside are immature stages of Diptera and Coleoptera [7–10].

Moth flies (Insecta: Diptera: Psychodidae) are commonly found in water bodies, as the majority of psychodid species develop in water during larval stages, with a few exceptions that develop in soil, dung, or fungi [11–13]. There are six subfamilies recognized worldwide, namely Bruchomyiinae, Horaiellinae, Phlebotomininae, Psychodinae, Sycoracinae, and Trichomyiinae; all of them except Horaiellinae are present in Europe [14,15]. Species in the subfamily Trichomyiinae develop inside tree holes or rotting wood and their larvae are considered xylophagous (from Greek “xylon”, meaning wood, and “faguein”, meaning eating) [5]. Species of Bruchomyiinae and Phlebotomininae have been reported developing on the ground and leaf litter feeding on decaying organic matter [15]. The larval stages of Sycoracinae species have been found developing in aquatic mosses and leaf litter [15]. On the contrary, species of the subfamily Psychodinae are commonly reported developing in water bodies such as ponds and streams, but there are a few genera present in Europe whose larvae develop inside of tree holes, namely Clognia, Clytocerus, Lepiseodina, Pneumia, Psychoda, and Telmatoscopus [13,16–25]. Although the larval development and habitat for some European species are known, the moth flies associated with water-filled tree holes have been poorly studied in Europe [25].

In the present study, we revise the available literature about the European species of the subfamily Psychodinae known to develop in dendrotelmata, with a special emphasis on the genera Lepiseodina and Telmatoscopus. Additionally, we study the dendrolimnetic moth flies collected in Germany resulting from the German Barcode of Life (GBOL) project [26] (www.bolgermany.de, accessed on 02 April 2022). As an outcome, we redescribe Lepiseodina latipennis (Sarà, 1953) from Germany (Nordrhein-Westfalen) and Italy (Sicily) based on new morphological and molecular data, and we synonymize Kriosheinoscopus Ježek, 2001 syn. nov. under Telmatoscopus Eaton, 1904. Furthermore, we change combination and synonymize Telmatoscopus vogneri (Salamanna, 1982) comb. nov. et syn. nov. under Telmatoscopus advena (Eaton, 1893), and we describe for the first time the female and eggs of Telmatoscopus advena (Eaton, 1893) based on morphological and molecular data. Moreover, we provide the first record of Lepiseodina latipennis (Sarà, 1953) and Telmatoscopus bartai Ježek 2004 comb. nov. from Germany. Furthermore, we provide the first COI (5′-end of the cytochrome c oxidase subunit I) sequences, also known as DNA barcode, for Telmatoscopus bartai (Ježek, 2001) comb. nov., Lepiseodina latipennis (Sarà, 1953), Lepiseodina rothschildi (Eaton, 1912), and Lepiseodina tristis (Meigen, 1830).

2. Material and methods

2.1. Geographic Scope

We follow the proposed geographic boundaries of Europe by de Jong et al. [27] with the following limits: East: Ural (E 60°), West: Atlantic Ocean (W 30°), South: Mediterranean (N 35°), and North: Atlantic Islands (N 82°).

2.2. Literature Records

Literature search was conducted by tracking references from known literature with the help of search engines (e.g., www.scholar.google.com, accessed on 5 January 2022) and scientific databases (e.g., www.jstor.org, www.scopus.com, www.webofscience.com, 5 January 2022) using the search keywords “Psychodidae, Psychodinae, dendrolimnetic, Diptera, water-filled tree holes”. Literature used for the study encompasses records since the description of the species to the most recent published works until the beginning of
2022, focusing on Psychodinae and dendrolimnetic studies. More than 100 items were analyzed; however, only 11 included records of dendrolimnetic Psychodinae (as listed in Table 1).

2.3. Sampling

Specimens were collected using Malaise traps during the years 2013–2021 as part of the German Barcode of Life (GBOL) project [26] (www.bolgermany.de, accessed on 2 April 2022). Specimens were preserved in 96% ethanol and stored at −20 °C until they were dissected for DNA extraction and preparing permanent slides. All sampled specimens are stored at the Zoologisches Forschungsmuseum Alexander Koenig, Bonn, Germany (ZFMK). Further material was bred from organic matter sampled from dendrotelmata, and, occasionally collected with a Malaise trap and directly collected in Italy, during 2010–2022 (AM).

Additional examined material is deposited in the following natural history collections mentioned in the text using their acronyms:

AM: Alessio Morelli private collection, Pianella (PE), Italy; later to be deposited at the Natural History Museum of Genova.
BNHM: British National History Museum, London, United Kingdom.
MNGD: Museo di Storia Naturale Giacomo Doria, Comune di Genova, Genova, Italy.
NMP: Národní Muzeum, Prague, Czech Republic.
ZFMK: Zoologisches Forschungsmuseum Alexander Koenig, Bonn, Germany.

2.4. Study of the Collected Material and Terminology

After lysis (see Genetics below), the specimens were cleared using NaOH 10%, dissected, and permanently mounted using Euparal (Waldeck GmbH & Co. KG, Division Chroma, Havinbecker Straße, Münster, Germany) as mounting medium, following the procedure detailed in Ibáñez-Bernal [28], with the modification that prior to the diaphanization process, we performed the dissection of the head, wings, and terminal abdominal segments to macerate them in NaOH 10% and continue with the procedure, while preserving the remaining tissue (thorax, legs, first abdominal segments) in ethanol for posterior DNA extraction. Additional material was macerated in KOH 10%, transferred in acetic acid 10%, and dehydrated in acetone 99%. Specimens were dissected in clove oil and parts mounted on microscope slides with Canada balm. For the additional material, some specimens were photographed to keep a quality image of the habitus.

We follow the general terminology proposed by Cumming et al. and Kvifte and Wagner [15,29], except for the male terminalia that we use the term “hypopods” for the posterior genitalic appendages, which have been treated in the literature as cercopodia or surstyli originating in the 9th abdominal segment, or 10th segment, or a combination of both, as proposed by Kvifte and Wagner [30]. For the female Terminalia we follow Kotrba [31]. Egg terminology follows De Almeida et al. [32].

In the examined material of each species, a code is provided for each examined specimen (e.g., ZFMK-DIP-000852020), and all the label information is found as Supplementary Material (Table S1).

2.5. Genetics

Specimens were processed at the ZFMK, where lysis and PCR were performed following the protocol and primers by [33,34]. After the PCR, samples were sent to Beijing Genomics Institute (BGI) for bidirectional sequencing. Raw data were curated manually using Geneious (v. 7.1.9). Final COI sequences were 658 bp long. All sequences will be publicly available at www.bolgermany.de. BOLD and genebank accession IDs can be found in the Supplementary Material.
3. Results

Based on our literature review, we report 13 species of Psychodinae associated with dendrotelmata belonging to six genera (Table 1), namely Clogmia [C. albipunctata (Williston, 1893)], Clytocerus [C. xylophilus Vaillant, 1983], Lepiseodina [L. latipennis (Sará, 1953), L. rothschildi (Eaton, 1912) and L. tristis (Meigen, 1830)], Pneumia [P. canescens (Meigen, 1804) and P. trivialis (Eaton, 1893)], Psychoda [P. alternata Say, 1824, P. cinerea Banks, 1894 and P. minuta Banks, 1894] and Telmatoscopus [T. advena (Eaton, 1893), T. bartai (Ježek, 2004) comb. nov., and T. thuringicus Beran, Doczkal, Pfister & Wagner, 2010]. Additionally, we list Telmatoscopus bartai (Ježek, 2004) comb. nov. as a potential dendrolimnetic species, giving arguments for this decision.

Table 1. Dendrolimnetic Psychodinae taxa reported for Europe, tree species where it was reported and references. Double asterisk (**) denotes a new record of tree species for the Psychodinae species.

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Tree Species</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clogmia albipunctata (Williston, 1893)</td>
<td>Oak (Quercus sp.), not specified</td>
<td>[35,36]</td>
</tr>
<tr>
<td>Clytocerus xylophilus Vaillant, 1983</td>
<td>Lime tree (Tilia sp.)</td>
<td>[37]</td>
</tr>
<tr>
<td>Lepiseodina tristis (Meigen, 1830)</td>
<td>Ash (Fraxinus sp.), beech (Fagus sp.), birch (Betula sp.), cherry (Prunus sp.), elm (Ulmus sp.), maple (Acer sp.), oak (Quercus sp.), mulberry (Morus sp.), lime (Tilia sp.), not specified, ** Populus nigra L.</td>
<td>[13,22,25,36,38]</td>
</tr>
<tr>
<td>Lepiseodina rothschildi (Eaton, 1912)</td>
<td>Maple (Acer sp.), oak (Quercus sp.), not specified</td>
<td>[9,13,38]</td>
</tr>
<tr>
<td>Lepiseodina latipennis (Sará, 1953)</td>
<td>** Maple tree (Acer sp.), ** Oak (Quercus sp.)</td>
<td></td>
</tr>
<tr>
<td>Pneumia canescens (Meigen, 1804)</td>
<td>Not specified</td>
<td>[18,25]</td>
</tr>
<tr>
<td>Pneumia trivialis (Eaton, 1893)</td>
<td>Not specified</td>
<td>[22,25]</td>
</tr>
<tr>
<td>Psychoda alternata Say, 1824</td>
<td>Not specified</td>
<td>[23]</td>
</tr>
<tr>
<td>Psychoda cinerea Banks, 1894</td>
<td>Apple (Malus sp.), oak (Quercus sp.), ** Hornbeam (Carpinus or Ostrya sp.)</td>
<td>[25]</td>
</tr>
<tr>
<td>Psychoda minuta Banks, 1894</td>
<td>Maple (Acer sp.), oak (Quercus sp.)</td>
<td>[25]</td>
</tr>
<tr>
<td>Telmatoscopus advena (Eaton, 1893)</td>
<td>Ash (Fraxinus sp.), birch (Betula sp.), elm (Ulmus sp.), oak (Quercus sp.), sycamore (Platanus sp.)</td>
<td>[13,22,25,38]</td>
</tr>
<tr>
<td>Telmatoscopus thuringicus Beran, Doczkal, Pfister &amp; Wagner, 2010</td>
<td>Not specified (assumption of development)</td>
<td>[5]</td>
</tr>
<tr>
<td>Telmatoscopus laurentii Freeman, 1953</td>
<td>Lime tree (Tilia sp.)</td>
<td>[39]</td>
</tr>
</tbody>
</table>

3.1. Key to the Males of European Psychodinae Genera Found in Dendrotelmata

**Differential diagnosis.** Adults of the subfamily Psychodinae can be easily differentiated from the adults of the exclusively xylophagous subfamily Trichomyiinae, which can also be found in tree holes, by the presence of an eye bridge in Psychodinae (absent in Trichomyiinae) and wing vein R with five branches in Psychodinae, with two longitudinal veins between radial and medial forks (vein R with four branches in Trichomyiinae, with one longitudinal vein between radial and medial forks).

1. Antenna with at least flagellomeres 2–10 nodiform, divided into a basal nod and a distal neck (Figure 1B,D) . . . 3
2. Cornicula (everted sac-shaped structures on the back surface of the head (presumed scent organs); also known as patagia) usually present; antennal scape more than three times longer than pedicel; flagellomere 1 with a distal brush of wavy setae (Figure 1C) . . . Clytocerus Eaton, 1904
- Cornicula always absent; antennal scape less than three times the length of pedicel; flagellomere 1 without a distal brush of wavy setae (Figure 1A) . . . Pneumia Enderlein, 1935
Figure 1. First antennal segments (left antenna) of: (A) *Pneumia trivialis* (Eaton, 1893), (B) *Telmatoscopus advena* (Eaton, 1893), (C) *Clytocerus ocellaris* (Meigen, 1818), (D) *Psychoda* sp. Abbreviations: asc = ascoids, flm = flagellomere, ped = pedicel, scp = scape. Scale (A–C) in millimeters (mm).
3. Ascoids with anterior and posterior branches (ascoids Y-shaped) (Figure 1D); hypopods of males with only one apical tenaculum ... *Psychoda Latreille, 1797*

- Ascoids with a single curved branch or carrying only anterior branches (ascoids digitiform and not Y-shaped) (Figure 1B); hypopods of males with four or more apical tenacula ... 4

4. Ascoids bifurcate (as in Kvifte & Wagner [15], Figure 16; also in Ibáñez-Bernal [40], Figures 55 and 56); tenacula distally knife-shaped and shorter than basal width of hypopods (as in Ibáñez-Bernal [40], Figure 58) ... *Clogmia Enderlein, 1937*

- Ascoids with a single digitiform or leaf-shaped branch, not bifurcate (Figure 1B); tenacula distally feathered and longer than the basal width of hypopods (Figures 2B, 3B, 4D, 5B, 6B, 7B, 8A,C, 9A–C, 10B and 11B) ... 5

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**Figure 2.** *Lepiseodina latipennis* (Sarà 1953), male genitalia adapted from the original description by Sarà (1953). (A). Gonocoxites and gonostyli. (B). Hypopods and hypoproct. Abbreviations: gnx = gonocoxite, gns = gonostyli, hpd = hypopod, epi = epiproct, ten = tenacula. No scale available based on original drawing.
Figure 3. *Lepiseodina rothschildi* (Eaton, 1912), male genitalia. (A). Hypandrium, gonocoxites and gonostylus. (B) Epandrium, hypopods, tenacula, hypoproct. (C) Aedeagus. Abbreviations: aed = aedeagus, con plt = condyle plate-like, eja = ejaculatory apodeme, epi = epiproct, gnx = gonocoxite gns = gonostylus, ten = tenacula, hyp = hypandrium. Scale (A–C) in millimeters (mm).
Figure 4. *Lepiseodina latipennis* (Sarà, 1953), male. (A) Head. (B) Wing. (C) Flagellomeres and ascoids. (D) Genitalia. Scale (A–D) in millimeters (mm).
Figure 5. Lepiseodina latipennis (Sarà, 1953), male genitalia. (A) Hypandrium, gonocoxites and gonostylus. (B) Epandrium, hypopods, tenacula, hypoproct. (C) Aedeagus. Abbreviations: aed = aedeagus, epa = epandrium, eja = ejaculatory apodeme, epi = epiproct, gnx = gonocoxite, gns = gonostylus, ten = tenacula, hpd = hypopods, hyp = hypandrium. Scale (A–C) in millimeters (mm).
Figure 6. *Lepiseodina latipennis* (Sarà, 1953), male. SEM pictures. (A) First flagellomere (3rd antennal segment), red circle higher magnification of sensilla. (B) Genitalia, light blue = hypandrium, orange = aedeagal sheath, green = gonocoxites, purple = gonostyli, red = aedeagus, dark blue = hypopods, yellow = tenacula. Scale (A,B) in micrometers (μm).
Figure 7. *Lepiseodina tristis* (Meigen, 1830), male genitalia. (A) Hypandrium, gonocoxites and gonostylus. (B) Epandrium, hypopods, tenacula, hypoproct. (C) Aedeagus. Abbreviations: aed = aedeagus, con plat = condyles plate-like, epa = epandrium, eja = ejaculatory apodeme, epi = epiproct, gnx = gonocoxite, gns = gonostylus, ten = tenacula, hpd = hypopods, hyp = hypandrium. Scale (A–C) in millimeters (mm).
Figure 8. Wings variability of: (A–D) Lepiseodina latipennis (Sarà 1953). (E,F) L. tristis (Meigen, 1830), (G,H) L. rothschildi (Eaton, 1912). Scale (A–H) in millimeters (mm).

5. Aedeagus asymmetrical, with ejaculatory apodeme broader than distal elements (Figures 3C, 4D, 5C, 6B and 7C); ascoids digitiform or S-shaped . . . Lepiseodina Enderlein, 1936

- Aedeagus symmetrical, with ejaculatory apodeme narrower than distal elements (Figures 12C and 13C); ascoids leaf-shaped or digitiform . . . Telmatoscopus Eaton, 1904
Figure 9. Genitalia of: (A) *Lepiseodina tristis* (Meigen, 1830), (B) *Lepiseodina rothschildi* (Eaton, 1912), (C) *Lepiseodina latipennis* (Sarà, 1953). Scale (A–C) in millimeters (mm), scale lines = 0.10 mm.
Figure 10. Genitalia of: (A) *Lepiseodina latipennis* (Sarà, 1953), (B) *Lepiseodina tristis* (Meigen, 1830). Blue arrows show differences in hypandrium, gonocoxite, and gonostyli. Scale (A,B) in millimeters (mm), scale lines = 0.10 mm.
Figure 11. Genitalia variability of *Lepiseodina latipennis* (Sarà, 1953): (A–C) Aedeagal complex in dorsal view. (D) same in posterodorsal view. (E) Habitus of *L. latipennis* (Sarà, 1953), (F) Habitus of *L. rothschildi* (Eaton, 1912). Scale (A–D) in millimeters (mm); without scale (E,F).

### 3.2. Systematic Assessment

**Genus Clogmia Enderlein, 1937**

*Clogmia* Enderlein, 1937: 87. Type species: *Psychoda albipennis* Williston (=*albipunctata* Williston).

**Diagnosis.** Scape less than 2.5 times its width; flagellomeres symmetrically nodiform, with ascoids variable in shape; flagellomere 14 with elongated apiculus tapering towards apex; wing with radial fork basal to median fork; ejaculatory apodeme Y-shaped, narrow in dorsal view; aedeagus symmetrical in dorsal view.
Figure 12. Telmatoscopus advena (Eaton, 1893), male genitalia. (A) Hypandrium, gonocoxites and gonostylus. (B) Epandrium, hypopods, tenacula, hypoproct. (C) Aedeagus. Abbreviations: epa = epandrium, epi = epiproct, eja = ejaculatory apodeme, gnx = gonocoxite, gns = gonostylus, hpd = hypopod, ten = tenacula, hyp = hypandrium. Scale (A–C) in millimeters (mm).
Figure 13. *Telmatoscopus bartai* (Ježek, 2004), male genitalia. (A) Hypandrium, gonocoxites and gonostylus. (B) Epandrium, hypopods, tenacula, hypoproct. (C) Aedeagus. Abbreviations: aed = aedeagus, epa = epandrium, epi = epiproct, eja = ejaculatory apodeme, gnx = gonocoxites, hpd = hypopod, hyp = hypandrium, ten = tenaculum. Scale (A–C) in millimeters (mm).

**Species present in Europe associated with dendrotelmata:** *C. albipunctata* (Williston, 1893) [35,36] (Table 1).

*Clogmia albipunctata* (Williston, 1893)
Psychoda albipunctata Williston, 1893: 113. Type locality: Cuba, La Havana.
Pericoma meridionalis Eaton, 1894: 194. Type locality: East Africa.
Psychoda snowii Haseman, 1907: 311. Type locality: USA, Texas, Galveston.
Psychoda legnothisa Spieser, 1909: 44. Type locality: Tanzania.
Psychoda erecta Curran, 1926: 102. Type locality: West Indies.
Psychoda nocturna Abreu, 1930: 115. Type locality: Not given, probably Canary Islands
(see Ibáñez-Bernal [40]).

Psychoda nocturna var. nigrithorax Abreu, 1930: 115. Type locality: not given, probably
Canary Islands.

Telmatoscopus haranti Mirouse, 1958: 93. Type locality: Midi de la France.
Telmatoscopus albipunctatus (Williston): [41] (p. 185).
Clogmia albipunctata (Williston): [42] (p. 42); 351 [11] (p. 351) (see Ibáñez-Bernal [40]).

Diagnosis. Antenna with symmetrically nodiform flagellomeres, each flagellomere
with a bifurcate ascoids; eyes separated by 1 facet diameter, eye bridge with 4 facet rows;
aedeagus symmetrical, ejaculatory apodeme straight and narrow in dorsal view; hypopods
with five-six tenacula, tenacula decreasing in size towards the apex of hypopod.

Examined material: ZFMK-DIP-00081299, ZFMK-DIP-00081508, ZFMK-DIP-00081514,
ZFMK-DIP-00081543, ZFMK-DIP-00081624, ZFMK-DIP-00081591, ZFMK-DIP-00082118
[ZFMK].

Distribution. This is one of the most widespread species of Psychodidae in the world,
highly invasive and synanthropic. In Europe, it is recorded in Azores Archipelago, Belgium,
Canary Islands, Corsica, Croatia, Czech Republic, Denmark, Finland, France, Germany,
Greece, Hungary, Italy, Luxemburg, Madeira Islands, Sardinia, Slovakia, Slovenia, Spain,
Sweden and United Kingdom [9,43–47].

Genus Clytocerus Eaton, 1904

Clytocerus Eaton, 1904: 59. Type species: Clytocerus ocellaris (Meigen, 1804: 44) by
subsequent monotypy of Malloch (1907) (see [48]).

Diagnosis: Cornicula (everted sac-shaped structures on the back surface of the head)
are usually present; eye bridge with 3–6 facet rows, separated by 1–3 facet diameters;
antennal scape elongate, three-six times longer than wide; flagellomere I and II fused to
form a compound flagellomere, with a distal brush of wavy setae [14].

Species present in Europe associated with dendrotelmata: C. xylophilus Vaillant,
1983 [37] (Table 1).

Clytocerus xylophilus Vaillant, 1983

Clytocerus xylophilus Vaillant, 1983: 355. Type locality: France.

Diagnosis. Eyes separated by 2 facet diameters, interocular suture absent. Antennal
scape 4.75 times longer than wide. Hypopods with 6–7 tenacula.

Examined material. None.

Distribution. France [43].

Remarks. Vaillant [37] mentions under the description of C. xylophilus that the larvae of
Clytocerus pulvereus Vaillant, 1983 can be found in the same substrate as C. xylophilus;
however, it is referring to the “black humus” found near a river (same substrate men-
tioned by Krek [49]), and later Vaillant [37] clarifies that only C. xylophilus is known as a
dendrolimnetic species.

Genus Lepiseodina Enderlein, 1937


Diagnosis. Frons and clypeus separated and not protruding over eye margin; scape
short, less than 2 times length of pedicel; flagellomeres symmetrically nodiform in dorsal
view, with a pair of digitiform sinuous ascoids; flagellomere 14 with elongated apiculus;
wing vein R2+3 not connected to R4; apex of R5 ending on, or below wing apex; gonocoxites
short, with dorsal projection undeveloped; ejaculatory apodeme narrow in dorsal view;
aedeagal complex asymmetric, with a parameral structure more or less sclerotized, connect-
ing aedeagus to the gonocoxal apodemes; hypopods with indistinctly fringed tenacula;
epandrium with a single foramen.
Species present in Europe associated with dendrotelmata: L. rothschildi (Eaton, 1912), L. latipennis (Sarà, 1953), and L. tristis (Meigen, 1830) [9,13,23,25,36,38] (Table 1).

**Lepiseodina latipennis** (Sarà, 1953)
Figures 2, 4, 5, 6, 7A, 8C, 9C, 10 and 11E.


*Telmatoscopus latipennis* Sarà: [50] (p. 53).

*Clognia latipennis* (Sarà): [51] (p. 60).

*Lepiseodina latipennis* (Sarà): [52] (p. 146).

**Diagnosis.** Male hypandrium broad along its entire length; gonocoxal alveoli distributed in the entire gonocoxal surface; gonostyli without alveoli on the base, twice the length of gonocoxites; epandrium not divided, with a single kidney-shaped foramen; aedeagus incurved; paramere not strongly sclerotized; hypoproct tongue-shaped.

**Differential diagnosis.** This species is closely related to *Lepiseodina tristis*, but it can be distinguished by the combination of the following characters: hypandrium broad along its entire length with convexity in the basal margin in *L. latipennis* (hypandrium narrow with a medial projection in the basal margin in *L. tristis*); gonocoxal alveoli distributed in the entire gonocoxal surface in *L. latipennis* (gonocoxal alveoli restricted to the apical margin in two or three irregular rows in *L. tristis*); gonostyli without alveoli on the base in *L. latipennis* (gonostyli with alveoli in the base in *L. tristis*); epandrium not divided in the apical margin, with a kidney-shaped foramen in *L. latipennis* (epandrium divided in the apical margin with a rounded foramen in *L. tristis*); aedeagus incurved in *L. latipennis* (aedeagus straight in *L. tristis*); parameres not strongly sclerotized and almost not visible in *L. latipennis* (parameres strongly sclerotized in *L. tristis*).

*Lepiseodina latipennis* can be differentiated from *Lepiseodina rothschildi* by the gonostyli twice the length of gonocoxites in *L. latipennis* (gonostyli less than half of the length of gonocoxite in *L. rothschildi*), apical margin of hypandrium convex in *L. latipennis* (apical margin of hypandrium concave in *L. rothschildi*), and parameres not strongly sclerotized and almost not visible in *L. latipennis* (parameres strongly sclerotized in *L. rothschildi*).

**Redescription.** Measurements in mm (mean, SD = 0.005, n = 2). Wing length 2.53 (2.20–3.30), wing width 0.98 (1.02–1.32). Head length 0.50 (0.48–0.59), head width 0.63 (0.53–0.65). Antennal segments, scape length: 0.11 (0.09–0.11), pedicel: 0.08 (0.70–0.90), post-pedicel: 0.10 (0.10–0.13), flagellomeres average length 0.11; Palpomeres 1: 0.10 (0.08–0.10), 2: 0.25 (0.20–0.27), 3: 0.23 (0.20–0.24), 4: 0.29 (0.24–0.29).

**Male.** Head: eye bridge with rows of four facets (rarely 3 facets); frontal patch undivided, with an irregular row of alveoli extending towards the interocular suture, the whole frontal patch together with the irregular row resemble the shape of a handbell; interocular suture as an inverted “v”. No alveoli of supra ocular setae are present. Labella bulbous, longer than wide, with 10–12 small setae. Antenna with scape cylindrical, 1.34 times longer than its width, 1.43 (1.2–1.4) times the length of pedicel; pedicel spherical; 14 nodiform flagellomeres with basal bulb and distal neck, apical flagellomere with long apiculus, apiculus almost half the length of the flagellomere (Figure 4A); antennal ascoids slightly flattened, digitiform, sinuous (S-shaped), total length about 1.88 the length of flagellomeres; flagellomeres with a sensilla (as shown in Figure 6A). Palpal segment proportions 1.0:2.63:2.37:3.03.

Thorax with no particular characters.

Wing: Length about 2.5 (2–2.5) times its width, variable in shape, with anterior margin more or less curved; membrane bare except on veins; hyaline with a slight infuscation on costal cell; Sc straight, ending at level of the origin of R_{2+3}; Origin of R_{3+3} not joining R_{4}, a little distal to the origin of M_{1+2}; origin of M_{1+2} broad and rounded; forks of R_{2+3} and M_{1+2} almost at the same level, the fork of R_{2+3} being slightly distal to M_{1+2}; R_{5} ending at wing apex; CuA ending in wing margin at the level of R_{2+3} fork.

Abdomen. Without any particular characters.

Genitalia. Hypandrium plate-like, broad, basal margin with medial projection widely rounded, distal margin almost straight; gonocoxites longer than wide, covered in alveoli
on almost all the surface; gonostyli about 1.5 times the length of gonocoxites, tapering towards apex, almost straight; gonocoxal apodemes poorly distinguishable in dorsal view; parameres forming a single slight sclerotized structure, pyriform, resembling the upper half of a bowling pin connecting like a bridge aedeagus to gonocoxal apodemes (as in Figures 4D, 5C, 6B, 9C and 10A); epandrium subrectangular, about twice wider than long with a single kidney-shaped foramen; hypopods slightly out curved, apex rounded, with 9–13 tenacula (as in Figures 4B, 5C and 9C); epiproct short, triangular and covered in pilosity; hypoproct broad and tongue-shaped, covered in pilosity, extending towards mid of hypopods. Aedeagus with ejaculatory apodeme digitiform, narrow 7.5 times longer than its width, about 1.5 times the length of gonocoxites; distiphallus about the same length of ejaculatory apodeme, incurred, extending towards the apex of gonostyli, both ejaculatory apodeme and distiphallus form a single complex, jointed with parameral structure and encircles by a membranous parameral sheath.

**Female.** Unknown.

**Remarks.** The holotype (slide mounted) is quite dark, making the observation of structures difficult. The head is dissected, and quite difficult to see clearly, one antenna is dissected and the other antenna is missing, one complete palpus is dissected, the other palpus is missing. The hypopods are dissected and placed apart from each other, one gonocoxites and gonostylus are dissected and placed apart from the genitalia, the remaining parts of the genitalia (aedeagal complex, one gonocoxites and gonostylus) are found together. One wing is dissected and separated from the thorax. On the original description Sarà (1953) mentions the shape of the hypoproct being trilobed (Figure 2), after examination of the holotype in the preparation the hypoproct looks indeed trilobed; however, we consider this a malformation on the slide itself, and not the natural shape of the hypoproct, all other examined material present a broad tongue-shaped hypoproct. On the original description Sarà abstained to illustrate or describe the aedeagal complex; however, after examination we are sure all our specimens belong to the same species.

**Biology.** Some specimens (No. 0190-0193) emerged from organic matter sampled from an *Acer sp.* dendrotelma (Figure 14), in a mixed submontan forest, additionally, the specimen used for the SEM pictures was collected in a Malaise trap next to dendrotelmata of an oak (*Quercus* sp.) tree, suggesting that also *L. latipennis* is a dendrolimnetic species. In Germany altitude ranges from 250–280 m a.s.l. (meters above sea level), while in Italy it has a range from 40–800 m a.s.l.

**Examined material.** *Holotype* examined [MNGD], ZFMK-DIP-00081595, ZFMK-DIP-00081552 [ZFMK], Specimen No. 0039, 0488, 0487, 0439, 0190, 0191, 0192, 0193 [AM].

**Distribution:** Previously known only from the type locality in Messina (Sicily, Italy). The records here reported are the first from Germany (Rheinland-Pfalz) and for northern and central Italy.

*Lepiseodina rothschildi* (Eaton, 1912)

Figures 3, 8G,H and 9B


*Telmatoscopus rotschildi* (Eaton): *Lapsus calami* of [50], followed by Salamanna in Dahl et al. [53] (see below).


*Clogmia rotschildi* (Eaton): Dahl et al. [53] (p. 33).


**Diagnosis.** Male gonostyli are half the length of gonocoxites; paramere strongly sclerotized, overlapping to the lateral branch of aedeagus; hypandrium W-shaped, broad on the entire surface with the apical margin concave.
Figure 14. (A,B) Habitat of Psychoda cinerea Banks, 1894, dendrotelma of hornbeam (Carpinus or Ostrya sp.). (C,D) Habitat of Lepiseodina tristis (Meigen, 1830), rotting tree hole with basal dendrotelma of Populus nigra. All in a mixed submontan forest (Foro Valley, central Italy).

Examined material: ZFMK-DIP-00081311, ZFMK-DIP-00081322, ZFMK-DIP-00081323, ZFMK-DIP-00081324, ZFMK-DIP-00081327, ZFMK-DIP-00081328, ZFMK-DIP-00081329, ZFMK-DIP-00081330, ZFMK-DIP-00081510, ZFMK-DIP-00081537, ZFMK-DIP-00081551, ZFMK-DIP-00081558, ZFMK-DIP-00081559, ZFMK-DIP-00081567, ZFMK-DIP-00081574,
ZFMK-DIP-00081623, ZFMK-DIP-00082122, ZFMK-DIP-00082126, ZFMK-DIP-00082147,
ZFMK-DIP-00082149 [ZFMK]. Specimens number: 13666, 13689, 13694, 13625, 13628, 13822,
18655, 18490, 18225, 19261, 19110, 16352, 16489, 18408, 19469, 17902, 21769, 12319, 12320,
21533, 21114 [NMP]. Specimens number: 0489 [AM].

**Distribution.** Austria, Belgium, Bulgaria, Czech Republic, Finland, France, Germany,
Ireland, Italy, Lithuania, Netherlands, Slovakia, Spain, United Kingdom [9,13,54,55].

*Lepiseodina tristis* (Meigen, 1830)
Figures 7, 8E,F, 9A and 10B

Psychoda tristis Meigen, 1830: 272. Type locality: Not specified, probably Belgium
or Germany.

Pericoma tristis (Meigen): [56] (p. 16).

Telmatoscopus tristis (Meigen): [58] (p. 170).

Lepiseodina tristis (Meigen): [42]) (p. 91); [52] (p. 146).

Clogmia tristis (Meigen): [53] (p. 33); [59] (p. 7).

**Diagnosis.** Male hypandrium narrow, with a small abrupt median projection; gono-
coxal alveoli restricted to two irregular lines on the apical surface; gonocoxites with a patch
of alveoli at base; two parameres strongly sclerotized.

Examined material: ZFMK-DIP-00081512, ZFMK-DIP-00081513, ZFMK-DIP-00081563,
ZFMK-DIP-00081583, DIP-ZFMK-00081622, ZFMK-DIP-00082119, ZFMK-DIP-00082124,
ZFMK-DIP-00082125, ZFMK-DIP-00082128, ZFMK-DIP-00082130, ZFMK-DIP-00082133,
ZFMK-TIS-2628581, ZFMK-TIS-2628542, ZFMK-TIS-2628588, ZFMK-TIS-2628587, ZFMK-
TIS-2628608 [ZFMK]. Specimen number: 24339, 13701, 13668, 13633, 13823, 20777, 20243,
20077, 20077, 20076, 20080, 20079, 20067, 3226, 33282, 34041, 34042, 34043,
34044, 20987, 21365, 23772, 24339 [NMP]. Specimen number: 0490, 0491 [AM].

**Distribution.** Algeria, Austria, Belgium, Croatia, Czech Republic, France (incl. Cor-
sica), Germany, Ireland, Italy, Lithuania, Slovakia, and United Kingdom [25,36,43,50,53,59,60].
In Italy, the species is known only from two old and uncertain records for the northern
and peninsular region [53,61]. The specimens here reported confirm the occurrence of this
species in the Italian peninsula.

**Key to the European adult males of Lepiseodina**

1. Gonostyli longer than gonocoxites . . . 2
   - Gonostyli about half the length of gonocoxites (Figure 3A) . . . *L. rothschildi*

2. Hypandrium narrow, with a small medial projection on basal margin (Figure 7A);
gonocoxites in gonocoxites restricted in two or three irregular rows in the apical margin
(Figures 9A and 10B); gonostyli with alveoli on the base (Figures 9A and 10B); two
parameres strongly sclerotized (Figure 7C) . . . *L. tristis*
   - Hypandrium broad in its entire length, without medial projection (Figure 5A); alveoli
in gonocoxites scattered in the whole surface (Figure 5A); gonostyli without alveoli on
the base (Figure 5A); one parameral structure not strongly sclerotized (Figure 5C) . . .
*L. latipennis*

**Genus Pneumia Enderlin, 1935**

*Pneumia* Enderlein, 1935: 247. Type species: *Pericoma palustris* Meigen, 1804 (see [62]).

**Diagnosis.** Eye bridge with 6 facet rows. Antennal flagellomeres barrel-shaped, basal
flagellomeres lacking spines or clusters of stiff setae; apical flagellomere with a digitiform
apical protuberance about as long as or longer than the basal part of the flagellomere
carrying it. Aedeagal complex symmetrical; gonostyli bulbous and elongate at the base,
tapering towards the apex.

**Species present in Europe associated with dendrotelmata:** *Pneumia canescens* (Meigen,
1818) and *Pneumia trivialis* (Eaton, 1893) [18,23,25] (Table 1).

**Pneumia canescens** (Meigen, 1818)
*Trichoptera canescens* Meigen, 1818: 45. Type locality: not given, probably Germany.
*Pericoma canescens* (Meigen): 20 (p. 156).
Pneumia canescens (Meigen): [63] (p. 124).

Diagnosis. Eye bridge with 5–6 facet rows: wing vein fork R2+3 with a backward projection (as in [11] (Figure 12)); hypopods with 10 tenacula; ejaculatory apodeme wider at the base than apex; aedeagal complex longer than gonocoxites, tapering towards apex.

Examined material. NMP-2687, NMP-2913, NMP-10787, NMP-17243 [NMP].

Distribution: Afghanistan, Armenia, Austria, Azerbaijan, Belgium, Bulgaria, China, Czech Republic, Denmark, France, Georgia, Germany, Greece, Hungary, Kyrgyzstan, Lithuania, Netherlands, Poland, Romania, Russia (Novosibirsk Region), Slovakia, Sweden, Turkey and United Kingdom [40,56,57].

Pneumia trivialis (Eaton, 1893)

Pericoma trivialis Eaton, 1893: 121. Type locality: Great Britain.

Diagnosis. Eye bridge with 7 facet rows; frons not extending beyond first palpal segment; all pal segments of similar width; fore tibia straight and not engrossed; wing vein fork R2+3 without a backward projection; hypopods with 7 or fewer tenacula; ejaculatory apodeme straight, not wider at base; aedeagal complex shorter than gonocoxites, not tapering towards apex, distal part of aedeagus smooth without wrinkles.


Distribution: Austria, Azerbaijan, Belgium, Bosnia–Herzegovina, Bulgaria, Croatia, Czech Republic, Denmark, Finland, France, Georgia, Germany, Hungary, Ireland, Netherlands, Norway, Poland, Portugal, Serbia, Slovakia, Slovenia, Spain, Sweden, Switzerland, Turkey, Ukraine and United Kingdom [43,55,64,65].

Genus Psychoda Latreille, 1797

Psychoda Latreille, 1796: 152. Type species: Tipula phalaenoides Linnaeus, 1758 by subsequent designation of Quate [41] (p. 191).

Diagnosis. Species with short vertex; labellum flattened and carrying digitiform setae; eyebridge without interocular suture; antenna with 12–14 flagellomeres, those beyond 11th always reduced in size and showing different types of fusion; flagellomeres nodiform with a neck (except apical flagellomeres); ascoids usually with three branches, two anterior and one posterior branch, Y-shaped or with four branches, shaped like a plus sign (+); aedeagus often asymmetrical; hypopods with a single apical tenaculum.

Species present in Europe associated with dendrotelmata: P. alternata Say, 1824, P. cinerea Banks, 1894 and P. minuta Banks, 1894. [23,25] (Table 1).

Psychoda alternata Say, 1824


Psychoda tripunctata Macquart, 1838: 85. Type locality: not given, probably France.

Psychoda marginipunctata von Roser, 1840: 50. Type locality: not given.

Psychoda sexpunctata Phillippi, 1865: 631. Type locality: not given.

Psychoda nocturna Haseman, 1907: 319. Type locality: USA, Missouri, Columbia.

Psychoda floridica Haseman, 1907: 324. Type locality: USA, Florida.

Psychoda bengalensis Brunetti, 1908: 371. Type locality: India, Calcutta and Simla.

Psychoda albimaculata Welch, 1912: 411. Type locality: USA, Illinois.

Psychoda dakotensis Dyar, 1926: 109. Type locality: USA, South Dakota.

Psychoda alternata var. marmosa Abreu, 1930: 123. Type locality: not given.

Psychoda alternata var. floridica Haseman: Johannsen, 1934: 25.

Psychoda cinerea Banks, 1894

Psychoda cinerea Banks, 1894: 331. Type locality: USA, New York, Sea Cliff, L. I.


Psychoda cinerea (Banks): [66] (p. 135); [80] (p. 100).

Psychoda (Tinearia) alternata Say: Bravo et al. [71] (p 5, 11).

Psychoda alternata Say: [40] (p. 97). [41] (p. 218) [72] (p. 216); [73] (p. 195); [74] (p. 16); [75] (p. 12); [76] (p 238); [77] (p. 67); [78] (p. 21); [79] (p. 48).

Psychoda minuta Banks, 1894

Psychoda minuta Banks, 1894: 331. Type locality: USA, New York, near Sea Cliff.

Psychoda marylandana Del Rosario, 1936: 111.

Psychoda spreta Tonnoir, 1940: 57.

Psychoda minuta (Banks): [52] (p. 56).

Psychoda minuta Banks, 1894: 331. Type locality: USA, New York, near Sea Cliff.

Psychoda marylandana Del Rosario, 1936: 111.

Psychoda spreta Tonnoir, 1940: 57.

Psychoda minuta (Banks): [52] (p. 56).

Diagnosis. Eyes separated by 1–3 facet diameters, without interocular suture; antenna with 13 flagellomeres, last three flagellomeres small, flagellomere 11–12 fused, flagellomere 13 smaller and partially fused to 12; labellum with one short and four long teeth and four setae; aedeagus asymmetrical, with on paramere thicker than aedeagus, ejaculatory apodeme broad posteriorly, longer than aedeagal complex; hypopods 2.3 times longer than gonostyli. [40].

Examined material. None.

Distribution. Cosmopolitan [40,55]. In Europe, it is present in Austria, Balearctic Islands, Belgium, Bulgaria, Croatia, Cyprus, Czech Republic, Denmark, Finland, France, Germany, Greece, Hungary, Ireland, Italy, Netherlands, Norway, Poland, Portugal, Romania, Sardinia, Slovakia, Slovenia, Spain, Sweden, Switzerland, and United Kingdom [55].

Psychoda cinerea Banks, 1894

Psychoda cinerea Banks, 1894: 331. Type locality: USA, New York, Sea Cliff, L. I.


Psychoda cinerea (Banks): [66] (p. 135); [80] (p. 100).

Diagnosis. Eyes separated by 1–2 facet diameters, without interocular suture; antenna with 14 flagellomeres, last 3 separated not fused, labellum with four terminal digitiform setae and up to 5 setiform setae; medial fork with basal swelling; aedeagal complex slightly S-shaped, asymmetrical, with unpaired bent paramere; aedeagus subtriangular, covering a squarish structure; ejaculatory apodeme triangular at base in dorsal view; hypandrium trapezoidal; hypopods as long as gonostyli, gradually narrowing toward the apex, but without basal swelling.

Examined material. Specimens number: No. 0480-4081 [AM] Reared from dendrotel-mata of a hornbeam (Carpinus or Ostrya sp.).

Distribution. Cosmopolitan [71]. In Europe, it is present in Austria, Azores archipelago, Belgium, Bosnia–Herzegovina, Bulgaria, Canary Islands, Corsica, Czech Republic, Denmark, Finland, France, Germany, Greece, Hungary, Ireland, Iceland, Italy, Luxembourg, Madeira Islands, Netherlands, Norway, Poland, Romania, Sardina, Serbia, Slovakia, Slovenia, Spain, Sweden, Switzerland, and United Kingdom [43,49].

Psychoda minuta Banks, 1894

Psychoda minuta Banks, 1894: 331. Type locality: USA, New York, near Sea Cliff.

Psychoda marylandana Del Rosario, 1936: 111.

Psychoda spreta Tonnoir, 1940: 57.

Psychoda minuta (Banks): [52] (p. 56).

Diagnosis. Eyes separated by 1 facet diameters, without interocular suture; antenna with 16 flagellomeres, last 4 flagellomeres reduced in size and fused; labellum with four terminal digitiform setae and two trichiform setae; aedeagus asymmetrical, flanked by two large triangular plates of morphologically unknown origin; gonostyli narrowing to tapered point in apical fifth; hypandrium narrow, not trapezoidal; hypopods much longer than gonostyli, with basal swelling.
Examined material. None.

Distribution. Holarctic. In Europe, it is present in Austria, Balearic Islands, Belgium, Bulgaria, Corsica, Cyprus, Czech Republic, Denmark, Finland, France, Germany, Greece, Hungary, Ireland, Italy, Lithuania, Madeira Islands, Netherlands, Norway, Romania, Sardinia, Slovakia, Slovenia, Spain, Sweden, Switzerland, and United Kingdom [25,43,59,81].

Genus Telmatoscopus Eaton, 1904

Telmatoscopus Eaton, 1904: 58. Type species: Pericoma advena Eaton, 1893, by designation of Quate [82].

Sciria Enderlein, 1935: 247. Type species: Pericoma advena Eaton, 1893, by original designation (see [83]).

Krivosheinoscopus Ježek, 2001: 57 Type species: Telmatoscopus ussuricus (Ježek, 2001) syn. nov.

Diagnosis (modified from Kvifte, [83]). Frons and clypeus separated and not protruding over eye margin; flagellomeres asymmetrically nodiform, with paired leaf-shaped to digitiform ascoids; flagellomere 14 with elongated apiculus; wing veins R\textsuperscript{2+3} not connected to R4; apex of R5 ending at wing apex; ejaculatory apodeme narrow in dorsal view and distally ending in two short branches with membranous connection to aedeagal complex; aedeagal complex symmetrical; aedeagal complex encapsulated in a parameral sheath; hypopods with indistinctly fringed tenacula; epandrium with a single foramen.

Remarks: Ježek [84] described Krivosheinoscopus based on five male specimens of the type species K. ussuricus Ježek, 2001 (now Telmatoscopus ussuricus comb. nov.) from the Russian Far East. Later, he described K. bartai Ježek, 2004 (now Telmatoscopus bartai comb. nov.) from the Czech Republic (Ježek 2004). Ježek [84] provided a diagnostic table of closely related telmatoscopoid genera, namely, Lepiseodina, Sciria (=Telmatoscopus), Iranotelmatoscopus, Krivosheinoscopus, Telmatoscopus auctt. (=Seoda) [84] (p. 60).

In the table, seven morphological characters were provided to differentiate Krivosheinoscopus from Sciria (=Telmatoscopus), they are numbered and discussed below.

1. The scape/pedicel proportion is 2:1 in Krivosheinoscopus (1:1 in Sciria); however, even while the proportion of scape/pedicel length proposed by Ježek [84] is 2:1 in T. ussuricus, this is not the case for T. bartai where it is 1:1.

2. The ascoids are very long, thin and coiled in Krivosheinoscopus (large, flat, leaf or hood-shaped in Sciria). This character is contradicted by Telmatoscopus laurenceti [38], which has digitiform ascoids and coiled, and these character states are polymorphic in Vaillantodes Wagner, 2001 and Panimerus Eaton, 1913. Intermediary forms also occur in some genera of Telmatoscopoids. Due to widespread polymorphism of this character we do not consider this a reliable genus-level character unless supported by other, independent lines of evidence.

3. The first palpal segment very short and keg-shaped in Krivosheinoscopus (long and cylindrical in Sciria). This character is variable inside the genus Telmatoscopus (e.g., long in Telmatoscopus advena and shorter in Telmatoscopus thuringicus Beran, Doczkal, Pfister & Wagner, 2010), therefore it is here considered as intrageneric variability, and thus not a diagnostic character.

4. The position of the radial and medial forks of wing venation, at the same distance in Krivosheinoscopus (medial fork distad to radial fork in Sciria). This is another character presenting as variable within the genus Telmatoscopus (e.g., medial fork distal in T. advena, medial fork basal in T. thuringicus, forks at the same level in Telmatoscopus bartai).

5. Two pairs of protuberances in the aedeagal complex in Krivosheinoscopus (one pair in Sciria). According to the homologization of [83], the absence of protruding parameres in Telmatoscopus are not due to absence of parameres, they are present as transverse sclerites within the aedeagal–parameral complex. The difference between T. advena, T. bartai and T. ussuricus in the parameres is a question of degree of development rather than a clear-cut presence/absence question, and intermediate cases occur in Nearctic species (e.g., T. patibulus Quate).
6. The hypoproct has a terminal projection, long and narrow in *Krivosheinoscopus* (without a terminal projection, triangular in *Sciria*). This character is contradicted by *Telmatoscopus bartai* comb. nov. where the hypoproct is triangular, and not thin and elongated.

With three shared characters between *Krivosheinoscopus* and *Telmatoscopus* (= *Sciria sensu* Ježek) including the wing vein $R_5$ ending at wing apex, vein $Cu$ ending distal to $M_{1+2}$ and, the ejaculatory apodeme laterally compressed (=basal apodeme of the aedeagal complex in Ježek) [84].

Furthermore, Ježek listed seven (five are commented here) morphological characters separating *Krivosheinoscopus* from *Seoda* Enderlein (considered as *Telmatoscopus* auctt. In Ježek [84]), including:

1. The scape/pedicel proportion, being 2:1 in *Krivosheinoscopus* (3-4:1 in *Seoda*); this remains a diagnostic difference.
2. First palpal segment short and keg-shaped in *Krivosheinoscopus* (long and cylindrical in *Seoda*). This character is variable inside *Telmatoscopus*, and is thus not a diagnostic character.
3. Vein $Cu$ ending distal to $M_{1+2}$ in *Krivosheinoscopus* (samel level or distad to $M_{1+2}$ in *Seoda*) yet again, a variable character;
4. Wing vein $R_5$ ending at wing apex in *Krivosheinoscopus* (ending below wing apex in *Seoda*), this character is diagnostic between *Seoda* and *Telmatoscopus* as pointed out in [83].
5. The ejaculatory apodeme laterally compressed in *Krivosheinoscopus* (dorso-ventrally compressed in *Seoda*) also a diagnostic character pointed out in [83].

Diagnostic morphological characters to separate *Telmatoscopus* from *Seoda* presented in [83] include single pair of digitiform to filiform ascoids (an additional ring of small setiform ascoids + the main pair of digitiform ascoids in *Seoda*); frons clearly separated from clypeus not protruding over the mesal margin of eyes in *Telmatoscopus* (frons fused with clypeaus and protruding over the mesal margin of eyes in *Seoda*); parameral sclerites not fused in *Telmatoscopus* (parameral sclerites fused in *Seoda*).

Our currently reformulated conscription of *Telmatoscopus* includes 10 species globally, namely *T. advena* (Eaton, 1893), Palaearctic; *T. bartai* (Ježek, 2004) comb. nov., Europe; *T. dendrophilus* Vaillant, 1983, USA; *T. frondeus* Tokunaga & Etsuko, 1955, Japan; *T. laurencii* Freeman, 1953, Europe; *T. pappi* (Wagner, 1979), Afghanistan; *T. patibulus* Quate, 1955, USA; *T. ussuricus* (Ježek, 2001) comb. nov., Russia; *T. tanegashimensis* (Ježek & Mogi, 1995), Japan; *T. thuringicus* Beran, Doczkal, Pfister & Wagner, 2010, Europe.

**Notes.** Previously, Kvitše, [83] included *Panimerus wagneri* Salamanna, 1982 as *Seoda*, however the inclusion of *T. wagneri* (= *T. advena*) comb. nov. et syn. nov. inside of *Telmatoscopus* is based on the male genital morphology presented in the original description, therefore the new combination from *Seoda* to *Telmatoscopus*. Additionally, the characters separating *T. wagneri* (= *T. advena*) syn. nov. and, *T. advena* are quite inconspicuous, however, we did not examine the holotype of *T. wagneri* (= *T. advena*) syn. nov., nonetheless, based on the original description and figures, we did not find any morphological characters that support them as separate species, and we treat *T. wagneri* as a new synonym of *T. advena*.

Additional characters and closer examination of the species *T. pappi*, *T. frondeus*, and, *T. tanegashimensis* is desirable as they share some characters with the genus *Lepiseodina*, however, these species are outside the geographical scope of this work, and therefore are included as *Telmatoscopus* following [83] until a broader revision of the genus is available.


**Telmatoscopus advena** (Eaton, 1893)

*Figures 12–14*

*Pericoma advena* Eaton, 1893: 127. Type locality: Great Britain.

*Telmatoscopus advena* (Eaton): [81] (pp. 205–209)

*Sciria advena* (Eaton): [85] (p. 87).

Panimerus advenus (Eaton): [50] (p. 80).
Panimerus havelkai Wagner, 1975: 1; (see [83]).
Panimerus wagneri Salamanna, 1982 syn. nov.
Telmatoscopus seguyi Vaillant, 1990: 378; (see [83]).

**Diagnosis.** Male. Ascoids broad, leaf-shaped and coiled; phallomeres incurved, symmetrical, less than half the length of ejaculatory apodeme; hypopods with tenacula restricted to apex.

**Female description** (Figures 15 and 16) based on the description of the male in [83] the female is similar to the male except: Head slightly longer than wide (0.5 mm wide, 0.6 mm length). Eye bridge with four facet rows, separated by 1.5 facet diameter; only first palpal segment present in examined material; apical antennal flagellomeres absent in examined material, ascoids s-shaped, coiled, thin (not broad leaf-shaped as in male). Wing length, wing width, length x times its width.

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Figure 15. *Telmatoscopus advena* (Eaton, 1983), female. (A) Head. (B) Genital chamber. (C) Egg. (D) Cerci. Scale (A–D) in millimeters (mm).
**Figure 16.** *Telmatoscopus advena* (Eaton, 1893), female. (A) First antennal segments. (B) Sternite 8 and genital chamber. (C) Egg. (D) Cerci. Abbreviations: AP = anterior pole, asc = ascoids, cerc = cerci, flm = flagellomere, gen chamb = genital chamber, ped = pedicel, PP = posterior pole, scp = scape, stn = sternite. Scale (A–D) in millimeters (mm).

Wing 2.7 times longer than wide (1 mm wide, 2.7 mm long), Sc long, ending at junction of R₄ + R₂₋₃. Infuscated in area between C and R₁.
Sternite 8 (Subgenital plate) about the same length and width (0.25 mm wide, 0.24 mm long), waisted in the middle; basal margin straight, apical margin covered in small setae and strongly concave in the middle. Cerci long (0.4 mm length), 1.6 times the length of the sternite 8, with 10 setae on the dorsal surface, the surface in the base of the cerci presents multiple plate-shaped protuberances, resembling the texture of crocodile skin. Subgenital lobes rectangular.

**Egg description.** (Mean length 0.32 mm, width 0.1 mm, n = 15) Ovoid, longer than wide (0.1 mm wide, 0.32 long). No Micropyle structure is visible in the anterior pole; however, it does seem to be a circular aperture where the micropyle could be located. Exochorion with both longitudinal and transversal single, flattened ridges that when joined, form cells (mainly rectangular or hexagonal) across the exochorion.


**Distribution.** Aegean Islands, Belgium, Finland, France, Germany, Ireland, Norway, Slovakia, United Kingdom [13,43,87].

**Telmatoscopus bartai** (Ježek, 2004) comb. nov.

Figure 13

*Krivosheinoscopus bartai* Ježek, 2004: 117, Type locality: Czech Republic: Bohemia or., Železné hory Mts Protected landscape area.

**Diagnosis.** Male. Ascoids digitiform, thin, long, and coiled; phallomeres symmetrical, out curved, more than half the length of ejaculatory apodeme; hypopods with tenacula restricted to apex.

**Examined material.** Holotype examined: Cat. No. 34243 [NMP]. ZFMK-DIP-00081598, ZFMK-DIP-00081597, ZFMK-DIP-00081599 [ZFMK].

**Distribution.** Czech Republic [88], Germany. Our three reported specimens are the first record of this species in Germany.

**Telmatoscopus laurencei** Freeman, 1953

Telmatoscopus laurencei Freeman, 1953: 71. Type locality: Great Britain. Herts, Harpenden, Rothamsted Experimental Station.

**Diagnosis.** Male. Ascoids digitiform, thin, coiled; phallomeres coiled, less than half the length of ejaculatory apodeme; hypopods with tenacula restricted to apex.

**Examined material.** None.

**Distribution.** Only known from Great Britain [13,39,43].

**Telmatoscopus thuringicus** Beran, Doczkal, Pfister & Wagner, 2010

Telmatoscopus thuringicus Beran, Doczkal, Pfister & Wagner, 2010: 63. Type locality: Germany, Thuringia, National Park Hainich, Weberstedt, Birkensee.

**Diagnosis.** Male. Ascoids are broad, leaf-shaped, tapering towards the apex and coiled; phallomeres out curved, less than half the length of ejaculatory apodeme; hypopods
with a cluster of approximately 30 tenacula at apex, and more tenacula scattered along the entire surface of hypopods.

**Examined material.** None.

**Distribution.** Only known from Germany [5].

**Key to the European adult males of Telmatscopus**

1. More than 30 tenacula; tenacula not restricted to the apical portion of hypopods, scattered in all the hypopods length (as in [5] (Figure 10)) . . . *T. thuringicus*
   - Less than 30 tenacula; tenacula restricted to the apical portion of hypopods and not scattered in the hypopods length . . . 2

2. Ascoids broad, almost as broad as the basal node of flagellomere carrying them (Figure 1B); aedeagus with parameres not coiled and strongly incurved (Figure 12C) . . . *T. advena*
   - Ascoids narrow, never broader than 1/3 of the width of the basal node of flagellomere carrying them; parameres coiled or out curved (Figure 13C) . . . 3

3. Parameres coiled with strong incurvation after the coil and apical tips hook-out curved (as in [38] (Figure 2D)); hypopods with less than 20 tenacula . . . *T. laurencei*
   - Parameres not coiled and out curved (Figure 13C); hypopods with more than 20 tenacula . . . *T. bartai*

4. **Discussion**

All of the above-mentioned species (also in Table 1) are recorded to be associated with dendrotelmata to a certain degree, whether they are specialized to complete their life cycle in this ecosystem or they are only opportunistic is a different matter. As mentioned by Oboňa and Ježek [25] the extreme variation of environmental conditions such as pH, temperature, frequent water loss with rapid flooding and, oxygen deficit can cause the death of non-specialized species, especially in larval stages that are using dendrotelmata as an irregular breeding/developing site, while specialized species can endure these harsh environmental variations thriving through all their life cycle.

The occurrence of *Clogmia albipunctata*, *C. xylophilus*, *Pneumia canescens*, *P. trivialis*, *Psychoda alternata*, *P. cinerea*, and *P. minutata* in dendrotelmata is rather incidental or highly understudied [23,25]. In other words, the presence of these species could be an extension of their regular development sites (e.g., small water bodies, streams, etc.) and adults happen to find water-filled tree holes that are a potential development site for their offspring, thus, they could develop in Dendrotelmata and survive, but their long-term usage of this site is not well documented. Further studies in different habitat could provide key information to better understand the ecological relationship between water-filled tree holes and the species of Psychodinae that develop in them. To the date, only a handful of studies specifically targeted moth flies in dendrotelmata.

*Clogmia albipunctata* is broadly distributed both in Europe and worldwide and is the most synanthropic species inside the Psychodidae fauna, it can be commonly found inside buildings, and sewage treatment plants in almost every city. The high synanthropy is shared with some species of the genus *Psychoda*, which are also often found in cities, including *P. alternata* and *P. cinerea*, this species can certainly adapt and develop in a wide range of habitats, including dendrotelmata, therefore, these species could be classified as generalists when it comes to sites for larval development and not bound to develop in water-filled tree holes. Some further *Psychoda* species may be found in dendrotelmata, but the development of most species is still unknown.

*Lepiseodina tristis*, *L. rothschildi*, *Telmatscopus advena* and *T. laurencei* are well-documented species that develop inside dendrotelmata (Table 1) with multiple records of both adults and larvae. On the contrary, *Telmatscopus thuringicus* and *T. bartai comb. nov.* are only assumed to be dendrolimnetic based on observations of the closely related species *T. advena* [5]. In the case of the herein described species *Lepiseodina latipennis*—some specimens collected in Italy
were reared from decaying organic matter collected in a dendrotelmata from a maple tree (*Acer sp.*), further specimens collected in Germany came from a Malaise Trap placed next to a water-filled tree whole in an Oak tree, thus proving that this species develop inside dendrotelmata as congeneric species.

5. Conclusions

In Europe, only 13 species of Psychodinae are known to develop inside water-filled tree holes, after our extensive record search we report that the Psychodinae species develop in 13 different tree species, including two new tree species in which no previous record of larval development was reported. We redescribed *Lepisodina latipennis* through holotype and new material examination, and we report it for the first time in Germany. We also report *Telmatoscopus bartai* comb. nov. for the first time in Germany, and we provide a generic discussion of *Telmatoscopus*. Water-filled tree holes (dendrotelmata) are usually associated with old trees which are commonly endangered through the forest management strategies applied in European countries, and they are becoming rare to find inside forests. To summarize, there is a gap in the knowledge of the ecological interactions inside the Psychodinae and their environment, further studies can potentially provide new information, new records and new interactions that would be beneficial to better understand the ecosystems. Furthermore, old tree individuals remain a key component in the forests, as they harbor high biodiversity that is still understudied and they should remain untouched until the natural decomposition takes place.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/d14070532/s1, Table S1: Examined material collection Data.

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