

## Article

# Specific and Intraspecific Diversity of Symphypleona and Neelipleona (Hexapoda: Collembola) in Southern High Appalachia (USA)

Caroline D. Dukes<sup>1</sup>, Frans Janssens<sup>2</sup>, Ernesto Recuero<sup>1</sup> and Michael S. Caterino<sup>1,\*</sup> 

<sup>1</sup> Department of Plant & Environmental Sciences, Clemson University, Clemson, SC 29634, USA

<sup>2</sup> Evolutionary Ecology Group, Department of Biology, University of Antwerp, 2020 Antwerp, Belgium

\* Correspondence: mcateri@clemson.edu

**Abstract:** Collembola, commonly known as springtails, are important detritivores, abundant in leaf litter and soil globally. Springtails are wingless hexapods with many North American species having wide distributions ranging from as far as Alaska to Mexico. Here, we analyze the occurrence and intraspecific diversity of springtails with a globular body shape (Symphypleona and Neelipleona), in southern high Appalachia, a significant biodiversity hotspot. The peaks of high Appalachia represent ‘sky islands’ due to their physical isolation, and they host numerous endemic species in other taxa. We surveyed globular Collembola through COI metabarcoding, assessing geographic and genetic diversity across localities and species. Intraspecific diversity in globular Collembola was extremely high, suggesting that considerable cryptic speciation has occurred. While we were able to associate morphospecies with described species in most of the major families in the region (Dicrytomidae, Katiannidae, Sminthuridae, and Sminthurididae), other families (Neelidae, and Arrhopalitidae) are in more pressing need of taxonomic revision before species identities can be confirmed. Due to poor representation in databases, and high intraspecific variability, no identifications were accomplished through comparison with available DNA barcodes.

**Keywords:** metabarcoding; Appalachian Mountains; cryptic species; springtails; soil biodiversity



**Citation:** Dukes, C.D.; Janssens, F.; Recuero, E.; Caterino, M.S. Specific and Intraspecific Diversity of Symphypleona and Neelipleona (Hexapoda: Collembola) in Southern High Appalachia (USA). *Diversity* **2022**, *14*, 847. <https://doi.org/10.3390/d14100847>

Academic Editors: Luc Legal, Bruno Bellini, Wanda Maria Weiner and Bruna Winck

Received: 26 July 2022

Accepted: 29 September 2022

Published: 7 October 2022

**Publisher’s Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



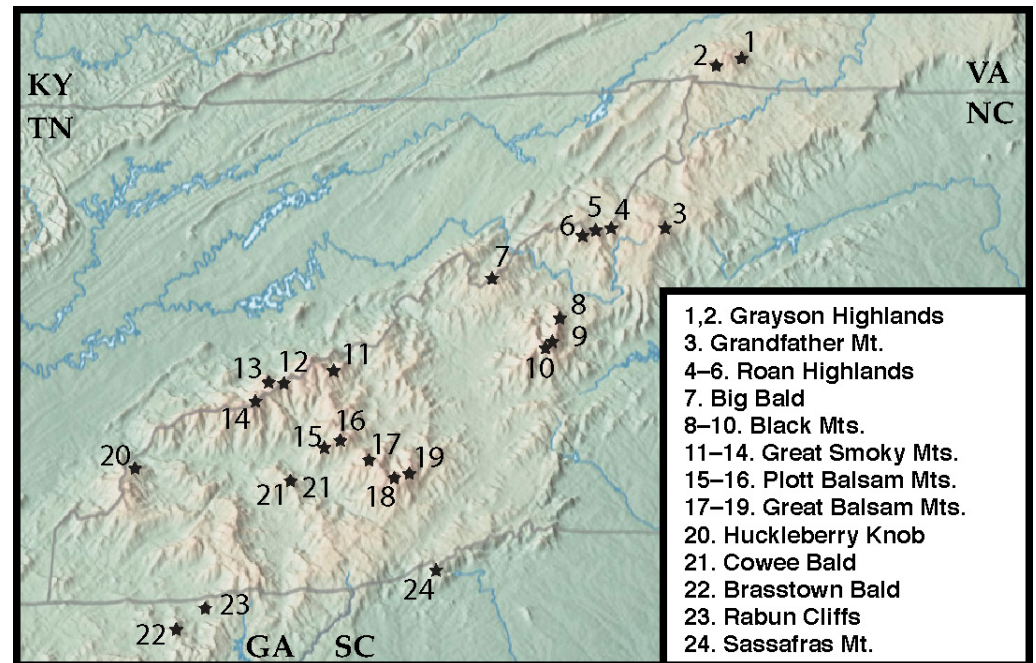
**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The Appalachian Mountains are one of the world’s oldest ranges, and they host an exceptionally diverse biota. The range extends from Alabama to southeastern Canada, and encompasses a wealth of natural communities. The Appalachian fauna has evolved over millennia of climatic fluctuations, with many elements believed to have persisted over tens of millions of years (e.g., Plethodontid salamanders; [1]). Lineages have diversified and adapted in response to these fluctuations, alternately retreating to and expanding from scattered refugia [2]. In the southern Appalachians and the Blue Ridge region, this is reflected in numerous short range endemic taxa, with distributions less than 1000 km<sup>2</sup> [3]. Some groups, especially small arthropods, such as Coleoptera [4] and Collembola [5], have very high diversity in the area and yet have received little taxonomic attention.

One of the most distinctive environments in the southern Appalachians is the high elevation Red spruce-Fraser fir (*Picea rubens* Sarg. & *Abies fraseri* (Pursh) Poir.) forest belt. These sky island forests are found only in the highest portions of southern Appalachia, in eastern Tennessee, western North Carolina, and southwestern Virginia, where elevations exceed 5500 ft. (1700 m) (see Figure 1). Widespread and more contiguous during glacial advances, these isolated forests now persist on a few dozen scattered peaks. These forests host numerous endemic arthropods (including *Trechus* ground beetles [6], *Geostiba* rove beetles [7], and *Adelopsis* fungus beetles [8,9], *Dasycerus* beetles, [10], the collembolan genus *Intricatonura* [11], and many others). Genetic diversity within many such lineages (e.g., *Hypochilus pococki* [12], and the federally endangered mygalomorph spider *Microhexura*

*montivaga* Crosby & Bishop [13]) has been shown to be high, revealing another dimension of cryptic diversity. This rich, restricted fauna is increasingly imperiled by threats from climate change [14–16] and invasive species [17,18].



**Figure 1.** Map of High Appalachia, with sampling sites numbered as in Table 1.

**Table 1.** Summary data for localities sampled.

Locality	Number on Figure 1	Morphosp Abbrev	State	Lat.	Long.	Elevation (m)
Mount Rogers	1	MRg	VA	36.6602	−81.5447	1746
Whitetop	2	WT	VA	36.6388	−81.6062	1682
Grandfather Mountain	3	GrM	NC	36.1118	−81.8105	1812
Grassy Ridge Bald	4	GRB	TN/NC	36.1001	−82.0818	1878
Roan High Bluff	5	RHB	NC	36.0933	−82.1447	1910
Roan High Knob	6	RHK	TN/NC	36.1045	−82.1224	1916
Big Bald	7	BgBld	TN/NC	35.9893	−82.4903	1681
Celo Knob	8	CK	NC	35.8527	−82.2487	1928
Big Tom	9	BT	NC	35.7798	−82.2599	2006
Mount Mitchell	10	MM	NC	35.7644	−82.2641	2037
Big Cataloochee Mountain	11	BCat	NC	35.6722	−83.1756	1876
Mount Kephart	12	MK	NC/TN	35.6311	−83.3895	1895
Newfound Gap (off Hwy 441)	13	Hwy	TN/NC	35.624	−83.4163	1394
Clingmans Dome	14	CD	TN/NC	35.5623	−83.5036	2025
Browning Knob	15	BrK	NC	35.463	−83.1319	1902
Mount Lyn Lowry	16	LL	NC	35.4635	−83.1107	1902
Richland Balsam	17	RB	NC	35.363	−82.989	1954
Mount Hardy	18	MHy	NC	35.3032	−82.9276	1865
Black Balsam Knob	19	BBK	NC	35.3258	−82.8777	1894
Huckleberry Knob	20	HKnb	NC	35.3216	−83.9929	1683
Cowee Bald	21	CB	NC	35.3287	−83.3366	1506
Brasstown Bald	22	BBld	GA	34.8763	−83.8107	1457
Rabun Cliffs	23	RC	GA	34.9707	−83.3008	1320
Sassafras Mt.	24	Sass	SC	35.0658	−82.7763	1083

Springtails, or the hexapod class Collembola, are important detritivores, contributing to decomposition of organic debris on the forest floor [19,20]. Their common name refers to a spring-operated jumping mechanism possessed by most, comprising an abdominal furca and retinaculum. The scientific name refers to the collophore, which is a unique abdominal appendage that characterizes all Collembola. Collembola are ubiquitous in leaf litter environments on forest floors worldwide [21], and the high elevations of the southern Appalachians are no exception [5,22]. Despite their small size, Collembola often have wide distributional ranges, species sometimes spanning continents or occurring across, for example, the Nearctic and Palearctic regions [23,24]. Whether this results from high dispersal rates through unknown mechanisms, or simply from coarse and inadequate taxonomic resolution is very unclear, but would suggest relatively low rates of endemism in otherwise distinctive faunas, like high Appalachia.

Here, we explore the taxonomic and genetic diversity within two orders of Collembola, the Symphypleona and Neelipleona, in the higher elevations of southern Appalachia. Members of these orders appear to have fused body segments, giving them a globular shape. These minute arthropods have received scant attention in the southeastern US, let alone in any specific subregions like high Appalachia. Bernard & Felderhoff [5] provided a brief review of the Collembola fauna of Great Smoky Mts National Park, but this did not focus on higher elevations, did not include a species list, and made only passing mention of Symphypleona or Neelipleona. Wray [25] did provide a species list for the Great Smoky Mountains, but the taxonomy has changed considerably since then, and numerous potentially occurring species have been described or separated out since (e.g., [26–34]). Direct information on the Symphypleona of the region can otherwise only be gleaned from general references in Christiansen & Bellinger's [22] Collembola of North America, most records in which are not resolved below the state or county level. Resources available nevertheless suggest as many as 67 described Symphypleona and Neelipleona species potentially occurring in the southern Appalachians (Dukes & Caterino, unpub.).

By applying an intensive COI metabarcoding approach (e.g., [35]), we simultaneously assess species level diversity of globular Collembola in southern high Appalachia, attempting to identify specimens by their barcodes, and species coherence, assessing the degree to which morphologically delimited species correspond to ones suggested by genetic data. Although COI can provide only preliminary insight into cryptic species diversity [36,37], species delimited on the basis of COI sequences can provide initial hypotheses for further testing using other genes and novel morphological characters (e.g., [38]), as well as helping to delimit evolutionarily significant units for conservation management [39,40].

## 2. Materials and Methods

As part of a larger inventory of leaf litter inhabiting arthropods, litter samples were obtained from 26 high elevation localities (>3300 ft or 1000 m) across 5 states (Virginia, North Carolina, Tennessee, South Carolina, Georgia) over the years 2018–2021 (see Figure 1 for general localities and Table 1 for details on each site). We visited most sites on two different dates, roughly in spring and fall timeframes. On each visit we took at least 3 leaf litter samples by sifting. Litter in most spruce-fir sites consists of deep needle litter, with minor components of deciduous leaves and fine woody debris. Litter was sifted down to the soil surface (or to a depth where litter was so decayed as to be indistinguishable from soil, where the interface was not a hard boundary), over an area of approximately one square meter, through an 8 mm mesh, until a bag of approximately 6 L was filled. Precise GPS coordinates were captured for each sample. Samples were processed in the lab using Berlese-Tullgren funnels, running subsamples until thoroughly dry, approximately 12 h per batch. Specimens were collected directly into 100% ethanol, and moved to –20 C storage after each subsample was complete. Springtail specimens were removed from bulk samples and sorted to morphospecies.

The analyses here include 204 Symphypleona and Neelipleona sequences plus an outgroup *Isotoma* (Isotomidae) sequence. These represent one individual of each mor-

phospecies from 41 sampling events (41 sets of samples from a given site/date). Full collecting data for each specimen extracted are available in Table S1. Multiple individuals of a putative morphospecies were only included from a site if they were collected on different dates. Specimens were tentatively identified using Christiansen & Bellinger's 'Collembola of North America' [22] and through comparisons to specimen photographs online that had been identified by specialists (e.g., collembola.org; [41]). Each specimen was imaged, subdivided or punctured to permit tissue digestion, and placed in a separate well in a 96-well plate. Images of morphospecies are archived on our lab Flickr page (<https://flickr.com/photos/183480085@N02/albums/72157720213462655>; accessed on 16 May 2022), identifiable by morphospecies code (site.visit.###, as given in Table S1). Tissues were digested with lysis buffer and proteinase K (Omega BioTek, Norcross, GA, USA), then the liquid fraction was removed to a new plate, with the voucher remains saved for archiving. The digested tissue mixture was extracted using Omega BioTek's MagBind HDQ Blood and Tissue kit on a Hamilton Microlab Star automated liquid handling system, eluting with 150 µL elution buffer.

Following digestion, remains of extracted specimens were recombined with any non-extracted body parts, labelled, assigned unique CUAC (Clemson University Arthropod Collection) identifiers, and curated into the CUAC. Unextracted representatives of morphospecies, if any, remain in bulk order-level samples, and are also permanently vouchered in the CUAC, as are unsorted residues (containing additional representatives of hyperabundant taxa, principally Acari and Collembola).

These analyses include sequences from three separate sequencing approaches. For one plate of extracts we amplified a 658 base pair region of the cytochrome oxidase one (COI) mitochondrial 'barcoding' gene using primers LCO1490 and HCO2198 (GGTCAACAAAT-CATAAAGATATTGG & TAAACTTCAGGGTGACCAAAAAATCA, respectively; [42]). These PCR products were run on an agarose gel to assess amplification success and sent for clean-up and Sanger sequencing to Psomagen (Rockville, MD, USA); amplicons were sequenced in both directions. This produced 64 of the sequences used here. The other specimens were sequenced using next generation platforms as 'mini-barcodes', a 421 bp fragment of the mitochondrial COI gene using the primers BF2-BR2 (GCHCCHGAYA-TRGCHTTYCC & TCDGGRTGNCCRAARAAYCA, respectively; [43]), corresponding to the downstream two-thirds of the standard barcoding region. Each well was tagged with a unique combination of forward and reverse 9 bp indexes, synthesized as part of the primer by Eurofins Genomics (Louisville, KY, USA). These indexes were derived from a list provided by Meier et al. [44], to allow multiplexed next-generation sequencing. All PCRs were conducted in 12.5 µL volumes (5.6 µL water, 1.25 µL Taq buffer, 1.25 µL dNTP mix [2.5 mM each], 0.4 µL MgCl [50 mM], 1.5 µL each primer, 0.05 µL Platinum Taq polymerase, 1 µL DNA template, with a 95 C initial denaturation for 5 min, followed by 35 cycles of 94 C (30 s), 50 C (30 s), 72 C (30 s), and a 5 min 72 C final extension on an Eppendorf Gradient Mastercycler.

For Illumina library preparation, PCR products were combined and purified using Omega Bio-Tek's Mag-Bind Total Pure NGS Kit, in a ratio of 0.7:1 (enriching for fragments > 300 bp). Illumina adapters and sequencing primers were ligated to PCR products using New England BioLab's Blunt/TA Ligase Master Mix. The amplicon + adapter library was again purified using Mag-Bind Total Pure NGS, and subsequently quantified using a Qubit fluorometer. This final library was sequenced on an Illumina MiSeq using a v.3 2 × 300 paired-end kit. Nanopore libraries were prepared using the ligation sequencing kit LSK-112 (Oxford Nanopore Technologies, Oxford, UK) and sequenced on a MinION using a v10.4 flowcell.

Sanger sequences were edited in Geneious (v8.1.8) by combining forward and reverse reads, confirming basecalls, and exporting as text. Illumina reads were processed with bbtools software package (<https://jgi.doe.gov/data-and-tools/bbtools/>; v38.87 [45]; accessed on 10 February 2022) to merge paired read ends, remove PhiX reads, trim Illumina adapters, filter reads for the correct size, remove reads with quality score < 30, cluster



sequences by similarity allowing 5 mismatches (~1%) and generate a final matrix in FASTA format. Nanopore reads were basecalled using the ‘super-accurate’ algorithm of Guppy (v6.1.2), then demultiplexed using ONTbarcoder v0.1.9 [46], with minimum coverage set at 5. FASTA files from all sequencing methods were trimmed to match the shorter 421 bp BF2-BR2 fragment, combined, and aligned with the online version of Mafft v7 [47] using the auto strategy.

Phylogenetic reconstructions were performed using Maximum likelihood (ML) and Bayesian inference (BI) methods, providing trees for assessment as to species coherence and identity. The ML analysis was done with W-IQ-TREE v2.0 [48,49], available at <http://iqtree.cibiv.univie.ac.at> (accessed on 9 August 2022). This program was used also to determine the best substitution model for our data. We set a perturbation strength of 0.4. and an IQ-TREE stopping rule value at 200. Branch support is based on an ultrafast bootstrap analysis [50], run with 1000 bootstrap replicates with a minimum correlation coefficient of 0.99. Bayesian analysis was performed using BEAST v1.10.4 [51], with a dataset including no outgroups, a relaxed lognormal molecular clock and a birth-death incomplete sampling speciation tree prior [52]. Given the absence of an adequate fossil record to calibrate a molecular clock for our data, we used an estimated substitution rate for COI of  $0.0169 \pm 0.0019$  [53], which has been the most commonly used in Collembola studies [54–57]. The analysis was run for  $30 \times 10^6$  generations sampling every 30,000, and repeated independently three times to assess the consistency of the results. We used Tracer v1.7 [58] to determine that effective sample sizes (ESS) of the generated statistics were higher than 200. Finally, we built a maximum clade credibility (MCC) tree using TreeAnnotator v1.10.4, excluding the first 2000 trees as burn-in, that was midpoint rooted in FigTree v1.4.0 (<http://tree.bio.ed.ac.uk/software/figtree/>; accessed on 5 September 2022).

For automated species delimitation we used five different single-locus delimitation methods. Automatic Barcode Gap Discovery (ABGD; [59]) and Assemble Species by Automatic Partitioning (ASAP; [60]) are both based on the characterization of barcode gaps from pairwise genetic distances. Two different implementations of the Poisson Tree Process (PTP) method [61] were also used, Bayesian PTP (bPTP) and multi-rate PTP (mPTP) [62]; this method is based on the detection of transitions in branching rates on a phylogenetic tree according to speciation and coalescent models. This is also true for the other method used, the General Mixed Yule-Coalescent model (GMYC) [63], that uses an ultrametric tree to estimate those rate transitions. ABGD analysis was performed using the web version (available at <https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html>; accessed on 16 May 2022) using Jukes-Cantor (JC69) genetic distances and setting a prior maximum divergence of intraspecific diversity (P) from 0.005 to 0.15, with a relative gap width (X) of 1 and a number of bins of 20. ASAP was run using the web version (available at <https://bioinfo.mnhn.fr/abi/public/asap/asapweb.html>; accessed on 16 May 2022), with Kimura 2-parameter [64] distances and a split probability of 0.01. For both PTP analyses we used our ML tree as input. We used the bPTP web server (available at <https://species.h-its.org>; accessed on 16 May 2022), running the analysis for  $5 \times 10^5$  generations, removing the outgroup from the tree and using a thinning of 500 and a burn-in of 0.2. The mPTP analysis was performed using the web server (available at <https://mptp.h-its.org/#/tree>; accessed on 16 May 2022), removing the outgroup from the tree. The GMYC web server (available at <https://species.h-its.org/gmyc/>; accessed on 16 May 2022) used the BEAST ultrametric tree as input, running both single and multi-threshold methods (see [65]). We examined clades corresponding to morphospecies to assess diversity across sampling sites. Any sequences for which we did not have an a priori identification were searched on the Barcoding of Life database and on GenBank through BlastN for tentative matches. However, this did not identify any additional sequences.

### 3. Results and Discussion

The Bayesian tree showing hypothesized species delimitations is shown in Figure 2 (outgroup removed), with the more readily identifiable species indicated. Our dated tree is available as Figure S1. Automated single-locus species delimitation methods yielded diverse results for our dataset. The most conservative method was mPTP, recovering a total of 43 distinct symphypleonan and neelipleonan species. By contrast, the greatest subdivision is observed with bPTP suggesting up to 90 putative species. GMYC resulted in 74 species using the single-threshold method and 77 with the multi-threshold method; the latter can show over-split in its results [65], so we discuss the single-threshold results. As for the “barcode gap” methods, ABCD suggested the existence of 77 species in our dataset, while the best partition in ASAP resulted in a total of 82 species. The best estimated intra-/interspecific distance threshold estimated by ASAP was at 7% (K2P-corrected distance). A histogram showing the distribution of all pairwise distances is included as Figure 3. All distances cited below are K2P distances, and are interpreted relative to this hypothesized threshold.

These results would seem to be a significant overestimate in at least some of the methods. Our own morphological identifications and morphospecies sorting would have suggested a more modest 25–30 species. Under all automated methods, some apparently morphologically uniform species were separated into multiple species. For example, *Ptenothrix atra* was split into at least 9 species. However, levels of divergence were also consistently very high for COI, reaching 18% uncorrected (22% corrected) distance between Grandfather Mt and other localities for *P. atra*, for example. Relatively high distances have been reported in other intraspecific studies of springtails [66–69]; Porco [66], for example, considering 14% (K2P) distance to represent a conservative intraspecific cutoff. Clearly rates of mitochondrial evolution are much higher in globular Collembola than in other arthropods commonly examined in the barcoding literature (e.g., [70–73]). However, it is nonetheless worth considering that, where highly divergent and strongly supported intraspecific lineages show geographical coherence, there may be considerable cryptic species diversity in the fauna. We would not consider COI alone sufficient basis for concluding that cryptic species were present, but it is certainly a hypothesis worth examining further with additional data.

Second, attempts to identify any of these sequences via DNA barcodes using a variety of algorithms against BOLD and GenBank databases failed completely. Between poor representation of these groups (and litter arthropods in general; Recuero & Caterino, in prep.) in public databases, and extremely high degrees of COI divergence within and across lineages, our Symphypleona and Neelipleona sequences were not sufficiently close to any publicly available sequences to strongly support identifications at any taxonomic level—no sequence was less than 5% different from anything available, and many very incorrect (to arthropod order) possibilities were only a few percent more distant. Submission of these sequences will aid in future efforts, but these problems are likely to plague litter and soil arthropod identification for the foreseeable future, until major investments are made in establishing comprehensive reference sequences across arthropod taxa and geographic areas.

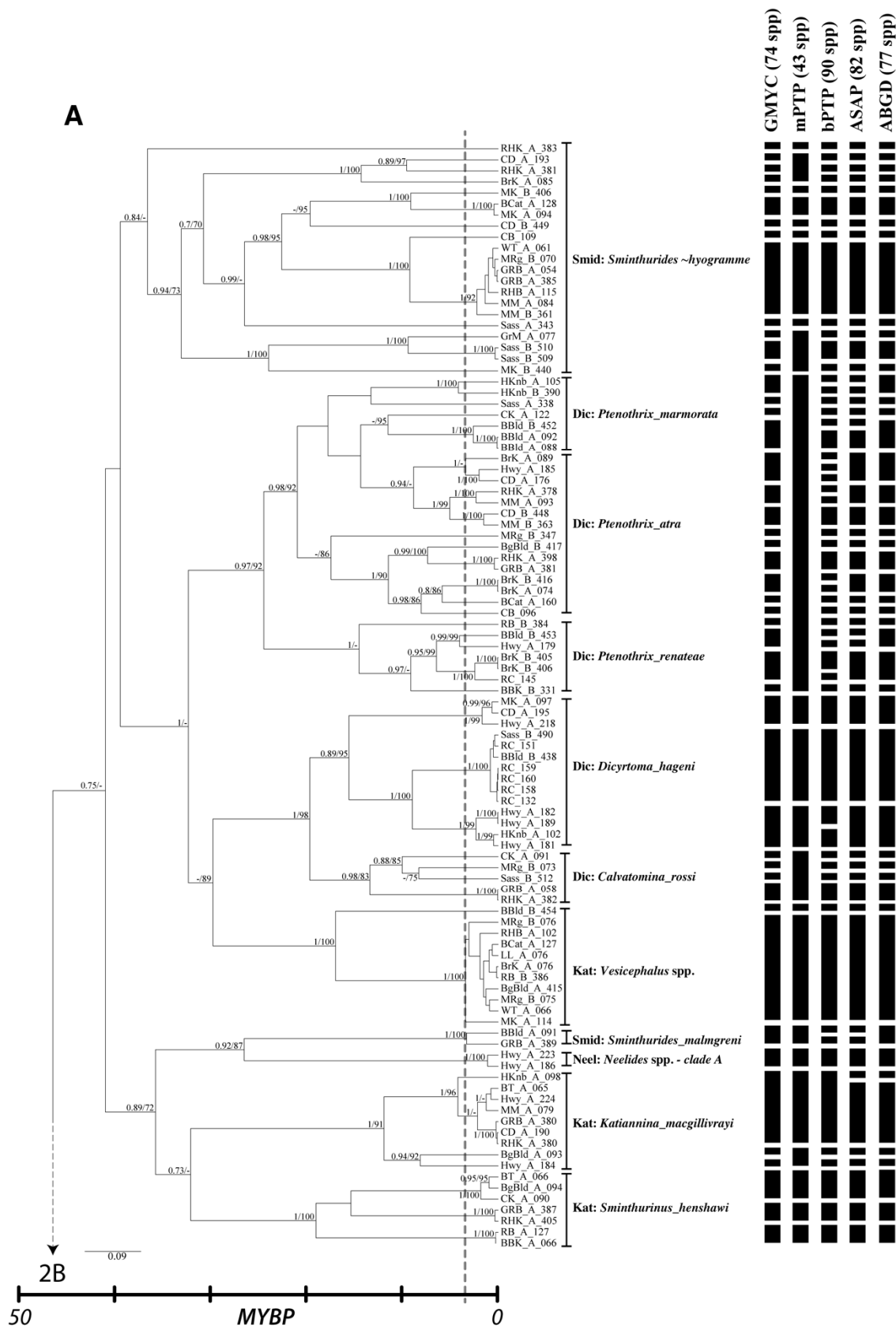
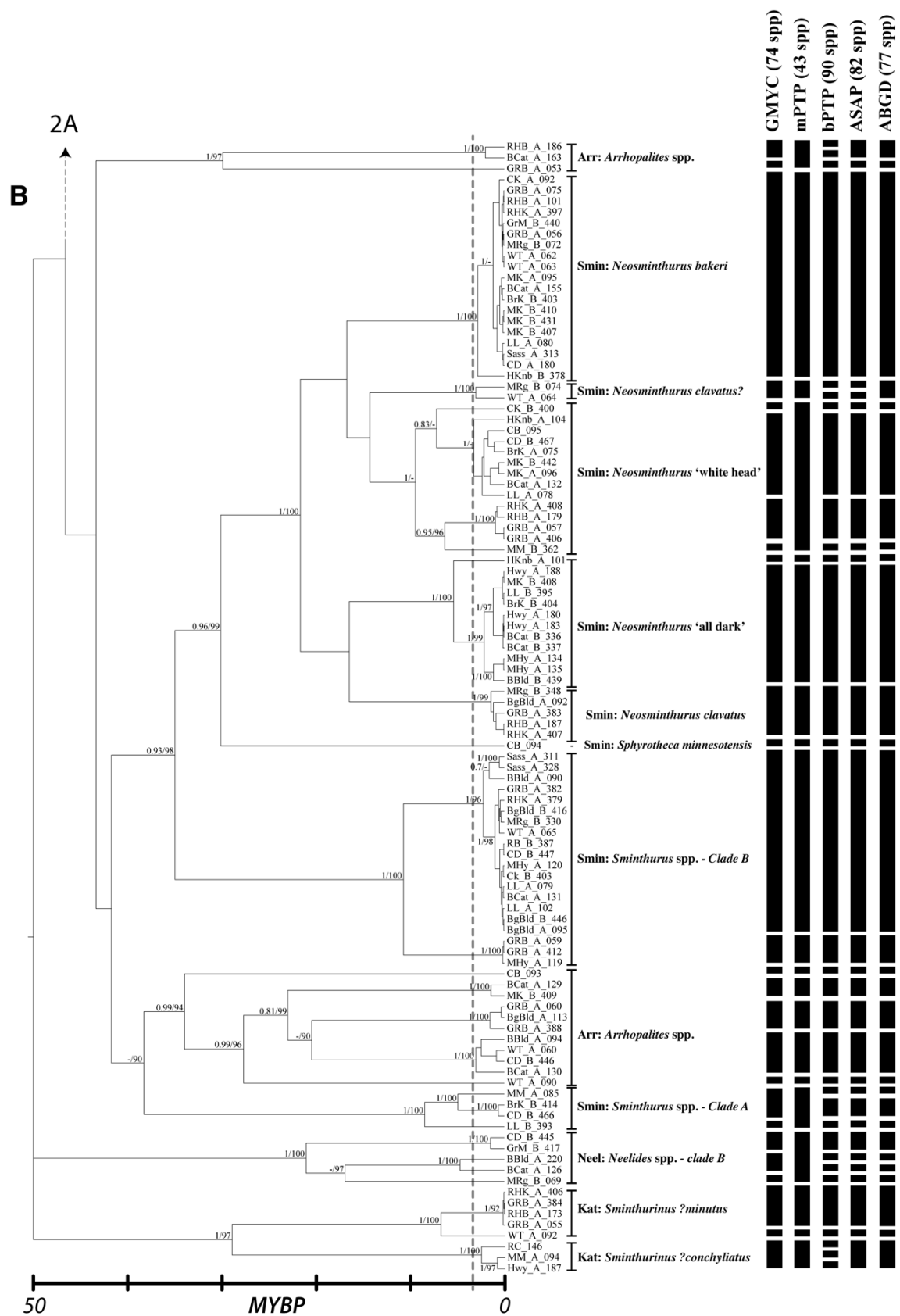
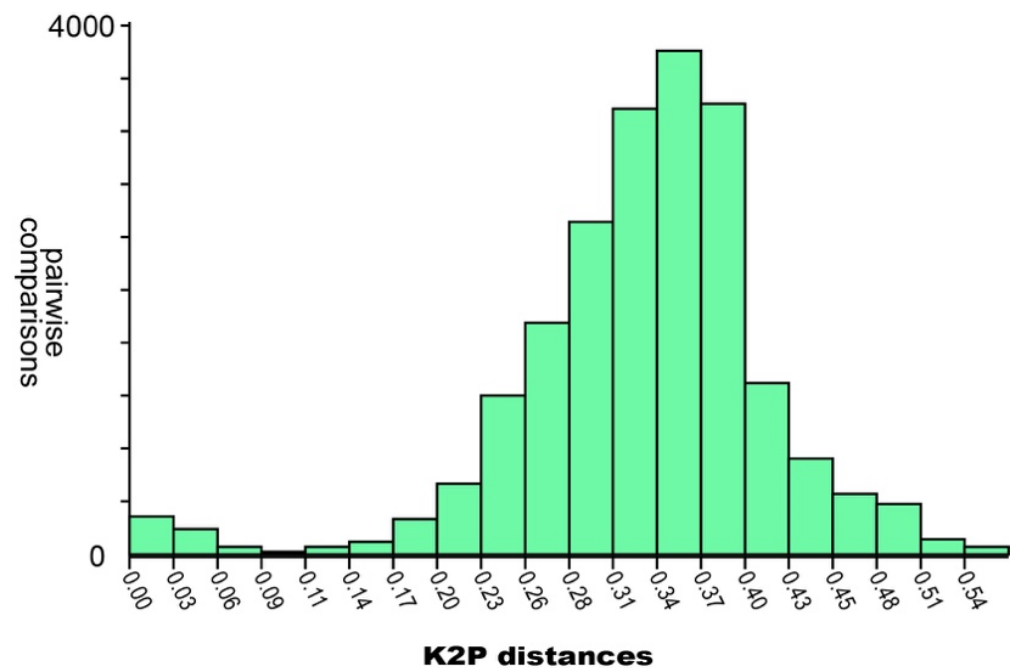


Figure 2. Cont.



**Figure 2. (A)** Time-calibrated phylogeny showing species delimitations Scale bar below is in million years before present. Dotted vertical line indicates the approximate 'intraspecific barcoding threshold', ~7% K2P distance or ~3MYBP. Numbers on branches indicate posterior probabilities/mL bootstrap values. **(B)** Phylogeny showing species delimitations.



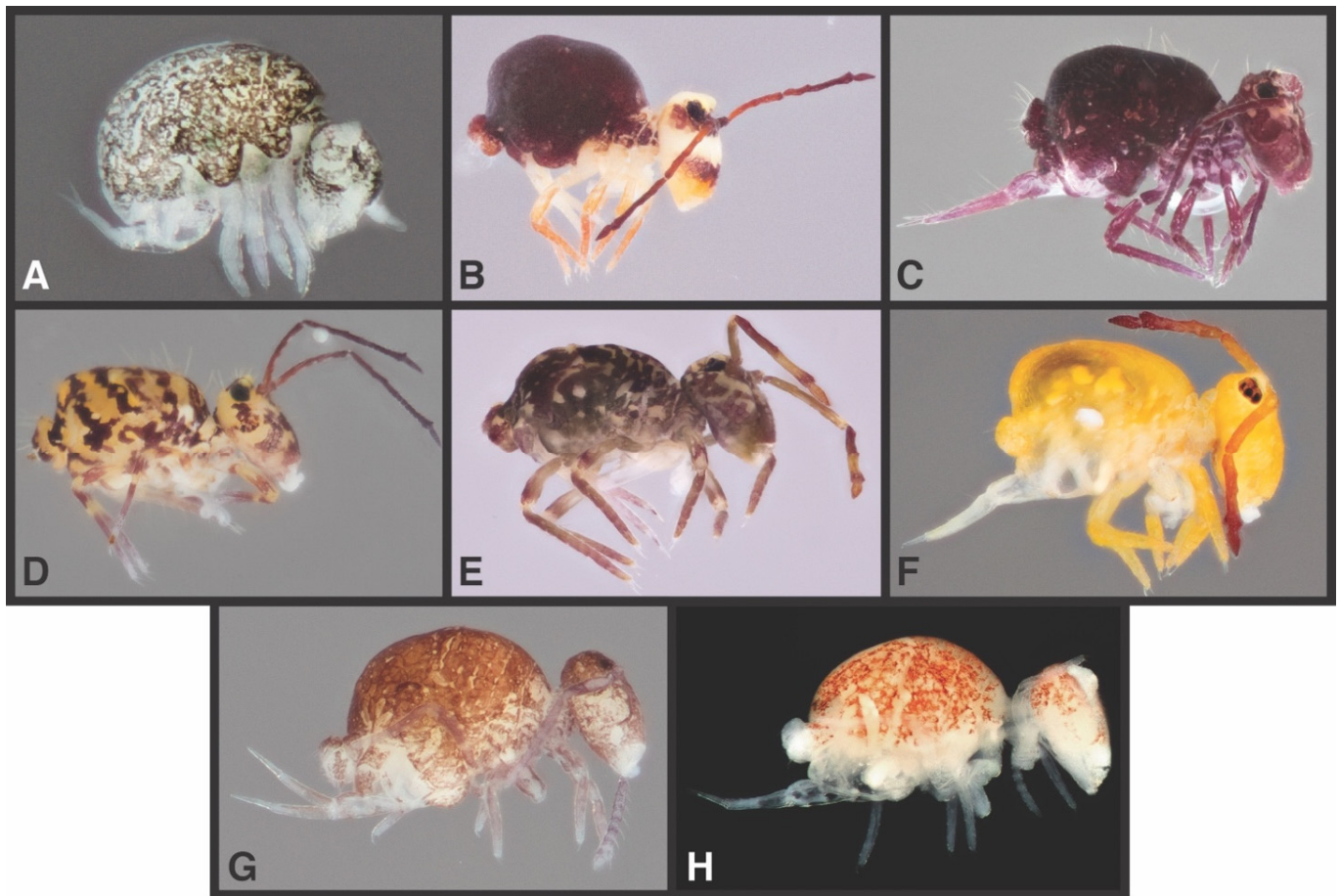


**Figure 3.** Histogram of all pairwise K2P distances, from ASAP.

### 3.1. *Neelipleona*

#### Neelidae

Neelidae were represented by seven individuals, all morphologically identified as *Neelides Caroli* (Figure 4A). These were resolved into two widely separated clades that are nearly 40% divergent. While this group is too poorly represented in our data to reach any serious conclusions about biogeographic relationships or taxonomy, delimitation analyses suggest anywhere from 2 to 5 species, with at least two of them approximately sympatric in the central Great Smoky Mountains (the ‘Hwy’ clade and one or more lineages from Clingmans Dome and Big Cataloochee Mt.) The larger lineage, which includes the latter individuals, comprises three highly divergent (>20%) lineages, with these two having as their closest relatives other individuals from distant localities, Brasstown Bald and Grandfather Mt, respectively. The third lineage includes only a single individual from Mt. Rogers in the northeast, and it seems reasonable to hypothesize that all three of these represent distinct species, giving a total of 4. Only two species of *Neelides* have been reported from southern Appalachia, *N. diana*e Christiansen and Bellinger, and *N. minutus* (Folsom) [22], although even they recognized the possibility that the latter might be a complex of species. Unfortunately, none of these vouchers is adequately preserved to seriously assess identities based on specific morphological characters. More material and comparisons with type specimens will be necessary to sort out just how many species we’ve sampled, and whether they correspond to described ones or not.



**Figure 4.** Photographs of Appalachian Neelipleona and Symphypleona. (A) Neelidae: *Neelides*. (B) Dicyrtomidae: *Dicyrtoma hageni*. (C) Dicyrtomidae: *Ptenothrix atra*. (D) Dicyrtomidae: *Ptenothrix marmorata*. (E) Dicyrtomidae: *Ptenothrix renateae*. (F) Dicyrtomidae: *Calvatomina rossi*. (G) Arrhopalitidae: *Arrhopalites* sp.1 (BBld.A.094). (H) Arrhopalitidae: *Arrhopalites* sp.2 (GRB.A.053).

### 3.2. Symphypleona

#### 3.2.1. Dicyrtomidae

We recovered at least five morphological species of Dicyrtomidae, *Dicyrtoma hageni* (Folsom) (f. *frontalis*), *Calvatomina rossi* (Wray), *Ptenothrix atra* (Linnaeus), *P. renateae* Snider, and *P. marmorata* (Packard). These are estimated to represent between 5 (mPTP) and 33 (bPTP) species by delimitation analyses. The lower estimate lumps several well-differentiated morphospecies together and can be disregarded, while all the other estimates finely subdivide each of the species of *Ptenothrix*. The reality certainly lies somewhere in between.

*Dicyrtoma hageni* (Figure 4B) was found only in localities southwest of the French Broad River, from Mount Kephart in the Smokies south to Brasstown Bald in north Georgia. These localities fall into three highly distinct genetic clades, and are reconstructed as three species by 4 of 5 delimitation analyses. The fact that we observe some sympatry (‘Hwy’ localities in the Great Smoky Mts) among these very divergent clades lends support the idea that there are indeed multiple cryptic species present. One of these clades includes only our southernmost localities, spanning north Georgia and South Carolina, none of which have any spruce-fir component. This suggests that some ecological differentiation might also have occurred.

*Ptenothrix renateae* (Figure 4E) was described from north Georgia and lower elevations of South Carolina, while our records extend this northward into the Great Balsam Mts. of southwestern North Carolina. bPTP results suggest that each of the six localities for

*P. renateae* represents a distinct species, which would initially seem to be an overestimate, as they cover no more than 100 linear km. However, no delimitation analyses support fewer than four species, and divergences among them mostly exceed ASAP's estimated threshold (ranging to 21% between the Richland Balsam and Black Balsam Knob populations, for example). *Ptenothrix atra* (Figure 4C) was found over a broad area from the higher parts of the Smokies (Big Cataloochee Mt.) in the west to Mt. Rogers in the northeast. However, these represent two deeply divergent clades that are not resolved as each other's closest relatives. Delimitation analyses subdivide these into 9 or more species. These larger clades are broadly sympatric, sharing a couple of localities (Browning Knob and Roan High Knob) where individuals are >21% divergent. So there seems clear evidence that multiple cryptic species are present. Even within each larger *P. atra* clade some structuring may be significant. For example, two examples from Clingmans Dome are resolved in highly divergent lineages (almost 13% between CD.A.176 and CD.B.448) with individuals from other localities interspersed. So, in this case, more than two cryptic species seems a reasonable hypothesis.

Individuals identified as *P. marmorata* (Figure 4D) fell out in two clades within a paraphyletic set of *P. atra* lineages, mostly from the southwestern portion of our sampling region, though one immature individual from Celo Knob in the Black Mts also resolved among these. Delimitation analyses would suggest that every locality sampled represented a distinct species, and divergences across the two major lineages is comparable to those in *P. atra*, well above estimated intraspecific thresholds.

Specimens identified as *Calvatomina rossi* (Figure 4F) were found from Sassafras Mt. SC to the Roan Highlands along the NC/TN border, and represent the first records for the region, having previously only been reported from Florida [29], Massachusetts, and Illinois [20]. All localities are lumped as one species by mPTP but divided into 4 by all other delimitation analyses. There is not obvious geographic signal in the relationships apart from identity between two Roan Mt localities (GRB and RHK).

### 3.2.2. Arrhopalitidae

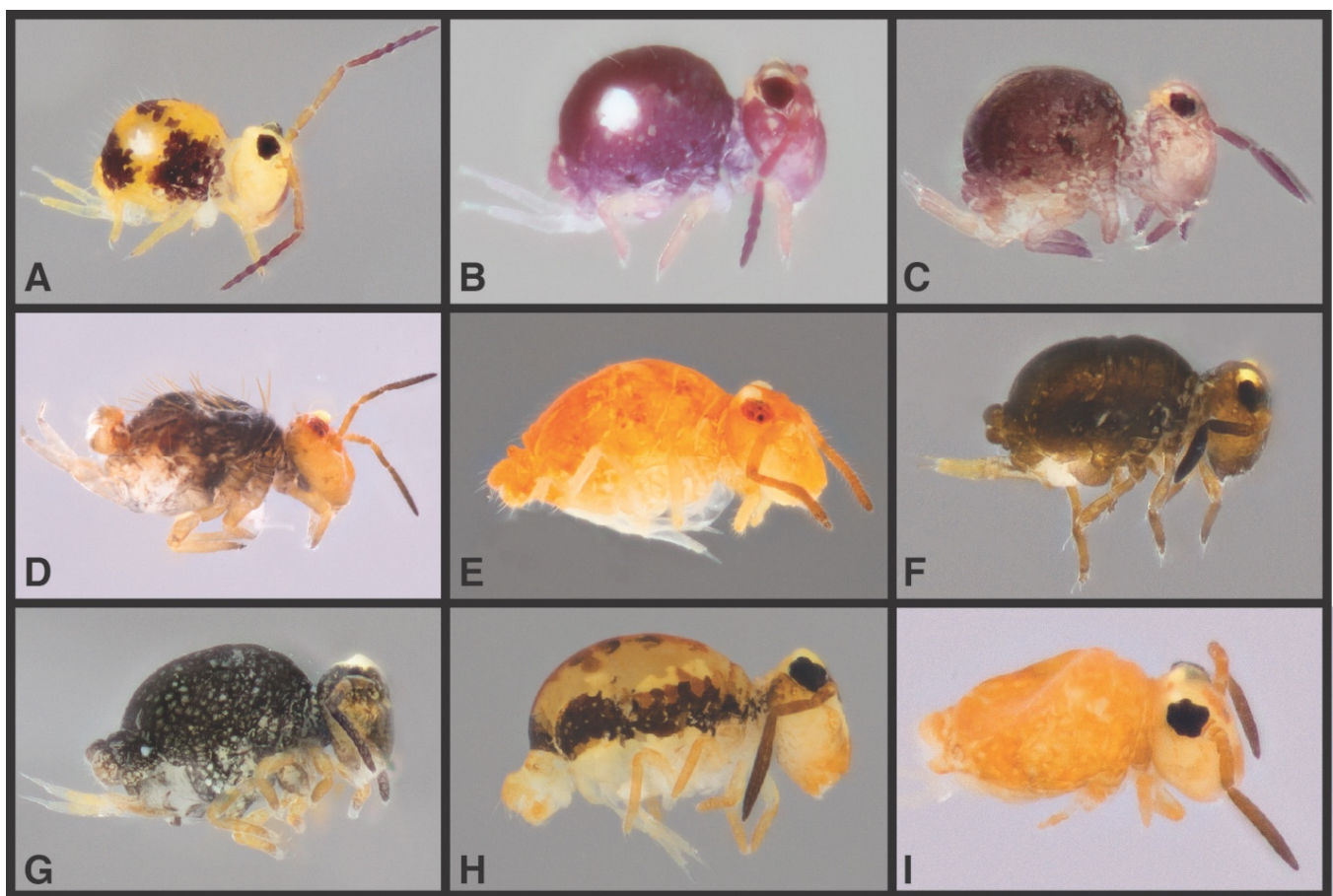
Arrhopalitidae (Figure 4G,H) is represented by 14 specimens, resolved into two independent lineages, with some apparently misplaced *Sminthurus* (Sminthuridae) close to one. All should represent the genus *Arrhopalites* (or the genus *Pygmarrhopalites* Vargovitsh, the status of which has been disputed by [74]), which contains 25 poorly defined nearctic species, with perhaps half of these expected to occur in the southern Appalachians. Delimitation analyses suggest that our sequences represent between 6 and 8 distinct species, and divergences among the lineages corresponding with geography would seem to support the higher end of this range. However, none are well-enough sampled to conclude much now. One larger cluster of 7 sequences (GRB.A.060 to BCat.A.130) comprises all darker blueish specimens with rather distinctive lighter patterning, but even across two subgroups here (e.g., GRB.A.388 vs. WT.A.060), divergences reach nearly 30%. Otherwise, darker and lighter rust-colored forms are intermingled on several very long branches. This family will need much more focused attention.

### 3.2.3. Sminthurididae

The family Sminthurididae was represented by 22 specimens of *Sminthurides* (one of which is possibly an immature *Sphaeridia*; Figure 5C). These included two individuals of the Holarctic *Sminthurides malmgreni*, from Brasstown Bald, GA, and Grassy Ridge Bald in the Roan Highlands, clustered together though about 8% divergent from each other, and hypothesized as distinct by just 2 of 5 delimitation techniques. This clade was far from the remaining *Sminthurides*, which mostly corresponded to *Sminthurides hyogramme* (Figure 5A), a distinctive species with blue stripes and a bright white lateral spot, among the most common Symphypleona encountered. These spanned our entire sampling range, from Sassafras Mt., SC in the south to Mt. Rogers, VA in the north (and Brasstown Bald in the west, though that individual did not sequence successfully). There is considerable variation



in specifics of color pattern among these, some of which may be meaningful. Darker individuals, especially with a darker head, from Sassafras Mt (Figure 5B) and Grandfather Mt (GrM.A.077) cluster together, conceivably representing *Sminthurides macnamarai* Folsom & Mills (as described in [22]). There is also a clade of several individuals with a complete ventral stripe below the white lateral spot (CD.B.449, MK.B.406, MK.A.094, BCat.A.128). The latter, however, also represents a series of relatively proximate localities in the Great Smoky Mts., nested among clusters of more typical coloration, so the significance for possible specific-level differentiation is not yet clear. A singleton from Roan High Knob (RHK.A.383) has a dark body and white head, and is >25% divergent from any others. Delimitation analyses subdivide this *S. hyogramme* clade into anywhere from 9 to 13 species. We would suggest the total is more likely in the range of 3–5 species based on morphological variation. However, further molecular data will be needed to test whether more truly cryptic species are present.



**Figure 5.** Photographs of Appalachian Symphyleona. (A) Sminthurididae: *Sminthurides hyogramme* (BgBld.B.440). (B) Sminthurididae: *Sminthurides macnamarai*? (Sass.B.509). (C) Sminthurididae: *Sphaeridia* sp. (D) Katiannidae: *Vesicephalus crossleyi*. (E) Katiannidae: *Katiannina macgillivrayi*. (F) Katiannidae: *Sminthurinus conchylitatus*? (G) Katiannidae: *Sminthurinus* nr *minutus*. (H) Katiannidae: *Sminthurinus henshawi similitorus*. (I) Katiannidae: *Sminthurinus henshawi aureus*.

### 3.2.4. Katiannidae

Katiannidae are represented by at least 4 species in three genera. A total of 11 individuals of the genus *Vesicephalus* (Figure 5D) were sequenced from ten localities, spanning our whole sampled range from the Grayson Highlands in the northeast to Brasstown Bald in the southwest. Delimitation analyses are unanimous in resolving these into exactly 2 species. Though represented by only a single individual, the Brasstown Bald specimen (BBld.B.454) has dark eyes and a fairly distinctive color pattern, and is more than 20%

divergent from all others. It is possible that our samples represent *V. longisetis* (Guthrie) and *V. crossleyi* Snider. However, the type of the former is poorly preserved, and described differences between the two are of dubious value [75]. Christiansen & Bellinger [22] report both species from the region, but also suggest that what they considered *V. longisetis* could represent multiple species. Further work will be needed to conclusively identify these as either of the described or possibly (the Brasstown Bald specimen) undescribed.

The monotypic genus *Katiannina* (Figure 5E) was represented by 9 specimens from 7 peaks, resolving into two deeply divergent lineages (separated by ~20%). These almost certainly represent at least two species, and the one lineage with just two individuals from quite distant localities ('Hwy', near Newfound Gap in Great Smoky Mountains National Park and Big Bald, northeast of the Asheville Depression) is itself subdivided by most delimitation analyses. In the larger clade there is no obvious geographic structuring among populations or lineages, with an individual from Clingmans Dome very similar to one from the Roan Highlands, and the rather proximate Big Bald and Roan Highlands representatives (only 35 km apart, both on same side of French Broad River valley) are separated as far as possible in the species' cluster. Whether any of these represents the sole described species *K. macgillivrayi* (Banks), described from New York, is questionable, as all are simply pale orange and lack the 'black stripe [ . . . ] from the eye running back to the anal tubercle' originally described (though that may be a condition of a feeding instar rather than a morphological character of the species) [76]. Another form known from lowland South Carolina that we have not yet sequenced seems different still, predominantly reddish with distinct, bright white dorsal spots. The genus clearly needs further taxonomic attention.

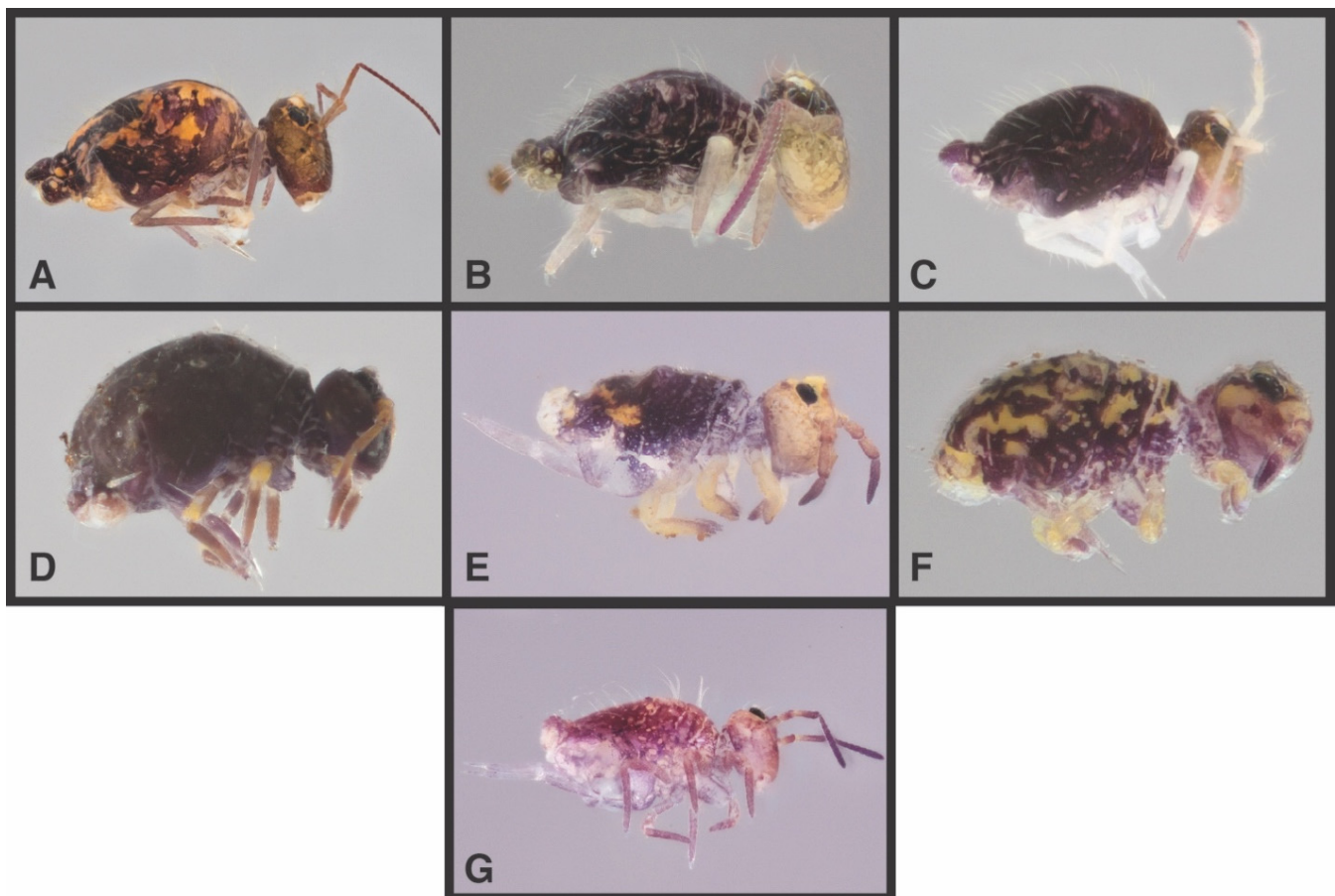
Fifteen individuals of three morphologically distinctive forms of *Sminthurinus* were sampled, though these did not all resolve together in the tree. Three individuals of a species near *S. conchylatus* Snider (spanning Rabun Cliffs, GA to Mount Mitchell, NC; Figure 5F) were resolved together, differing by at most 8.4%, only barely above the ASAP estimated threshold. These were sister to a lineage of 5 individuals of a species near *S. minutus* MacGillivray (Figure 5G), all collected from NE of the French Broad River valley, most from the Roan Highlands. These did cluster tightly together (<1% difference) relative to the one individual from Whitetop Mt. in Virginia (~17%). These (including the Whitetop specimen) seem likely to be distinct species from *S. minutus*, all possessing a distinctive, complete white cap between the eyes, and a mostly dark-colored head, where in *S. minutus* two separate white spots seem always to be present, separated by an orange wedge, with the rest of the head relatively light-colored. Christiansen & Bellinger [22] considered *S. minutus* to be a potential synonym of *S. quadrimaculatus* (Ryder), but we agree with their admitted possibility that these represent a species cluster in need of subdivision. Lastly, we obtained 7 sequences for *Sminthurinus henshawi*, 5 striped individuals representing what has been termed a form 'similitortus' (Figure 5H), and 2, a deeply divergent monophyletic sister to those, the form 'aureus', lacking longitudinal blue stripes (Figure 5I). The two 'aureus' individuals, both from the Great Balsam Mts. (RB and BBK) differ by less than 1%. The 'similitortus' types form two highly divergent clusters (>25%), one from the Roan Highlands (2 individuals < 1% different) and one with individuals from the Black Mts (Big Tom and Celo Knob, 6% different) and Big Bald (~4.5% from either of the Black Mts. individuals). Big Bald is about equidistant from either the Blacks or the Roan Highlands, so its much closer relationship to the former is surprising.

### 3.2.5. Sminthuridae

Finally, the family Sminthuridae, which formerly contained nearly all Symphypleona species (e.g., [77], is represented here by three genera. As many as 7 or 8 described species of *Sminthurus* should occur in the region, which are said to be largely indistinguishable based on external color patterns. Our first of 3 lineages ('clade A') of these (resolved as sister to the larger *Arrhopalites* clade) contains 4 individuals, mostly distinctively patterned (aside from one immature) with dark blue and strongly contrasting white dorsal stripes (Figure 6A). These range from Clingmans Dome in the west to Mount Mitchell in the



northeast (and probably includes very similar as-yet-unsequenced specimens from Mt Rogers and Whitetop even further north). Delimitation results subdividing these into 2 or 3 distinct species are difficult to evaluate, though distances among lineages do exceed estimated thresholds. In the other major lineage ('Clade B'), a tight cluster of three individuals (MHy.A.119, GRB.A.059, and GRB.A.412; (Figure 6B) seems to represent a distinct species, as supported by all delimitation analyses. Unfortunately 2 of these individuals are immature, so it is impossible to assess meaningful morphological consistencies among them. The other lineage in clade B includes 17 individuals ranging across the region from Brasstown Bald to Mt. Rogers. These exhibit remarkably low divergences among those considered here (<5%), and almost certainly do represent a single species. Assessing its morphology to attempt to identify it is complicated by the surprisingly high incidence of immatures. Mature individuals (like Sass.A.339) have most of the body dark blue, with numerous, small, obscure lighter spots, and a round, distinctively green cheek patch below the eye (Figure 6C). However, this can be clearly seen in only a handful of specimens. It is possible that this corresponds to *S. bivittatus* Snider, in which a 'gena with dark green polygons forming rosettes' is described.



**Figure 6.** Photographs of Appalachian Sminthuridae. (A) *Sminthurus* sp. (CD.B.466). (B) *Sminthurus* sp. (GRB.A.412). (C) *Sminthurus bivittatus*? (Sass.A.339). (D) *Neosminthurus* sp. 'all dark' (HKnb.A.101). (E) *Neosminthurus* sp. 'white head' (CB.095). (F) *Neosminthurus bakeri* (LL.A.080). (G) *Sphyrothea minnesotensis*.

A single individual of *Sphyrothea minnesotensis* (Figure 6G) was found at Cowee Bald, a non-spruce-fir peak at a slightly lower elevation (~1500 m). Despite its name, this species has previously been recorded through much of the eastern Nearctic, from Minnesota to Ontario to Louisiana, though never specifically from higher parts of Appalachia.

*Neosminthurus* represents the last, very commonly collected genus. Specimens have relatively short antennae, and a lightly debris-cloaked appearance, often appearing to have retained parts of previous molts on the body. There are three named species in the region, and it's likely that all are represented among our more than 50 individuals sequenced, though delimitation analyses suggest between 6 and 10 species. Resolution of exactly what clades corresponds to what species is not entirely straightforward. The largest lineage, spanning the whole range from Huckleberry Knob, NC (HKnb.B.378) and Sassafras Mt., SC (Sass.A.313), to Whitetop Mt, VA (WT.A.063), comprises mottled individuals with each antennomere apically darkened (Figure 6F), characters that correspond well to *Neosminthurus bakeri* Snider. Divergences within this cluster are relatively shallow, no more than about 2%. Another large clade (almost divided into north and south clades, apart from the 'misplaced' Celo Knob CK.B.400 individual) consists of individuals exhibiting generally white heads in mature individuals (e.g., BCat.A.132 and CB.095; (Figure 6E). Divergences between these subgroups are around 13%, though they are nearly as high within, particularly in the north group, at least comparing Mount Mitchell with the Roan Highlands group. One small clade from northeast of the Asheville depression (GRB.A.383, RHK.A.407, etc.) corresponds to *N. clavatus*, with voucher specimens exhibiting the diagnostically flattened dorsal setae. These individuals are all dark, head and body. Members of a smaller clade from further north, (MRg.B.074 + WT.A.064) also have the flattened clavate setae, but have light colored heads, suggesting homoplasy in dorsal setal morphology. Lastly, a larger clade of 12 individuals found only south of the Asheville depression (HKnb.A.101, Hwy.A.180, etc.; Figure 6D) are entirely dark, body and head, like *N. clavatus*. However, these have narrower, cylindrical dorsal setae, and would not be assignable to that species. *N. bakeri* is the only described species in the region that lacks 'clavate' body setae, but given the deep divergences, lack of monophyly and broadly sympatric distributions of the clades exhibiting that morphology, it is clear that there is more than one species involved. Most likely the 'all dark' clade from western North Carolina and north Georgia represents something undescribed, as does the 'white head' lineage (or lineages). However, further work will be needed to test this possibility.

#### 4. Conclusions

This work represents a significant step forward in the integrative systematic study of globular Collembola in the southeastern US. At the simplest level, we have broadened the known distributions for a number of poorly documented species, and begun to reveal some meaningful biogeographic patterns in some. More significantly, these data reveal extraordinary levels of intraspecific diversity in nearly all unambiguously identifiable species, as has become typical in Collembola intraspecific work [37,67–69,78–80], indicating long residence times for these in the region, and high potential for the presence of cryptic species. While we do not see much basis for the high levels of splitting that most automated delimitation methods suggested, almost all morphologically well-defined species contain highly divergent, geographically coherent clades that would qualify at least as evolutionarily significant units (ESUs) and as candidates for separate conservation consideration. More comprehensive sampling of different genomic regions, particularly nuclear genes, and inclusion of more individuals from within and beyond this region will be necessary before conclusions about cryptic endemics can be supported.

The attempted use of DNA barcodes to identify globular Collembola failed completely; none of our sequences was a close enough match in any public database to confidently support an identification. The closest matches were often correct to family, and occasionally even to species. However, even in such cases the similarities were never greater than 90%, often with not much worse matches (~82–85%) being to members of different genera, families, or even incorrect hexapod orders (Coleoptera, Hemiptera, and others). Clearly these databases have a long way to go in representation of more obscure animal groups to be up to the task of molecular identification.

Regarding the workflow presented here, methods for high-throughput generation of sequence data for studying arthropod biodiversity have been evolving rapidly. The data analyzed here represent a mix of ‘traditional’ Sanger sequencing techniques and two next-generation approaches, Illumina and Nanopore sequencing. After using all these, we agree with Srivathsan et al. [46] in endorsing the Nanopore approach for its relative ease and cost-effectiveness. Despite its allegedly higher error rates, the high degree of replication and ability to sort through these with software such as ONT barcoder effectively neutralizes this concern, and not needing to devote significant flow cell space to control sequences (as required on Illumina’s MiSeq with low diversity libraries) ensure that yields are maximized. One additional consideration, associated with our use of 96 well plates and liquid handling robots for parts of the DNA extraction procedure, is the loss of numerous (nearly half) of the voucher specimens. This problem is probably worse for Collembola than for any other arthropod group we’ve worked with due to their very thin cuticle and their becoming completely transparent during digestion. Single tube extractions, though much slower, would probably recover a greater proportion of voucher specimens. Regardless, we would emphasize the importance of photographing specimens of minute arthropods before extracting them assuming relatively high rates of destruction and loss.

Symphyleona and Neelipleona represent a diverse and, for their sizes, quite charismatic group of litter arthropods. Their relative neglect by the broader community can only be attributed to their minuteness and limited taxonomic resources. Large scale biodiversity assessments using molecular methods stand to revolutionize our understanding of this and other dark taxa [81,82], and we hope that this contribution helps underscore the potential. Threatened areas and faunas, like the high elevations of southern Appalachia, include large numbers of such species that through ignorance risk extinction before we’re even aware of their existence. They desperately deserve the attention of conservation biologists, but careful taxonomic revision, accurate species delimitation, and useable identification resources are necessary before they can be practically considered in such planning.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/d14100847/s1>, Table S1: (Dukes et al.) All specimen data for material used in this study, including localities, voucher codes, morphospecies codes, and GenBank accession numbers; Figure S1: Time calibrated ultrametric phylogram, with estimated dates (in million years before present) for each node and 95% confidence intervals shown.

**Author Contributions:** C.D.D., E.R. and M.S.C. conducted the fieldwork, and generated the data. C.D.D., F.J. and M.S.C. carried out identifications. All authors analyzed the data and participated in drafting the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was funded by the CAFLS Undergraduate Research Initiative (to CD), The U.S. National Science Foundation (Award DEB-1916263 to MSC) and the Clemson University Experiment Station (SC-1700596 to MSC). We also acknowledge the support of the John and Suzanne Morse Endowment for Arthropod Biodiversity. This paper represents Technical Contribution No. 7085 of the Clemson University Experiment Station.

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** All data generated during this study are available on GenBank (sequence data; Accession #s OP021206-OP021410) or the Clemson University Arthropod Collection’s search page on the SCAN (Symbiota Collections of Arthropods Network) database website (occurrence data).

**Acknowledgments:** Thank you to the North Carolina State Parks, Great Smoky Mountains National Park, and Blue Ridge Parkway National Park for collecting permits, and to Monica Martin for facilitating access to Big Bald. For assistance with field work we thank Frank Etzler, Curt Harden, Patricia Wooden, Adam Haberski, Roy Kucuk, Laura Vasquez-Velez, Laary Cushman, Paul Marek, Michael Ferro, and Will Kuhn. We thank three anonymous reviewers for comments that improved the manuscript.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Barnes, R.; Clark, A.T. Sixty-Five million years of change in temperature and topography explain evolutionary history in eastern North American plethodontid salamanders. *Am. Nat.* **2017**, *190*, E1–E12. [\[CrossRef\]](#) [\[PubMed\]](#)
- Crespi, E.J.; Rissler, L.J.; Browne, R. Testing Pleistocene refugia theory: Phylogeographical analysis of *Desmognathus wrighti*, a high-elevation salamander in the southern Appalachians. *Mol. Ecol.* **2003**, *12*, 969–984. [\[CrossRef\]](#)
- Means, J.C.; Marek, P.E. Is geography an accurate predictor of evolutionary history in the millipede family Xystodesmidae? *PeerJ* **2017**, *5*, e3854. [\[CrossRef\]](#)
- Carlton, C.; Bayless, V. Documenting Beetle (Arthropoda: Insecta: Coleoptera) Diversity in Great Smoky Mountains National Park: Beyond the Halfway Point. *Southeast. Nat.* **2007**, *6*, 182–193. [\[CrossRef\]](#)
- Bernard, E.C.; Felderhoff, K.L. Biodiversity explosion: Collembola (springtails) of Great Smoky Mountains National Park. *Southeast Nat.* **2007**, *1*, 175–182. [\[CrossRef\]](#)
- Barr, T.C. Revision of Appalachian *Trechus* (Coleoptera: Carabidae). *Brimleyana* **1979**, *2*, 29–75.
- Gusarov, V.I. A revision of nearctic species of the genus *Geostiba* Thomson, 1858. *Zootaxa* **2002**, *81*, 1–88. [\[CrossRef\]](#)
- Peck, S.B. A systematic revision and the evolutionary biology of the Ptomaphagus (Adelops) beetles of North America with emphasis on cave-inhabiting species (Leiodidae). *Bull. Mus. Comp. Zool.* **1973**, *145*, 29–162.
- Peck, S.B. Systematics and evolution of forest litter *Adelopsis* in the southern Appalachians (Coleoptera: Leiodidae; Catopinae). *Psyche A J. Entomol.* **1978**, *85*, 355–382. [\[CrossRef\]](#)
- Wheeler, Q.D.; McHugh, J.V. A New Southern Appalachian Species, *Dasycerus Bicolor* (Coleoptera: Staphylinidae: Dasycerinae), from Declining Endemic Fir Forests. *Coleopt. Bull.* **1994**, *48*, 265–271.
- Smolis, A.; Bernard, E.C. *Intricatonura fjellbergi*, a new peculiar genus and species of Neanurini (Collembola: Neanuridae: Neanurinae) from Great Smoky Mountains National Park. *Florida Entomol.* **2017**, *100*, 725–730. [\[CrossRef\]](#)
- Keith, R.; Hedin, M. Extreme mitochondrial population subdivision in southern Appalachian paleoendemic spiders (Araneae: Hypochilidae: *Hypochilus*), with implications for species delimitation. *J. Arachnol.* **2012**, *40*, 167–181. [\[CrossRef\]](#)
- Hedin, M.; Carlson, D.; Coyle, F. Sky island diversification meets the multispecies coalescent—Divergence in the spruce-fir moss spider (*Microhexura montivaga*, Araneae, Mygalomorphae) on the highest peaks of southern Appalachia. *Mol. Ecol.* **2015**, *24*, 3467–3484. [\[CrossRef\]](#)
- Moskwik, M. Recent elevational range expansions in plethodontid salamanders (Amphibia: Plethodontidae) in the southern Appalachian Mountains. *J. Biogeogr.* **2014**, *41*, 1957–1966. [\[CrossRef\]](#)
- Thomas, S.M.; Hedin, M. Natural history and distribution of the enigmatic southern Appalachian opilionid, *Fumontana deprehendor* Shear (Laniatores: Triaenonychidae), with an assessment of morphological variation. *Zootaxa* **2006**, *1242*, 21–36.
- Ulrey, C.; Quintana-Ascencio, P.F.; Kauffman, G.; Smith, A.B.; Menges, E.S. Life at the top: Long-term demography, microclimatic refugia, and responses to climate change for a high-elevation southern Appalachian endemic plant. *Biol. Conserv.* **2016**, *200*, 80–92. [\[CrossRef\]](#)
- White, P.B.; Van de Gevel, S.L.; Soule, P.T. Succession and disturbance in an endangered red spruce—Fraser fir forest in the southern Appalachian Mountains, North Carolina, USA. *Endanger. Species Res.* **2012**, *18*, 17–25. [\[CrossRef\]](#)
- Jenkins, M.A. Impact of the Balsam Woolly Adelgid (*Adelges piceae* Ratz.) on an *Abies fraseri* (Pursh) Poir. Dominated Stand near the Summit of Mount LeConte, Tennessee. *Castanea* **2003**, *68*, 109–118.
- Rusek, J. Biodiversity of Collembola and their functional role in the ecosystem. *Biodivers. Conserv.* **1998**, *7*, 1207–1219. [\[CrossRef\]](#)
- Filho, L.C.I.O.; Baretta, D. Why should we care about edaphic springtails? *Sci. Agrar.* **2016**, *17*, 21–40. [\[CrossRef\]](#)
- Hopkin, S.P. *Biology of the Springtails*; Oxford University Press: Oxford, UK, 1997.
- Christiansen, K.; Bellinger, P. *The Collembola of North America, North of the Rio Grande*; Grinnell College: Grinnell, IA, USA, 1998.
- Folsom, J.W. The distribution of Holarctic Collembola. *Psyche* **1901**, *9*, 159–162. [\[CrossRef\]](#)
- Ávila-Jiménez, M.L.; Coulson, S.J. A holarctic biogeographical analysis of the collembola (Arthropoda, Hexapoda) unravels recent post-glacial colonization patterns. *Insects* **2011**, *2*, 273–296. [\[CrossRef\]](#) [\[PubMed\]](#)
- Wray, D.L. Collembola of the Great Smoky Mountains. *J. Tenn. Acad. Sci.* **1963**, *38*, 85–86.
- Snider, R.J. *Sminthurus bivittatus*, new species from the southeastern United States (Collembola: Sminthuridae). *Florida Entomol.* **1985**, *68*, 574–582. [\[CrossRef\]](#)
- Snider, R.J. *Dicyrtoma* (Ptenothrix) *renateae*, New Species from the Savannah River Plant and Georgia (Collembola: Dicyrtominae). *Florida Entomol.* **1985**, *68*, 561–566. [\[CrossRef\]](#)
- Snider, R.J. Redescription of *Sminthurus floridanus* Macgillivray, 1893 (Collembola: Sminthuridae). *Florida Entomol.* **1982**, *65*, 221–227. [\[CrossRef\]](#)
- Snider, R.J. A new species of Ptenothrix and records from the southeastern United States (Dicyrtomidae: Collembola). *Florida Entomol.* **1990**, *73*, 248–257. [\[CrossRef\]](#)
- Snider, R.J. *Denisiella lithophila*, a new species from a granite outcrop in Georgia (Collembola: Sminthuridae). *Florida Entomol.* **1988**, *71*, 124–129. [\[CrossRef\]](#)
- Snider, R.J. *Sminthurus carolinensis*, new species from South Carolina (Collembola: Sminthuridae). *Florida Entomol.* **1981**, *64*, 417–424. [\[CrossRef\]](#)
- Snider, R.J. *Sminthurus fischeri*, new species from Georgia (Collembola: Sminthuridae). *Florida Entomol.* **1982**, *65*, 321–326. [\[CrossRef\]](#)



33. Snider, R.J. *Dicyrtoma (Ptenothrix) castanea*, New Species from the Savannah River Plant (Collembola: Dicyrtominae). *Florida Entomol.* **1985**, *68*, 582–587. [[CrossRef](#)]
34. Snider, R.J. *Vesicephalus crossleyi*, New Species from the Savannah River Plant and Georgia (Collembola: Sminthurinae). *Florida Entomol.* **1985**, *68*, 567–574. [[CrossRef](#)]
35. Hebert, P.D.N.; Penton, E.H.; Burns, J.M.; Janzen, D.H.; Hallwachs, W. Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly *Astraptes fulgerator*. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 14812–14817. [[CrossRef](#)] [[PubMed](#)]
36. DeSalle, R.; Goldstein, P. Review and interpretation of trends in DNA barcoding. *Front. Ecol. Evol.* **2019**, *7*, 302. [[CrossRef](#)]
37. Porco, D.; Bedos, A.; Greenslade, P.; Janion, C.; Skarżyński, D.; Stevens, M.I.; Jansen Van Vuuren, B.; Deharveng, L. Challenging species delimitation in Collembola: Cryptic diversity among common springtails unveiled by DNA barcoding. *Invertebr. Syst.* **2012**, *26*, 470–477. [[CrossRef](#)]
38. Deharveng, L. Recent advances in Collembola systematics, 6th International Seminar on Apterygota, Siena, Italy, 2002. *Pedobiologia* **2004**, *48*, 415–433. [[CrossRef](#)]
39. Hutama, A.; Dahrudin, H.; Busson, F.; Sauri, S.; Keith, P.; Hadiaty, R.K.; Hanner, R.; Suryobroto, B.; Hubert, N. Identifying spatially concordant evolutionary significant units across multiple species through DNA barcodes: Application to the conservation genetics of the freshwater fishes of Java and Bali. *Glob. Ecol. Conserv.* **2017**, *12*, 170–187. [[CrossRef](#)]
40. Dincă, V.; Runquist, M.; Nilsson, M.; Vila, R. Dispersal, fragmentation, and isolation shape the phylogeography of the European lineages of *Polyommatus (Agrodiaetus) ripartii* (Lepidoptera: Lycaenidae). *Biol. J. Linn. Soc.* **2013**, *109*, 817–829. [[CrossRef](#)]
41. Bellinger, P.F.; Christiansen, K.; Janssens, F. Checklist of the Collembola of the World. Available online: [www.collembola.org](http://www.collembola.org) (accessed on 16 May 2022).
42. Folmer, O.; Black, M.; Hoeh, W.; Lutz, R.; Vrijenhoek, R. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotechnol.* **1994**, *3*, 294–299.
43. Elbrecht, V.; Leese, F. Validation and Development of COI Metabarcoding Primers for Freshwater Macroinvertebrate Bioassessment. *Front. Environ. Sci.* **2017**, *5*, 11. [[CrossRef](#)]
44. Meier, R.; Wong, W.; Srivathsan, A.; Foo, M. \$1 DNA barcodes for reconstructing complex phenomes and finding rare species in specimen-rich samples. *Cladistics* **2016**, *32*, 100–110. [[CrossRef](#)] [[PubMed](#)]
45. Bushnell, B.; Rood, J.; Singer, E. BBMerge—Accurate paired shotgun read merging via overlap. *PLoS ONE* **2017**, *12*, e0185056. [[CrossRef](#)]
46. Srivathsan, A.; Lee, L.; Katoh, K.; Hartop, E.; Kutty, S.N.; Wong, J.; Yeo, D.; Meier, R. ONTbarcoder and MinION barcodes aid biodiversity discovery and identification by everyone, for everyone. *BMC Biol.* **2021**, *19*, 217. [[CrossRef](#)]
47. Katoh, K.; Rozewicki, J.; Yamada, K.D. MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. *Brief. Bioinform.* **2017**, *20*, 1160–1166. [[CrossRef](#)]
48. Nguyen, L.T.; Schmidt, H.A.; Von Haeseler, A.; Minh, B.Q. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol. Biol. Evol.* **2015**, *32*, 267–274. [[CrossRef](#)] [[PubMed](#)]
49. Trifinopoulos, J.; Nguyen, L.T.; von Haeseler, A.; Minh, B.Q. W-IQ-TREE: A fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Res.* **2016**, *44*, W232–W235. [[CrossRef](#)] [[PubMed](#)]
50. Minh, B.Q.; Nguyen, M.A.T.; Von Haeseler, A. Ultrafast approximation for phylogenetic bootstrap. *Mol. Biol. Evol.* **2013**, *30*, 1188–1195. [[CrossRef](#)]
51. Suchard, M.A.; Lemey, P.; Baele, G.; Ayres, D.L.; Drummond, A.J.; Rambaut, A. Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evol.* **2018**, *4*, vey016. [[CrossRef](#)] [[PubMed](#)]
52. Stadler, T. On incomplete sampling under birth-death models and connections to the sampling-based coalescent. *J. Theor. Biol.* **2009**, *261*, 58–66. [[CrossRef](#)]
53. Papadopoulou, A.; Anastasiou, I.; Vogler, A.P. Revisiting the insect mitochondrial molecular clock: The mid-Aegean trench calibration. *Mol. Biol. Evol.* **2010**, *27*, 1659–1672. [[CrossRef](#)] [[PubMed](#)]
54. Katz, A.D.; Taylor, S.J.; Davis, M.A. At the confluence of vicariance and dispersal: Phylogeography of cavernicolous springtails (Collembola: Arrhopalitidae, Tomoceridae) codistributed across a geologically complex karst landscape in Illinois and Missouri. *Ecol. Evol.* **2018**, *8*, 10306–10325. [[CrossRef](#)]
55. Lukić, M.; Delić, T.; Pavlek, M.; Deharveng, L.; Zagmajster, M. Distribution pattern and radiation of the European subterranean genus *Verhoeffiella* (Collembola, Entomobryidae). *Zool. Scr.* **2020**, *49*, 86–100. [[CrossRef](#)]
56. Ding, Y.H.; Yu, D.Y.; Guo, W.B.; Li, J.N.; Zhang, F. Molecular phylogeny of *Entomobrya* (Collembola: Entomobryidae) from China: Color pattern groups and multiple origins. *Insect Sci.* **2019**, *26*, 587–597. [[CrossRef](#)] [[PubMed](#)]
57. Collins, G.E.; Hogg, I.D.; Convey, P.; Sancho, L.G.; Cowan, D.A.; Lyons, W.B.; Adams, B.J.; Wall, D.H.; Green, T.G. Genetic diversity of soil invertebrates corroborates timing estimates for past collapses of the West Antarctic Ice Sheet. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 22293–22302. [[CrossRef](#)] [[PubMed](#)]
58. Rambaut, A.; Drummond, A.J.; Xie, D.; Baele, G.; Suchard, M.A. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Syst. Biol.* **2018**, *67*, 901–904. [[CrossRef](#)]
59. Puillandre, N.; Lambert, A.; Brouillet, S.; Achaz, G. ABGD, Automatic Barcode Gap Discovery for primary species delimitation. *Mol. Ecol.* **2012**, *21*, 1864–1877. [[CrossRef](#)]
60. Puillandre, N.; Brouillet, S.; Achaz, G. ASAP: Assemble species by automatic partitioning. *Mol. Ecol. Resour.* **2021**, *21*, 609–620. [[CrossRef](#)]



61. Zhang, J.; Kapli, P.; Pavlidis, P.; Stamatakis, A. A general species delimitation method with applications to phylogenetic placements. *Bioinformatics* **2013**, *29*, 2869–2876. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Kapli, P.; Lutteropp, S.; Zhang, J.; Kobert, K.; Pavlidis, P.; Stamatakis, A.; Flouri, T. Multi-rate Poisson tree processes for single-locus species delimitation under maximum likelihood and Markov chain Monte Carlo. *Bioinformatics* **2017**, *33*, 1630–1638. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Pons, J.; Barraclough, T.G.; Gomez-Zurita, J.; Cardoso, A.; Duran, D.P.; Hazell, S.; Kamoun, S.; Sumlin, W.D.; Vogler, A.P. Sequence-based species delimitation for the DNA taxonomy of undescribed insects. *Syst. Biol.* **2006**, *55*, 595–609. [\[CrossRef\]](#)
64. Kimura, M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.* **1980**, *16*, 111–120. [\[CrossRef\]](#) [\[PubMed\]](#)
65. Fujisawa, T.; Barraclough, T.G. Delimiting species using single-locus data and the generalized mixed yule coalescent approach: A revised method and evaluation on simulated data sets. *Syst. Biol.* **2013**, *62*, 707–724. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Porco, D.; Skarżyński, D.; Decaëns, T.; Hebert, P.D.N.; Deharveng, L. Barcoding the Collembola of Churchill: A molecular taxonomic reassessment of species diversity in a sub-Arctic area. *Mol. Ecol. Resour.* **2014**, *14*, 249–261. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Katz, A.D.; Giordano, R.; Soto-Adames, F.N. Operational criteria for cryptic species delimitation when evidence is limited, as exemplified by North American *Entomobrya* (Collembola: Entomobryidae). *Zool. J. Linn. Soc.* **2015**, *173*, 818–840. [\[CrossRef\]](#)
68. Cicconardi, F.; Nardi, F.; Emerson, B.C.; Frati, F.; Fanciulli, P.P. Deep phylogeographic divisions and long-term persistence of forest invertebrates (Hexapoda: Collembola) in the North-Western Mediterranean basin. *Mol. Ecol.* **2010**, *19*, 386–400. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Cicconardi, F.; Fanciulli, P.P.; Emerson, B.C. Collembola, the biological species concept and the underestimation of global species richness. *Mol. Ecol.* **2013**, *22*, 5382–5396. [\[CrossRef\]](#)
70. Meier, R.; Zhang, G.; Ali, F. The Use of Mean Instead of Smallest Interspecific Distances Exaggerates the Size of the “Barcoding Gap” and Leads to Misidentification. *Syst. Biol.* **2008**, *57*, 809–813. [\[CrossRef\]](#) [\[PubMed\]](#)
71. Zhang, H.; Bu, W. Exploring Large-Scale Patterns of Genetic Variation in the COI Gene among Insecta: Implications for DNA Barcoding and Threshold-Based Species Delimitation Studies. *Insects* **2022**, *13*, 425. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Hebert, P.D.N.; Cywinska, A.; Ball, S.L.; deWaard, J.R. Biological identifications through DNA barcodes. *Proc. Biol. Sci.* **2003**, *270*, 313–321. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Ratnasingham, S.; Hebert, P.D.N. A DNA-Based Registry for All Animal Species: The Barcode Index Number (BIN) System. *PLoS ONE* **2013**, *8*, e66213. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Zeppelini, D. Phylogeny of *Arrhopalites* s.l. (Collembola: Symphypleona: Arrhopalitidae): Testing the monophyly of the recently erected genera *Arrhopalites* s.s. and *Pygmarrhopalites*. *Invertebr. Syst.* **2011**, *25*, 91–105. [\[CrossRef\]](#)
75. Baquero, E.; Snider, R.; Jordana, R. A revision of the spring-tail genus *Vesicephalus* Richards (Collembola, Symphypleona, Sminthurinae). *Syst. Entomol.* **2006**, *31*, 633–647. [\[CrossRef\]](#)
76. Banks, N. Two new Sminthurids. *J. N. Y. Entomol. Soc.* **1897**, *5*, 33–34.
77. Borrer, D.J.; Triplehorn, C.A.; Johnson, N.F. *An Introduction to the Study of Insects*; Saunders College Publishing: Philadelphia, PA, USA, 1989.
78. Zhang, B.; Chen, T.W.; Mateos, E.; Scheu, S.; Schaefer, I. Cryptic species in *Lepidocyrtus lanuginosus* (Collembola: Entomobryidae) are sorted by habitat type. *Pedobiologia* **2018**, *68*, 12–19. [\[CrossRef\]](#)
79. Porco, D.; Potapov, M.; Bedos, A.; Busmachiu, G.; Weiner, W.M.; Hamra-Kroua, S.; Deharveng, L. Cryptic diversity in the ubiquitous species *Parisotoma notabilis* (Collembola, Isotomidae): A long-used chimeric species? *PLoS ONE* **2012**, *7*, e46056. [\[CrossRef\]](#)
80. Mateos, E.; Winkler, D.; Riutort, M.; Álvarez-Presas, M. New morphological and molecular data reveal an important underestimation of species diversity and indicate evolutionary patterns in European *Lepidocyrtus* (Collembola: Entomobryidae). *Invertebr. Syst.* **2021**, *35*, 471–492. [\[CrossRef\]](#)
81. Hartop, E.; Srivathsan, A.; Ronquist, F.; Meier, R. Towards large-scale integrative taxonomy (LIT): Resolving the data conundrum for dark taxa. *Syst. Biol.* **2022**, syac033. [\[CrossRef\]](#)
82. Page, R.D.M. DNA barcoding and taxonomy: Dark taxa and dark texts. *Philos. Trans. R. Soc. B Biol. Sci.* **2016**, *371*, 20150334. [\[CrossRef\]](#)