

MDPI

Article

# Diversity of Helminths of Insectivorous Mammals (Mammalia: Eulipothyphla) from Large Forest Protected Areas of the Middle Volga Region (European Russia)

Nadezhda Yu. Kirillova <sup>1</sup>, Alexander A. Kirillov <sup>1</sup>, Alexander B. Ruchin <sup>2</sup> and Alexander I. Fayzulin <sup>1,\*</sup>

- Laboratory for Zoology and Parasitology, Institute of Ecology of Volga River Basin of RAS, Samara Federal Research Center of RAS, Togliatti 445003, Russia; nadinkirillova2011@yandex.ru (N.Y.K.); parasitolog@yandex.ru (A.A.K.)
- <sup>2</sup> Joint Directorate of the Mordovia State Nature Reserve and National Park "Smolny", Saransk 430005, Russia; ruchin.alexander@gmail.com
- \* Correspondence: alexandr-faizulin@yandex.ru

Abstract: Insectivores (Eulypotiphla) are a substantial component of Russian forest ecosystems. The parasites of these animals also form an important part of natural biocenoses and act as one of the factors in the formation of biodiversity. The Mordovia Nature Reserve and National Park "Smolny" are large, forested areas located in the center of European Russia. We studied the helminth fauna of insectivores in these protected areas in 2018-2022. In total, using the method of complete helminthological necropsy, we examined 478 individuals of shrews, moles, and hedgehogs and recorded 34 species of parasitic worms, i.e., 8 trematode, 7 cestode, 1 acanthocephalan, and 18 nematode species. The most diverse helminth fauna was found in Sorex araneus (22 species). The composition of helminths in S. isodon (12), Neomys fodiens (9), Sorex minutus, and Erinaceus roumanicus (8 species each) turned out to be less diverse. The lowest species diversity of helminths was observed in Neomys milleri (3) and Talpae europaea (2 species). Taking into account the newly obtained data, we conducted a review of the helminth diversity in shrews, hedgehogs, and moles in the Middle Volga region. According to our literature data, the helminth fauna of insectivores in this region consists of 52 species, including 14 cestodes, 13 trematodes, 22 nematodes, and 3 acanthocephalans. Most of them belong to the Palearctic faunal complex (36 species). The helminth fauna of insectivores in the studied protected areas was compared with the helminth fauna of micromammals in other areas of the Middle Volga region. Our comparative analysis showed a high and average degree of similarity in the helminth fauna within individual species and genera of Eulipotyphla.

**Keywords:** protected areas; parasitic worms; shrews; moles; hedgehogs; biodiversity; overview; Volga region



Citation: Kirillova, N.Y.; Kirillov, A.A.; Ruchin, A.B.; Fayzulin, A.I. Diversity of Helminths of Insectivorous

Mammals (Mammalia: Eulipothyphla) from Large Forest Protected Areas of the Middle Volga Region (European Russia). *Diversity* 2024, 16, 307. https://doi.org/10.3390/d16050307

Academic Editor: Chang-Bae Kim

Received: 16 April 2024 Revised: 9 May 2024 Accepted: 17 May 2024 Published: 20 May 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

# 1. Introduction

The species diversity of small mammals in biocenoses is usually low. In terms of the number of species, they constitute hundredths of the total biodiversity of ecosystems, but their importance in natural communities is extremely diverse. This is determined by their high level of vital processes as well as their great activity [1–5]. Mammals are the main components of food chains and participate in the cycling of a significant amount of substances in terrestrial ecosystems [1,6–9].

Small mammals, such as Eulipotyphla and Rodentia, play a key role in regulatory processes that ensure the sustainability of natural ecosystems [6,7,10–16]. Widespread insectivores are an important component of Russian forest ecosystems. The significant role of insectivores in biocenoses is associated mainly with the regulation of the abundance of invertebrates. Thus, shrews consume up to 60–70% of the biomass of soil invertebrates [11,12,17–19]. Due to their high mobility, insectivores make a significant

Diversity 2024, 16, 307 2 of 24

contribution to the cycle of matter and energy in forest ecosystems [20]. Shrews, in particular, can carry micromycete spores on their fur, thereby promoting the decomposition of litter [21,22]. In addition, insectivorous mammals are the definitive and/or paratenic hosts of parasitic worms and participate in the circulation of the helminths of other vertebrates [23–25].

Helminths are one of the factors in the formation of global biodiversity [26–28]. In this regard, data on helminth communities are necessary to assess changes in the structure of the biocenosis, the functioning of food chains, and biodiversity [29–31].

The helminth fauna of insectivores has been studied in different parts of Europe. Thus, the most detailed studies were carried out in Germany, where 69 species of helminths were identified in 7 species of insectivores [32–37]. In addition, in Poland and Belarus, 64 helminth species were each found in 7 host species [36,38–51]. In Bulgaria, 62 species of helminths have been recorded in 8 species of insectivorous mammals [52,53], and in the territory of the former Czechoslovakia, 67 species of helminths have been recorded in 8 host species [36,54–59]. Regular helminthological studies of Eulipotyphla were also carried out in Spain [60–67], France [34,68–71], Switzerland [34,72–75], Hungary [76,77], Lithuania [78–80], Finland [81–84], Ukraine [85–88], and Moldova [89–93]. Data on the helminth communities in Eulipotyphla hosts were also obtained from the UK [94–98], Ireland [99], Belgium [100], Denmark [101], Portugal [102,103], Italy [104–106], Austria [107,108], Greece [109], Serbia [110], and Romania [111]. As a rule, studies were devoted either to individual taxonomic groups of helminths or to individual species or genera of hosts. Thus, the helminth fauna of hedgehogs [25,36,101] and moles [112] in Europe has been studied quite fully. Among shrews, the most common species, *Sorex araneus*, is the most studied [113]. In recent decades, reviews of the helminth fauna of individual species or genera of insectivores, as well as molecular genetic studies of cestodes from shrews, have appeared [25,36,101,112–119].

Despite the wide distribution of insectivorous mammals and the long history of studying their helminth fauna, in European Russia, there are still areas where the helminths of insectivores are almost unstudied. Systematic and long-term studies were carried out only on the territory of the Komi Republic [120,121], Karelia [122–127], and the Samara region [128–132]. Several studies of the helminths of various insectivorous mammals have been conducted on the territory of Tatarstan [133,134], Chuvashia [133], Mari El [133], Astrakhan [135,136], Vologda [137], Voronezh [138,139], Kursk [140,141], Leningrad [142], Nizhny Novgorod [143–145], and Tver [146] regions.

The article by Shaldybin [147] is the first and only study of the helminths of vertebrates in the Mordovia Nature Reserve. The author found 23 species of parasitic worms in four species of insectivores. This information was subsequently incorporated into review studies of the helminths of vertebrates in this protected area [148,149]. Such studies have not been carried out on the territory of the National Park "Smolny".

Here we present a study of the species diversity of parasitic worms in insectivorous mammals from the protected areas of Mordovia, with an overview of the helminth fauna of shrews, moles, and hedgehogs inhabiting the Middle Volga region (European Russia).

# 2. Materials and Methods

### 2.1. Study Area Description

The helminth fauna of insectivorous mammals was studied on the territory of the Federal State Budgetary Institution "Reserved Mordovia" ("Zapovednaya Mordovia"), which includes the Mordovia Nature Reserve and the National Park "Smolny". Both protected areas are located on the territory of the Republic of Mordovia and represent large forest areas.

The territory of the Mordovia Nature Reserve is located in the northwestern part of the Republic of Mordovia and occupies an area approximately within the range of  $54^{\circ}42' - 54^{\circ}56'$  north latitude and  $43^{\circ}04' - 43^{\circ}36'$  east longitude. The territory of the Mordovia Nature Reserve represents natural ecosystems of southern woodlands (southern taiga), located on

Diversity 2024, 16, 307 3 of 24

the border of mixed and broad-leaved forests and forest steppe and occupying the right bank of the Moksha River [150]. From the north and east, the territory of the reserve is limited by the Satis River (a tributary of the Moksha River) and the Arga River, which flows into the Satis. From the south, the forest steppe directly approaches the border of the reserve. The territory of the reserve is rich in small rivers, streams, and lakes, such as Pushta, Arga, Bolshaya Chernaya, Malaya Chernaya, Picherki, and Inorskoye. During dry years, rivers and streams in the reserve may dry up. The hydrological regime of small rivers often largely depends on beaver dams, which contribute to the flooding of large areas [151,152]. The fauna of the Mordovia Nature Reserve consists of 299 species, including 10 species of insectivores, i.e., northern white-breasted hedgehog Erinaceus roumanicus Barrett-Hamilton, 1900; European hedgehog Erinaceus europaeus Linnaeus, 1758; lesser white-toothed shrew Crocidura suaveolens (Pallas, 1811); common shrew Sorex araneus Linnaeus, 1758; Eurasian pygmy shrew Sorex minutus Linnaeus, 1766; masked shrew Sorex caecutiens Laxmann, 1788; taiga shrew Sorex isodon Turov, 1924; Eurasian water shrew Neomys fodiens Pennant, 1771; Mediterranean water shrew Neomys milleri Mottaz, 1907; and European mole Talpa europaea Linnaeus, 1758 [150,153].

The National Park "Smolny" is located in the northeastern part of the Republic of Mordovia and occupies an area approximately within the range of 54°43′–54°53′ north latitude and 45°04′–45°37′ east longitude. The National Park is located on the border of mixed and broad-leaved forests and forest steppe [154]. The protected area is characterized by a fairly dissected relief with an extensive network of ravines in the northeastern part. The rivers of the park belong to the Sura River basin and include the small rivers Kalysha and Yazovka, as well as many streams. The lakes are concentrated mainly in the floodplain of the Alatyr River in the south of the National Park "Smolny" [152]. The diversity of natural conditions determines the richness of the flora and fauna of the National Park "Smolny". The fauna of this protected area consists of 282 species of animals, including 9 species of Eulipothyphla, i.e., *E. roumanicus*, *E. europaeus*, *C. suaveolens*, *S. araneus*, *S. minutus*, *S. caecutiens*, *S. isodon*, *N. fodiens*, and *T. europaea* [154–157].

## 2.2. Parasite Collection and Examination

Parasitic worms in insectivores were studied during the 2018–2022 field seasons. The material for parasitological research was obtained as a result of work on counting the number of small mammals. They are counted regularly according to research topics of the Mordovia Nature Reserve and the National Park "Smolny", approved by the Ministry of Natural Resources and Ecology of Russia. In total, 478 mammals of eight species were examined on the territory of the Mordovia Nature Reserve and the National Park "Smolny" (Table 1). Sampling sites for shrews, moles, and hedgehogs are presented in Table S1.

Species	Mordovia Nature Reserve	National Park "Smolny"
Sorex araneus	118	247
Sorex minutus	64	12
Sorex isodon	12	_
Neomys fodiens	3	3
Neomys milleri	1	_
Crocidura suaveolens	3	_
Talpa europaea	1	6
Erinaceus roumanicus	3	5
Total	205	273

**Table 1.** List of insectivorous mammals examined in protected areas of Mordovia.

Data on the helminth fauna of shrews were additionally obtained when studying the abundance of myomorph rodents in protected areas of Mordovia. Shrews were captured using spring metal snap traps ( $120 \times 55$  mm). Trap lines with 20 snap traps, at 10 m intervals, were installed along the banks of small rivers and streams, forest edges, fields,

Diversity **2024**, 16, 307 4 of 24

and meadows. Pieces of rye bread fried in sunflower seed oil were placed in the snap traps. Capturing was carried out for 5 days in each locality. Some of the shrew specimens were obtained during the study of insects using pitfall traps and were kindly provided by entomologists of the Mordovia Nature Reserve. We also examined roadkill specimens of *E. roumanicus* and moles that died naturally and were provided by local residents.

The animals were examined using a complete helminthological necropsy, according to Ivashkin et al. [158] and Anikanova et al. [125]. Parasitic worms were recovered and fixed in 70% ethanol. Flukes and flatworms were stained with aceto-carmine, cleared with clove oil, and then mounted in Canada balsam. Nematodes were cleared with lactic acid and mounted in glycerin jelly [125,158,159]. Identification of parasites was carried out at the Laboratory for Zoology and Parasitology of the Institute of Ecology of the Volga Basin of the Russian Academy of Sciences (Togliatti).

# 2.3. Data Analysis

To characterize helminth infection in insectivorous mammals, the following indices were used: infection prevalence (P, %) and mean abundance (MA). The dominance of species in the helminth fauna was determined using the Palia–Kovnatsky index (D) [160]. Parasite dominance groups were considered as follows: 10–100—dominants, 1–10—subdominants, 0.001–1.000—adominants.

The similarity of helminth compositions was evaluated using Morisita's overlap index ( $C_m$ ). The degree of similarity is as follows: 0–0.33—low; 0.34–0.66—medium; 0.67—1—high. Statistical data processing was performed using the PAST 2.16 [161] and Microsoft Excel 2003 software packages.

#### 3. Results

3.1. Parasitic Worms of Insectivorous Mammals in the Protected Areas of Mordovia (European Russia)

In total, 34 species of helminths were found in shrews, hedgehogs, and moles from two protected areas of Mordovia, including 8 digenean, 7 cestode, 1 acanthocephalan, and 18 nematode species. On the territory of the Mordovia Nature Reserve (MNR), 28 species of helminths were recorded in insectivorous mammals, and 31 species of parasites were found on the territory of the National Park "Smolny" (NP "Smolny") (Table 2).

**Table 2.** Parasite–host checklist of helminths from insectivorous mammals in the protected areas of Mordovia.

Species	Host	Site	MNR P, %	MNR P. % MA		olny" MA
Family Dilepididae Dilepis undula (Schrank, 1788), larvae	Sorex araneus	small intestine	1.7	0.1	<b>P,</b> %	0.4
Family Hymenolepididae Hymenolepis erinacei (Gmelin, 1789) (=Rodentolepis erinacei (Gmelin, 1789), Rodentolepis steudeneri (Janicki, 1904))	Erinaceus roumanicus	small intestine	66.7 (in 2 of 3 examined)	36.3	80.0 (in 4 of 5 examined)	8.0
Ditestolepis diaphana (Cholodkowsky, 1906)	Sorex araneus Sorex isodon Sorex minutus	small intestine	28.8 41.7 50.8	2.1 1.7 1.3	21.1 41.7	1.1 1.6
Monocercus arionis Siebold, 1850 (=Molluscotaenia crassiscolex (von Linstow, 1890))	Sorex araneus Sorex isodon Sorex minutus	small intestine	78.0 58.3 34.9	3.4 2.5 0.8	74.5 - 75.0	5.8 3.5
Neoskrjabinolepis schaldybini Spasskii, 1947	Sorex araneus Sorex isodon	small intestine	28.8 8.3	1.0 0.3	25.1	1.8
Staphylocystis brusatae (Vaucher, 1971)	Crocidura suaveolens	small intestine	66.7 (in 2 of 3 examined)	4.0	-	
Vigisolepis spinulosa (Cholodkowsky, 1906)	Sorex araneus	small intestine	-		5.7	0.2
Family Brachylaimidae Brachylaima fulvum Dujardin, 1843 (=Brachylaemus oesophagei Shaldybin, 1953)	Sorex araneus Sorex isodon	small intestine	4.2 16.7	0.9 0.3	3.2	0.1

Diversity **2024**, 16, 307 5 of 24

 Table 2. Cont.

Species	Host	Site	MNR		NP "Smol	
Species		Site	P, %	MA	P, %	MA
Family Pananistidas	Sorex araneus Sorex isodon		6.8 8.3	0.2 0.1	5.0	0.2
Family Panopistidae Pseudoleucochloridium soricis (Soltys, 1952) (=Leucochloridium skrjabini	Sorex minutus	stomach, small	1.6	0.02		
(Shaldybin, 1953))	Neomys fodiens	intestine	100 (in 3	3.7	66.7 (in 2 of 3	1.7
	iveomys joutens		examined)	3.7	examined)	1.7
	Erinaceus				40.0 (in 2	
Family Strigeidae	roumanicus	fat tissue between	-		of 5	1.2
Strigea strigis (Schrank, 1788), mtc.	Talpa europaea	esophagus and trachea	_		examined) 1.6	0.03
	, ,				40.0 (in 2	
Family Echinostomatidae Isthmiophora melis (Schrank, 1788) (=Euparyphium melis (Schrank, 1788))	Erinaceus roumanicus	small intestine	_		of 5	1.4
isumiophora mens (Schlank, 1700) (-Euparyphiam mens (Schlank, 1700))	Tournations				examined)	
Family Omphalometridae	N		33.3 (in 1	1.0	66.7 (in 2	0.7
Neoglyphe locellus (Kossack, 1910) (=Opistioglyphe oschmarini Shaldybin, 1953)	Neomys fodiens	small intestine	of 3 examined)	1.0	of 3 examined)	0.7
Neoglyphe sobolevi Schaldybin, 1953	Sorex araneus	small intestine	5.9	0.8	1.6	0.5
14cographic soboleof Schalacybiri, 1755		Sittan intestine				
	Sorex araneus Sorex isodon		21.2 16.7	0.9 0.9	7.0	0.2
	Sorex minutus		34.9	1.4	58.3	1.2
Rubenstrema exasperatum (Rudolphi, 1819)	Neomys fodiens	stomach	66.7 (in 2 of 3	1.0	_	
	1 veomys jouiens		examined)	1.0		
	Neomys milleri		in one examined	1.0	_	
	C		cxanimica		1.0	0.02
	Sorex araneus		33.3 (in 1		1.2	0.03
Rubenstrema opisthovitellinus (Soltys, 1954).	Neomys fodiens	small intestine	of 3	1.0	_	
			examined)			
Family Centrorhynchidae	Sorex araneus Sorex isodon	liver	0.8 8.3	0.02	1.2%	0.02
Centrorhynchus aluconis (Müller, 1780), larvae						
Family Capillariidae  Aonchotheca erinacei (Rudolphi, 1819) (=Capillaria erinacei (Rudolphi, 1819))	Erinaceus roumanicus	small intestine	100 (in 3 examined)	7.7	100 (in 5 examined)	24.8
Tionenometa erimaet (Rudolpin, 1017) (-eapitaria erimaet (Rudolpin, 1017))					схиниси)	
Aonchotheca exigua (Dujardin, 1845)	Sorex minutus	atomach	6.4	0.2	100 (in 3	
Tionenometa exigua (Dujarant, 1043)	Neomys fodiens	stomach	_		examined)	1.7
	Sorex araneus		2.5	0.1	9.3	0.4
Aonchotheca kutorii (Ruchljadeva, 1964)	Neomys fodiens	stomach	_		100 (in 3 examined)	2.3
Caladium canisicala (Valcacaya at Nichigani 1924) (-Hanaticala canisicala	Coway avanous		E 1	2 2	2.8	0.1
Calodium soricicola (Yokogava et Nishigori, 1924) (=Hepaticola soricicola (Yokogava et Nishigori, 1924))	Sorex araneus Sorex isodon	Liver parenchyma	5.1 8.3	2.3 0.8	2.0	0.1
	Sorex araneus	mucous membrane	3.4	0.2	17.4	0.5
Eucoleus oesophagicola (Soltys, 1952) (=Thominx blarinae (Ogran, 1953))	Sorex isodon	of esophagus	8.3	1.2	_	
	Sorex araneus		12.7	0.8	15.0	0.9
	Sorex isodon		50.0	3.9	_	
Liniscus incrassatus Diesing, 1851 (=Capillaria incrassata (Diesing, 1851))	Neomys fodiens	bladder	33.3 (in 1 of 3	0.7	_	
	,	bidder	examined)			
	Neomys milleri		in one examined	1.0	-	
Family Soboliphymidae						
Soboliphyme soricis Baylis et King, 1932	Sorex araneus	stomach	0.8	0.01	5.3	0.1
T 1 M					40.0 (in 2 of	
Family Metastrongylidae Crenosoma striatum Zeder, 1800	Erinaceus roumanicus	bronchi	_		`5	6.0
Crenosonia stratam Zeaci, 1000	704774077				examined)	
Paracrenosoma skrjabini (Pologentsev, 1935)	Sorex minutus	bronchi	1.6	0.03		
Family Heligmosomidae	Sorex araneus		70.3	4.0	71.7	10.8
Longistriata paradoxi Schaldybin, 1964	Sorex isodon Sorex minutus	small intestine	75.0 55.6	7.7 1.9	75.0	3.2
	Seres minutes			1./		U.2
	Neomys fodiens		100 (in 3 examined)	10.3	-	
Longistriata neomi Lubarskaja, 1962	Neomys milleri	small intestine	in one	3.0	_	
			examined			
Tricholinstowia linstowi Travassos, 1918 (=Longistriata vigisi Petrov et Savinov,	Talpa europaea	small intestine	in one	20.0	50.0 (in 3 of 6	15.8
1959)	тигри ситориси	oman micsine	examined	20.0	examined)	15.0
	Sorex araneus		11.0	0.3	11.7	0.5
		mesentery, serous	8.3	0,2	_	0
Family Anisakidae	Sorex isodon					
Family Anisakidae Porrocaecum depressum (Zeder, 1800), juveniles		integument of gastrointestinal	in one	11.0	83.3 (in 5 of 6	15.7
	Talpa europaea		in one examined	11.0	of 6 examined)	15.7
		gastrointestinal		0.02	of 6	0.1

Diversity **2024**, 16, 307 6 of 24

TO 1 1		$\sim$	
Tabl	e 2.	Con	t.

Species	Host	Site	MNR P, %	MA	NP "Smo P, %	lny" MA
Family Physalopteridae	Erinaceus roumanicus	stomach (adults)	66.7 (in 2 of 3 examined)	7.0	100 (in 5 examined)	9.4
Physaloptera clausa Rudolphi, 1819	Sorex araneus	gastric mucosa (juveniles)	5.1	0.1	1.2	0.01
	Sorex araneus Sorex minutus		3.2	0.1	2.0	0.1
Pseudophysaloptera soricina Baylis, 1934 (=Physaloptera soricina (Baylis, 1934))	Neomys fodiens	stomach	-		33.3 (in 1 of 3 examined)	0.3
Family Spirocercidae Physocephalus sexalatus (Molin, 1860), juveniles	Erinaceus roumanicus	walls of stomach and small intestine	-		80.0 (in 4 of 5 examined)	61.8
Species insertae sedis Agamospirura minuta (Sharpilo, 1963), juveniles	Erinaceus roumanicus	walls of stomach and small intestine	-		40.0 (in 2 of 5 examined)	1.0

Most of the parasites found in insectivores were represented by adult forms (26 species). The larval stages of seven helminth species were also identified. Nematodes of the species *Physaloptera clausa* were represented by both adult and larval stages.

The studied insectivorous mammals are obligatory hosts of the majority of identified helminths (28 species). The same mammalian species serve as paratenic hosts for four helminth species (*S. strigis, Ph. sexalatus, A. minuta*, and *C. aluconis*) and abortive hosts for two species (*H. truncata* and *D. undula*). For the last two species, insectivores represent a so-called parasitic impasse.

The helminth fauna of *S. araneus* in the "Reserved Mordovia" includes 22 species of parasites. Of these, 11 helminth species (*D. diaphana*, *M. arionis*, *N. schaldybini*, *V. spinulosa*, *P. soricis*, *Neoglyphe sobolevi*, *R. exasperatum*, *R. opisthovitellinus*, *A. kutorii*, *S. soricis*, and *L. paradoxi*) are host-specific parasites of shrews of the family Soricidae. The cestode *D. diaphana* is a host-specific parasite of *Sorex* spp., and the five helminths (*B. fulvum*, *C. soricicola*, *L. incrassatus*, *E. oesophagicola*, and *P. soricina*) are generalist species that parasitize a variety of soricomorphs. The larvae of *P. depressum* are a common parasite of shrews, moles, and hedgehogs. The larvae of *D. undula*, *S. strigis*, *Ph. clausa*, *H. truncata*, and *C. aluconis* are occasional parasites of the common shrew. The total infection rate of *S. araneus* with helminths was 100%, MA = 21.3.

The helminth fauna of *S. araneus* is dominated by nematodes, represented by both adult (7 species) and larval forms (3 species). The infection rate of shrews with nematodes reaches 76.7%, 11.5. The trematode fauna of *S. araneus* includes six species; the total infection with digeneans was 17.5%, 1.3. Among them, five species of trematodes are represented by adult worms, and only one species (*S. strigis*) occurs at the larval stage. A total of five species of cestodes were identified in *S. araneus*, of which four are represented by the mature stage and one species (*D. undula*) is represented exclusively by juvenile individuals. The total infection with cestodes was 97.0%, 8.5. Of the acanthocephalans, larvae of only one species were recorded—*C. aluconis* (0.8%, 0.01).

According to the Palia–Kovnatsky dominance index, the helminth fauna of *S. araneus* was dominated by *L. paradoxi* (D = 28.7) and *M. arionis* (17.9). The subdominants were the cestodes *D. diaphana* (1.6) and *N. schaldybini* (1.9). The remaining 19 species of parasitic worms were classified as adominants. In two protected areas of Mordovia, eighteen shared species of helminths were found in shrews, and four species of parasites (*V. spinulosa*, *S. strigis*, *R. opistovitellinus*, and *P. soricina*) were found only in *S. araneus* from the National Park "Smolny".

The helminth fauna of *S. minutus* includes eight species of parasites, which are represented by adult individuals. All of them are host-specific parasites of insectivores from the family Soricidae. The total infection rate of *S. minutus* was 100%, 6.3. Like in *S. araneus*, the species composition of helminths in *S. minutus* is dominated by nematodes (4 species). The total infection of the pygmy shrew with nematodes was 64.0%, 2.4. Cestodes and trema-

Diversity **2024**, 16, 307 7 of 24

todes in the helminth fauna of *S. minutus* are represented by two species each. Infection of *S. minutus* with cestodes was 78.7%, 2.5, and with trematodes—41.3%, 1.4.

The composition of dominant and subdominant helminth species in the pygmy shrew turned out to be the same as in *S. araneus*, but the order of dominance was different. The helminth fauna of *S. minutus* was dominated by *D. diaphana* (10.2) and *L. paradoxi* (19.6); the subdominants were *R. exasperatum* (8.6) and *M. arionis* (8.0). The remaining four parasite species were adominants.

Four species of helminths (*D. diaphana, M. arionis, R. exasperatum,* and *L. paradoxi*) were found in *S. minutus* in both studied protected areas of Mordovia. The remaining four species (*P. soricis, P. soricina, A. exigua,* and *P. skrjabini*) were found in the pygmy shrew only in the Mordovia Nature Reserve.

*Sorex isodon* was studied only in the Mordovia Nature Reserve, where 12 species of helminths, also typical of *S. araneus*, were found. The total infection rate of *S. isodon* with parasites reaches 100%, 19.6. The helminth fauna of *S. isodon*, like other shrew species, was dominated by nematodes (5 species). The infection rate of *S. isodon* with these parasites was 100%, 13.8. Cestodes and trematodes are represented by three species each. Infection with cestodes was 83.3%, 4.4, and with digeneans—41.7%, 1.3. Acanthocephalans are represented by larvae of one species, *C. aluconis* (8.3%, 0.2). Among the helminths of *S. isodon*, the nematode *L. paradoxi* (29.4) dominated. The subdominants were the cestodes *M. arionis* (7.4), *D. diaphana* (3.6), and the nematode *L. incrassatus* (10.0). The remaining eight species of helminths were adominants.

The helminth fauna of *N. fodiens* includes nine species of parasites, represented by adult worms (Table 1). Six species (*R. exasperatum*, *R. opisthovitellinus*, *P. soricis*, *A. exigua*, *A. kutorii*, and *P. soricina*) are host-specific parasites of shrews from the family Soricidae, while the trematode *N. locellus* and the nematode *L. neomi* are associated with representatives of the genus *Neomys*. The nematode *L. incrassatus* parasitizes a wide range of soricomorphs. All six examined water shrews were infected with helminths (100%, 12.2). Five species of nematodes were found in all studied *N. fodiens* individuals (MA = 7.7). Trematodes in *N. fodiens* were represented by four species and were found in all shrews (100%, 4.5). In both protected areas, four helminth species (*P. soricis*, *N. locellus*, *R. opisthovitellinus*, and *A. kutorii*) were recorded for the water shrew. The nematodes *P. soricina* and *A. exigua* were found in *N. fodiens* only in the National Park "Smolny", while the trematode *R. exasperatum*, the nematodes *L. incrassatus*, and *L. neomi* were recorded only in the Mordovia Nature Reserve. The most common helminth species identified in *N. fodiens* was the trematode *P. soricis*, found in five of six shrews studied (83.3%, 2.7).

Three species of parasitic worms were found in the only studied specimen of *N. milleri* in the Mordovia Nature Reserve, i.e., *R. exasperatum*, *L. incrassatus*, and *L. neomi* (a total of five helminth specimens). In three studied individuals of *C. suaveolens* from the Mordovia Nature Reserve, one parasite species was recorded, the cestode *S. brusatae*, which is a host-specific parasite of the lesser white-toothed shrew.

The helminth fauna of *T. europaea* is represented by two species of nematodes, i.e., *T. linstowi* and *P. depressum*, juv. *Tricholinstowia linstowi* is a specific monohostal parasite of the European mole. At the same time, *P. depressum*, juv. is a common parasite of insectivorous mammals. The total infection rate of moles with parasites was 85.7%, 31.4. These nematode species were found in moles in both protected areas. The most common parasite found in moles is *P. depressum*, juv. (85.7%, 15.0).

The helminth fauna of *E. roumanicus* includes eight species of helminths. Of these, four species (*H. erinacei*, *A. erinacei*, *C. striatum*, and *Ph. clausa*) are host-specific parasites of hedgehogs. The remaining four helminth species are facultative or occasional parasites of hedgehogs (*I. melis*, *S. strigis*, mtc., *Ph. sexalatus*, juv., and *A. minuta*, juv.). In our study, all eight hedgehogs were infected with helminths (100%, 90.3). In the helminth community of *E. roumanicus*, nematodes predominate both in the number of species (4) and in terms of infection rates. Roundworms were found in all hedgehog specimens (MA = 69.9). Nematodes are represented by both adults (*A. erinacei*, *P. clausa*) and larval

Diversity **2024**, 16, 307 8 of 24

forms (*Ph. sexalatus*, *A. minuta*). Trematodes in *E. roumanicus* are represented by two species and were found in two of eight examined hedgehogs (25.0%, 1.8). The trematode *I. melis* was recorded at the mature stage, and *S. strigis* was recorded at the metacercarial stage. Cestodes were represented by only one species—*H. erinacei*. In both protected areas, *E. roumanicus* has three species of parasites (*H. erinacei*, *A. erinacei*, and *P. clausa*). The remaining five helminth species (*I. melis*, *A. minuta*, juv., *C. striatum*, *S. strigis*, mtc., and *Ph. sexalatus*, juv.) were found in hedgehogs only in the National Park "Smolny".

No shared parasites were identified in all the species of insectivores we studied. Only *S. araneus* shares parasite species with other animals studied. The largest host range was recorded for the trematode *R. exasperatum* (five species). The trematode *P. soricis* and the nematode *Liniscus incrassatus* each use four species of insectivores as hosts. Three host species each were found for *D. diaphana, M. arionis, L. paradoxi, P. soricine,* and *P. depressum,* juv. The host range of 11 species of helminths (*B. fulvum, R. opisthovitellinus, S. strigis,* mtc., *N. schaldybini, A. exigua, A. kutorii, C. soricicola, E. oesophagicola, L. neomi, P. clausa,* and *C. aluconis,* larvae) includes two species of insectivores. The remaining 15 species of parasitic worms are each found in only one host.

Only one of the 34 species of helminths found in insectivores from the protected areas of Mordovia has veterinary significance as a causative agent of dangerous helminthiasis. This is a nematode, *Ph. sexalatus*, juv., that causes physocephalosis in wild boars and domestic pigs.

# 3.2. List of Helminths of Insectivorous Mammals in the Middle Volga Region

Previous studies of parasitic worms in insectivorous mammals were carried out in six of the 11 regions of the Middle Volga region, i.e., Chuvashia, Mari El, Mordovia, Tatarstan, Nizhny Novgorod, and Samara regions (Figure 1).



**Figure 1.** Map of the Middle Volga region showing regions with cases of detection of helminth species in insectivorous mammals (dark red—more than 30 species of helminths detected; red—10 or more helminth species; pink—less than 10 species; white—no helminthological studies). The numbers show the number of helminth species identified.

Taking into account the results of our study, the helminth fauna of insectivores includes 52 parasite species found in 9 species of soricomorphs and hedgehogs; these are 14 cestode, 13 trematode, 22 nematode, and 3 acanthocephalan species (Table 3).

**Table 3.** List of helminths found in insectivorous mammals of the Middle Volga region (European Russia).

Helminth Species	D	Hosts	Locality	References
Family Hymenolepididae Hymenolepis erinacei (Gmelin, 1789)	Palaearctic	Erinaceus roumanicus	Mordovia (MNR, NP "Smolny") Samara region (Zhiguli Reserve)	this study [128,130–132]
Monocercus arionis Siebold,	Delegantia	Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147], this study
1850	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
No. 1 i di colori di di dici		Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147], this study
Neoskrjabinolepis schaldybini Spasskii, 1947	Palaearctic	Sorex araneus	Nizhny Novgorod region	[145]
•		Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Ditestolepis diaphana	Delegantic	Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147], this study
(Cholodkowsky, 1906)	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Insectivorolepis infirma Zarnowski, 1955	Palaearctic	Sorex araneus, Sorex minutus	Mordovia (MNR)	[147]
Neomylepis magnirostellata (Baer, 1931)	Palaearctic	Neomys fodiens	Mordovia (MNR)	[147]
Pseudobothrialepis		Sorex araneus, Sorex minutus	Mordovia (MNR)	[147]
mathevossianae Schaldybin, 1957	Palaearctic	Sorex araneus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Soricinia soricis (Baer, 1928)	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Spasskylepis ovaluteri Schaldybin, 1964	Palaearctic	Sorex araneus, Sorex minutus, Neomys fodiens	Mordovia (MNR)	[147]
Staphylocystis brusatae (Vaucher, 1971)	Europe	Crocidura suaveolens	Mordovia (MNR), Samara region (NP "Samarskaya Luka")	[128,130], this study
Staphylocystis furcata (Stieda, 1862)	Palaearctic	Sorex araneus, Sorex minutus	Mordovia (MNR), Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132,147]
Staphylocystoides stefanskii (Zarnowski, 1954)	Palaearctic	Sorex araneus	Mordovia (MNR)	[147]
Vigisolepis spinulosa		Sorex araneus, Sorex minutus	Mordovia (MNR, NP "Smolny")	[147], this study
(Cholodkowsky, 1906)	Palaearctic	Sorex araneus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Family Dilepididae Dilepis undula (Schrank, 1788), larvae	Palaearctic	Sorex araneus	Mordovia (MNR, NP "Smolny")	this study
Family Brachylaimidae Brachylaima fulvum Dujardin,	Palaearctic	Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147,162], this study
1843		Sorex araneus	Nizhny Novgorod region	[145]

 Table 3. Cont.

Helminth Species	D	Hosts	Locality	References
Ityogonimus talpae (Goeze, 1782)	Palaearctic	Talpa europaea	Nizhny Novgorod region	[144]
Family Panopistidae Pseudoleucochloridium soricis (Soltys, 1952)	Palaearctic	Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147,162], this study
Family Opisthorchiidae Metorchis bilis (Braun, 1890)	Holarctic	Neomys fodiens	Mordovia (MNR)	[147]
Family Echinostomatidae Isthmiophora melis (Schrank, 1788)	Holarctic	Erinaceus roumanicus	Mordovia (NP "Smolny")	this study
Family Omphalometridae Neoglyphe locellus (Kossack, 1910)	Holarctic	Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147,162], this study
		Sorex araneus, Sorex minutus	Mordovia (MNR, NP "Smolny")	[147,162], this study
Neoglyphe sobolevi Schaldybin, 1953	Holarctic	Sorex araneus Sorex araneus, Sorex minutus	Nizhny Novgorod region Samara region (NP "Samarskaya Luka")	[145] [128–132]
		Sorex araneus, Sorex isodon, Sorex minutus, Neomys fodiens, Neomys milleri	Mordovia (MNR, NP "Smolny")	[147,162], this study
Rubenstrema exasperatum (Rudolphi, 1819)	Holarctic	Sorex araneus Sorex araneus, Sorex minutus	Nizhny Novgorod region Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[145] [128–132]
Rubenstrema opisthovitellinus (Soltys, 1954)	Palaearctic	Sorex araneus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[162], this study
Omphalometra desmanae (Sobolev, Maschkov et Maschkov, 1939)	Europe	Desmana moschata	Nizhny Novgorod region Mordovia (MNR) Tatarstan Samara region	[163] [147] [164] [165]
Family Diplostomidae Alaria alata (Goeze, 1782), mesocercaria	Cosmopolitan	Sorex araneus	Samara region (NP "Samarskaya Luka")	[128–130]
Family Strigeidae Strigea strigis (Schrank, 1788), metacercaria	Palaearctic	Erinaceus roumanicus, Sorex araneus	Mordovia (NP "Smolny")	this study
Holostephanus desmanae	Europe	Desmana moschata	Nizhny Novgorod region	[166]
Family Capillariidae		Sorex araneus Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147], this study
Aonchotheca kutorii (Ruchljadeva, 1964)	Palaearctic	Sorex araneus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Aonchotheca erinacei (Rudolphi, 1819)	Palaearctic	Erinaceus roumanicus	Mordovia (MNR, NP "Smolny") Samara region (Zhiguli Reserve)	[128,130–132], this study
		Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	this study
Aonchotheca exigua (Dujardin, 1845)	Europe	Sorex minutus, Crocidura suaveolens	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128,130–132]
Calodium soricicola (Yokogava		Sorex araneus, Sorex isodon	Mordovia (MNR, NP "Smolny")	this study
et Nishigori, 1924)	Palaearctic	Sorex araneus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]

 Table 3. Cont.

Helminth Species	D	Hosts	Locality	References	
Eucoleus oesophagicola (Soltys, 1952)	Palaearctic	Sorex araneus, Sorex isodon, Neomys fodiens Sorex araneus, Erinaceus roumanicus, Talpa europaea	Mordovia (MNR, NP "Smolny") Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[147], this study	
		Sorex araneus, Sorex isodon, Neomys fodiens, Neomys milleri	Mordovia (MNR, NP "Smolny")	[147], this study	
Liniscus incrassatus Diesing, 1851	Palaearctic	Neomys fodiens Sorex araneus	Tatarstan Samara region (Zhiguli Reserve, NP "Samarskaya	[134] [128–132]	
		Sorex uruneus	Luka")	[120 102]	
Nematoideum talpae Siebold, 1850	Palaearctic	Talpa europaea	Samara region (Zhiguli Reserve)	[128,130–132]	
Family Soboliphymidae	<b>D</b> .1	Sorex araneus, Neomys fodiens	Mordovia (MNR, NP "Smolny")	[147], this study	
Soboliphyme soricis Baylis et King, 1932	Palaearctic	Sorex araneus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]	
Family Metastrongylidae Crenosoma striatum Zeder, 1800	Palaearctic	Erinaceus roumanicus	Mordovia (NP "Smolny"), Samara region (Zhiguli Reserve), Nizhny Novgorod region	[128,130–132,143], this study	
Paracrenosoma skrjabini (Pologentsev, 1935)	Palaearctic	Sorex minutus	Mordovia (MNR), Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128,130–132], this study	
Family Heligmosomidae		Sorex araneus, Sorex isodon, Sorex minutus	Mordovia (MNR, NP "Smolny")	[147], this study	
Longistriata paradoxi Schaldybin, 1964	Palaearctic	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Longistriata codrus Thomas, 1953	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]	
Longistriata neomi Lubarskaja, 1962	Palaearctic	Neomys fodiens Neomys fodiens, Neomys milleri	Tatarstan Mordovia (MNR)	[134] [147], this study	
Tricholinstowia linstowi Travassos, 1918	Palaearctic	Talpa europaea	Mordovia (MNR, NP "Smolny"), Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128,130–132], this study	
Tricholinstowia talpae (Morgan, 1928)	Palaearctic	Talpa europaea	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128,130–132]	
		Talpa europaea Sorex araneus, Sorex isodon, Talpa europaea	Chuvashia, Mari El Mordovia (MNR, NP "Smolny")	[133] this study	
Family Anisakidae Porrocaecum depressum (Zeder,	Cosmopolitan	Neomys fodiens Talpa europaea	Tatarstan	[133,134]	
1800), juveniles	Comoponum	Talpa europaea	Nizhny Novgorod region	[144]	
		Sorex araneus, Talpa europaea, Erinaceus roumanicus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]	
Family Habronematidae Hadjelia truncata (Creplin, 1825), juveniles	Palaearctic	Sorex araneus	Mordovia (MNR, NP "Smolny")	this study	
Family Physalopteridae  Physaloptera clausa Rudolphi,	Holarctic	Erinaceus roumanicus (adults), Sorex araneus (larvae)	Mordovia (MNR, NP "Smolny"), Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128,130–132], this study	
1819		Erinaceus roumanicus	Nizhny Novgorod region	[143]	

Table 3. Cont.

Helminth Species	D	Hosts	Locality	References
Pseudophysaloptera soricina	***	Sorex araneus, Sorex minutus, Neomys fodiens	Mordovia (MNR, NP "Smolny") Samara region (Zhiguli	this study
Baylis, 1934	Holarctic	Sorex araneus, Sorex minutus	Reserve, NP "Samarskaya Luka")	[128–132]
Family Spiruridae Spirura talpae (Gmelin, 1790)	Europe	Talpa europaea	Tatarstan	[133]
Family Spirocercidae Physocephalus sexalatus (Molin, 1860), juveniles	Cosmopolitan	Erinaceus roumanicus	Mordovia (NP "Smolny")	this study
Species insertae sedis Agamospirura minuta (Sharpilo, 1963), juveniles	Palaearctic	Erinaceus roumanicus	Mordovia (NP "Smolny")	this study
Family Centrorhynchidae		Sorex araneus, Sorex isodon	Mordovia (MNR)	this study
Centrorhynchus aluconis (Müller, 1780), larvae	Palaearctic	Sorex araneus, Sorex minutus	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]
Family Oligacanthorhynchidae <i>Oligacanthorhynchus citilli</i> (Rudolphi, 1806)	Palaearctic	Crocidura suaveolens	Samara region (NP "Samarskaya Luka")	[128,130]
Family Moniliformidae Moniliformis moniliformis (Bremser, 1811), larvae	Cosmopolitan	Sorex araneus, Sorex minutus, Crocidura suaveolens	Samara region (Zhiguli Reserve, NP "Samarskaya Luka")	[128–132]

Note: D-distribution.

In the Middle Volga region, the main part of the helminth fauna of insectivores consists of host-specific parasite species (42 species). Occasional and facultative parasites of insectivores are represented by nine species, i.e., *D. undula* (larvae), *M. bilis*, *A. alata* (msc.), *S. strigis* (mtc.), *Ph. clausa* (juv.) for *S. araneus*, *Ph. sexalatus* (juv.), *H. truncata* (juv.), *C. aluconis* (larvae), *O. citilli*, and *M. moniliformis* (larvae).

Shrews, moles, and hedgehogs are the definitive hosts of 42 species of parasitic worms. These insectivorous mammals serve as intermediate and/or paratenic hosts for nine species of helminths, i.e., *D. undula*, *A. alata*, *S. strigis*, *P. depressum*, *H. truncata*, *Ph. clausa* (juv.) (for *S. araneus*), *Ph. sexalatus*, *A. minuta*, *C. aluconis*, and *M. moniliformis*. Adult worms of *Ph. clausa* are host-specific parasites of hedgehogs. *Sorex araneus* serves as the paratenic host of the larval stage of this nematode.

The majority of helminths (40 species) recorded in insectivores have an indirect life cycle. Their definitive or intermediate/paratenic hosts are shrews, moles, and hedgehogs. Parasites with a direct life cycle include only 12 species of nematodes from the families Capillariidae and Heligmosomidae.

In the insectivores of the Middle Volga region, nematodes, which are represented by nine families, have the greatest species richness. Of these, the family Capillariidae (seven species) is the most represented. Five of the identified species belong to the family Heligmosomidae, and two species each belong to the families Metastrongylidae and Physalopteridae. The families Anisakidae, Habronematidae, Soboliphymidae, Spiruridae, and Spirocercidae are represented by only one species.

Trematodes in the studied insectivores are representatives of seven families. Among them, the family Omphalometridae (five species) predominates. Two species belong to the family Brachylaimidae. The remaining four families of trematodes (Diplostomidae, Panopistidae, Echinostomatidae, and Strigeidae) in the helminth fauna of insectivores are represented by one species each. Cestodes and acanthocephalans are the least represented helminths in the insectivores of the Middle Volga region. The observed tapeworms belong to only two families, i.e., the Hymenolepididae (13 species) and the Dilepididae (1). Three

Diversity 2024, 16, 307 13 of 24

species of acanthocephalans recorded in the shrews of the Middle Volga region belong to the families Centrorhynchidae, Moniliformidae, and Oligacanthorhynchidae.

Despite the high species richness of helminths (42 species) found in insectivores in the protected areas of Mordovia, it has not yet reached its maximum. In total, 52 species of parasitic worms are currently known for 9 studied species of insectivores in the Middle Volga region (Table 3). In the Samara region, six species of these small mammals were studied, of which 33 species of helminths were registered. In particular, in the Zhiguli Nature Reserve and the National Park "Samarskaya Luka", we previously found 28 species of helminths each. The helminth fauna of the Nizhny Novgorod region turned out to be much poorer, since among four insectivorous hosts, 10 species of parasites were identified here. The least studied is the helminth fauna of insectivores in Tatarstan (5 species), as well as in Chuvashia and Mari El, where one parasite species (*P. depressum*, juv.) was found in the common mole (Table 3).

Most species of helminths (36) found in insectivores of the Middle Volga region belong to the Palearctic faunal complex (Table 3). Seven species (*R. exasperatum*, *P. soricina*, *M. bilis*, *I. melis*, *N. locellus*, *N. sobolevi*, and *P. clausa*) have a Holarctic distribution. Four species of helminths (*A. alata*, *P. depressum*, *Ph. sexalatus*, and *M. moniliformis*) are cosmopolitan. The distribution of five species (*O. desmanae*, *H. desmanae*, *S. brusatae*, *A. exigua*, and *S. talpae*) is limited to Europe.

Three of the 52 species of helminths found in insectivorous mammals in the Middle Volga region are of veterinary importance as causative agents of dangerous helminthiases. These are the trematodes *A. alata* and *M. bilis* and the nematode *Ph. sexalatus*, juv. that cause alariosis (affects canids and mustelids), metorchosis (piscivorous animals), and physocephalosis (wild boars and domestic pigs), respectively.

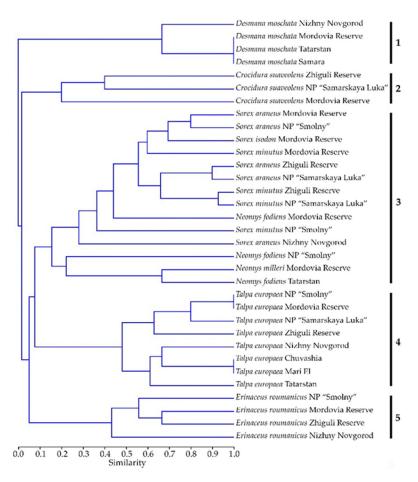
We conducted a comparative analysis of the species structure of helminths in insectivores from different areas of the Middle Volga region. A dendrogram of the similarity of the helminth fauna of various insectivorous species is presented in Figure 2.

As a result of clustering, the studied insectivores were divided into five groups with the closest species composition of helminths. The first single-species group consists of parasites of *D. moschata*. In this group, complete similarity of the helminth fauna was revealed for *D. moschata* from the Samara region, Tatarstan, and the Mordovia Nature Reserve. The highest degree of similarity in the composition of helminths was found in *D. moschata* from the Nizhny Novgorod region and other territories (0.67).

The second group is also single-species and is represented by the helminth fauna of *C. suaveolens*. In this group, an average degree of similarity in the composition of helminths was revealed for the lesser white-toothed shrew from the Zhiguli Nature Reserve and the National Park "Samarskaya Luka" (0.40). The helminth fauna of *C. suaveolens* from the Mordovia Nature Reserve is also close to them (0.40).

The third group is the largest and has five species. It includes the parasitic fauna of shrews of the genera *Sorex* and *Neomys*. A high degree of similarity here was revealed for *S. minutus* from the Zhiguli Reserve and the NP "Samarskaya Luka" (0.93), *S. araneus* from the Zhiguli Reserve and the NP "Samarskaya Luka" (0.90), *S. araneus* from the Mordovia Reserve and the NP "Smolny" (0.80), *S. minutus* and *S. araneus* from the Mordovia Reserve (0.72), *S. araneus* from the NP "Smolny" and *S. isodon* from the Mordovia Reserve (0.71), *S. araneus* from the NP "Smolny" and the NP "Samarskaya Luka" (0.70), *S. araneus* and *S. isodon* from the Mordovia Reserve (0.69), *S. araneus* from the NP "Smolny" and the Zhiguli Reserve (0.68), *S. araneus* from the Zhiguli Reserve and the NP "Samarskaya Luka" (0.68), *S. araneus* from the Zhiguli Reserve and the Mordovia Reserve (0.67), *N. fodiens* from Tatarstan and *N. milleri* from the Mordovia Reserve (0.67). The remaining pairs in the third group showed an average and low degree of similarity.

Diversity 2024, 16, 307 14 of 24



**Figure 2.** Dendrogram of the similarity of the helminth fauna of insectivores from different areas of the Middle Volga region, obtained using the Morisita index (UPGMA). 1–5—groups with the most similar helminth fauna. Correlation coefficient: r = 0.935.

The fourth group is formed by the helminth fauna of *T. europaea* from different regions of the Middle Volga region. This group is divided into two subgroups. The first subgroup is formed by helminths of moles from protected areas of Mordovia and the Samara region. Here, a complete similarity of helminths was revealed for *T. europaea* from the NP "Smolny" and the Mordovia Reserve. A high degree of similarity of the helminth fauna was noted in moles from the NP "Smolny" and the NP "Samarskaya Luka", from the Mordovia Reserve and the NP "Samarskaya Luka" (0.80 each), from the Zhiguli Reserve, and from the NP "Samarskaya Luka" (0.75). The second subgroup is formed by helminths of moles from the Nizhny Novgorod region, Chuvashia, Mari El, and Tatarstan, where complete similarity of the helminth fauna was revealed for Chuvashia and Mari El. A high degree of similarity was revealed for Tatarstan, Chuvashia, and Mari El (0.67 each).

The fifth group consists of the helminth fauna of hedgehogs from different regions of the Middle Volga region. The greatest similarity in this group was found between the Zhiguli Reserve and the Mordovia Reserve (0.67).

#### 4. Discussion

Previously, 23 species of helminths were known to parasitize insectivorous mammals in Mordovia [148]. As a result of our research, 19 species of parasites were added to this list. Thus, the list of helminths of insectivores in Mordovia now includes 42 species.

Analysis of the helminth fauna of insectivores from both protected areas of Mordovia showed that nematodes are characterized by the greatest species richness (18 species) (Table 2). The identified nematode species have different life cycles and different localizations in the host body. Host-specific nematodes (*A. kutorii*, *A. erinacei*, *A. exigua*, *C. soricicola*,

Diversity **2024**, 16, 307 15 of 24

*E. oesophagicola, L. incrassatus, L. paradoxi, L. neomi*, and *T. linstowi*) develop directly, without the involvement of intermediate hosts. Infection with these parasites occurs through accidental ingestion of nematode eggs and infective larvae in close contact with the soil litter. In addition, infection can occur when insectivores feed on earthworms, which are reservoir hosts for these geohelminths [121,167,168].

Nine species of helminths found in the insectivores studied are parasites with an indirect life cycle. Being one of the main food items for shrews and moles, earthworms serve as intermediate hosts for the nematodes *S. soricis* and *P. depressum* [169–172]. Infection of shrews and hedgehogs with nematodes *Ph. clausa*, *P. soricina*, *Ph. sexalatus*, and *H. truncata* occurs when animals feed on beetles of the families Tenebrionidae and Scarabaenidae, which are intermediate hosts of these parasites [55,172–174]. The development of *C. striatum*, like, apparently, that of *P. skrjabini*, occurs with the participation of terrestrial gastropods (slugs and snails), through which hedgehogs and shrews become infected with nematodes [175]. Thus, gastropods *Radix balthica* (Linnaeus, 1758), *Morlina glabra* (Rossmassler, 1835), *Arion circumscriptus* Johnston, 1828, *Succinea putris* Linnaeus, 1758, *Urticicola umbrosus* (Pfeiffer, 1828), *Malacolimax tenellus* (Muller, 1774) and *Tandonia rustica* (Millet, 1843) are known as intermediate hosts of this nematode species [176].

Nematodes have different localizations in the bodies of their hosts. Most of these helminths parasitize in various parts of the gastrointestinal tract, i.e., the esophagus (*E. oesophagicola*), stomach (*Ph. clausa*, *P. soricina*, *S. soricis*), small intestine (*A. kutorii*, *A. erinacei*, *A. exigua*, *L. paradoxi*, *L. neomi*, *T. linstowi*, *H. truncata*, juv.), and liver (*C. soricicola*). Nematodes are also found in the bladder (*L. incrassatus*), lungs (*C. striatum*, *P. crenosoma*), and walls of the esophagus, stomach, and intestines (*P. depressum*, juv., *Ph. sexalatus*, juv., *A. minuta*, juv.).

The trematode fauna of insectivores in protected areas of Mordovia is represented by eight species (Table 2). Infection occurs exclusively when animals eat invertebrates, which are intermediate hosts of these parasites. Thus, the trematodes *B. fulvum* and *P. soricis* use terrestrial gastropods as intermediate hosts [44,177].

In North America, the intermediate host of the trematode *N. locellus* is the freshwater mollusk *Galba* (*Bakerilymnaea*) *bulimoides* (Lea, 1841) [178]. Metacercariae develop in gastropods, larvae, and adults of aquatic and semi-aquatic insects of the Coleoptera, Neuroptera, and Diptera [55,179,180]. The development of other representatives of the family Omphalometridae (*N. sobolevi*, *R. exasperatum*, and, apparently, *R. opisthovitellinus*) occurs with the participation of gastropods of the genera *Lymnaea* and *Planorbarius*. *Culex pipiens* Linnaeus, 1758 mosquito larvae, have been reported as second intermediate hosts [181,182]. *Isthmiophora melis* develops with the involvement of the gastropod *Lymnaea stagnalis* (Linnaeus, 1758). The second intermediate hosts of this parasite are amphibians and freshwater fish [183–185].

Infection of shrews and hedgehogs with the metacercariae of *S. strigis* probably occurs when the animals feed on amphibians, which are the second intermediate host of this parasite [186]. Infection of insectivores with this trematode species is less likely through aquatic gastropods, intermediate hosts of strigeids [25,186].

Most trematodes are localized in the gastrointestinal tract of insectivores. Five digenean species (*B. fulvum*, *I. melis*, *N. locellus*, *N. sobolevi*, and *R. opisthovitellinus*) parasitize only in the small intestine, and *R. exasperatum*—only in the stomach of shrews. *Pseudoleucochloridium soricis* was found in both the stomach and small intestine of shrews. Only the metacercariae of *S. strigis* are localized in the adipose tissue around the trachea and esophagus.

The cestode fauna of insectivores in the protected areas of Mordovia includes seven species (Table 2). Like trematodes, all cestode species have an indirect life cycle. Shrews and hedgehogs become infected with species such as *N. schaldybini*, *D. diaphana*, *V. spinulosa*, and *H. erinacei* by feeding on the beetles, which serve as intermediate hosts for these tapeworms [55,187]. The development of *H. erinacei* cysticercoids can occur both in the intestinal villi of the definitive hosts (hedgehogs) and in the small intestine, where they

Diversity 2024, 16, 307 16 of 24

grow to maturity [188], with the participation of intermediate hosts—beetles of the genera *Oiceoptoma, Nicrophorus*, and *Geotrupes* [187]. The development of *M. arionis* occurs with the participation of terrestrial mollusks of the genus *Succinea* [125,189]. Infection of shrews with the cestode *D. undula* probably occurred when they ate the intermediate hosts of the cestode—oligochaetes [125]. The presence of acanthocephalan larvae of *C. aluconis* in shrews indicates that the animals consume the intermediate hosts of this parasite—beetles of the families Tenebrionidae and Scarabaeidae [190,191].

Findings of larval forms of helminths (*S. strigis*, *P. depressum*, *Ph. sexalatus*, and *C. aluconis*) in insectivores indicate the important role of these small mammals in the spread and circulation of parasitic worms of carnivores and birds of prey. Hedgehogs and shrews are paratenic hosts of these parasites.

In our opinion, the findings of larvae of the cestode *D. undula* and the nematode *H. truncata* in the intestines of shrews are cases of transit parasitism. These parasites do not reach maturity in the intestine of shrews and leave the host body without development.

Agamospirura minuta is an occasional parasite of insectivores. This juvenile nematode is a specific parasite of reptiles, usually parasitizing the slowworm Anguis fragilis Linnaeus, 1758, and less commonly other lizards and snakes [192]. The definitive hosts of this parasite are unknown. Lewin [193] found similar nematode larvae in reptiles in Poland and identified them as Protostrongylidae sp. Further studies using molecular genetic methods are required to accurately identify this parasite.

Despite the fact that the various species of insectivores under consideration often live in the same habitats, the strict specificity of their parasites to the species and genera of hosts is revealed. Our study showed that no shared parasite species was found among the studied hosts. The highest species richness of helminths was observed in the most numerous and eurytopic shrew, *S. araneus* (22 species). It is this species of shrew that shares parasites with all other studied insectivores. Compared to *S. araneus*, all other insectivores have a poorer composition of helminths. Apparently, this is due to the small sample size of the animals examined. In addition, the formation of helminth fauna is influenced by the lifestyle of the host. For example, *C. suaveolens* prefers to settle in open, dry habitats where the likelihood of infection with helminths is low. The underground lifestyle of *T. europaea* limits the range of food items as possible intermediate hosts of helminths, which reduces the diversity of the parasitic fauna. Due to its lifestyle, *E. roumanicus* has a specific composition of parasites. The enrichment of its helminth fauna occurs due to occasional parasites characteristic of other vertebrates [25].

It should be noted that all the species of insectivores we studied, due to their diet, behavioral characteristics, and lifestyle throughout their range, are characterized by a high infection with helminths [115,125,129,194,195].

According to our data and previous records, there are 52 species of parasites in the helminth fauna of insectivores in the Middle Volga region. A comparative analysis of the species composition of helminths in insectivores from different areas of the Middle Volga region showed both significant similarity in the helminth fauna of insectivorous species as well as its originality in a certain research area.

In *Sorex* spp., the majority of helminth species recorded are common and widespread parasites of shrews, occurring throughout their range. This explains the high and average degree of similarity of the helminth fauna of representatives of the genus *Sorex* from different areas of the Middle Volga region (Figure 2).

The helminth fauna of *E. roumanicus* in the Middle Volga region includes nine species, of which four were found only in the National Park "Smolny". Therefore, the similarity of the helminth fauna of hedgehogs in the NP "Smolny" with the helminth fauna in other regions is less. Only two species of helminths have been registered in hedgehogs of the Nizhny Novgorod region, which determines the low similarity of its helminth fauna with other regions (Figure 2).

The uniqueness of the helminth fauna of *N. fodiens* from the National Park "Smolny" is due to the presence of nematodes *A. exigua* and *P. soricina* (Tables 2 and 3). The water

Diversity 2024, 16, 307 17 of 24

shrew from the NP "Smolny" also lacks a number of parasites found in *N. fodiens* from the Mordovia Nature Reserve. This fact determines the low degree of similarity of the helminth fauna of water shrews from the NP "Smolny", the Mordovia Nature Reserve, and Tatarstan. We found an average degree of similarity in the composition of helminths in *Sorex* and *Neomys* species (Figure 2). At the same time, of the 19 species of parasites found in *N. fodiens* of the Middle Volga region, 14 are common with *S. araneus*, and 8 species each are shared with *S. isodon* and *S. minutus*.

Only four species of helminths have been registered in *C. suaveolens*, including one species in the Mordovia Nature Reserve and three species in both protected areas of the Samara region (Table 3). No shared species of helminths were identified in the lesser white-toothed shrew from the Zhiguli and Mordovia Nature Reserves. For *C. suaveolens* from the Zhiguli Nature Reserve and the National Park "Samarskaya Luka", as well as from the Mordovia Nature Reserve and the National Park "Samarskaya Luka", only one shared species of helminths was noted. Therefore, *C. suaveolens* from the Zhiguli Nature Reserve and the National Park "Samarskaya Luka" have an average degree of similarity (Figure 2).

In *D. moschata*, only two species of specific trematodes were noted, of which *O. desmanae* was found in four regions, which ensures complete similarity of the helminth community in the Mordovia Nature Reserve, Tatarstan, and the Samara region (Table 3, Figure 2).

In total, six species of helminths have been registered in *T. europaea* in the Middle Volga region (Table 3). Only two species of parasites were found in moles from the Mordovia Nature Reserve and the National Park "Smolny". This determines the complete similarity of their helminth fauna. One species (*P. depresum*, juv.) was found in moles in all studied areas of the Middle Volga region. This fact determines the complete similarity of the helminth fauna of *T. europaea* from Chuvashia and Mari El, where only this species of helminth was found in moles.

Differences in the helminth composition of insectivores from different areas of the Middle Volga region are mainly due to the degree of study of the helminth fauna of various Eulipotiphla species (Figure 2). On the other hand, differences in the helminth fauna of insectivores may depend on abiotic and biotic factors at each specific study site, such as microclimatic conditions of habitats, species composition and abundance of invertebrates (intermediate and/or paratenic hosts of helminths), and population density of insectivorous mammals.

## 5. Conclusions

As a result of our research, the list of helminths in insectivorous mammals in Mordovia was replenished with 19 species of parasites and currently includes 42 species. For the first time, we registered the nematodes *A. exigua* and *P. soricina* in *N. fodiens* from European Russia, as well as the trematode *Rubenstrema opisthovitellinus* in *S. araneus* from the Volga basin. The first data on the helminths of *S. isodon* in the Volga basin have been obtained.

According to our research and previous records, the helminth fauna of insectivores of Eulipotyphla from the Middle Volga region includes 52 species, i.e., 14 cestodes, 13 trematodes, 22 nematodes, and 3 acanthocephalans. Most of them belong to the Palearctic faunal complex (36 species). Among all insectivorous species, the richest helminth fauna was found in *S. araneus* (31 species), *S. minutus* (20), and *N. fodiens* (19). The composition of helminths is less diverse in *S. isodon* (12), *E. roumanicus* (9), and *T. europaea* (6). The least studied soricomorphs also have the fewest recorded parasite species. These species include *C. suaveolens* (4 species), *N. milleri* (3), and *D. moschata* (2).

Forty of the 52 helminth species found in the insectivores of Eulipotyphla have an indirect life cycle. Shrews, moles, and hedgehogs serve as definitive or intermediate/paratenic hosts for these parasites. The twelve species of nematodes observed are geohelminths.

Insectivorous mammals are the definitive hosts of 42 species of parasitic worms. For nine species of helminths, these animals serve as intermediate and/or paratenic hosts. In

Diversity 2024, 16, 307 18 of 24

addition, the adult worm of *Ph. clausa* is a specific parasite of hedgehogs, and *S. araneus* serves as the paratenic host of the larval stage of this nematode. The bulk of the helminth fauna of insectivores consists of host-specific species (42). Facultative and occasional parasites are represented by nine species.

A comparative analysis of the helminth fauna of various insectivorous species in the Middle Volga region showed a high and average degree of similarity among the helminth communities within individual species and genera of Eulipotyphla.

**Supplementary Materials:** The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/d16050307/s1. Table S1: Sampling sites of helminths of insectivorous mammals, with coordinates.

**Author Contributions:** Conceptualization, A.B.R., A.A.K. and N.Y.K.; methodology, A.A.K., A.I.F. and N.Y.K.; software, A.A.K. and A.I.F.; validation, A.A.K. and N.Y.K.; formal analysis, A.A.K. and N.Y.K.; investigation, A.A.K. and N.Y.K.; resources, A.A.K., A.B.R., N.Y.K. and A.I.F.; data curation, A.A.K., N.Y.K. and A.B.R.; writing—original draft preparation, A.A.K., N.Y.K. and A.I.F.; writing—review and editing, N.Y.K., A.A.K. and A.B.R.; visualization, A.A.K., N.Y.K. and A.B.R.; supervision, A.A.K., N.Y.K. and A.B.R.; project administration, N.Y.K., A.I.F. and A.B.R.; funding acquisition, A.I.F. and A.B.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research has no external funding.

**Institutional Review Board Statement:** The study was approved by the Bioethics Committee of the Institute of Ecology of the Volga River Basin of RAS (registration number: 2/24; 26 March 2024). Our research was conducted in compliance with ethical standards for the humane treatment of animals in accordance with the recommended standards described in the Directive of the European Parliament and of the Council of the European Union of September 22, 2010, "On the protection of animals used for scientific purposes" (EU Directive 2010/63/EU).

**Data Availability Statement:** The data presented in this study are available upon request from the corresponding author.

**Acknowledgments:** Our study was carried out on the research topic No. 1023062000002-6-1.6.20;1.6.19 of the Institute of Ecology of the Volga River Basin of RAS, "Terrestrial vertebrates of the Middle Volga region and adjacent territories and their parasitic worms: ecological, faunistic, and biological aspects of the organization and functioning of communities against the background of natural and anthropogenic changes", and within the framework of state assignment 1-22-31-1 of the Ministry of Natural Resources and Ecology of the Russian Federation.

Conflicts of Interest: The authors declare no conflicts of interest.

## References

- 1. Sinclair, A.R.E. The role of mammals as ecosystem landscapers. *Alces* 2003, 39, 161–176.
- 2. Lacher, T.E., Jr.; Davidson, A.D.; Fleming, T.H.; Gomez-Ruiz, E.P.; McCracken, G.F.; Owen-Smith, N.; Peres, C.A.; Vander Wall, S.B. The functional roles of mammals in ecosystems. *J. Mamm.* **2019**, *100*, 942–964. [CrossRef]
- 3. Kirilyuk, A.V.; Kirilyuk, V.E.; Minaev, A.N. Daily activity patterns of wolves in open habitats in the Dauria ecoregion, Russia. *Nat. Conserv. Res.* **2021**, *6*, 95–109. [CrossRef]
- 4. Sokolova, N.A.; Litvinova, E.M.; Sorokin, P.A.; Hernandez-Blanco, J.A. How Eurasian otters (*Lutra lutra*) use suboptimal habitats? Space use dynamics in forest streams of Central Russian Upland. *Nat. Conserv. Res.* **2022**, *7*, 54–65. [CrossRef]
- 5. Coutinho, P.D.F.; Matte, A.L.L.; Bezerra, B. Manatee watching is widespread and seasonally affected in Northeast Brazil: A case of the Endangered *Trichechus manatus manatus* (Sirenia: Trichechidae). *Nat. Conserv. Res.* **2023**, *8*, 61–75. [CrossRef]
- 6. Hayward, G.F.; Phillipson, J. Community structure and functional role of small mammals in ecosystems. In *Ecology of Small Mammals*; Stoddart, D.M., Ed.; Chapman and Hall: London, UK; Wiley: New York, NY, USA, 1979; pp. 135–211. [CrossRef]
- 7. Bazhenov, Y.A. Ecology of bat species in the arid region of the Daurian steppe at the peak of drought. *Nat. Conserv. Res.* **2021**, *6*, 42–49. [CrossRef]
- 8. Martínez-Sotelo, J.; Sanchez-Jasso, J.M.; Ibarra-Zimbron, S.; Sanchez-Nava, P. Zoonotic intestinal parasites in free-ranging dogs (*Canis lupus familiaris*): A risk to public health in a Mexican Protected Area. *Nat. Conserv. Res.* **2022**, 7, 21–31. [CrossRef]
- 9. Kaicheen, S.S.; Mohd-Azlan, J. Community structures of mid-sized to large-bodied mammals in tropical lowland and lower montane forests in Gunung Pueh National Park, Western Sarawak, Borneo. *Nat. Conserv. Res.* **2022**, *7*, 70–79. [CrossRef]
- 10. Strann, K.; Yoccoz, N.; Ims, R. Is the heart of Fennoscandian rodent cycle still beating? A 14-year study of small mammals and Tengmalm's owls in northern Norway. *Ecography* **2002**, 25, 81–87. [CrossRef]

11. Elkinton, J.S.; Liebhold, A.M.; Muzika, R.-M. Effects of alternative prey on predation by small mammals on gypsy moth pupae. *Pop. Ecol.* **2004**, *46*, 171–178. [CrossRef]

- 12. Davidson, A.D.; Detling, J.; Brown, J.H. Ecological roles and conservation challenges of social, burrowing, herbivorous mammals in the world's grasslands. *Front. Ecol. Environ.* **2012**, *10*, 477–486. [CrossRef] [PubMed]
- 13. Kollberg, I.; Bylund, H.; Huitu, O.; Björkman, C. Regulation of forest defoliating insects through small mammal predation: Reconsidering the mechanisms. *Oecologia* **2014**, *176*, 975–983. [CrossRef] [PubMed]
- 14. Krebs, C.J.; Boonstra, R.; Boutin, S.; Sinclair, A.R.E.; Smith, J.N.M.; Gilbert, B.S.; Martin, K.; O'Donoghue, M.; Turkington, R. Trophic dynamics of the boreal forests of the Kluane region. *Arctic* **2014**, *67*, 71–81. [CrossRef]
- 15. Bobretsov, A.V. Population Ecology of Small Mammals in Plain and Mountain Landscapes of the North-East of the European Part of Russia; KMK: Moscow, Russia, 2016; pp. 3–381.
- 16. Yakimova, A.E.; Gaidysh, I.S. The species composition and abundance of terrestrial small mammals in the Finnish-Russian Friendship Nature Reserve. *Nat. Conserv. Res.* **2021**, *6*, 127–136. [CrossRef]
- 17. Churchfield, S. The Natural History of Shrews; Comstock Pub. Associates: New York, NY, USA, 1990; pp. 3–178.
- 18. Khodasheva, K.S.; Eliseeva, V.I. *Shrews in the Ecosystems of the Central Forest-Steppe of the Russian Plain*; Science: Moscow, Russia, 1992; pp. 3–212.
- 19. Liebhold, A.M.; Elkinton, J.; Williams, D.; Muzika, R. What causes outbreaks of the gypsy moth in North America? *Pop. Ecol.* **2000**, 42, 257–266. [CrossRef]
- 20. Rutovskaya, M.V.; Aleksandrov, A.N.; Podshivalina, V.N.; Soboleva, A.S.; Glushenkov, O.V. Habitat conditions of *Desmana moschata* (Talpidae, Eulipotyphla, Mammalia) in the buffer zone of the Prisurskiy State Nature Reserve (Russia). *Nat. Conserv. Res.* 2020, 5, 36–46. [CrossRef]
- 21. Shchipanov, N.A.; Alexandrov, D.Y.; Alexandrova, A.V. Distribution of spores of microscopic fungi of small mammals. *Zool. Zhurn.* **2006**, *85*, 101–113.
- 22. Alexandrov, D.Y. Distribution of Microscopic Fungal Spores on the Fur of Small Mammals. Ph.D. Thesis, Institute of Problems of Ecology and Evolution of RAS, Moscow, Russia, 2008; 172p.
- 23. Shimalov, V.V. Helminth fauna in insectivores of canal bamks in meliorared territories. Parazitologiia 2007, 41, 201–205. [PubMed]
- 24. Romashov, B.V.; Odoevskaya, I.M.; Romashova, N.B.; Golubova, N.A. Ecology of trichinellosis transmission in the Voronezh State Nature Reserve and adjacent areas, Russia. *Nat. Conserv. Res.* **2021**, *6*, 1–15. [CrossRef]
- 25. Kirillov, A.A.; Kirillova, N.Y.; Ruchin, A.B. Helminths of *Erinaceus roumanicus* (Eulipotyphla, Erinaceidae) in Mordovia (Russia) with an overview of helminth fauna of *Erinaceus* spp. inhabiting the Palaearctic region. *Diversity* **2022**, *14*, 165. [CrossRef]
- 26. Price, P.W. Evolutionary Biology of Parasites; Princeton University Press: Princeton, NJ, USA, 1980; pp. 3–237.
- 27. Poulin, R.; Morand, S. Parasite Biodiversity; Smithsonian Institution Press: Washington, DC, USA, 2004; pp. 3–216.
- 28. Horwitz, P.; Wilcox, B. Parasites, ecosystems and sustainability: An ecological and complex systems perspective. *Int. J. Parasitol.* **2005**, *35*, 725–732. [CrossRef] [PubMed]
- 29. Poulin, R. Phylogeny, ecology and the richness of parasite communities in vertebrates. Ecol. Monogr. 1995, 65, 283–302. [CrossRef]
- 30. Lafferty, K.D.; Allesina, S.; Arim, M.; Briggs, C.J.; De Leo, G.; Dobson, A.P.; Dunne, J.A.; Johnson, P.T.; Kuris, A.M.; Marcogliese, D.J.; et al. Parasites in food webs: The ultimate missing links. *Ecol. Lett.* **2008**, *11*, 533–546. [CrossRef] [PubMed]
- 31. Marcogliese, D.J. Parasites: Small players with crucial roles in the ecological theater. EcoHealth 2004, 1, 151–164. [CrossRef]
- 32. Stammer, H.J. Die Parasiten deutscher kleinsauger verhandlungen der Deutschen zoologischen gesselschaft in Erlangen. *Zool. Anz.* **1956**, *19*, 362–390.
- 33. Brendow, V. Ein beitrag zur trematodenfauna der Soricidae im raume gieben sowie im Naturpark Hoher Vogelsberg. *Z. Parasitenk.* 1970, 33, 282–313. [CrossRef] [PubMed]
- 34. Vaucher, C. Les cestodes, parasites de Soricidae d'Europe. Etude anatomique, revision taxonomique et biologie. *Rev. Suisse Zool.* 1971, 78, 1–113. [CrossRef]
- 35. Pecher, W.T.; Mokhtari-Derakhshan, F.; Böckeir, W. Epidemiologische Bemerkungen zur Helminthen fauna von zwei Spitzmausarten (*Sorex araneus* L. 1758, *S. minutus* L., 1766) mit unterschiedlichen ökologischen Ansprüchen. *Mitt. Osterr. Ges. Trop. Med. Parasitol.* 1996, 18, 183–190.
- 36. Pfaffle, M.P.; Bolfíkova, B.C.; Hulva, P.; Petney, T. Different parasite faunas in sympatric populations of sister hedgehog species in a secondary contact zone. *PLoS ONE* **2014**, *9*, e114030. [CrossRef]
- 37. Lambert, D. *Parasitosen und Mykosen des Igels*, 2nd ed.; Pro Igel e.V., Arbeitskreis Igelschutz Berlin e.V.: Berlin, Germany, 2019; pp. 3–149.
- 38. Soltys, A. The helminthes of common shrew (*Sorex araneus* L.) of the National Park of Bialowieza (Poland). *Ann. Univ. Marie Curie Sklodowska* **1952**, *5*, 165–202.
- 39. Soltys, A. Helmintofauna of Soricidae in the Bialowieza National Park. Acta Parasitol. Polon. 1954, 16, 353-402.
- 40. Zarnowski, E. Parasitic worms of forest micromammalians (Rodentia and Insectivora) of the environment of Pulawy (district Lublin). I. Cestoda. *Acta Parasitol. Polon.* **1955**, *8*, 127–168.
- 41. Zarnowski, E. Parasitic worms of forest micromammalians (Rodentia and Insectivora) of the environment of Pulawy (district Lublin). II. Trematoda. *Acta Parasitol. Polon.* **1960**, *3*, 279–368.
- 42. Furmaga, F. Materials to the helminth fauna of hedgehogs Erinaceus roumanicus Barrett-Hamilton. Acta Parasitol. 1961, 9, 441–445.

43. Kisielewska, K. Circulation of tapeworms in *Sorex araneus* L. in biocenosis of Bialowieza National Park. *Acta Parasitol. Polon.* **1961**, 9, 331–369.

- 44. Pojmanska, T. Investigations on the occurrence and biology of trematodes of *Sorex araneus araneus* L. in Bialowieza National Park. *Acta Parasitol. Polon.* **1961**, *9*, 305–330.
- 45. Mizgajska-Wiktor, H.; Jarosz, W.; Pilacinska, B.; Dziemian, S. Helminths of hedgehogs, *Erinaceus europaeus* and *E. roumanicus* from Poznan region, Poland–coprological study. *Wiad. Parazytol.* **2010**, *56*, 329–332. [PubMed]
- 46. Merkusheva, I.V.; Bobkova, A.F. *Helminths of Domesticated and Wild Animals in Belarus*; Nauka i Tehnika: Minsk, Belarus, 1981; pp. 3–120.
- 47. Shimalov, V.V.; Shimalov, V.V. Helminth fauna of the European mole (*Talpa europea* Linnaeus, 1758) in Bielorussian Polesie. *Parasitol. Res.* **2001**, 87, 790–791. [CrossRef] [PubMed]
- 48. Shimalov, V.V. Monitoring of the helminth of insectivorous of meliorative canal banks in Belorussian Polesie. *Parazitologiia* **2012**, 46, 472–478.
- 49. Shimalov, V.V. Helminth parasites of the Eurasian water shrew (*Neomys fodiens* Pennant, 1771) in South-West Belarus. *J. Parasit. Dis.* **2017**, 41, 602–604. [CrossRef]
- 50. Bychkova, E.I.; Akimova, L.N.; Degtyarik, S.M.; Yakovich, M.M. *Helminths of Vertebrates and Man in Belarus*; Belarusskaya Nauka Publishing House: Minsk, Belarus, 2017; pp. 3–316.
- 51. Shimalov, V.V.; Zhukovsky, A.T. The Bicolored Shrew (*Crocidura leucodon* Hermann, 1780) and its helminth fauna on the reclamation systems of the Brest Polesie. *Proc. Brest Univ.* **2019**, *2*, 55–59.
- 52. Genov, T. Helminths of Insectivores and Rodents in Bulgaria; Bulgarian Academy of Sciences: Sofia, Bulgaria, 1984; pp. 3–348.
- 53. Vasileva, G.P.; Tkach, V.V.; Genov, T. Two new hymenolepidid species (Cestoda, Hymenolepididae) from water shrews *Neomys fodiens* Pennant (Insectivora, Soricidae) in Bulgaria. *Acta Parasitol.* **2005**, *50*, 56–64.
- 54. Prokopic, J. Faunistic study of helminths of shrews from the genus Neomys. Acta Soc. Zool. Boh.-Slov. 1957, 21, 44–64.
- 55. Prokopic, J. Vysledky helmintologickeho vyzkumu nasich jezku. Vest. Ceskosl. Zool. Spol. 1957, 21, 97–111.
- 56. Prokopic, J. The parasitic helminths of insectivora in CSR. Cesk. Parasitol. 1959, 6, 87–134.
- 57. Meszaros, F.; Stollmann, A. Results of complex parasitological investigations on small mammals in the Cergov Mountains (Western Carpathians, Czechoslovakia). *Misc. Zool. Hung.* **1984**, 2, 7–11.
- 58. Murai, E.; Meszaros, F. Helminths from small mammals in the Cergov Mountains (Western Carpatians, Czecoslovakia). *Misc. Zool. Hung.* **1984**, *2*, 17–28.
- 59. Slapeta, J.R. Endoparasites of hedgehogs in the Czech Republic. Veterinarstvi 1999, 49, 248–252.
- 60. Mas-Coma, S. Metastrongylides parasites des Soricides d'Europe. Ann. Parasitol. 1977, 52, 447–456. [CrossRef]
- 61. Cordero del Campillo, M.; Castanon Ordonez, L.; Reguera Feo, A. *Index-Catalogue of Iberian Parasites of Animals*, 2nd ed.; Universidad de Leon, Secretariado de Publicaciones: Leon, Spain, 1994; pp. 3–650.
- 62. Fuentes, M.V.; Cerezuela, A.M.; Galan-Puchades, M.T. A helmithological survey of small mammals (insectivores and rodents) in the Serra Calderona mountains (Valencian Community, Spain). *Res. Rev. Parasitol.* **2000**, *60*, 25–35.
- 63. Mas-Coma, S.; Esteban, J.G.; Fuentes, M.V.; Bargues, M.D.; Valero, M.A.; Galan-Puchades, M.T. Helminth parasites of small mammals (insectivores and rodents) on the Pityusic Island of Eivissa (Balearic Archipelago). *Res. Rev. Parasitol.* **2000**, *60*, 41–49.
- 64. Feliu, C.; Blasco, S.; Torres, J.; Miquel, J.; Casanova, J.C. On the helminthfauna of *Erinaceus europaeus* Linnaeus, 1758 (Insectivora, Erinaceidae) in the Iberian Peninsula. *Res. Rev. Parasitol.* **2001**, *61*, 31–37.
- 65. Portoles, E.; Granel, P.; Esteban, J.G.; Cabaret, J. Helminth associations in white-toothed shrews *Crocidura russula* (Insectivora: Soricidae) from the Albufera Natural Park, Spain. *J. Parasitol.* **2004**, *90*, 572–578. [CrossRef] [PubMed]
- 66. Ribas, A.; Casanova, J.C. Helminth parasites of *Talpa europaea* (Insectivora, Talpidae) in southwestern Europe. *Acta Parasitol.* **2005**, 50, 161–167.
- 67. Galan-Puchades, M.T.; Mas-Coma, S.; Valero, M.A.; Fuentes, M.V. First data on the helminth community of the smallest living mammal on Earth, the Etruscan pygmy shrew, *Suncus etruscus* (Savi, 1822) (Eulipotyphla: Soricidae). *Animals* **2021**, *11*, 2074. [CrossRef] [PubMed]
- 68. Bernard, L. Quelques especes d'helminthes de micromammiferes recoltes en France et en Espagne. Vie Milieu 1961, 2, 125–149.
- 69. Euset, L.; Jourdane, J. Helminthes parasites des micromammiferes des Pyrenees-Orientales. I, cestodes de *Neomys fodiens* (Schreber). *Bull. Soc. Neuchatel. Sci. Nat.* **1968**, *91*, 31–42.
- 70. Jourdane, J. Deux nouvelles especes de Trematodes parasites des Soricinae dans les Pyrenees. *Ann. Parasitol.* **1973**, *58*, 667–676. [CrossRef]
- 71. Quentin, J.C. Nematodes parasites de musareignes en Sologne. Symbioses 1986, 18, 265–279.
- 72. Joyeux, C.; Baer, J.G. Quelques helminthes nouveaux et peu connus de la musaraigne, *Crocidura russula* Herm. (Premiere partie, Trematodes et Cestodes). *Rev. Suiss. Zool.* **1936**, 43, 25–50.
- 73. Joyeux, C.; Baer, J.G. Quelques helminthes nouveaux et peu connus de la musaraigne, *Crocidura russula* Herm. (Deuxieme partie, Nematodes et Acanthocephales). *Rev. Suiss. Zool.* **1937**, *44*, 27–40.
- 74. Vaucher, C.; Hunkeler, P. Contribution to the study of cestodes and trematodes parasites of micromammals in Switzerland I. *Bull. Neuchatel Soc. Nat. Sci.* **1967**, *90*, 161–184.
- 75. Liesegang, A.; Lehmann, M.C. Haufigkeit von krankheit und abgangsursachen bei igeln. *Schweiz. Arch. Tierheil.* **2003**, 145, 589–591. [CrossRef] [PubMed]

- 76. Edelenyi, B. Saugwürmer in einheimischen Kleinsäugern II. Ann. Hist.-Nat. Mus. Nat. Hung. 1965, 57, 217–222.
- 77. Murai, E.; Meszaros, F.; Sey, O. On parasitic helminths of mammals living in the environs of Lake Balaton. *Parasitol. Hung.* **1992**, 25, 23–36.
- 78. Žąsityte, R.; Grikieniene, J. Some data on endoparasites of common mole in Lithuania. *Acta Zool. Lituan.* **2002**, *12*, 403–409. [CrossRef]
- 79. Žąsityte, R. Cestodes of shrews (Insectivora, Soricidae) from Lithuania. Ekologija 2001, 3, 19–22.
- 80. Binkiene, R. Helminth fauna of shrews (Sorex spp.) in Lithuania. Acta Zool. Lituan. 2006, 16, 241–245. [CrossRef]
- 81. Haukisalmi, V. Intestinal helminth communities of Sorex shrews in Finland. Ann. Zool. Fenn. 1989, 26, 401–409.
- 82. Haukisalmi, V.; Henttonen, H. Distribution patterns and microhabitat segregation in gastrointestinal helminths of *Sorex* shrews. *Oecologia* **1994**, *97*, 236–242. [CrossRef]
- 83. Haukisalmi, V. Checklist of tapeworms (Platyhelminthes, Cestoda) of vertebrates in Finland. *ZooKeys* **2015**, *533*, 1–61. [CrossRef] [PubMed]
- 84. Rautio, A.; Isomursu, M.; Valtonen, A.; Hirvela-Koski, V.; Kunnasranta, M. Mortality, diseases and diet of European hedgehogs (*Erinaceus europaeus*) in an urban environment in Finland. *Mammal Res.* **2016**, *61*, 161–169. [CrossRef]
- 85. Bychovskaya-Pavlovskaya, I.E.; Vysotzkaya, S.O.; Kulakova, A.P. Trematodes of small mammals from Transkarpathian region. *Parazitologiia* **1970**, *4*, 25–33.
- 86. Melnichenko, E.D.; Panasenko, N.A. To the helminth fauna of some insectivorous mammals of the Middle Dnieper region. *Vestn. Zool.* **1979**, *5*, 79–81.
- 87. Iskova, N.I.; Sharpilo, V.P.; Sharpilo, L.D.; Tkach, V.V. Catalogue of Helminths of Vertebrates in the Ukraine. Trematodes of Terrestrial Vertebrates; Naukova Dumka: Kiev, Ukraine, 1995; pp. 3–93.
- 88. Vysotskaya, S.O. Helminths of small mammals from Transcarpathian region (The East Carpathians, Ukraine). *Parazitologiia* **1997**, 31, 346–353.
- 89. Andreyko, O.F.; Spassky, A.A. Nematodes of insectivorous mammals from Prut-Dniester interfluve. *Par. Anim. Plant.* **1969**, 4, 44–59.
- 90. Andreyko, O.F. Trematodes of insectivores (Insectivora) in Moldavia. Par. Anim. Plant. 1970, 5, 3–17.
- 91. Andreyko, O.F.; Spassky, A.A. Cestodes of insectivorous mammals in Moldavia. Par. Anim. Plant. 1970, 5, 44-59.
- 92. Andreyko, O.F. Overview of helminth fauna of insectivores (Insectivora) of the USSR and adjacent countries. *Par. Anim. Plant.* **1973**, *9*, 3–34.
- 93. Andreyko, O.F. Parasites of Mammals in Moldavia; Știința: Chisinau, Moldova, 1973; pp. 3–185.
- 94. Lewis, J.W. Studies on the helminth parasites of voles and shrews from Wales. J. Zool. 1968, 154, 313–331. [CrossRef]
- 95. Lewis, J.W. Helminth parasites of British rodents and insectivores. Mammal Rev. 1987, 17, 81–93. [CrossRef]
- 96. Boag, B.; Fowler, P. The prevalence of helminth parasites from the hedgehog *Erinaceus europaeus* in Great Britain. *J. Zool.* **1988**, 215, 379–382. [CrossRef]
- 97. Bunnell, T. The incidence of disease and injury in displaced wild hedgehogs (Erinaceus europaeus). Lutra 2001, 44, 3–14.
- 98. Gaglio, G.; Allen, S.; Bowden, L.; Bryant, M.; Morgan, E.R. Parasites of European hedgehogs (*Erinaceus europaeus*) in Britain: Epidemiological study and coprological test evaluation. *Eur. J. Wildl. Res.* **2010**, *56*, 839–844. [CrossRef]
- 99. Haigh, A.; O'Keeffe, J.; O'Riordan, R.M.; Butler, F. A preliminary investigation into the endoparasite load of the European hedgehog (*Erinaceus europaeus*) in Ireland. *Mammalia* **2013**, 78, 103–107. [CrossRef]
- 100. Bernard, J. Liste des nematodes parasites des micromarnmiferes de la faune Beige. *Ann. Parasitol. Hum. Comp.* **1961**, *6*, 775–784. [CrossRef]
- 101. Rasmussen, S.L.; Hallig, J.; van Wijk, R.E.; Petersen, H.H. An investigation of endoparasites and the determinants of parasite infection in European hedgehogs (*Erinaceus europaeus*) from Denmark. *Int. J. Parasitol. Par. Wildl.* **2021**, *16*, 217–227. [CrossRef] [PubMed]
- 102. Casanova, J.C.; Miquel, J.; Fons, R.; Molina, X.; Feliu, C.; Mathias, M.L.; Torres, J.; Libois, R.; Santos-Reis, M.; Collares-Pereira, M.; et al. On the helminthfauna of wild mammals (Rodentia, Insectivora and Lagomorpha) in Azores archipelago (Portugal). *Vie Milieu/Life Environ.* **1996**, *46*, 253–259.
- 103. Fuentes, M.V.; Saez, S.; Trellis, M.; Cruz, J.; Sarmento, P.; Casanova, J.C.; Torres, J.; Feliu, C.; Esteban, J.G. Helmintofauna of small mammals (Insectivora, Rodentia) collected in the Serra da Malcata (Portugal). *Rev. Iber. Parasitol.* **2003**, *63*, 89–92.
- 104. Milazzo, C.; Casanova, J.C.; Aloise, G.; Ribas, A.; Cagnin, M. The helminth community of *Talpa romana* (Thomas, 1902) (Insectivora, Talpidae) in southern Italy. *Parasitol. Res.* **2002**, *88*, 979–983.
- 105. Poglayen, G.; Giannetto, S.; Scala, A.; Garippa, G.; Capelli, G.; Scaravelli, D.; Brianti, E.; Reeve, N.J. Helminths found in hedgehogs (*Erinaceus europaeus*) in three areas of Italy. *Vet. Rec.* **2003**, 152, 22–24. [CrossRef] [PubMed]
- 106. Mariacher, A.; Santini, A.; Del Lesto, I.; Tonon, S.; Cardini, E.; Barone, A.; Eleni, C.; Fichi, G.; Perrucci, S. Endoparasite infections of the European hedgehog (*Erinaceus europaeus*) in Central Italy. *Animals* **2021**, *11*, 3171. [CrossRef] [PubMed]
- 107. Prokopic, J.; Mahnert, V. Über Helminthen der Kleinseuger (Insectivora, Rodentia) Tirols (Osterreichs). *Ber. Naturwiss.-Med. Ver. Innsbr.* 1970, *58*, 143–154.
- 108. Hoerweg, C. The helminth fauna of the common shrew, *Sorex araneus* L., from two sites near Vienna, Austria. *Bull. Scandinav. Soc. Parasitol.* **2000**, *10*, 112.

Diversity 2024, 16, 307 22 of 24

109. Liatis, T.K.; Monastiridis, A.A.; Birlis, P.; Prousali, S.; Diakou, A. Endoparasites of wild mammals sheltered in wildlife hospitals and rehabilitation centres in Greece. *Front. Vet. Sci.* **2017**, *4*, 220. [CrossRef]

- 110. Pavlovic, I.; Savic, B. Helminth fauna of the northern white-breasted hedgehog (*Erinaceus roumanicus*) in Serbia. *J. Parasit. Dis.* **2017**, *41*, 605–606. [CrossRef]
- 111. Chiriak, E.; Hamar, M. Contributions a la connaissance des helminthes des petits mammiferes (Rongeurs, Insectivores) de la Roumanie. *Acta Parasitol. Polon.* **1966**, *14*, 61–72.
- 112. Ribas, A.; Casanova, J.C. Helminth fauna of Talpa spp. in the Palaearctic Realm. J. Helminthol. 2006, 80, 1-6. [CrossRef]
- 113. Nikonorova, I.A.; Binkiene, R.; Bugmyrin, S.V.; Ieshko, E.P. Helminth fauna of the common shrew *Sorex araneus* L. in the European part of the species range. *Parazitologiia* **2018**, 52, 41–69.
- 114. Haukisalmi, V.; Hardman, L.M.; Foronda, P.; Feliu, C.; Laakkonen, J.; Niemimaa, J.; Lehtonen, J.T.; Henttonen, H. Systematic relationships of hymenolepidid cestodes of rodents and shrews inferred from sequences of 28S ribosomal RNA. *Zool. Scr.* **2010**, 39, 631–641. [CrossRef]
- 115. Binkiene, R.; Kontrimavichus, V.; Hoberg, E.P. Overview of the cestode fauna of European shrews of the genus *Sorex* with comments on the fauna in *Neomys* and *Crocidura* and an exploration of historical processes in post-glacial Europe. *Helminthologia* **2011**, 48, 207–228. [CrossRef]
- 116. Binkiene, R.; Kornienko, S.A.; Tkach, V.V. *Soricinia genovi* n. sp. from *Neomys fodiens* in Bulgaria, with redescription of *Soricinia globosa* (Baer, 1931) (Cyclophyllidea: Hymenolepididae). *Parasitol. Res.* **2015**, *114*, 209–218. [CrossRef] [PubMed]
- 117. Kornienko, S.A.; Binkiene, R. Redescription and systematic position of *Soricinia tripartita* Żarnowski, 1955 (Cestoda: Cyclophyllidea), a cestode species parasitic in shrews of the genus *Sorex*, including erection of *Gulyaevilepis* gen. n. *Fol. Parasitol.* **2014**, *61*, 141–147. [CrossRef]
- 118. Neov, B.; Vasileva, G.P.; Radoslavov, G.; Hristov, P.; Littlewood, D.T.J.; Georgiev, B.B. Phylogeny of hymenolepidid cestodes (Cestoda: Cyclophyllidea) from mammalian hosts based on partial 28S rDNA, with focus on parasites from shrews. *Parasitol. Res.* **2019**, *118*, 73–88. [CrossRef] [PubMed]
- 119. Binkiene, R. Specific and non-specific cestodes of shrews (Eulipotyphla: Soricidae) in Europe with a description of *Coronacanthus longicirrosus* n. sp (Cestoda: Hymenolepididae). *Syst. Parasitol.* **2021**, *98*, 321–332. [CrossRef]
- 120. Yushkov, V.F. Helminth fauna of mammals (insectivores, predators, lagomorphs and rodents) of the Komi ASSR. *Tr. Gel'mintologicheskoi Lab.* **1971**, 22, 232–248.
- 121. Yushkov, V.Y. *Helminths of Mammals. Fauna of the European North-East of Russia*; Nauka: St. Petersburg, Russia, 1995; Volume 3, pp. 3–202.
- 122. Novikov, M.V. The trematodes and cestodes of *Sorex araneus* L. in Valaam island (Ladoga Lake, USSR). *Mem. Inst. Oswaldo Cruz* **1992**, 87, 155–160. [CrossRef]
- 123. Anikanova, V.S.; Ieshko, E.P.; Bugmyrin, S.V.; Borodina, K.A. Peculiarities of biotopic distribution of cestodes from the common shrew *Sorex araneus* in southern Karelia. *Parazitologiia* **2003**, *37*, 479–487. [PubMed]
- 124. Anikanova, V.S.; Ieshko, E.P.; Boyko, N.S. Helminths of shrews of genus *Sorex* (Soricidae, Insectivora) from the Kandalaksha Reserve. *Parazitologiia* **2005**, *39*, 559–568.
- 125. Anikanova, V.S.; Bugmyrin, S.V.; Ieshko, E.P. *Methods of the Collection and Studies of Helminths of Small Mammals*; Karelian Scientific Center of RAS: Petrozavodsk, Russia, 2007; pp. 3–145.
- 126. Anikanova, V.S.; Ieshko, E.P.; Bugmyrin, S.V. Dynamics of the helminth fauna in the common shrew (*Sorex araneus* L.) from cut-over lands of different age in Karelia. *Parazitologiia* **2009**, *43*, 79–89.
- 127. Nikanorova, I.A.; Kutenkov, A.P.; Bugmyrin, S.V.; Ieshko, E.P. Helminths of the common shrew *Sorex araneus* in the Kivach Nature Reserve. *Proc. Karel. Sci. Cent. RAS* **2019**, *1*, 106–111. [CrossRef]
- 128. Kirillova, N.Y. Helminth fauna of insectivorous mammals (Insectivora) of Samarskaya Luka. *Proc. Sam. Sci. Cent. RAS* **2004**, *6*, 334–340.
- 129. Kirillova, N.Y.; Kirillov, A.A. Helminth fauna of the common shrew *Sorex araneus* L. (Soricidae) from Samarskaya Luka. *Parazitologiia* **2007**, 41, 392–398. [PubMed]
- 130. Kirillova, N.Y. *Helminths in Small Mammals from the Samarskaya Luka (Fauna and Ecology)*; Lambert Academic Publishing: Saarbrucken, Germany, 2011; pp. 3–251.
- 131. Kirillova, N.Y.; Kirillov, A.A. Overview of helminths in small mammals in the Zhiguli State Reserve. *Nat. Conserv. Res.* **2017**, 2, 24–37. [CrossRef]
- 132. Kirillov, A.A.; Kirillova, N.Y.; Krasnobaev, Y.P.; Vekhnik, V.P. *Parasitic Worms of Small Mammals of Zhiguli State Reserve. Flora and Fauna of Reserves*; Committee of RAS for the Conservation of Biological Diversity: Moscow, Russia, 2017; Volume 128, 77p.
- 133. Troitskaya, A.A. Helminth fauna of wild fur animals in the Tatarskaya ASSR. Proc. Soc. Natur. 1960, 63, 335–358.
- 134. Lubarskaya, O.D. On the nematode fauna of Neomys fodiens. Zool. Zhurn. 1962, 41, 833-838.
- 135. Dubinin, E.B. Parasite fauna of myomorph rodents and its changes in the Volga delta. Parasitol. Misc. 1953, 15, 252–301.
- 136. Ivanov, V.M. Monitoring, Structural Changes and Ecological Features of the Trematode Fauna of Vertebrates in the Volga Delta and North of the Caspian Sea. Ph.D. Thesis, Institute for Parasitology of RAS, Moscow, Russia, 2003; 323p.
- 137. Petrov, A.M.; Savinov, V.A. Helminth fauna of moles in the Vologda region. Misc. Vologda Res. Vet. Stat. 1956, 3, 107-115.
- 138. Romashov, B.V. Capillariids (Nematoda, Capillariidae) from the bladder of shrews: *Capillaria incrassata* and *C. konstantini* sp. n. Zool. Zhurn. **1999**, 78, 929–938.

139. Romashova, N.B. Retrospective analysis of the helminth fauna of mammals in the Khopersky Nature Reserve. In *The State of Specially Protected Areas in the European Part of Russia*; Voronezh State University: Voronezh, Russia, 2005; pp. 418–423.

- 140. Vlasov, E.A. First data on the helminth fauna of the shrews (Eulipotyphila, Soricidae) from Central Chernozem State Nature Reserve. In *Contemporary Problems of Zoology and Parasitology: Proceedings of the VII International Scientific Conference Voronezh, Russia, 10 April 2015*; VSU Publishing House: Voronezh, Russia, 2015; pp. 32–38.
- 141. Vlasov, E.A. Helminths of Wild Mammals of the Central Chernozem Reserve (Fauna, Ecology, Pathogenic Significance). Ph.D. Thesis, All-Russian Scientific Research Institute for Fundamental and Applied Parasitology of Animals and Plants, Moscow, Russia, 2017; 166p.
- 142. Vasilyev, V.V. Parasites fauna of rodents and insectivorous from vicinity of Leningrad. *Proc. Leningrad Univ. Biol. Ser.* **1949**, 101, 73–80.
- 143. Fedoseev, S.D. To the nematode fauna in mammals from the Gorky region. Proc. Gorky St. Ped. Inst. 1937, 1, 121–133.
- 144. Morosov, F.N. Helminth fauna of moles in Gorky region. Proc. Gorky St. Ped. Inst. 1957, 19, 27–29.
- 145. Kostyunin, V.M. Materials for the study of the trematode fauna of the common shrew in the Nizhny Novgorod region. In *Zoological Studies of Regions of Russia and Adjacent Territories*; Zryanin, V.A., Ed.; Novosibirsk State Pedagogical University: Novosibirsk, Russia, 2002; pp. 124–125.
- 146. Petrov, A.M.; Savinov, V.A. To the helminth fauna in moles in the Kalinin region. *Proc. All-Union Inst. Helminthol.* **1959**, *6*, 160–166.
- 147. Shaldybin, L.S. Helminth fauna of mammals in Mordovia State Nature Reserve. Proc. Mord. St. Nat. Res. 1964, 2, 135–180.
- 148. Ruchin, A.B.; Kirillov, A.A.; Chikhlyaev, I.V.; Kirillova, N.Y. *Parasitic Worms of Land Vertebrates of the Mordovia Nature Reserve. Flora and Fauna of Reserves*; Committee of RAS for the Conservation of Biological Diversity: Moscow, Russia, 2016; Volume 124, pp. 3–72.
- 149. Kirillova, N.Y.; Ruchin, A.B.; Kirillov, A.A.; Chikhlyaev, I.V.; Alpeev, M.A. Overview of helminths in land vertebrates from the Mordovia Nature Reserve, European Russia. *Nat. Environ. Pollut. Technol.* **2023**, 22, 1667–1690. [CrossRef]
- 150. Artaev, O.N.; Ruchin, A.B.; Bugaev, K.E.; Grishutkin, G.F.; Potapov, S.K.; Spiridonov, S.N. *Vertebrates of the Mordovia State Nature Reserve. Flora and Fauna of Reserves*; Committee of RAS for the Conservation of Biological Diversity: Moscow, Russia, 2012; Volume 120, pp. 3–64.
- 151. Grishutkin, O.G. Patterns of the distribution of swamps depending on the absolute marks of the relief on the territory of the Mordovia State Nature Reserve. *Proc. Mord. St. Nat. Res.* **2013**, *11*, 259–263.
- 152. Artaev, O.N.; Grishutkin, O.G. Lakes of Mordovia. Mord. Res. 2014, 6, 20-23.
- 153. Artaev, O.N.; Smirnov, D.G. Bats (Chiroptera; Mammalia) of Mordovia: Specific structure and features of distribution. *Nat. Conserv. Reserve* **2016**, *1*, 38–51. [CrossRef]
- 154. Grishutkin, G.F.; Lapshin, A.S.; Spiridonov, S.N.; Artaev, O.N.; Ruchin, A.B.; Kuznetsov, V.A.; Andreychev, A.V. *Vertebrate Animals of National Park "Smolny"*. Flora and Fauna of Reserves; Committee of RAS for the Conservation of Biological Diversity: Moscow, Russia, 2013; pp. 3–56.
- 155. Kirillova, N.Y.; Kryštufek, B.; Kirillov, A.A.; Ruchin, A.B.; Grishutkin, G.F. The first record of Microtus subterraneus (de Sélys-Longchamps, 1836) (Rodentia, Cricetidae) for Mordovia, Russia. *Acta Biol. Sib.* **2019**, *5*, 145–149. [CrossRef]
- 156. Smirnov, D.G.; Kirillova, N.Y.; Kirillov, A.A.; Ruchin, A.B.; Vekhnik, V.A. New records of *Nyctalus leisleri* (Kuhl, 1817) and *Myotis nattereri* (Kuhl, 1817) (Mammalia: Chiroptera: Vespertilionidae) from National Park "Smolny" and its surroundings, Republic of Mordovia. *J. Threat. Taxa* 2022, 14, 21553–21560. [CrossRef]
- 157. Kirillova, N.Y.; Kirillov, A.A.; Ruchin, A.B.; Alpeev, M.A.; Smirnov, D.G.; Vekhnik, V.P. Occurrence of Small Mammals in Mordovia State Nature Reserve and National Park "Smolny" (European Russia), Version 1.1. Occurrence Dataset. Joint Directorate of the Mordovia State Nature Reserve and National Park "Smolny": Copenhagen, Denmark, 2024. [CrossRef]
- 158. Ivashkin, V.M.; Kontrimavichus, V.L.; Nasarova, N.S. *Methods of the Collection and Studies of Helminths of Land Mammals*; Nauka: Moscow, Russia, 1971; pp. 3–123.
- 159. Zander, R.H. Four water-soluble mounting media for microslides. *Phytoneuron* **2014**, 32, 1–4.
- 160. Bakanov, A.I. *Quantitated Estimation of Dominance in Ecological Communities*; All-Union Institute of Scientific and Technical Information (VINITI): Borok, Russia, 1987; 64p, No. 8593–B87.
- 161. Hammer, O.; Harper, D.A.T.; Ryan, P.D. PAST: Paleontological statistics software package for education and data analysis. *Palaeontol. Electron.* **2001**, *4*, 9. Available online: http://palaeo-electronica.org/2001\_1/past/issue1\_01.htm (accessed on 24 December 2023).
- 162. Artyukh, E.S. Helminth fauna of beneficial and harmful wild mammals (rodents, insectivores and bats) if the Middle Trans-Volga region. *Proc. Kuyb. Agricult. Inst.* **1950**, *10*, 31–39.
- 163. Kirillova, N.Y.; Kirillov, A.A.; Vekhnik, V.A.; Ruchin, A.B. Trematodes of Small Mammals (Soricomorpha, Erinaceomorpha, Chiroptera, Rodentia) in the Middle-Volga Region (European Russia). Occurrence Dataset. Version 1.6. 2023. Available online: <a href="https://www.gbif.org/dataset/25dee5e7-4e48-49a3-987f-a6799c9ed568">https://www.gbif.org/dataset/25dee5e7-4e48-49a3-987f-a6799c9ed568</a> (accessed on 4 March 2024).
- 164. Sobolev, A.A.; Mashkov, V.V.; Mashkov, N.V. New trematode Skrjabinomerus desmanae. Proc. Gorky St. Ped. Inst. 1939, 4, 83-84.
- 165. Troitskaya, A.A. To the study of the helminth fauna of wild animals of the Middle Volga region and the Bashkirskaya ASSR. *Proc. All-Union Res. Inst. Anim. Raw Mat. Fur.* **1967**, 21, 266–277.
- 166. Sobolev, A.A.; Mashkov, V.V.; Mashkov, N.V. New trematode species from Russian desman. *Proc. Gorky St. Ped. Inst.* **1940**, *5*, 57–60.

Diversity 2024, 16, 307 24 of 24

- 167. Romashov, B.V. Life cycle of nematoda Capillaria erinacei (Capillariidae). Helminthologia 1981, 17, 181-189.
- 168. Majeed, S.; Morris, P.; Cooper, J. Occurrence of the lungworms *Capillaria* and *Crenosoma* spp. in British hedgehogs (*Erinaceus europaeus*). *J. Comp. Pathol.* 1989, 100, 27–36. [CrossRef] [PubMed]
- 169. Mozgovoy, A.A. To the study of nematodes of the genus *Porrocaecum* Railliet et Henry, 1912. *Proc. Helminthol. Lab. USSR* **1949**, 2, 42–49.
- 170. Spassky, A.A. On the question of the development cycle of anisakids (Ascaridata, Anisakidae). *Proc. Helminthol. Lab. USSR* **1952**, 6, 72–73.
- 171. Karmanova, E.M. *Dioctophymidea in Animals and Humans and the Diseases they Cause. Essentials of Nematodology*; Nauka: Moscow, Russia, 1968; Volume 20, pp. 3–262.
- 172. Ryzhikov, K.M.; Gvozdev, E.V.; Tokobaev, M.M.; Shaldybin, L.C.; Matsaberidze, G.V.; Merkusheva, I.V.; Nadtochiy, E.V.; Khokhlova, I.G.; Sharpilo, L.D. *Keys to the Helminths of Rodents in the USSR Fauna, Nematodes and Acanthocephalans*; Nauka: Moscow, Russia, 1979; pp. 3–272.
- 173. Ryzhikov, K.M. On the question of paratenic parasitism of the pig nematode *Physocephalus sexalatus* (Molin, 1860). *Proc. Helminthol. Lab. USSR* **1952**, *6*, 139–141.
- 174. Chabaud, A.G. Sur le cycle evolutif des spirurides et de nematodes ayant une biologie comparable. *Ann. Parasitol. Hum. Comp.* **1954**, *29*, 206–249. [CrossRef] [PubMed]
- 175. Grewal, P.S.; Grewal, S.K.; Tan, L.; Adams, B.J. Parasitism of molluscs by nematodes: Types of associations and evolutionary trends. *J. Nematol.* **2003**, *35*, 146–156. [PubMed]
- 176. Barus, V.K.; Blazek, K. The life cycle and the pathogenicity of the nematode *Crenosoma striatum* (Zeder, 1800). Fol. Parasitol. 1971, 18, 215–226.
- 177. Jourdane, J. Recherches sur le cycle biologique de *Pseudoleucochloridium soricis* (Soltys, 1952) dans les Pyrenees. *Ann. Parasitol.* **1976**, *50*, 421–432. [CrossRef]
- 178. Macy, R.W.; Moore, D.J. The life cycle of *Opisthioglyphe locellus* Kossack, 1910 with a redescription of the species. *Transact. Amer. Microscop. Soc.* **1958**, 77, 396–403. [CrossRef]
- 179. Bock, D. The life cycle of *Opisthioglyphe locellus* Kossack, 1910 (Trematoda: Plagiorchiidae) a parasite of shrews (Soricidae). *Zeit. Parasitenk.* **1982**, *67*, 155–163. [CrossRef]
- 180. Sharpilo, V.P.; Iskova, N.I. Trematodes. Plagiorchiata. Fauna of Ukraine; Naukova Dumka: Kiev, Ukraine, 1989; pp. 3–280.
- 181. Busta, J.; Nasincova, V. Developmental cycle of *Rubenstrema exasperatum* (Rudolphi, 1819) (Trematoda: Omphalometridae). *Fol. Parasitol.* **1991**, *38*, 209–215.
- 182. Nasincova, V.; Busta, J.; Krasnolobova, T.A. Contribution to the developmental cycle and taxonomy of *Neoglyphe sobolevi* Schaldybin 1953 (Trematoda: Omphalometridae). *Fol. Parasitol.* **1989**, *36*, 313–319.
- 183. Faltynkova, A.; Nasinkova, V.; Kablaskova, L. Larval trematodes (Digenea) of the great pond snail, *Lymnaea stagnalis* (L.), (Gastropoda, Pulmonata) in Central Europe: A survey of species and key to their identification. *Parasite* **2007**, *14*, 39–51. [CrossRef]
- 184. Kirillova, N.Y.; Kirillov, A.A. Trematodes (Trematoda) of small mammals in the Middle Volga region. *Parazitologiia* **2009**, 43, 225–239. [PubMed]
- 185. Radev, V.; Kanev, I.; Khrusanov, D.; Fried, B. Reexamination of the life cycle of *Isthmiophora melis* (Trematoda: Echinostomatidae) on material from southeast Europe. *Parazitologiia* **2009**, 43, 445–453. [PubMed]
- 186. Sudarikov, V.E.; Shigin, A.A.; Kurochkin, Y.V.; Lomakin, V.V.; Sten'ko, R.P.; Yurlova, N.I. *Metacercariae of Trematodes—Parasites of Freshwater Aquatic Organisms in Central Russia*; Nauka: Moscow, Russia, 2002; pp. 3–298.
- 187. Procopic, J. The life cycle of the cestode Rodentolepis erinacei (Gmelin, 1789). Fol. Parasitol. 1971, 18, 27–32.
- 188. Joyeux, C. Recherches sur le cycle evolutif d'Hymenolepis erinacei (Gmelin, 1789). Annales Par. Hum Comp. 1927, 5, 20–26. [CrossRef]
- 189. Ishigenova, L.A.; Kornienko, S.A. Development of cysticercoids in *Monocercus arionis* (Cestoda, Dilepididae). *Zool. Zhur.* **2013**, 92, 1–6.
- 190. Kabilov, T.K. Some data on the larval forms of helminths of vertebrate animals among beetles of pasture biotopes of the Fergana region. In *Ecology and Biology of Animals of Uzbekistan*; Gan, E.I., Ed.; Fan: Tashkent, Uzbekistan, 1969; pp. 65–68.
- 191. Khokhlova, I.G. Acanthocephals of Terrestrial Vertebrates of the USSR Fauna; Nauka: Moscow, Russia, 1986; pp. 3–277.
- 192. Sharpilo, V.P. Parasitic Worms of Reptiles of Fauna of the USSR; Naukova Dumka: Kiev, Ukraine, 1976; pp. 3–287.
- 193. Lewin, J. Parasitic worms in a slowworm (*Anguis fragilis* L.) population from the Bieszczady Mountains (Poland). *Acta Parasitol. Polon.* **1990**, 35, 207–215.
- 194. Kornienko, S.A.; Dokuchaev, N.E.; Odnokurtsev, V.A. Cestodes of shrews in Yakutia. In *Proceedings of the Center for Parasitology*; Center for Parasitology of the Institute of Ecology and Evolution: Moscow, Russia, 2018; Volume 50, pp. 104–106.
- 195. Sheykina, Z.V.; Zhigileva, O.N. Helminthes of shrews (Soricidae) in the Malaya Sosva Nature Reserve (Russia). *Nat. Conserv. Res.* **2018**, *3*, 28–36. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.