



# Article A Contribution to Knowledge of *Russula* Section *Ingratae* (Russulales, Russulaceae) in China: Two New Taxa and Amended Descriptions of One Known Species

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**Abstract:** *Russula*, one of the main genera of Russulaceae (Russulales), is both ecologically and economically important. It is a large and complex genus including eight subgenera. Although many studies have focused on *Russula* in China, the diversity still remains incompletely understood. In the present study, several collections of *Russula* subgen. *Heterophyllidiae* sect. *Ingratae* from China were studied based on morphology and molecular phylogenetic analyses. Three species were recognized, two of them described as new, *R. fujianensis* and *R. oraria*, and one of them already a known taxon, *R. rufobasalis*. Detailed descriptions, color photos of fresh basidiomata, and line-drawings of microstructures of the three species are presented.

Keywords: molecular phylogeny; morphology; new taxon; taxonomy



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# 1. Introduction

*Russula* Pers. (Russulaceae, Russulales) is a highly diverse group with about 2000 species worldwide. Nearly all species of the genus are typical ectomycorrhizal fungi, which play an important role in maintaining biodiversity in forest ecosystems [1–3]. Due to the importance of ecological value, extensive studies about *Russula* have been performed. Recently, the genus has been divided into eight subgenera: *Archaeae* Buyck and V. Hofst., *Brevipedum* Buyck and V. Hofst., *Compactae* (Fr.) Bon, *Crassotunicatae* Buyck and V. Hofst., *Glutinosae* Buyck and X.H. Wang, *Heterophyllidiae* Romagn., *Malodorae* Buyck and V. Hofst., and *Russula* Pers [4,5]. Among them, subgen. *Heterophyllidiae*, especially sect. *Ingratae* (Quél.) has received much attention. The section is characterized by a pileus with yellow to brown tints, a margin with obvious radial tuberculate striate, a presence of distinctly fetid or acrid odour, and basidiospores inamyloid to partly amyloid in the suprahilar area [6–9]. In China, species diversity of sect. *Ingratae* is abundant, with 14 taxa confirmed to distribute across the country [9–16]. Moreover, the poisonousness of the section has attracted great interest in the country. For example, collections identified as *R. senecis* S. Imai were reported to be poisonous, which can cause gastroenteritis [17–19].

Although many species of sect. *Ingratae* have been described in China [9–16], with more field investigations, more novel species of the section will be uncovered. During field investigations in subtropical and tropical China, several *Russula* collections were made. These specimens were examined using morphology and molecular phylogenetic analyses, and were eventually confirmed to represent two novel species and one previously known species of sect. *Ingratae*. Thus, the three taxa were described/redescribed herein, aiming to contribute to new taxonomic knowledge of sect. *Ingratae*.

# 2. Materials and Methods

# 2.1. Morphological Studies

Fresh basidiomata were photographed in the field in daylight and their macroscopic characteristics were measured and recorded. Specimens were dried at 50-60 °C, then deposited in the Fungal Herbarium of Hainan Medical University (FHMU) (Index Herbariorum), Haikou City, Hainan Province, China. Color codes referred to Kornerup and Wanscher [20]. Templates of descriptions and terminologies of micromorphological characters were obtained from Adamčík et al. [21]. The pileipellis section taken from the pileus between the center and margin, and the stipitipellis from the middle part along the longitudinal axis of the stipe, were also observed [22]. Spore ornamentation density estimated from scanning electron microscopy pictures followed Adamčík and Marhold [23]. The hymenial cystidia density estimates referred to Buyck [24]. The pileipellis ortho- or metachromatic reactions were examined in Cresyl Blue after Buyck [25]. Sulfovanillin (SV) was used to observe color changes of cystidia contents [26]. Observations and measurements of microscopic features were made in 1% Congo Red, five percent potassium hydroxide (KOH), or Melzer's reagent. Basidiospore measurements exclude ornamentation and apiculus. The basidiospores were examined using a TM4000Plus scanning electron microscope (SEM). All the microscopic structures were drawn by free-hand. The number of measured basidiospores is given as n/m/p, where "n" represents the total number of basidiospores measured from "m" basidiomata of "p" specimens. Dimensions of basidiospores are presented as (a-)b-e-c(-d), where the "b-c" represents a minimum of 90% of the measured values (5th to 95th percentile), and extreme values (a and d), whenever present (a < 5th percentile, d > 95th percentile), are in parentheses, "e" refers to the average length/width of basidiospores. "Q" refers to the length/width ratio of basidiospores; "Qm" refers to the average "Q" of basidiospores and is given with standard deviation.

#### 2.2. Molecular Procedures

Plant Genomic DNA Kit (CWBIO, Beijing, China) was used to extracted total genomic DNA from silica gel-dried collections according to the manufacturer's instructions. Primer pairs used for amplification were: nuc 28S rDNA D1-D2 domains (28S) with LR0R/LR5 [27,28], nuc rDNA region including the internal transcribed spacers 1 and 2, along with the 5.8S rDNA (ITS) with ITS5/ITS4 [29]. PCR reactions were performed with 4 min initial denaturation at 95 °C, followed by 35 cycles of denaturation at 94 °C for 30 s, annealing at the appropriate temperature (50 °C for 28S and ITS) for 30 s, extension at 72 °C for 120 s and a final extension at 72 °C for 7 min. Amplified PCR products were purified using the DNA Purification Kit (TIANGEN, Beijing, China) according to the manufacturer's instructions, and then directly sequenced using a BigDye terminator v3.1 Kit and an ABI 3730xl DNA Analyzer (Guangzhou Branch of BGI, Guangzhou, China) with the same primers used for PCR amplification. DNA sequences were compiled with BioEdit v7.0.9 [30], and then deposited in GenBank (Table 1).

**Table 1.** Taxa, vouchers, locations, and GenBank accession numbers of DNA sequences used in this study. The new sequences are in bold.

Taxon	Voucher	Locality	GenBank Accession Nos.		D (
			ITS	285	Reference
Russula aff. amoenolens	HMJAU37318	Heilongjiang, NE China	KY357332	_	[31]
R. ahmadii	LAH 35004	Pakistan	KT834638	KX911834	[32]
R. ahmadii	LAH 18081013	Pakistan	KU535609	_	[32]
R. amerorecondita	F (PGA17-017)	USA	MN130066	_	[21]
R. amerorecondita	SH120828	USA	MN130067	_	[21]
R. amoenolens	46372 (AH)	Spain	MK105626	MK105707	[33]
R. amoenolens	TUB nl27.9.95.6	Germany	AF418615	AF325295	[34]

Taxon

GenBank Accession Nos.

Tawar	Very alson	Locality		
Taxon	Voucher	Locality -	ITS	28S
R. arunii	CUH:AM270	India	KY450661	KY946733
R. arunii	CUH:AM261	India	KR872619	KY946732
к. игинн	(holotype)	intena	101/2017	R1740752
R. blennia	569/BB08.066	Madagascar	MH545687	KU237556
	(holotype)			
R. benghalensis	CUH AM654	India	MK414583	MK496844
R. catillus	(holotype) SFC20120725-52	Popublic of Voros	KX574685	
K. cututus	SFC20120725-32 SFC20120827-01	Republic of Korea	KA374003	
R. catillus	(holotype)	Republic of Korea	KX574686	_
R. catillus	SFC20120919-35	Republic of Korea	KX574688	_
R. catillus	LHJ150915-19	Guangdong, southern China	MK860690	_
R. cerolens	F36	USA	JN681168	
R. cf. pseudolepida	BPL247	USA	KT933962	KT933821
R. clavulus	RITF3300	Guizhou, SW China	MN317302	MN325079
R. clavulus	RITF3216	Guizhou, SW China	MN317303	MN325080
R. clavulus	RITF3204	Yunnan, SW China	MN317304	MN325081
R. clavulus	RITF3334	Yunnan, SW China	MN317301	MN325078
	(holotype)			
R. compacta	BPL242	USA	KT933960	KT933819
<i>R. compacta</i>	HFJAU0301	Jiangxi, SE China	MN258682	
R. echidna	HO 593336	Australia	MN130079	
R. echidna	HO 593337	Australia	MN130080	
R. elastica	3/BB 06.009	Madagascar	—	KU237451
R. ferruginea subsp. ferruginea	HUA Corrales935	Colombia	MZ604289	MZ604284
R. ferruginea subsp.			N/7/04001	N7(0400)
ferruginea	HUA Corrales1019	Colombia	MZ604291	MZ604286
<i>R. ferruginea</i> subsp.	HUA Corrales944	Colombia	MZ604288	MZ604283
ferruginea	(holotype)	Colonibia	WIZ004200	1012.004203
<i>R. ferruginea</i> subsp.	UCH A28	Panama	MZ604292	MZ604287
panamensis	(holotype)	i ununu	112001292	1012001207
R. flavobrunnea var.	71//BB 06.050	Madagascar		KU237468
violaceotincta				
R. fluvialis	KUO (JR8666)	Finland	MN130084	MN130125
R. fluvialis	(holotype) KUO (JR8313)	Finland	MN130085	MN130126
R. foetens	HMJAU 32234	Heilongjiang, NE China	KX095018	
R. foetens	TUB hue124	Germany	AF418613	AF325299
R. foetens	GENT FH-12-277	Germany	KT934016	KT933877
R. foetentoides	LAH 04081023	Pakistan	HE647707	
R. foetentoides	LAH 13081034	Pakistan	HE647708	_
R. foetentula	156	USA	KJ834623	_
R. fragrantissima	98	Italy	KJ530751	_
2 0	N.K. Zeng1449	,	•	ON1010201
R. fujianensis	(ELIN (LI002)	Fujian, SE China	ON332048	ON318391

Fujian, SE China

USA

USA

Guangdong, southern China

Guangdong, southern China

ON332049

MN130088

MN130089

MH168575

MH168574

ON318392

\_\_\_\_

\_\_\_\_

\_\_\_\_

Locality

# Table 1. Cont.

Voucher

R. fujianensis

(FHMU993) N.K. Zeng1431

(FHMU975)

(holotype)

F (PGA17-008)

F (PGA1-910)

K15052626

(holotype)

K16053119

# R. fujianensis

R. garyensis R. garyensis R. gelatinosa R. gelatinosa Reference

[35] [35]

[36]

[37] [38] [38] [38] Unpublished Unpublished

[39] [15]

[15] [15] [15]

[39] Unpublished [21] [21] [40]

[40]

[40]

[40]

[40]

[40]

[21] [21]

Unpublished [34] [39] [41] [41] [42] [42]

This study

This study

[21]

[21]

[12]

[12]

Taxon	Voucher	Locality	GenBank Accession Nos.		
			ITS	28S	- Reference
R. granulata	PC BB2004-226	USA	EU598192	_	Unpublished
R. granulata	PC BB2004-225	USA	EU598190	_	Unpublished
R. granulata	HMAS252604	Jilin, NE China	KF850414	_	[43]
R. granulata	BPL272	USA	KT933971	KT933832	[39]
R. grata	HMJAU 38008	Heilongjiang, NE China	KY681444	_	[31]
R. griseobrunnea	JAC13201	New Zealand	MW683824	MW683661	Unpublished
e e	N.K. Zeng4888				-
R. hainanensis	(FHMU4860) N.K. Zeng4899	Hainan, southern China	—	OM280996	[16]
R. hainanensis	(FHMU4855) (holotype)	Hainan, southern China	OM280999	OM280997	[16]
R. heterophylla	UE20.08.2004-2 IB 1997/0787	Sweden	DQ422006	_	[44]
R. hortensis	(holotype)	Italy	HG798528	—	Unpublished
R. illota	MICH73719	France	KF245509	—	Unpublished
R. illota	UE26.07.2002-3	Sweden	DQ422024	—	[34]
R. inamoena	107	Italy	KJ834597		[42]
R. inamoena	109	Italy	KJ834595	_	[42]
R. indocatillus	HGAS-MF009917	Guizhou, SW China	MN649191	_	[9]
R. indocatillus	HGAS-MF009903 AG 17-1571	Guizhou, SW China	MN649192	—	[9]
R. indocatillus	(holotype)	India	MN581483	—	[45]
R. indocatillus	AG 18-1653	India	MN581165	—	[45]
R. indosenecis	AG-21-04A	India	OL701254	—	[46]
R. indosenecis	AG-21-06A (holotype)	India	OL701269	—	[46]
R. insignis	HMAS267740	Heilongjiang, NE China	KF850404		[9]
R. laurocerasi	FH12178	Germany	KT933988	KT933849	[39]
R. laurocerasi	TUB nl1348	Germany	AF418614	_	[34]
R. laurocerasi	E00290534	UK	KF245532	_	Unpublished
R. livescens	F0177	China	GU371295	_	[47]
R. maguanensis	XHW4765	Yunnan, SW China	MH724918	MH714537	[48]
R. mistiformis	JC170305NR	Spain	MK105677	_	[33]
R. mistiformis	AMC H-69	Spain	MK105680		[33]
R. multilamellula	LHJ14092423	Guizhou, SW China	MT791347	MT793120	[15]
	(holotype)				
R. multilamellula	wuxingliang03 1176/S. Adamcik	Guizhou, SW China	MT791348	MT793121	[15]
R. mustelina	09.88	Slovakia	—	KU237596	[40]
R. mutabilis	DPL10654	USA	KF810137		[49]
R. mutabilis	BHI-F384a KD 16-30	USA	MF161239	—	[50]
R. obscuricolor	(holotype)	India	MF805816	—	[51]
R. obscuricolor	KD 16-22	India	MF805817	_	[51]
R. oleifera	254/BB 98.024	Burundi	_	KU237490	[40]
R. ombrophila	86	Spain	KF971694	_	[42]
R. oraria	N.K. Zeng4829 (FHMU5374)	Hainan, southern China	ON332052	OQ380695	This study
R. oraria	N.K. Zeng4823 (FHMU5355) (holotype)	Hainan, southern China	ON332053	OQ380696	This study
R noctinata	2010BT02	Cormony	KF318081	_	[42]
R. pectinata R. pectinata	2010BT02 2010BT48	Germany Germany	KF318081 KF318082	_	[42]
		Yunnan, SW China		_	[42]
R. pectinatoides	HMAS251202 NYS2303.1	USA	JX425405	_	[43]
R. pectinatoides	JMV800654 (BCN)		KU640189 MK105691		[42] [33]
R. pila	JIVI V 000034 (DUN)	Spain	IVIN100091	WIX103730	[33]

# Table 1. Cont.

# Table 1. Cont.

Taxon	Voucher	Locality -	GenBank Accession Nos.		
			ITS	28S	Reference
R. pilosella	BRI-H5974	Australia	EU019941		[52]
R. pseudocatillus	K16042406 (holotype)	Guangdong, southern China	MK049974	_	[14]
R. pseudocatillus	K15060706	Guangdong, southern China	MK049975	_	[14]
R. pseudociliata	545/BB 08.061	Madagascar	MH545688	KU237537	[40]
R. pseudopectinatoides	HMAS265020	Xizang, SW China	KM269079		[11]
	HMAS251523	Ũ			
R. pseudopectinatoides	(holotype)	Xizang, SW China	KM269077	—	[11]
R. pseudopectinatoides	RITF1353 AG 20-062	China	MK860689	_	[13]
R. pseudosenecis	(holotype)	India	OL461233	—	[46]
R. pseudosenecis	AG 21-073	India	OL461234	—	[46]
R. pulverulenta	4-1144IS79	USA	AY061736	—	[53]
R. pulverulenta	PC BB2004-245	USA	EU598186		Unpublished
R. punctipes	K17052318	Guangdong, southern China	MH168576	_	[12]
R. punctipes	K16051001	Guangdong, southern China	MH168577	_	[12]
R. recondita	92	Switzerland	KJ530750	—	[42]
R. rufobasalis	H17052204 (holotype)	Guangdong, southern China	MH168570	_	[12]
R. rufobasalis	H15060622	Guangdong, southern China	MH168567	_	[12]
R. rufobasalis	N.K. Zeng1416 (FHMU961)	Fujian, SE China	ON332050	ON318393	This study
R. rufobasalis	N.K. Zeng1270 (FHMU827)	Fujian, SE China	ON332051	ON318394	This study
R. senecis	CUH AM102	India	KP142981	_	[54]
R. senecis	SFC20110921-18	Republic of Korea	KX574698	_	[38]
R. siamensis	Watling 28784	Thailand	AB206535	_	[55]
R. straminella	HGAS-MF009920	Guizhou, SW China	MN649194		[9]
Destauration 11 -	HGAS-MF009922				
R. straminella	(holotype)	Guizhou, SW China	MN649195	_	[9]
R. straminella	HGAS-MF009925	Guizhou, SW China	MN649189	—	[9]
R. subfoetens	HMJAU 38006	Heilongjiang, NE China	KY681430	—	[31]
R. subpectinatoides	HBAU15023 (holotype)	Jiangsu, eastern China	MW041163	_	[9]
R. subpectinatoides	HBAU15024	Jiangsu, eastern China	MW041164	_	[9]
R. subpectinatoides	HBAU15025	Jiangsu, eastern China	MW041164 MW041165	—	[9]
R. subpectinatoides	HBAU15026	Jiangsu, eastern China	MW041165	_	[9]
,	RITF2616				
R. subpunctipes	(holotype)	Hubei, central China	MK860692	MK860695	[13]
R. subpunctipes	RITF2617	Hubei, central China	MK860693	MK860696	[13]
R. substriata	WJ292	Yunnan, SW China	MH724919	MH714538	[48]
R. substriata	XHW4749	Yunnan, SW China	MH724920	MH714539	[48]
R. substriata	XHW4766 (holotype)	Yunnan, SW China	MH724921	MH714540	[48]
R. substriata	XHW4767	Yunnan, SW China	MH724922	MH714541	[48]
R. substriata R. substriata	XHW4785	Yunnan, SW China	MH724923	MH714542	[48]
R. succinea	HGAS-MF 009909	Guizhou, SW China	MN649196		[9]
R. succinea	HGAS-MF 009904	Guizhou, SW China	MN649188	_	[9]
	(holotype)				
R. succinea	HGAS-MF 009906	Guizhou, SW China	MN649198	_	[9]
R. succinea	HGAS-MF 009915	Guizhou, SW China	MN649190		[9]
R. variata	BPL241	USA	KT933959	KT933818	[39]
R. ventricosipes	PC 0142480	USA	KY800364	—	[56]
R. vinaceocuticulata	PDD 64246	New Zealand	GU222258		Unpublished
<i>Russula</i> sp.	LHJ170913-01	Guangdong, southern China	MK860691	MK860694	[13]

SW = Southwestern, SE = Southeastern, NE = Northeastern.

## 2.3. Dataset Assembly

Twelve DNA sequences (6 of 28S, and 6 of ITS) from 6 collections were newly generated. Edited sequences were submitted to GenBank; the GenBank accession numbers of 28S and ITS are provided in Table 1, which are identified in boldface. For the concatenated dataset, newly generated 28S and ITS sequences were aligned with sequences of sect. *Ingratae* with greater representativeness as well as availability from previous studies and GenBank (Table 1). *Russula blennia* Buyck, *R. compacta* Frost and *R. griseobrunnea* McNabb were chosen as outgroup. MUSCLE was used to align the 28S and ITS sequences individually to test for phylogenetic conflict [57]. Then, the sequences of the different genes were concatenated using Phyutility v2.2 for further analyses [58].

#### 2.4. Phylogenetic Analyses

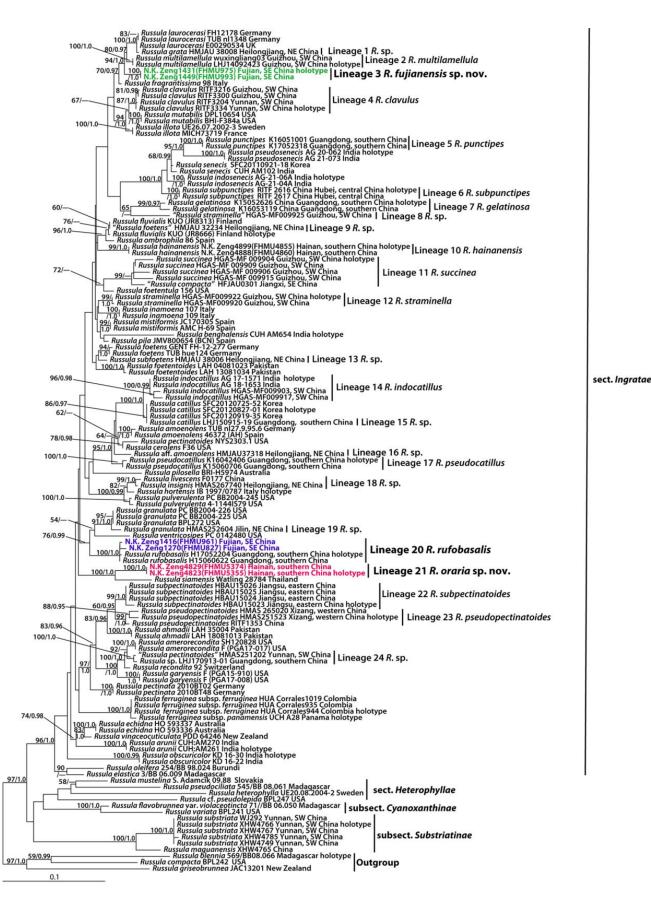
Both Maximum Likelihood (ML) and Bayesian Inference (BI) were employed for phylogenetic analysis. Maximum Likelihood of the phylogenetic relationships was conducted with the program RAxML 7.2.6 running 1000 replicates combined with an ML search [59]. Bayesian analysis with MrBayes 3.1 implementing the Markov Chain Monte Carlo (MCMC) technique and parameters predetermined with MrModeltest 2.3 was performed [60,61]. The best-fit likelihood models for 28S and ITS were SYM + I + G and HKY + I + G, respectively. Bayesian analysis was repeated for 17 million generations, and sampled every 100. Trees sampled from the first 25% of the generations were discarded as burn-in, and Bayesian posterior probabilities (PP) were then calculated for a majority consensus tree of the retained Bayesian trees. Runs were terminated once the average standard deviation of split frequencies decreased below 0.01.

## 3. Results

#### 3.1. Molecular Data

Sixty-three species of sect. *Ingratae*, four species of sect. *Heterophyllae*, two species of subsect. *Cyanoxanthinae*, and two species of subsect. *Substriatinae* were used for the construction of the phylogram of *Russula* sect. *Ingratae* (Figure 1). The topologies of the phylogenetic trees based on a single gene (28S and ITS) were identical, suggesting that the phylogenetic signals present in the different gene fragments (28S and ITS) were not in conflict. The two-locus data set (28S + ITS) consisted of 134 taxa and 1532 nucleotide sites. The topologies of phylogenetic trees generated from ML and BI analyses were identical, though statistical support for some branches showed slight differences. The phylogram with branch lengths inferred from the 28S and ITS dataset with RAxML is shown with the support values (Figure 1).

The current molecular data indicate that the Chinese specimens of sect. *Ingratae* were grouped into 24 independent lineages (1-24) (Figure 1). We were particularly interested in lineages 3, 20, and 21, which included our new collections. Lineage 3 had strong statistical support (BS = 100, PP = 1.0), and included two specimens (FHMU975 and FHMU993) from southeastern China; lineage 20 consisted of the holotype (H17052204) and one specimen (H15060622) of *R. rufobasalis* from southern China, and two collections (FHMU827 and FHMU961) from southeastern China, with high statistical support (BS = 100, PP = 1); lineage 21, which had strong statistical support (BS = 100, PP = 1) comprising two specimens (FHMU5355 and FHMU5374) from southern China (Figure 1).



**Figure 1.** A phylogram of *Russula* sect. *Ingratae* inferred from a two-locus (rDNA 28S and ITS) dataset using RAxML. BS  $\geq$  50% and PP  $\geq$  0.95 are indicated above or below the branches as RAxML BS/PP. SW = Southwestern, SE = Southeastern, NE = Northeastern. Newly generated sequences are in color.

# 3.2. Taxonomy

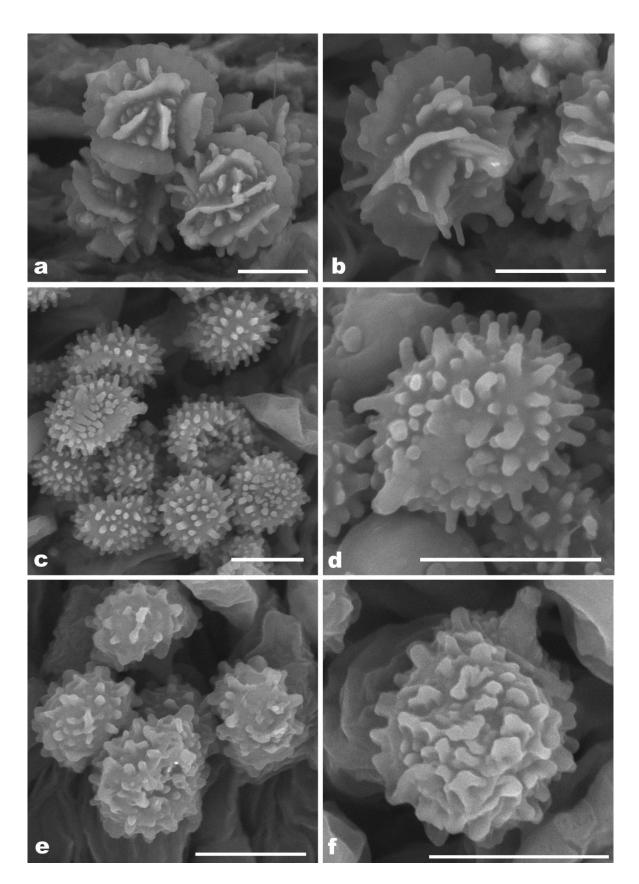
*Russula fujianensis* N.K. Zeng, Y.X. Han & Zhi Q. Liang, sp. nov. (Figures 2a,b, 3a,b, 4 and 5).

MycoBank: MB 845634.

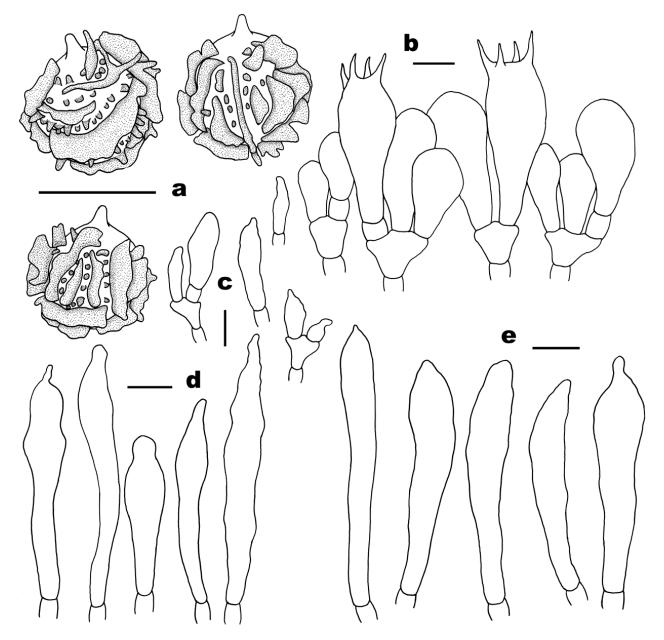
**Etymology**—Latin "*fujianensis*", refers to Fujian Province, China, holotype locality. **Diagnosis**—Differs from closest species of sect. *Ingratae* by a pale yellowish-brown pileus, a long, white stipe (ca. 6.5–12 cm), basidiospores usually with large wings (up to 2.0 μm), never forming a reticulum, a hymenophore with fewer pleurocystidia (less than 700/mm<sup>2</sup>), cystidia turning to yellowish brown in SV, and it is associated with trees of Fagaceae.



**Figure 2.** Basidiomata of *Russula* sect. *Ingratae* species. (**a**,**b**) *R. fujianensis* (**a**) from FHMU975, holotype; (**b**) from FHMU993); (**c**,**d**) *R. oraria* (**c**) from FHMU5355, holotype; (**d**) from FHMU5374); (**e**,**f**) *R. rufobasalis* (FHMU961). Bars: 1 cm. Photos: N.K. Zeng.



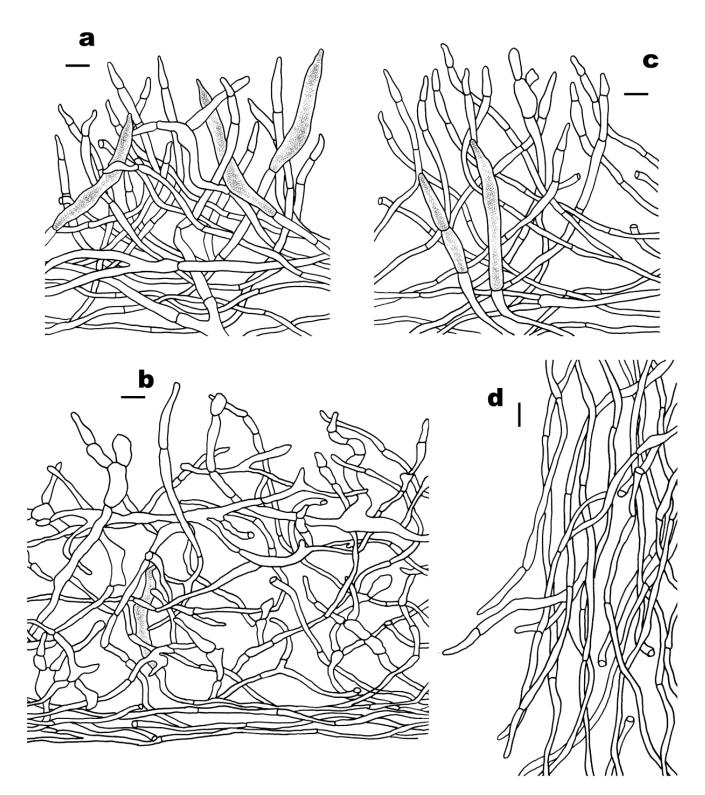
**Figure 3.** Basidiospores of *Russula* sect. *Ingratae* species from herbarium materials under SEM. (**a**,**b**) *R. fujianensis* (FHMU975, holotype); (**c**,**d**) *R. oraria* (FHMU5355, holotype); (**e**,**f**) *R. rufobasalis* (FHMU827). Bars: 5 μm. Photos: Y.X. Han.



**Figure 4.** Microscopic features of *Russula fujianensis* (FHMU975, holotype). (**a**) Basidiospores. (**b**) Basidia and basidiola. (**c**) Marginal cells. (**d**) Pleurocystidia. (**e**) Cheilocystidia. Bars: 10 μm. Drawings by Y.X. Han.

**Holotype**—CHINA. Fujian Province: Zhangping County, Xinqiao Town, Chengkou Village, elev. 350 m, 18 August 2013, *N.K. Zeng1431* (FHMU975). GenBank accession numbers: 28S = ON318392, ITS = ON332049.

**Description**—*Basidiomata* medium-sized. *Pileus* 5–7 cm diameter, spherical at first, then subhemispherical, applanate, center slightly depressed; surface slightly sticky, pale yellowish-brown (1B3), pale brown (5D4) to dark brown (5E5), margin with obvious radial tuberculate-striate; context 4–5 mm thick at the center of the pileus, white (2A1), unchanging in color when injured. *Hymenophore* lamellate, adnate; lamellae about 5 mm in height, crowded, occasionally forked, white (2A1), unchanging in color when injured, edge even; lamellulae rare, concolorous with lamellae. *Stipe* 6.5–12 × 1–1.8 cm, central, subcylindrical to cylindrical, hollow, annulus absent; surface white, sometimes tined with peal brown (5B3) to peal yellowish-brown (1B4). *Taste* acrid. *Odour* fetid. *Spore print* not obtained.



**Figure 5.** Microscopic features of *Russula fujianensis* (FHMU975, holotype). (**a**) Pileipellis at pileus margin. (**b**) Pileipellis at middle part between center and margin of pileus. (**c**) Pileipellis at pileus centre. (**d**) Stipitipellis. Bars: 10 μm. Drawings by Y.X. Han.

*Basidiospores* (excluding ornamentation) [40/6/2] 7–8.1–8.5 (–9) × 6.5–7.6–8 (–8.5) µm, Q = 1-1.14 (–1.15),  $Qm = 1.07 \pm 0.04$ , globose to subglobose, ornamentation composed of large, moderately distant to dense ((5–) 6–8 in a 3 µm diameter circle) amyloid warts or wings, 0.9–2.0 µm high, isolated or occasionally fused (0–3 fusions in the circle); wings nearly encircling the spores, mixed with numerous short, blunt, isolated warts, line connec-

tions absent; suprahilar spot inamyloid and indistinct. *Basidia* (35–) 39.5–45–51.5 (–54)  $\times$ (10–) 14.5–15.5–16 (–17) μm, hyaline in KOH, thin-walled (up to 0.4 μm), clavate, 4-spored; sterigmata 4–7  $\mu$ m, slightly tortuous, sometimes straight; basidiola clavate, ca. 7.5–15  $\mu$ m wide. *Pleurocystidia* dispersed, less than 700/mm<sup>2</sup>, (35–) 46–52–60  $\times$  (6.5–) 7.5–8.5–10  $\mu$ m, narrowly clavate to clavate, apically often obtuse or acute, sometimes fusiform, occasionally with 2.5–4  $\mu$ m long appendage, thin-walled (up to 0.4  $\mu$ m); contents abundant granulose, yellowish in Congo Red, turning to yellowish brown in SV. Cheilocystida (45-) 49.5-52- $62 \times (6-)$  7.5–9.5–11.5 µm, narrowly clavate to subcylindrical, apex obtuse or mucronate, sometimes with 1.5-4 µm long appendage, thin-walled (up to 0.3 µm); contents granulose, yellowish in Congo Red, turning to yellowish brown in SV. Lamellae edges fertile. Marginal *cells* (6–) 10.5–16.5–20.5  $\times$  3.5–4–5 (–6.5)  $\mu$ m, clavate or subcylindrical, usually shorter than basidiola, thin-walled (up to 0.4 µm). Lamellar trama mainly composed of sphaerocytes measuring up to 49  $\mu$ m in diameter, hyaline in KOH, thick-walled (1–1.5  $\mu$ m). *Pileipellis* orthochromatic in Cresyl Blue, not sharply delimited from the underlying context, a trichoderm 120-200 µm thick, two-layered, weakly gelatinized; composed of suprapellis (50–100 µm thick) and subpellis (70–100 µm thick). Suprapellis composed of erect to suberect hyphae 3–7 µm in diameter, thin-walled (up to 0.3 µm). Subpellis composed of horizontally oriented, dense, 2.5–7 µm wide intricate hyphae. Acid-resistant incrustations absent. Hyphal terminations near the pileus margin composed of 1–3 unbranched cells, not flexuous, thin-walled (up to 0.3  $\mu$ m); terminal cells (9–) 12–18–23  $\times$  4–4.5–5  $\mu$ m, mainly clavate, occasionally subcylindrical, apexobtuse, rarely mucronate; subterminal cells often wider and longer, rarely branched. Hyphal terminations on the middle part between center and margin of pileus 1–3 cells, less flexuous and unbranched, terminal cells (13–) 18–21.5–24  $\times$ (4–) 4.5–5–7 μm, mainly clavate, occasionally subcylindrical, apexobtuse, rarely mucronate; subterminal cells often wider and longer, occasionally branched. Hyphal terminations near the pileus centre 1–3 cells, not flexuous; shorter terminal cells 8–14–20  $\times$  3–4–4.5 (–5)  $\mu$ m, subcylindrical, apically obtuse; subterminal cells often wider and longer, rarely branched. *Pileal trama* made up of hyphae up to 41  $\mu$ m in diameter, thick-walled (up to 1  $\mu$ m), hyaline to pale yellowish in KOH. Pileocystidia near the pileus margin always long, one-celled, (55–)  $63-70-89 \times 7-8-8.5 \mu$ m, cylindrical to clavate, apex occasionally obtuse or usually mucronate, contents yellow in Congo Red, turning to yellowish brown in SV. Pileocystidia near the pileus centre narrower cylindrical to clavate, 1–2 celled, (26–) 45–57–65  $\times$  5–5.5–6  $\mu$ m, contents granulose, yellow in Congo Red, turning to yellowish brown in SV. Cystidioid hyphae in subpellis and context, contents granulose. Stipitipellis a cutis composed of hyphae slightly thick-walled (up to 0.7 µm), 2–6 µm wide, hyaline in KOH; terminal cells 12–28  $\times$  4–5.5 µm, subcylindrical or subclavate. *Stipe trama* mainly composed of sphaerocytes measuring up to 43 µm in diameter, hyaline to pale yellowish in KOH, thick-walled (1.5–2 μm). *Clamp connections* absent in all tissues.

Habitat—Gregarious or fascicled on the ground in forests dominated by *Castanopsis* kawakamii Hayata.

Known distribution—Eastern China (Fujian Province).

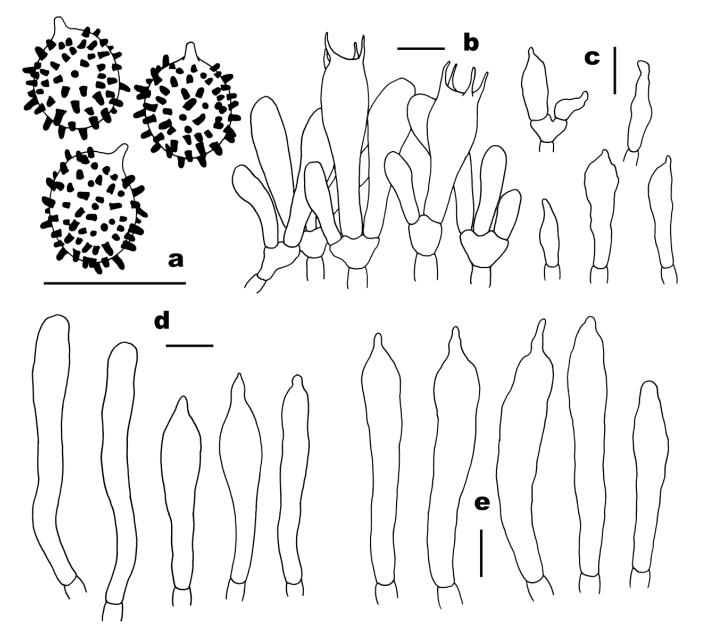
Additional specimen examined—China. Fujian Province: Zhangping County, Xinqiao Town, Chengkou Village, elev. 350 m, 20 August 2013, *N.K. Zeng*1449 (FHMU993).

**Notes**—Morphologically, some Asian members of section *Ingratae*, viz., *R. gelatinosa* Y. Song and L.H. Qiu, *R. punctipes* Singer, and *R. subpunctipes* J. Song share similar winged basidiospores with our new species. However, *R. gelatinosa*, originally described from Guangdong Province of southern China, has higher wings (up to 3.5  $\mu$ m high) on basid-iospores, short, blunt ridges and warts forming an incomplete network between basid-iospores wings, and suprapellis composed of short, inflated to globular subterminal and terminal elements [12]; *R. punctipes*, originally described from Hunan Province of central China, has a smaller basidioma, a shorter stipe (3.5–5.5 cm long), higher wings (up to 3  $\mu$ m high) on basidiospores wings, suprapellis mainly composed of inflated subterminal cells [12]; *R. subpunctipes*, originally described from Hubei Province of central China, has

a pileus cracking into patches towards the margin, higher wings (up to  $2.5 \,\mu$ m high) on basidiospores, and suprapellis mainly composed of inflated subterminal cell [13].

Phylogenetically, *R. fujianensis* is closely related to *R. multilamellula* B. Chen and J. F. Liang (Figure 1). However, *R. multilamellula*, originally described from southwestern China, has a stipe with brownish spot at the base, smaller basidiospores measuring (5.6–) 6.3–6.9–7.4 (–8) × (5–) 5.8–6.4–7 (–7.3)  $\mu$ m, a hymenophore with more abundant pleurocystidia (ca. 1000/mm<sup>2</sup>), and it is associated with trees of Pinaceae [15].

Russula oraria N.K. Zeng, Y.X. Han and Zhi Q. Liang, sp. nov. (Figures 2c,d, 3c,d, 6 and 7).



**Figure 6.** Microscopic features of *Russula oraria* (FHMU5355, holotype). (a) Basidiospores. (b) Basidia and basidiola. (c) Marginal cells. (d) Pleurocystidia. (e) Cheilocystidia. Bars: 10 μm. Drawings by Y.X. Han.

MycoBank: MB 845636.

Etymology—Latin "oraria", refers to the new species growing near the coast.

**Diagnosis**—Differs from closest species of *R*. sect. *Ingratae* by a white pileus with pale yellow to pale yellowish brown at center, basidiospores with small, dense, cylindric

to subcylindrical warts (0.5–1  $\mu$ m high), never forming a reticulum, cystidia turning to yellowish brown in SV, and it is associated with trees of Dipterocarpaceae.

**Holotype**—China. Hainan Province: Wanning County, Shimei Bay, elev. 5 m, 28 August 2020, *N.K. Zeng*4823 (FHMU5355). GenBank accession numbers: ITS = ON332053.

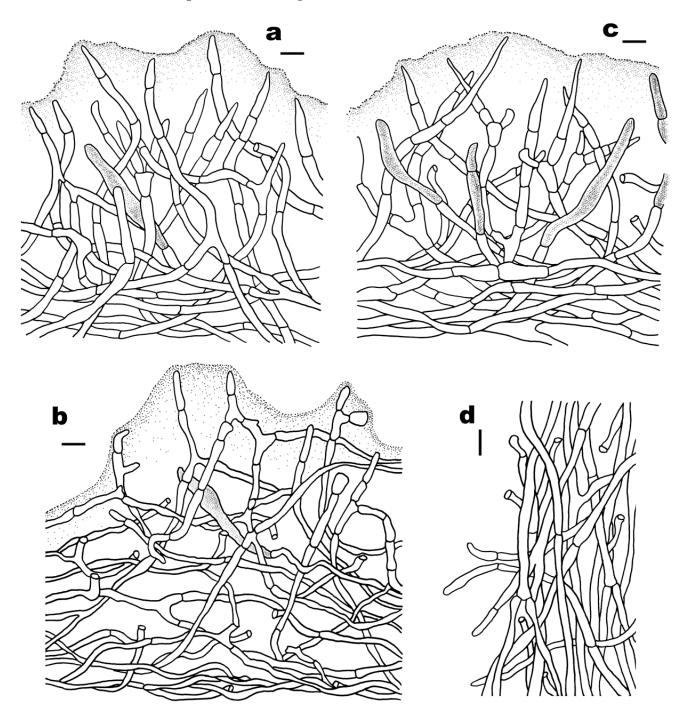


Figure 7. Microscopic features of *Russula oraria* (FHMU5355, holotype). (a) Pileipellis at pileus margin.
(b) Pileipellis at middle part between center and margin of pileus. (c) Pileipellis at pileus centre.
(d) Stipitipellis. Bars: 10 μm. Drawings by Y.X. Han.

**Description**—*Basidiomata* small-sized. *Pileus* 3.5–4 cm diameter, broadly convex to applanate, center slightly depressed, margin occasionally cracked; surface dry, white (2A1), pale yellow (3B2) to pale yellowish-brown (1B4) at center, margin with obvious radial tuberculate-striate; context about 2 mm thick at the center of the pileus, white (3A1),

unchanging in color when injured. *Hymenophore* lamellate, adnate; lamellae 4–5 mm in height, occasionally forked, white (3A1), unchanging in color when injured; lamellulae common, concolor with lamellae. *Stipe* 40–45 × 6–8 mm, central, subcylindric to cylindric, hollow, annulus absent; surface dry, white (3A1). *Taste* acrid. *Odour* fetid. *Spore print* not obtained.

Basidiospores (excluding ornamentation) [40/4/2] 6–6.6–7 (–7.5)  $\times$  5–5.4–6  $\mu$ m, Q = 1.08-1.3 (-1.4),  $Qm = 1.24 \pm 0.11$ , subglobose to broadly ellipsoid, ornamentation of relatively small, dense to very dense (9-14 in a 3 µm diameter circle) amyloid cylindric to subcylindrical warts, 0.5–1 µm high, isolated or rarely fused (0–2 fusions in the circle), line connections absent, never forming a reticulum; suprahilar spot inamyloid and indistinct. Basidia (28–) 29.5–32–36  $\times$  7–9–10 (–11.5) µm, hyaline in KOH, thin- to slightly thick-walled (0.4–0.5 µm), clavate, 4-spored; sterigmata 4–6 µm, slightly tortuous, sometimes straight; basidiola cylindric, then narrowly clavate, ca. 4.5-9.5 µm wide. Pleurocystidia moderately numerous, ca.  $1000/\text{mm}^2$ , (40.5–) 45–52–60.5 × 6–7–8 (–8.5) µm, narrowly clavate to subcylindrical, apex often obtuse or acute, sometimes moniliform, occasionally with  $1-5 \ \mu m$  long appendage, slightly thick-walled (up to 0.5  $\mu m$ ); contents abundant granulose, yellowish in Congo Red, turning to yellowish-brown in SV. Cheilocystida 40.5–52–55.5 (-61)  $\times$  7–8.5–9 µm, clavate to subcylindrical, apex obtuse or mucronate, sometimes with 3–7 µm long appendage, thin-walled (up to  $0.4 \,\mu$ m); contents abundant granulose, yellowish in Congo Red, turning to yellowish-brown in SV. Lamellae edges fertile. Marginal cells (13-) 14.5–19–25  $\times$  (4–) 4.5–6–8  $\mu$ m, clavate or subcylindrical, usually shorter than basidioles, thin-walled (up to  $0.4 \mu m$ ). Lamellar trama mainly composed of sphaerocytes measuring up to 39  $\mu$ m in diameter, hyaline in KOH, thick-walled (1–1.5  $\mu$ m). *Pileipellis* orthochromatic in Cresyl Blue, not sharply delimited from the underlying context, an ixotrichoderm 75–150  $\mu$ m thick, two-layered, gelatinized; composed of suprapellis (50–100  $\mu$ m thick) and subpellis (35–70 µm thick). Suprapellis composed of erect to suberect hyphae 1.5–7.5 µm in diameter, thin-walled (up to 0.4 µm). Subpellis composed of horizontally oriented, dense, 3–9 µm wide intricate hyphae. Acid-resistant incrustations absent. Hyphal terminations near the pileus margin composed of 1–3 unbranched cells, not flexuous, thin-walled (up to 0.4  $\mu$ m); terminal cells (14–) 15–20–24.5  $\times$  3.5–4–5 (–5.5)  $\mu$ m, narrowly subcylindrical or tapering upward; subterminal cells often wider, occasionally branched. Hyphal terminations on the middle part between center and margin of pileus 1–3 cells, unbranched and not flexuous; terminal cells 10–13–17.5 (–19)  $\times$  (3–) 3.5–4–5  $\mu$ m, subcylindrical, apex obtuse; subterminal cells often wider, occasionally branched. Hyphal terminations near the pileus center 1–3 cells, unbranched and not flexuous; terminal cells (10–) 14.5–17–38 (–40)  $\times$  3–4–5 (–5.5)  $\mu$ m, mainly clavate, occasionally subcylindrical; subterminal cells often shorter, unbranched. *Pileal trama* composed of hyphae up to 50  $\mu$ m in diameter, thick-walled (1–2  $\mu$ m), hyaline in KOH. Pileocystidia near the pileus margin one-celled, (40.5–)  $48-56-70 \times 5.5-7.5-8$  (-8.5)  $\mu$ m, cylindrical to clavate, apex usually obtuse, contents granulose, yellow in Congo Red turning to yellowish-brown in SV. *Pileocystidia near the pileus centre* cylindrical to clavate, 1–2 celled, (12.5–) 20–47–67.5  $\times$  4.5–5–6 (–7)  $\mu$ m, contents granulose, yellow in Congo Red turning to yellowish-brown in SV. Cystidioid hyphae in subpellis and context, contents granulose. Stipitipellis a cutis, composed of hyphae thin-walled (up to 0.4 µm), 3–8 µm wide, pale yellowish in KOH; terminal cells  $10-30 \times 3.5-5 \,\mu$ m, subcylindrical or subclavate. *Stipe trama* mainly composed of sphaerocytes measuring up to 46 µm in diameter, hyaline in KOH, thick-walled (1–1.5  $\mu$ m). *Clamp connections* absent in all tissues.

Habitat—Gregarious on the ground in forests dominated by *Vatica mangachapoi* Blanco. Known distribution—Southern China (Hainan Province).

Additional specimen examined—China. Hainan Province: Wanning County, Shimei Bay, elev. 5 m, 28 August 2020, *N.K. Zeng4829* (FHMU5374).

**Notes**—Morphologically, some Asian members of section *Ingratae*, viz., *R. ahmadii* Jabeen, Razaq, Niazi, I. Ahmad and Khalid, *R. catillus* H. Lee, M.S. Park and Y.W. Lim, *R. clavulus* B. Chen and J.F. Liang, *R. natarajanii* K. Das, J.R. Sharma and Atri, *R. obscuricolor* K. Das, A. Ghosh and Buyck, *R. pseudocatillus* F. Yuan and Y. Song, *R. pseudopectinatoides*  G.J. Li and H.A. Wen, R. straminella G.J. Li and C.Y. Deng, R. subpectinatoides G.J. Li and Q.B. Sun, and R. succinea G.J. Li and C.Y. Deng are similar to R. oraria. However, R. ahmadii has a darker pileus, and larger basidiospores (5.6–) 6.1-9.2 (–9.4)  $\times$  (5.4–) 5.7-6.6 (–7.5)  $\mu$ m with ornamentations composed of lower, irregular warts (up to  $0.3 \mu m$  high), sometimes forming a partial reticulum, and it is associated with trees of Pinaceae [32]; R. catillus has basidiospores with lower ornamentations composed of mostly conical warts (up to  $0.8 \,\mu m$ high), sometimes forming a partial reticulum, and it is associated with trees of Fagaceae [38]; R. clavulus has a larger basidioma (pileus 7–9 cm in diameter), larger basidiospores measuring (5.6–) 7.0–7.8–8.6 (–9.0)  $\times$  (5.4–) 6.5–7.2–8.0 (–8.3) µm with ornamentations composed of larger, cylindric warts (up to 1.6 µm high), often forming short or long ridges, and it is associated with trees of Fagaceae [15]; R. natarajanii has a larger basidioma (pileus 6–10.5 cm in diameter), larger basidiospores measuring  $6.5-8.8 \times 5.8-7.1 \mu m$  with ornamentations composed of minute warts and ridges (up to 0.4 µm high), forming an incomplete reticulum, and it is associated with trees of Fagaceae [62]; R. obscuricolor has a stipe with blood red coloration at stipe base, basidiospores with ornamentations composed of conical warts and ridges (up to 0.9 µm high), forming a partial to incomplete reticulum, and it is associated with trees of Fagaceae [51]; R. pseudocatillus has larger basidiospores measuring 7–7.9–8.6  $(-9.2) \times (5.1-) 5.5-6.1-6.6$  (-6.7) µm with ornamentations composed of higher, conical to cylindrical warts (up to 1.2 μm high), cystidia negative in SV [14]; R. pseudopectinatoides has basidiospores with ornamentations composed of lower, conical warts (0.2–0.5 µm high), forming a partial reticulum, and it is associated with trees of Pinaceae [11]; R. straminella has a darker brown pileus, a stipe turning brownish-yellow when injured, basidiospores with ornamentations composed of conical warts ( $0.7-1 \mu m$  high), forming a partial reticulum, a hymenophore with fewer pleurocystidia (less than 500/mm<sup>2</sup>), and it is associated with trees of Pinaceae; *R. subpectinatoides* has a darker brown pileus, a context slowly turning pale ochre when injured, basidiospores with ornamentations composed of lower, long ridges (0.3–0.5  $\mu$ m high), forming an incomplete to complete reticulum, a hymenophore with fewer pleurocystidia (less than 200/mm<sup>2</sup>), and it is associated with trees of Pinaceae; R. succinea has a stipe turning cream to pale ochre when injured, basidiospores with ornamentations composed of subcylindrical warts, forming an incomplete reticulum, and it is associated with trees of Pinaceae [9].

Phylogenetically, *R. oraria* is closely related to *R. siamensis* Yomyart, Piap., Watling, Whalley and Sihan. (Figure 1), originally described from Thailand. However, the latter has a smaller basidioma (pileus 2.8 cm in diameter), a dark brownish beige to pale ochraceous grey pileus with flakes or squamules, and larger basidiospores measuring  $8-9 \times 6-7 \mu m$  [55].

*Russula rufobasalis* Y. Song and L.H. Qiu, Cryptog. Mycol. 39 (3): 352, 2018 (Figures 2e,f, 3e,f, 8 and 9).

MycoBank: MB 824892.

**Description**—*Basidiomata* small-sized. *Pileus* 3–3.7 cm diameter, subhemispherical at first, then applanate, center slightly depressed; margin occasionally cracked; surface barely sticky, pale yellowish-brown (1B5), brown (5C5) to dark brown (5C8), margin with obvious radial tuberculate-striate; context about 2 mm thick at the center of the pileus, white (1A1), unchanging in color when injured. *Hymenophore* lamellate, adnate; lamellae 2 mm in height, distant, not forked, edge sparsely serrate, white (2A1), unchanging in color when injured; lamellulae rare. *Stipe* 30–35 × 5–6 mm, central, subcylindrical to cylindrical, hollow, annulus absent; surface white (3A1), sometimes tined with pale yellowish-brown (1B5), base reddish. *Taste* mild. *Odour* indistinct. *Spore print* not obtained.

*Basidiospores* (excluding ornamentation) [40/2/2] 5–6–6.5 (–7) × 4.5–5.2–6 µm, Q = 1–1.3, Qm = 1.15 ± 0.09, globose to broadly ellipsoid, ornamentation of relatively small, moderately distant to dense ((6–) 7–8 (–10) in a 3 µm diameter circle)) amyloid warts, 0.4–0.8 µm high, isolated or occasionally fused (0–3 fusions in the circle), small crests and ridges forming an incomplete reticulum, connected by occasional line connections ((0–) 1–3 in the circle); suprahilar spot inamyloid, distinct. *Basidia* (20.5–) 25–35–41 × 5–6–7.5(–8) µm, hyaline in

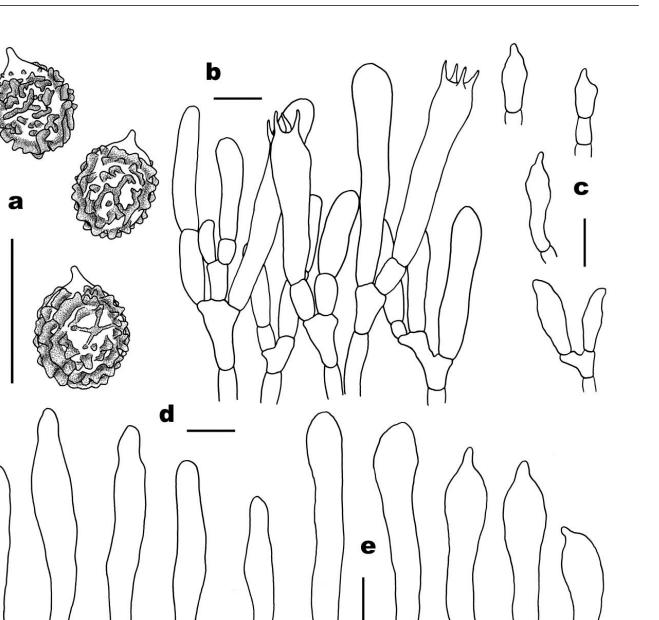
KOH, thin-walled (up to  $0.4 \mu m$ ), narrowly clavate to subcylindrical, 4-spored; sterigmata 3–5 µm, slightly tortuous, sometimes straight; basidiola cylindrical, then narrowly clavate, ca. 4.5–8  $\mu$ m wide. *Pleurocystidia* abundant, ca. 3200/mm<sup>2</sup>, (40–) 45–51.5–57 (–60) × (6–) 6.5–7– $8.5 \,\mu$ m, narrowly clavate to clavate, apically often obtuse, sometimes with mucronate apices, appendage 2.5-4 µm long, slightly thick-walled (up to 0.6 µm); contents abundant granulose, yellowish in Congo Red, turning to brownish in SV. Cheilocystida (32–) 46–50–58.5  $(-70) \times (7-)$  7.5–8–8.5 µm, narrowly clavate to subcylindrical, apically obtuse or mucronate, sometimes with 3–5 µm long appendage, thin-walled (up to 0.4 µm); contents granulose, yellowish in Congo Red, turning brownish in SV. Lamellae edges fertile. Marginal cells (10-) 13–16.5–20  $\times$  4.5–5–6 (–6.5) µm, clavate or subcylindrical, usually shorter than basidioles thin-walled (up to 0.4 µm). Lamellar trama mainly composed of sphaerocytes measuring up to 30  $\mu$ m in diameter, hyaline in KOH, thick-walled (1–1.5  $\mu$ m). *Pileipellis* orthochromatic in Cresyl Blue, not sharply delimited from the underlying context, a trichoderm 200–400 µm thick, two-layered, weakly gelatinized; composed of suprapellis (80–130  $\mu$ m thick) and subpellis (150–300 µm thick). Suprapellis composed of erect to suberect hyphae 2.5–7 µm in diameter, slightly thick-walled ( $0.4-0.5 \mu m$ ). Subpellis composed of frequently undulatesinuous intricate hyphae (2.5–7 µm in diameter). Acid-resistant incrustations absent. Hyphal terminations near the pileus margin composed of 1–3 unbranched cells, not flexuous, slightly thick-walled (0.4–0.5  $\mu$ m); terminal cells (7–) 12–18–26  $\times$  3–4.5–5  $\mu$ m, narrowly subcylindrical or tapering upward; subterminal cells often wider, occasionally branched. Hyphal *terminations on the middle part between center and margin of pileus* 1–3 cells, unbranched and not flexuous; terminal cells (14–)18–20–25  $\times$  2.5–3–3.5  $\mu$ m, subcylindrical; subterminal cells often wider and longer, rarely branched. Hyphal terminations near the pileus centre 1–3 cells, sometimes branched and not flexuous; terminal cells 12–18–24.5 (–27)  $\times$  3–4–5  $(-5.5) \mu m$ , subcylindrical or tapering; subterminal cells often wider and longer, occasionally branched. Pileal trama composed of hyphae up to 35 μm in diameter, thick-walled (up to 1  $\mu$ m), hyaline in KOH. *Pileocystidia near the pileus margin* one-celled, (30–) 45–54–61.5  $\times$ 4–5–6 μm, cylindrical to clavate, apex usually obtuse or occasionally mucronate, contents granulose, yellow in Congo Red, turning to yellowish-brown in SV. Pileocystidia near the *pileus centre* cylindrical to clavate, one-celled, (49–) 53.5–61–80  $\times$  3.5–4–5.5 µm, contents yellow in Congo Red, turning to yellowish-brown in SV. Cystidioid hyphae in subpellis and context, contents granulose. Stipitipellis a cutis composed of hyphae slightly thick-walled (up to  $0.5 \mu m$ ),  $2.5-8 \mu m$  wide, hyaline to pale yellowish in KOH; terminal cells 15–40  $\times$  4–5.5 µm, subcylindrical or subclavate. *Stipe trama* mainly composed of sphaerocytes measuring up to 40  $\mu$ m in diameter, hyaline in KOH, thick-walled (up to 1  $\mu$ m). *Clamp* connections absent in all tissues.

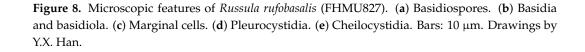
Habitat—Solitary on the ground in forests dominated by *Pinus massoniana* Lamb or *Castanopsis kawakamii* Hayata.

**Known distribution**—Southern China (Guangdong Province) and eastern China (Fujian Province) [12].

**Specimens examined**—China. Fujian Province: Zhangping County, Xinqiao Town, Chengkou Village, elev. 80 m, 17 August 2013, *N.K. Zeng1416* (FHMU961); Zhangping County, Fuzhige Park, elev. 80 m, 24 July 2013, *N.K. Zeng1270* (FHMU827).

**Notes**—*Russula rufobasalis* was originally described from Guangdong Province of southern China [12]. In the present study, it was also found to be distributed in Fujian Province, southeastern China. The species was redescribed according to our new specimens; it is characterized by a pale yellowish-brown, brown to dark brown pileus, lamellate edges sparsely serrate, a stipe with reddish base, basidiospores with small crests and ridges (0.4–0.8  $\mu$ m high), forming an incomplete reticulum, and it is associated with both trees of Pinaceae and Fagaceae. The previous description indicated the basidioma of *R. rufobasalis* is larger (3–6 cm), and the lamellulae are frequent [12], whereas the newly collected specimens have smaller basidiomata (3–3.7 cm) and rare lamellulae.





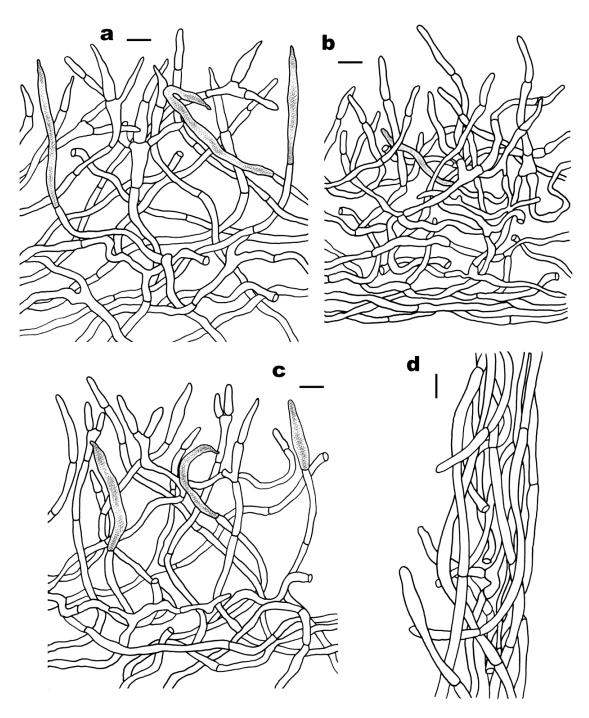


Figure 9. Microscopic features of *Russula rufobasalis* (FHMU827). (a) Pileipellis at pileus centre.
(b) Pileipellis at middle part between center and margin of pileus. (c) Pileipellis at pileus margin.
(d) Stipitipellis. Bars: 10 μm. Drawings by Y.X. Han.

# 4. Discussion

4.1. Species Diversity of Sect. Ingratae in China

High species diversity of sect. *Ingratae* in China was revealed in this study, and 24 lineages of the section were identified (Figure 1). Two lineages (lineages 3 and 21) were described as new species: *R. fujianensis* and *R. oraria*; thirteen lineages (lineages 2, 4, 5, 6, 7, 10, 11, 12, 14, 17, 20, 22 and 23) represent known species: *R. multilamellula*, *R. clavulus*, *R. punctipes* Singer, *R. subpunctipes* J. Song, *R. gelatinosa* Y. Song and L. H. Qiu, *R. hainanensis* N.K. Zeng, Y.X. Han and Zhi Q. Liang, *R. succinea*, *R. straminella*, *R. indocatillus* A. Ghosh, K. Das and R.P. Bhatt, *R. pseudocatillus* F. Yuan and Y. Song, *R. rufobasalis*, *R. subpectinatoides*, and

*R. pseudopectinatoides* [9,11–16]; other lineages (1, 8, 9, 13, 15, 16, 18, 19, and 24) also included Chinese specimens wait to be further identified, combing with morphological evidence. Aside from the aforementioned species/lineages, four additional taxa, viz., *R. foetens* Pers., *R. guangdongensis* Z.S. Bi and T.H. Li, *R. laurocerasi* Melzera, and *R. senecis* have also been described/reported from China [10,63,64]. *Russula guangdongensis*, originally described from southern China, should be examined with DNA sequences and more collections, while the collections identified as *R. foetens*, *R. laurocerasi*, and *R. senecis* in China should be re-evaluated.

# 4.2. Species Delimitation of Sect. Ingratae in China

Although the diagnosis of sect. *Ingratae* is relatively straightforward, species within the section are difficult to distinguish. This is explained by the lack of molecular phylogenetic studies as well as morphological convergence documented in this group in previous studies [65]. With the wide application of molecular phylogenetic analyses, the diversity of species in sect. Ingratae was revealed in China (Figure 1). In order to delimit the phylogenetic species, we should pay much attention to the ornamentations of basidiospores, which are more reliable characters for species of sect. Ingratae. For example, basidiospores of R. clavulus, R. indocatillus, R. multilamellula, R. pseudocatillus, and R. subpunctipes are absent of reticula [9,12,14,15]; those of R. gelatinosa, R. hainanensis, R. pseudopectinatoides, R. punctipes, R. rufobasalis, R. straminella, and R. succinea often form incomplete reticulums; R. subpectinatoides has basidiospores with incomplete to complete reticula [9,11,12,16]. Moreover, five ornamentation types, viz., cone, cylinder, crest, ridge, and wing were observed; for example, basidiospores ornamentations of R. indocatillus and R. straminella mostly composed of conical warts [9], R. subpectinatoides mostly composed of long ridges [9], R. clavulus and R. multilamellula composed of cylindric warts and ridges [15], R. subpunctipes composed of cylindrical warts and wings [13], R. hainanensis composed of crests and ridges [16], R. pseudocatillus composed of conical to cylindrical warts [14], R. pseudopectinatoides composed of conical warts and ridges [11], R. succinea composed of crests and ridges [9], R. gelatinosa and R. punctipes composed of cylindrical warts, ridges and wings [12], R. rufobasalis composed of subcylindrical warts, crests and ridges [12]. In addition, the height of ornamentations is also a useful character: R. gelatinosa, R. punctipes, and R. subpunctipes have basidiospores with high ornamentations (>2 µm) [12,13]; that of *R. clavulus*, *R. fujianensis*, *R. multilamellula*, *R. pseudocatillus*, and *R. succinea* is moderately high  $(1-2 \mu m)$  [9,14,15]; while that of R. hainanensis, R. indocatillus, R. oraria, R. pseudopectinatoides, R. rufobasalis, R. straminella, and *R. subpectinatoides* is low (<1 µm) [9,11,12,16]. As to our new species, viz., *R. fujianensis* and R. oraria are absent of reticula; R. oraria is mostly composed of cylindric warts, while *R. fujianensis* is composed of cylindrical warts and wings.

Besides ornamentations of basidiospores, the ecological preferences should also be noted; for example, *R. clavulus* and *R. subpunctipes* are associated with Fagaceae [13,15], while *R. multilamellula*, *R. pseudopectinatoides*, *R. straminella*, *R. subpectinatoides*, and *R. succinea* are associated with Pinaceae [9,11,15]. Some species, e.g., *R. indocatillus* and *R. rufobasalis* can be associated with both Pinaceae and Fagaceae [9,45]. More particularly, *R. hainanensis* is associated with Dipterocarpaceae [16]. As to our new species, *R. fujianensis* and *R. oraria* are associated with Fagaceae and Dipterocarpaceae, respectively.

#### 4.3. Phylogenetic Relationships and Geographic Divergence of Chinese Sect. Ingratae

Recent phylogenetic studies have provided new perspectives into the phylogeny and geography of sect. *Ingratae* [9,15,16]. Besides revealing two new species of sect. *Ingratae*, our molecular data have also contributed to other knowledge of this group (Figure 1). The molecular data indicated that the affinities of sect. *Ingratae* species between northeastern China and Europe are obvious, two species-level lineages (1 and 9) occuring in the two regions (Figure 1). Similar patterns have been documented in other fungi [66–70]. Moreover, we also noted that some Chinese species are closely related to the North America taxa; for example, one collection identified as *R. granulata* (lineage 19) from northeastern China

is affiliated with one taxon labelled as *R. granulata* from USA; lineage 24 from China is closely related to one species identified as *R. amerorecondita* also from USA (Figure 1). Our study did not identify disjunct populations of the same purported taxon in the two regions (Figure 1). In addition, we also noted that *R. indocatillus* occurs both in China and India (Figure 1).

# 5. Conclusions

In this study, three species of *Russula* sect. *Ingratae* are recognized in China; two of them are described as new: *R. fujianensis* and *R. oraria*, and one of them is an already known taxon: *R. rufobasalis*.

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#### References

- Buyck, B.; Thoen, D.; Watling, R. Ectomycorrhizal fungi of the Guinea-Congo region. Proceedings of the Royal Society of Edinburgh. Sect. B Biol. Sci. 1996, 104, 313–333. [CrossRef]
- 2. Yamashita, S.; Hijii, N. The role of fungal taxa and developmental stage of mushrooms in determining the composition of the mycophagous insect community in a Japanese forest. *Eur. J. Entomol.* 2007, *104*, 225–233. [CrossRef]
- Looney, B.P.; Meidl, P.; Piatek, M.J.; Miettinen, O.; Martin, F.M.; Matheny, P.B.; Labbé, J.L. Russulaceae: A new genomic dataset to study ecosystem function and evolutionary diversifcation of ectomycorrhizal fungi with their tree associates. *New Phytol.* 2018, 218, 54–65. [CrossRef]
- 4. Buyck, B.; Zoller, S.; Hofstetter, V. Walking the thin line ... ten years later: The dilemma of above- versus below-ground features to support phylogenies in the Russulaceae (Basidiomycota). *Fungal Divers.* **2018**, *89*, 267–292. [CrossRef]
- Buyck, B.; Wang, X.H.; Adamcíková, K.; Caboň, M.; Jančovičová, S.; Hofstetter, V.; Adamčík, S. One step closer to unravelling the origin of *Russula*: Subgenus *Glutinosae* subg. nov. *Mycosphere* 2020, *11*, 285–305. [CrossRef]
- 6. Shaffer, R.L. North American russulas of the subsection Foetentinae. Mycologia 1972, 64, 1008–1053. [CrossRef]
- 7. Singer, R. The Agaricales in Modern Taxonomy, 4th ed.; Koeltz Scientific Books: Königstein, Germany, 1986; pp. 1–981.
- 8. Sarnari, M. Monografia Illustrate de Genere Russula in Europa; Fondazione Centro Studi Micologici: Trento, Italy, 1998; pp. 1–800.
- Li, G.J.; Li, S.M.; Buyck, B.; Zhao, S.Y.; Xie, X.J.; Shi, L.Y.; Deng, C.Y.; Meng, Q.F.; Sun, Q.B.; Yan, J.Q.; et al. Three new *Russula* species in sect. *Ingratae* (Russulales, Basidiomycota) from southern China. *MycoKeys* 2021, 84, 103–139. [CrossRef]
- 10. Bi, Z.S.; Li, T.H. A preliminary note on *Russula* species from Guangdong, with a new species and a new variety. *Guihaia* **1986**, *6*, 193–199.
- 11. Li, G.J.; Zhao, D.; Li, S.F.; Wen, H.A. *Russula chiui* and *R. pseudopectinatoides*, two new species from southwestern China supported by morphological and molecular evidence. *Mycol. Prog.* **2015**, *14*, 1–14. [CrossRef]
- 12. Song, Y.; Buyck, B.; Li, J.W.; Yuan, F.; Zhang, Z.W.; Qiu, L.H. Two novel and a forgotten *Russula* species in sect. *Ingratae* (Russulales) from Dinghushan Biosphere Reserve in southern China. *Cryptogamie Mycol.* **2018**, *39*, 341–357. [CrossRef]
- 13. Song, J.; Chen, B.; Liang, J.F.; Li, H.J.; Wang, S.K.; Lu, J.K. Morphology and phylogeny reveal *Russula subpunctipes* sp. nov., from southern China. *Phytotaxa* **2020**, *459*, 16–24. [CrossRef]
- 14. Yuan, F.; Song, Y.; Buyck, B.; Li, J.W.; Qiu, L.H. *Russula viridicinnamomea* F. Yuan & Y. Song, sp. nov. and *R. pseudocatillus* F. Yuan & Y. Song, sp. nov., two new species from southern China. *Cryptogamie Mycol.* **2019**, *40*, 45–56. [CrossRef]
- 15. Chen, B.; Song, J.; Zhang, J.H.; Liang, J.F. Morphology and molecular phylogeny reveal two new species in *Russula* sect. *Ingratae from China. Phytotaxa* **2021**, *525*, 109–123. [CrossRef]

- 16. Han, Y.X.; Liang, Z.Q.; Jiang, S.; Zeng, N.K. *Russula hainanensis* (Russulaceae, Russulales), a new species from tropical China. *Phytotaxa* **2022**, *552*, 035–050. [CrossRef]
- 17. Mao, X.L. Poisonous mushrooms and their toxins in China. Mycosystema 2006, 25, 345–363.
- Chen, Z.H.; Zhang, P.; Zhang, Z.G. Investigation and analysis of 102 mushroom poisoning cases in Southern China from 1994 to 2012. *Fungal Divers.* 2013, 64, 123–131. [CrossRef]
- 19. Tolgor, B.; Bao, H.Y.; Li, Y. A revised checklist of poisonous mushrooms in China. *Mycosystema* **2014**, *33*, 517–548.
- 20. Kornerup, A.; Wanscher, J.H. Taschenlexikon der Farben. 3. Aufl.; Muster-Schmidt Verlag: Göttingen, Germany, 1981; p. 242.
- 21. Adamčík, S.; Looney, B.; Caboň, M.; Jančovičová, S.; Adamčíková, K.; Avis, P.G.; Barajas, M.; Bhatt, R.P.; Corrales, A.; Das, K.; et al. The quest for a globally comprehensible *Russula* language. *Fungal Divers.* **2019**, *99*, 369–449. [CrossRef]
- 22. Han, Y.X.; Liang, Z.Q.; Zeng, N.K. Notes on four species of *Russula* subgenus *Heterophyllidiae* (Russulaceae, Russulales) from southern China. *Front. Microbiol.* 2023, 14, 1140127. [CrossRef]
- Adamčík, S.; Marhold, K. Taxonomy of the Russula xerampelina group. I. Morphometric study of the Russula xerampelina group in Slovakia. Mycotaxon 2000, 76, 463–480.
- 24. Buyck, B. The study of microscopic features in Russula 2. Sterile cells of the hymenium. Russulales News 1991, 1, 62–85.
- Buyck, B. Valeur taxonomique du bleu de crésyl pour le genre Russula. Bull. Soc. Myc. Fr. 1989, 105, 1–6.
- Caboň, M.; Eberhardt, U.; Looney, B.; Hampe, F.; Kolařík, M.; Jančovičová, S.; Verbeken, A.; Adamčík, S. New insights in *Russula* subsect. *Rubrinae*: Phylogeny and the quest for synapomorphic characters. *Mycol. Prog.* 2017, *16*, 877–892. [CrossRef]
- 27. Vilgalys, R.; Hester, M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J. Bacteriol.* **1990**, 172, 4238–4246. [CrossRef] [PubMed]
- James, T.Y.; Kauff, F.; Schoch, C.; Matheny, P.B.; Hofstetter, V.; Cox, C.; Celio, G.; Gueidan, C.; Fraker, E.; Miadlikowska, J.; et al. Reconstructing the early evolution of the fungi using a six gene phylogeny. *Nature* 2006, 443, 818–822. [CrossRef]
- White, T.J.; Bruns, T.; Lee, S.; Taylor, J.W. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenies. In PCR Protocols: A Guide to Methods and Applications; Innis, M.A., Gelfand, D.H., Sninsky, J.J., White, T.J., Eds.; Academic Press: New York, NY, USA, 1990; pp. 315–322.
- Hall, T.A. BioEdit: A user-friendly biological sequence alignment editor and analyses program for Windows 95/98/NT. Nucleic Acids Symp. Ser. 1999, 41, 95–98. [CrossRef]
- Liu, X.L.; Tolgor, B.; Wang, X.H. Species diversity of *Russula* from the Greater and Lesser Hinggan Mountains in Northeast China. *Mycosystema* 2017, 36, 1355–1368. [CrossRef]
- 32. Jabeen, S.; Razaq, A.; Niazi, A.R.K.; Ahmad, I.; Grebenc, T.; Khalid, A.N. *Russula ahmadii* (Basidiomycota, Russulales), a new species in section *Ingratae* and its ectomycorrhiza from coniferous forests of Pakistan. *Phytotaxa* 2017, 321, 241–253. [CrossRef]
- Vidal, J.M.; Alvarado, P.; Loizides, M.; Konstantinidis, G.; Chachuła, P.; Mleczko, P.; Moreno, G.; Vizzini, A.; Krakhmalnyi, M.; Paz, A.; et al. A phylogenetic and taxonomic revision of sequestrate Russulaceae Mediterranean and temperate Europe. *Persoonia* 2019, 42, 127–185. [CrossRef]
- 34. Eberhardt, U. Molecular kinship analyses of the agaricoid Russulaceae: Correspondence with mycorrhizal anatomy and sporocarp features in the genus *Russula*. *Mycol. Prog.* **2002**, *1*, 201–223. [CrossRef]
- Crous, P.W.; Wingfield, M.J.; Burgess, T.I.; Hardy, G.E.; Barber, P.A.; Alvarado, P.; Barnes, C.W.; Buchanan, P.K.; Heykoop, M.; Moreno, G.; et al. Fungal planet description sheets: 558–624. *Persoonia* 2017, *38*, 240–384. [CrossRef]
- Wang, X.H.; Das, K.; Horman, J.; Antonin, V.; Baghela, A.; Chakraborty, D.; Hembrom, M.E.; Nakasone, K.; Ortiz-Santana, B.; Vizzini, A.; et al. Fungal biodiversity profiles 51–60. *Cryptogam. Mycol.* 2018, 39, 211–257. [CrossRef]
- Yuan, H.S.; Lu, X.; Dai, Y.C.; Hyde, K.D.; Kan, Y.H.; Kušan, I.; He, S.H.; Liu, N.G.; Sarma, V.V.; Zhao, C.L.; et al. Fungal diversity notes 1277–1386: Taxonomic and phylogenetic contributions to fungal taxa. *Fungal Divers.* 2020, 104, 1–266. [CrossRef]
- Lee, H.; Park, M.S.; Jung, P.E.; Eimes, J.A.; Seok, S.J.; Lim, Y.W. Re-evaluation of the taxonomy and diversity of *Russula* section *Foetentinae* (Russulales, Basidiomycota) in Korea. *Mycoscience* 2017, *58*, 351–360. [CrossRef]
- 39. Looney, B.P.; Ryberg, M.; Hampe, F.; Sánchez-García, M.; Matheny, P.B. Into and out of the tropics: Global diversification patterns in a hyperdiverse clade of ectomycorrhizal fungi. *Mol. Ecol.* **2016**, *25*, 630–647. [CrossRef]
- Buyck, B.; Eyssartier, G.; Armada, F.; Corrales, A.; Hembrom, M.E.; Rossi, W.; Bellanger, J.M.; Das, K.; Dima, B.; Ghosh, A.; et al. Fungal Biodiversity Profiles 111–120. *Cryptogam. Mycol.* 2022, *43*, 23–61. [CrossRef]
- Razaq, A.; Ilyas, S.; Khalid, A.N.; Niazi, A.R. Russula foetentoides (Russulales, Russulaceae)—A new species from Pakistan. Sydowia 2014, 66, 289–298. [CrossRef]
- 42. Melera, S.; Ostellari, C.; Roemer, N.; Avis, P.G.; Tonolla, M.; Barja, F.; Narduzzi-Wicht, B. Analysis of morphological, ecological and molecular characters of *Russula pectinatoides* Peck and *Russula praetervisa* Sarnari, with a description of the new taxon *Russula recondita* Melera & Ostellari. *Mycol. Prog.* 2017, *16*, 117–134.
- 43. Li, G.J. Taxonomy of Russula from China. Ph.D. Thesis, University of Chinese Academy of Sciences, Beijing, China, 2014.
- 44. Buyck, B.; Hofstetter, V.; Eberhardt, U.; Verbeken, A.; Kauff, F. Walking the thin line between *Russula* and *Lactarius*: The dilemma of *Russula* subsect. *Ochricompactae*. *Fungal Divers*. **2008**, *28*, 15–40.
- 45. Ghosh, A.; Das, K.; Bhatt, R.P.; Hembrom, M.E. Two new species of genus *Russula* from Western Himalaya with morphological details and phylogenetic estimations. *Nova Hedwig.* **2020**, *111*, 115–130. [CrossRef]
- Ghosh, A.; Buyck, B.; Das, K.; Chakraborty, D. Two new Asian species of *Russula* sect. *Ingratae with unique basidiospore features for* subg. *Heterophyllidiae. Eur. J. Taxon.* 2022, 847, 104–120. [CrossRef]

- 47. Xie, X.D.; Liu, P.G.; Yu, F.Q. Species diversity of russuloid mycorrhizae forming fungi on *Pinus yunnanensis* seedlings and the mycorrhizal morphology. *Acta Bot. Yunnanica* 2010, 32, 211–220. [CrossRef]
- 48. Wang, J.; Buyck, B.; Wang, X.H.; Tolgor, B. Visiting *Russula* (Russulaceae, Russulales) with samples from southwestern China finds one new subsection of *R*. subg. *Heterophyllidia* with two new species. *Mycol. Prog.* **2019**, *18*, 771–784. [CrossRef]
- 49. Looney, B.P. Molecular annotation of type specimens of *Russula* species described by W.A. Murrill from the southeast United States. *Mycotaxon* **2014**, *129*, 255–268. [CrossRef]
- 50. Haelewaters, D.; Dirks, A.C.; Kappler, L.A.; Mitchell, J.K.; Quijada, L.; Vandegrift, R.; Buyck, B.; Pfister, D.H. A preliminary checklist of fungi at the Boston Harbor Islands. *Northeast. Nat.* **2018**, *25*, 45–77. [CrossRef]
- 51. Das, K.; Ghosh, A.; Chakraborty, D.; Li, J.W.; Qiu, L.H.; Baghela, A.; Halama, M.; Hembrom, M.E.; Mehmood, T.; Parihar, A.; et al. Fungal biodiversity profiles 31–40. *Cryptogamie Mycol.* **2017**, *38*, 353–406. [CrossRef]
- 52. Lebel, T.; Tonkin, J.E. Australasian species of Macowanites are sequestrate species of *Russula* (Russulaceae, Basidiomycota). *Aust. Syst. Bot.* **2007**, *20*, 355–381. [CrossRef]
- 53. Miller, S.L.; Buyck, B. Molecular phylogeny of the genus *Russula* in Europe with a comparison of modern infrageneric classifications. *Mycol. Res.* **2002**, *106*, 259–276. [CrossRef]
- 54. Khatua, S.; Dutta, A.K.; Acharya, K. Prospecting *Russula senecis*: A delicacy among the tribes of West Bengal. *PeerJ* 2015, *3*, e810. [CrossRef]
- 55. Yomyart, S.; Piapukiew, J.; Watling, R.; Whalley, A.J.S.; Sihanonth, P. *Russula siamensis*: A new species of annulate *Russula* from Thailand. *Mycotaxon* **2006**, *95*, 247–254.
- 56. Buyck, B.; Duhem, B.; Das, K.; Jayawardena, R.S.; Niveiro, N.; Pereira, O.L.; Prasher, I.B.; Adhikari, S.; Albertó, E.O.; Bulgakov, T.S.; et al. Fungal biodiversity profiles 21–30. *Cryptogam. Mycol.* **2017**, *38*, 101–146. [CrossRef]
- 57. Edgar, R.C. MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 2004, 32, 1792–1797. [CrossRef] [PubMed]
- 58. Smith, S.A.; Dunn, C.W. Phyutility: A phyloinformatics tool for trees, alignments and molecular data. *Bioinformation* **2008**, *24*, 715–716. [CrossRef] [PubMed]
- Stamatakis, A. RAxML-VI-HPC: Maximum likelihood based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 2006, 22, 2688–2690. [CrossRef] [PubMed]
- 60. Huelsenbeck, J.P.; Ronquist, F. Bayesian analysis of molecular evolution using MrBayes. In *Statistical Methods in Molecular Evolution*; Nielsen, R., Ed.; Springer: New York, NY, USA, 2005; pp. 183–226. [CrossRef]
- 61. Nylander, J.A.A. *MrModeltest 2.3. Program Distributed by the Author;* Evolutionary Biology Center, Uppsala University: Uppsala, Sweden, 2004.
- 62. Das, K.; Sharma, J.R.; Atri, N.S. Russula in Himalaya 3: A new species of subgenus Ingratula. Mycotaxon 2006, 95, 271–275.
- 63. Chou, W.N.; Wang, Y.Z. Nine species of *Russula* (Basidiomycotina) new to Taiwan. *Taiwania* 2005, 50, 93–100.
- 64. Wu, X.L.; Dai, Y.C.; Li, T.H.; Yang, Z.L.; Song, B. Fungi of Tropical China; Science Press: Beijing, China, 2010; pp. 1–477.
- 65. Bazzicalupo, A.L.; Buyck, B.; Saar, I.; Vauras, J.; Carmean, D.; Berbee, M.L. Troubles with mycorrhizal mushroom identification where morphological differentiation lags behind barcode sequence divergence. *Taxon* **2017**, *66*, 791–810. [CrossRef]
- 66. Tang, L.P.; Hao, Y.J.; Cai, Q.; Tolgor, B.; Yang, Z.L. Morphological and molecular evidence for a new species of *Rhodotus* from tropical and subtropical Yunnan, China. *Mycol. Prog.* **2013**, *13*, 45–53. [CrossRef]
- 67. Xie, H.J.; Tang, L.P.; Mu, M.; Fan, Y.G.; Jiang, S.; Su, M.S.; Liang, Z.Q.; Zeng, N.K. A contribution to knowledge of *Gyroporus* (Gyroporaceae, Boletales) in China: Three new taxa, two previous species, and one ambiguous taxon. *Mycol. Prog.* **2022**, *21*, 71–92. [CrossRef]
- Zhang, Y.Z.; Lin, W.F.; Buyck, B.; Liang, Z.Q.; Su, M.S.; Chen, Z.H.; Zhang, P.; Jiang, S.; An, D.Y.; Zeng, N.K. Morphological and phylogenetic evidences reveal four new species of *Cantharellus* subgenus *Cantharellus* (Hydnaceae, Cantharellales) from China. *Front. Microbiol.* 2022, *13*, 900329. [CrossRef]
- 69. Zhao, H.; Zhou, M.; Liu, X.Y.; Wu, F.; Dai, Y.C. Phylogeny, divergence time estimation and biogeography of the genus *Onnia* (Basidiomycota, Hymenochaetaceae). *Front. Microbiol.* **2022**, *13*, 907961. [CrossRef] [PubMed]
- 70. Aoki, W.; Bergius, N.; Kozlan, S.; Fukuzawa, F.; Okuda, H.; Murata, H.; Ishida, T.A.; Vaario, L.M.; Kobayashi, H.; Kalmiş, E.; et al. New findings on the fungal species *Tricholoma matsutake* from Ukraine, and revision of its taxonomy and biogeography based on multilocus phylogenetic analyses. *Mycoscience* 2022, 63, 197–214. [CrossRef]

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