

1 Supplementary Material

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3 Table 1: Samples extracted in this study.

Sample	Absorbance ratio 260/280
<i>E.logifolia</i> root	1.77 ± 0.04
P1	1.67 ± 0.02
P2	1.60 ± 0.08
P3	1.72 ± 0.02
P4	1.56 ± 0.04
P5	1.21 ± 0.06
P6	0.79 ± 0.05
<i>C.sinensis</i> leaf	1.83 ± 0.02
P7	1.88 ± 0.01
P8	1.86 ± 0.00
P9	1.82 ± 0.01
Coffea bean	1.61 ± 0.03
P10	0.93 ± 0.23
P11	1.14 ± 0.51

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5 *E.longifolia* root, P1,P2,P3,P4, *C.sinensis* leaf, P7,P8 and P9 exhibit good DNA quality and were successfully  
 6 amplified using both rbcLa and ITS2 primers. However for P5, P6, P10 and P11, they yield a very low DNA  
 7 quality which hindered PCR amplification and HRM analysis.

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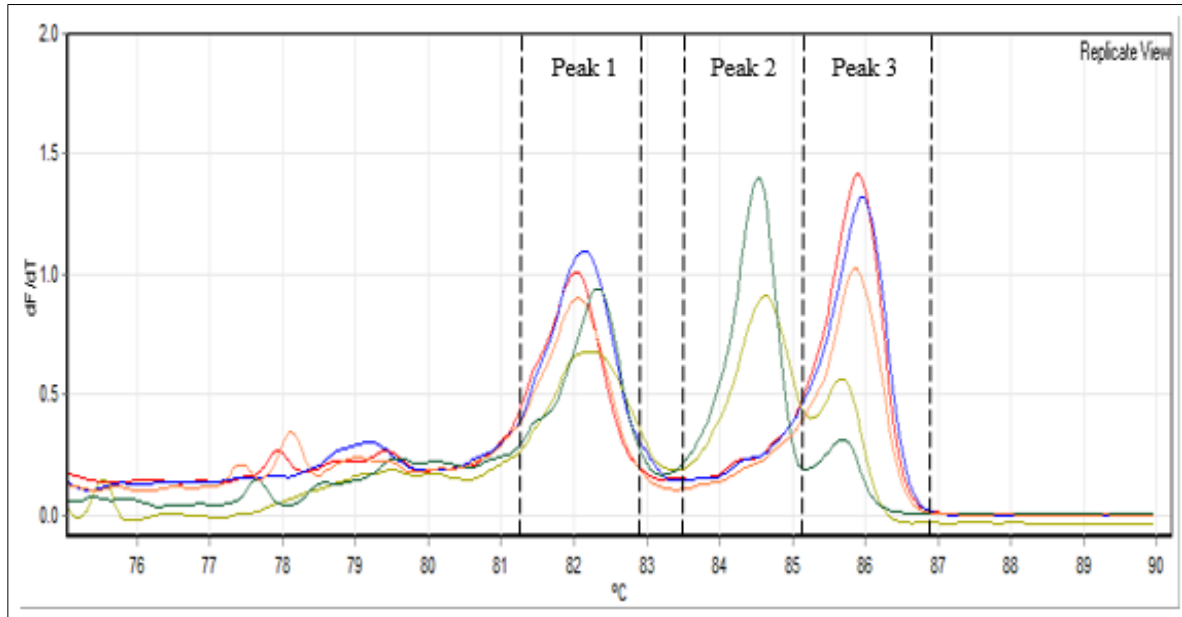
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Figure 1: Melt curve peak of *E. longifolia* and four herbal products amplified using *rbcLa* primer

Table 2: Melting temperature  $T_m$  (°C) of *E. longifolia* root and four herbal products amplified using *rbcLa* primer.

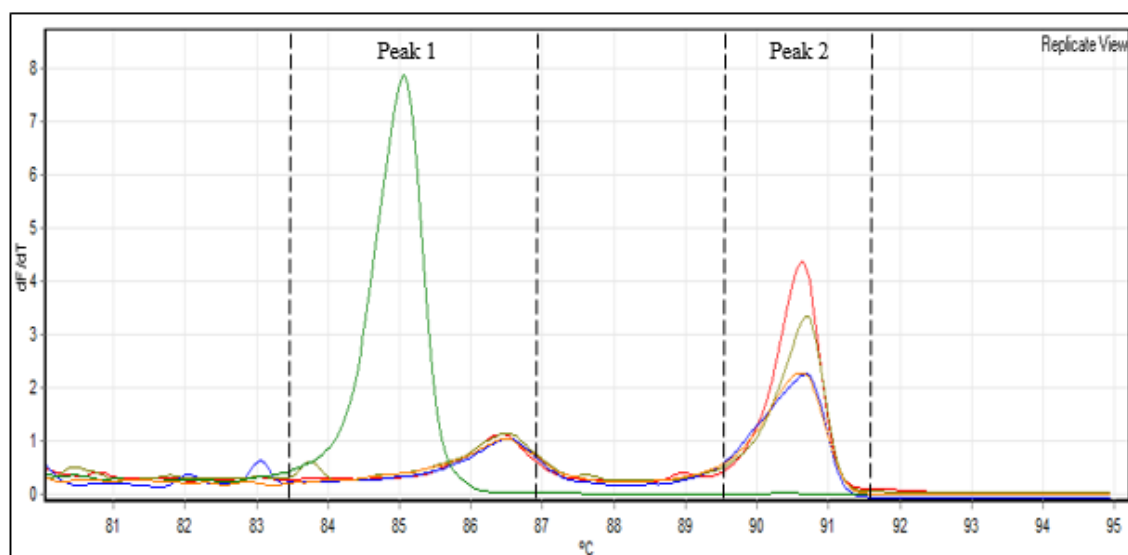
Sample	Peak 1 $T_m \pm SD$	Peak 2 $T_m \pm SD$	Peak 3 $T_m \pm SD$
<i>E. longifolia</i>	82.04 ± 0.02	-	85.90 ± 0.00
P1	82.14 ± 0.06	-	85.97 ± 0.02
P2	82.24 ± 0.26	84.64 ± 0.16	85.69 ± 0.13
P3	82.33 ± 0.04	84.53 ± 0.00	85.69 ± 0.02
P4	82.06 ± 0.04	-	85.87 ± 0.05

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27 Figure 2: Melt curve peak of *E. longifolia* and four herbal products amplified using *rbcl*a primer

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29 Table 3: Melting temperature  $T_m$  (°C) of *E. longifolia* root and four herbal products amplified using ITS2 primer.

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Sample	Peak 1 $T_m \pm SD$	Peak 2 $T_m \pm SD$
<i>E. longifolia</i>	$86.43 \pm 0.04$	$90.62 \pm 0.00$
P1	$86.51 \pm 0.01$	$90.69 \pm 0.01$
P2	$86.50 \pm 0.00$	$90.69 \pm 0.01$
P3	$85.06 \pm 0.01$	-
P4	$86.47 \pm 0.09$	$90.65 \pm 0.21$

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