



Review

Titanium Dioxide: Structure, Impact, and Toxicity

Anca Diana Racovita

Department of Chemistry, Faculty of Science, University of Warwick, Coventry CV4 7AL, UK;
diana.racovita@warwick.ac.uk

Abstract: Titanium dioxide, first manufactured a century ago, is significant in industry due to its chemical inertness, low cost, and availability. The white mineral has a wide range of applications in photocatalysis, in the pharmaceutical industry, and in food processing sectors. Its practical uses stem from its dual feature to act as both a semiconductor and light scatterer. Optical performance is therefore of relevance in understanding how titanium dioxide impacts these industries. Recent breakthroughs are summarised herein, focusing on whether restructuring the surface properties of titanium dioxide either enhances or inhibits its reactivity, depending on the required application. Its recent exposure as a potential carcinogen to humans has been linked to controversies around titanium dioxide's toxicity; this is discussed by illustrating discrepancies between experimental protocols of toxicity assays and their results. In all, it is important to review the latest achievements in fast-growing industries where titanium dioxide prevails, while keeping in mind insights into its disputed toxicity.

Keywords: titanium dioxide; nanotoxicity; industrial applications; photocatalysis; food preservative; sunscreen agent; health controversies

Citation: Racovita, A.D. Titanium Dioxide: Structure, Impact, and Toxicity. *Int. J. Environ. Res. Public Health* **2022**, *19*, 5681. <https://doi.org/10.3390/ijerph19095681>

Received: 30 March 2022

Accepted: 3 May 2022

Published: 6 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Titanium dioxide (TiO₂) is a white powder extensively used to decontaminate water and food, ensuring environmental and industrial safety, while also serving to protect the skin against harmful radiation [1–5]. To better understand how this metal oxide functions, it is relevant to describe its polymorphic crystal structure [1,2].

1.1. Polymorphism

Titanium dioxide exists in three phases: as rutile [1], anatase [1], and brookite [2]. These crystal phases assemble as octahedra, where six oxygen anions are shared by three titanium (IV) cations [2], hence the formula TiO_{6/3}, which equals TiO₂. While expanding in a three-dimensional space, these octahedral units arrange and distort differently for each polymorph, which leads to distinct patterns of crystallinity [2]. As such, the three polymorphs differ in shape, structure [1–3], density [1], and refractive index [1]. Rutile has a comparatively higher structural stability [1,2,4–6], given that transitions of this phase during synthesis and use are rare [1]. The metastable anatase and brookite can be thermally restructured into the more thermodynamically stable rutile, depending on the mineral's industrial purpose [4,5]. Brookite is a rarely encountered crystal phase and challenging to manufacture in industry [2].

1.2. Optical Properties

Titania (Figure 1) is a white powder with a high refractive index that averages values of 2.7 in rutile and 2.5 in anatase [1,6]. The refractive index is a measure of the metal oxide's ability to scatter photons [1]. In this sense, rutile reflects light more efficiently compared to anatase and brookite, which marks rutile as an optimal pigment [1]. Interestingly,

the metal oxide has a dual behaviour towards light, by scattering and absorbing ultraviolet radiation, both highly desired properties in photocatalysis [1–3,7].



Figure 1. Reagent-grade TiO_2 powder (anatase, 99% purity).

Under ultraviolet (UV) light, the metal oxide is a powerful semiconductor, capable of catalysing redox reactions due to its photoelectric properties [1,2,6,8–10]. These relate to titania's band structure as depicted in Figure 2. The energy gap (E_G) is the region between the valence band, where the lower energy levels are occupied by valence electrons, and the conduction band, where higher energy levels can be filled by more electrons [5,8]. Within a band gap, there are no energy levels for accommodating electrons [2,5]. The value of E_G is either equal to or smaller than the energy of ultraviolet photons [5,8]. The region can be associated with a barrier for electron transfer—for higher E_G values, stronger UV light must be absorbed by electrons in the valence band to ensure their excitation to energy levels within the conduction band [2,5]. Specifically, titanium dioxide displays E_G values of 3.1 eV for rutile and 3.3 eV for anatase [11]. By illuminating TiO_2 with ultraviolet light, electrons transit from their ground state in the valence band to an excited state in the conduction band (e^-_{CB}), giving rise to positive holes in the valence band (h^+_{VB}) [2,5,8,9]. After this event, molecules adsorbed onto the catalyst's surface, such as water and oxygen, can undergo redox reactions [2,5,8,9]. Oxidation occurs at the valence band, through h^+_{VB} electron acceptors, whereas reduction takes place at the conduction band, with the aid of elevated photoelectrons, e^-_{CB} [2,5,8,9].

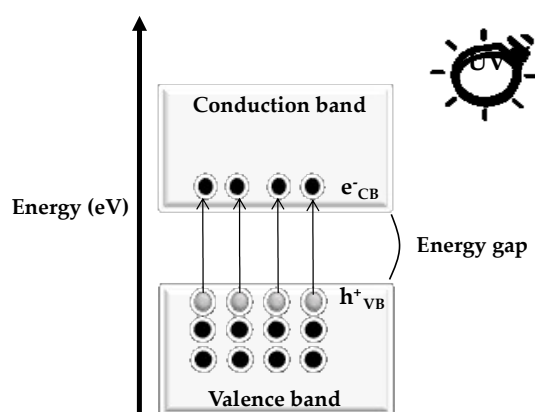


Figure 2. Photoelectrochemical phenomena at TiO_2 exposed to ultraviolet (UV) light. Adapted from reference [8].

1.3. Nanoarchitecture Achievements

Titanium dioxide nanoparticles are part of the top five nanoparticles used in industry [12], owing to their versatility in applications—as photocatalysts [5], in pharmaceuticals [13,14], processed foods [15–18] and household products [13,17,19], cosmetic white pigments [17,18], fabrics [18], and paints, and sunscreens [19].

Compared to microparticles, titanium dioxide nanoparticles display enhanced catalytic activity [3,5]. This is because a decrease in size leads to an increased surface area available for catalysis [5,10,12,20,21]. Recent breakthroughs have been achieved in medicinal applications of photocatalysis, by testing nano-titania as an anticancer agent [14,20,21]. Balachandran et al. reported that irradiated TiO₂ particles below 20 nm are an efficient “photo-killer” of pulmonary cancer cells [20]. Valence band holes, with their strong oxidant character, lead to the formation of reactive oxygen species; these will interact with defective cells, causing significant intracellular damage, to finally induce their necrosis [20,21].

Modern breakthroughs are also seen in nanobiotechnology. Although many synthetic routes have been designed for nano-TiO₂, their cost is significant and often associated with environmental hazards [22–25]. In contrast, the “green” syntheses of nano-TiO₂ from plants and seeds extracts have been extensively researched, as they prove to be safer, cost-effective, and less toxic [23–25]. In general, these methods require TiO₂ precursors, such as titanium isopropoxide [24] or titanium trichloride [25], which are centrifuged with natural extracts in aqueous solutions [23–25]. Interestingly, nanoparticle formation is accelerated by stabilizing interactions with these natural biocomponents [24]. Lingaraju et al. recently tailored the synthesis of anatase titania nanoparticles from fungal biomass [13]. The publication highlights an improved activity of UV-irradiated nano-TiO₂ against the proliferation of microbial pathogens [13]. Moreover, the metal oxide’s cytotoxic character was assessed, by monitoring the induction of apoptosis in lung and breast cancer cells [13]. Another medicinal attribute observed is the oxide’s role as an anticoagulant, by limiting the formation of blood clots and preventing heart and brain damage [13]. While the interaction mechanisms with cells have yet to be explored, the novel synthesis was facile, cost-effective, and environmentally benign [13].

Titania’s nanoform is also valuable in preventing skin cancer caused by overexposure to ultraviolet radiation [26,27]. TiO₂ nanoparticles scatter UV photons more efficiently than microparticles [26]. This ability enhances the sun protection factor (SPF) of sun creams, a measure of dermal shielding against photodamage [28–30]. Moreover, nano-titania’s photoprotective behaviour [28], coupled with its ability to preserve aliments [31], have seen a multitude of applications in the food industry [15,16,31].

Given the complexity of titania’s features, this review highlights the impact of the metal oxide’s optical properties on the environmental safety sector, and on the pharmaceutical and food industries. Further on, the focus will be on growing concerns in the scientific community regarding titanium dioxide’s nanotoxicity. Then, the discrepancies between toxicity assays will be elaborated on.

2. Environmental Decontaminant

2.1. Chemical and Biological Warfare Agents

There are countless contemporary threats to environmental and human safety. In this regard, chemical and biological warfare agents (CBWA) have been extensively used as weapons of mass destruction in last century’s war acts, but also in modern times [32–34]. Clearing the environment from these toxins has improved in the 21st century, owing to the semiconductor properties of metal oxides such as TiO₂ [32]. Its ability to generate reactive oxygen species (ROS) in aqueous suspensions has been employed in biological decontamination against ubiquitous Gram-negative pathogens, such as *Escherichia coli* [9] and *Vibrio harveyi*, by cleaving the bacterial membrane and harming intracellular components [35].

Moreover, ROS have been researched for their potential to decompose nerve agent simulants, especially organophosphates, which abound in aqueous environments [32,33].

In this field, doping techniques have seen useful applications by enhancing the photoelectrochemical properties of TiO_2 [2,3,36,37]. As experimented by Shen et al., the technique incorporates metal ion impurities of different electronegativities and sizes into titanium dioxide's structure [36,37]. These impurities lower the band gap value, E_G' , ergo increasing the catalyst's reactivity, as depicted in Figure 3 [2,3,36,37]. As a result, suspensions of the metal oxide's nanoform co-doped with transition metal cations, such as germanium (Ge^{4+}) and zirconium (Zr^{4+}), display superior efficiency in degrading chemical warfare agents, compared to pure titania [36]. Shen et al. continued their research on a different titania suspension, doped with Zn^{2+} cations instead of Zr^{4+} [37]. The degradation of toxins was feasible at temperatures below 0°C , which facilitated catalysis under extreme environmental conditions [37].

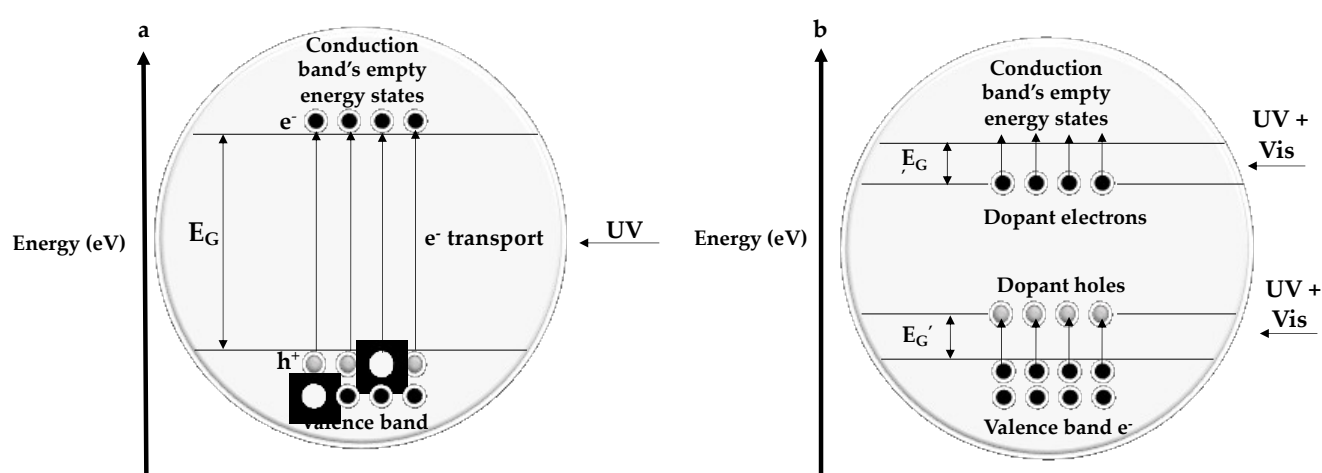
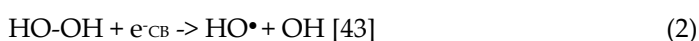


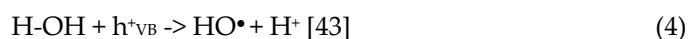
Figure 3. The effect of dopants on the band gap's electronic structure and E_G . (a) Band structure of pure titanium dioxide. (b) Formation of additional donor and acceptor energy levels as a result of doping. E_G = energy gap. Adapted from reference [3].

Another recent design in this field is represented by water-suspended magnesium micromotors coated with Au-doped titanium dioxide. When irradiated with UV light, the dispersed catalyst generates ROS from water molecules and inactivates *Bacillus globigii*, a simulant for the biological warfare agent *Bacillus anthracis* [32]. The same micromotors were found to decompose organophosphate nerve agents into nonhazardous fragments, indicating an advantageous dual use [32]. The catalytic activity against these warfare agents is enhanced by doping TiO_2 with Au nanoparticles, efficiently separating e_{CB} and h_{VB} to prevent electron–hole recombinations [32].

2.2. Large-Scale Water Decontamination

Degrading persistent chemical pollutants into nonhazardous fragments has also been extensively applied in large-scale water decontamination [38–42]. Various mechanisms of action have been suggested for titanium dioxide [9,43–46]. The consensus is that once the catalyst is illuminated with UV light of higher energy than its E_G , molecules that adsorbed on the metal oxide interact with h_{VB} , or e_{CB} [2,3,5,8,38], generating a steady flux of hydroxyl radicals (HO^\bullet), as seen in Equation 1–4 [3,9,38,42,43,45]. These radicals can break chemical bonds of molecular impurities, which therefore become non-polluting [9,42,44].





Water potability remains a growing concern worldwide, due to the detrimental impacts of chemical spills on human health and environmental balance [3,46]. Over 200 publications in the last 4 years relate to discovering novel methods for water decontamination. This could be motivated by the increasing number and complexity of chemical pollutants [3,5], but also by the persistence of synthetic toxins [46] and medicinal drugs [3] that have already been disposed of in water.

Recent innovations in this field involve the catalytic action of titanium dioxide in nanoform, based on its structural versatility. Even though the catalyst is highly efficient under UV light, there is scientific interest to enhance the compound's reactivity in the visible spectra, by merging it with carbon-based materials [2,3,39,47]. Notably, Silva et al. improved nano-TiO₂'s morphology with graphene oxides and carbon nanotubes through thermal treatments [39]. The results have revealed an increased surface area available for catalysis and a reduced frequency of recombination between electron-hole pairs [39], which ensures that more redox sites are active [2,39,47]. The novel approach enhances the efficiency of TiO₂ upon degrading organic pollutants from both aqueous and aerial media, under both UV and visible light [39].

Leshuk et al. outlined a more facile preparation method for restructured titania, termed "floating photocatalyst", whereby TiO₂ nanoparticles are mobilised onto a glass microsphere, as seen in Figure 4 [41]. The hollow beads float at random at the air–water interface because they exhibit a lower density than water [41]. Additionally, they can be reused for multiple catalytic cycles, compared to the conventional use of a catalyst slurry [3,41]. This facilitates a cost-effective decontamination, as only a small number of glass beads is required for each water treatment [3,41].

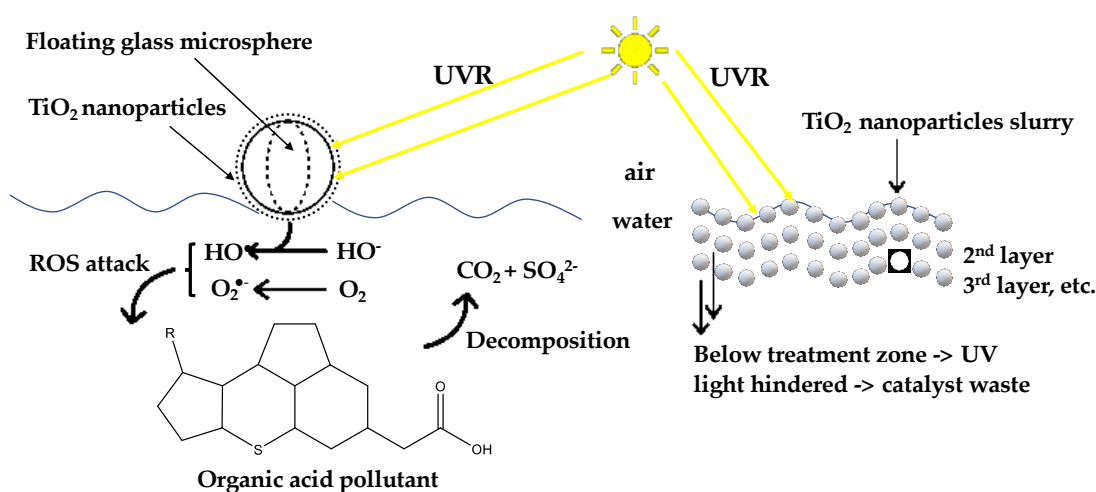


Figure 4. Floating photocatalyst at the air–water interface vs. conventional titania slurry. UVR = ultraviolet radiation. ROS = reactive oxygen species. HO = hydroxyl, O₂ = oxygen, CO₂ = carbon dioxide. SO₄²⁻ = sulphate. Adapted from ref. [41].

Both syntheses provide structural designs of titanium dioxide with prolonged activity, small costs, and recyclability features [39,41]. This renders the catalyst more efficient than commercially available decontaminants at degrading organic toxins such as benzene [39], dyes [39], and organic acids [41]. One factor that cannot be controlled is the intensity of sunlight in polluted environments.

3. Sunscreen Efficacy of TiO₂

3.1. UV Radiation: Exposure Knowledge and Impact

Sun tanning is popular in modern times, not only for cosmetic purposes, but also for medicinal ones [48]. The exposure of skin cells to ultraviolet radiation is beneficial due to the production of vitamin D₃ in the body [48] via a complex biosynthesis illustrated in Figure 5. Numerous publications relate vitamin D [49] and its synthetic analogues [50–52] to the inhibition of different types of cancer, such as hepatocellular [51], prostate cancer [52,53], and leukaemia [53]. Vitamin D is also responsible for normal levels of calcium [51] and phosphorus [54] in the body, thus maintaining bone strength [48,54] and preventing osteoporosis, a cause of bone fractures [54]. In contrast to the health benefits of tanning, there is a discrepancy between societal awareness of sunburn hazards and preventing them [55]. It has been reported that aesthetic trends and herd mentality take precedence over peoples' risk assessments of photodamage [55].

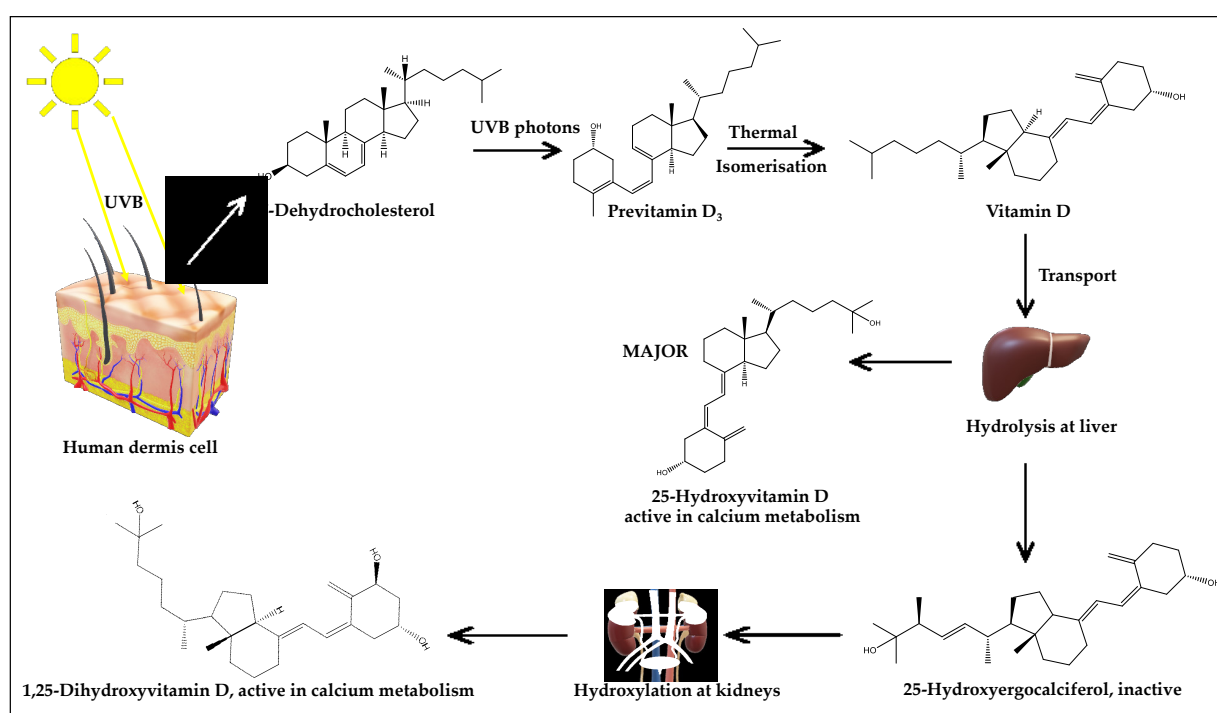


Figure 5. Vitamin D synthesis. Adapted from reference [48].

Sunburns thus remain a known carcinogen for skin cells [29,55,56]. In this context, there is a high demand for stable, photoactive chemicals termed sunscreens [57], which shield skin cells from the harmful impacts of ultraviolet radiation [29,55–59].

3.2. TiO₂ in Sun Cream Formulations

In general, sun creams consist of organic UV absorbers, whose action is complemented by inorganic UV scatterers [57–62]. When combined, dermal photoprotection is enhanced [57,58]. Their combination is necessary because one UV filter alone cannot shield against both ultraviolet B (UVB) and ultraviolet A (UVA) radiation [57,58]. These sunscreens are solubilised with emollients [61] and mixed with fragrances [62], preservatives [62], and many other stabilising agents. The industrial aim is to design cosmetic formulations that are commercially attractive [30,58], safe, and free from allergens [59].

Titanium dioxide is an inorganic sunscreen, authorised by the United States Food and Drug Administration [57] and commercialised under the name Eusolex®2000 [30]. The metal oxide scatters UV photons [1,60], thus operating as a shield for harmful radiation

[60,63,64]. However, the dual reactivity of titania towards sunlight is problematic in the sunscreen industry, given that light absorption can activate the catalyst and lead to redox reactions between h^+_{VB} or e^-_{CB} [9] and the surrounding elements within the sun cream [60,63,64]. Any structural changes in the formulation matrix are undesirable due to their impact on the cream's stability, safety, and photoprotective function [60,63,64]. To prevent photocatalysis, an extensive body of literature is dedicated to weakening titanium dioxide's catalytic performance whilst preserving its light-scattering attribute [60,63–68]. Thus far, this has been achieved by coating the nanomaterial's surface with an unreactive shell, such as aluminium oxide (Al_2O_3) [60], zirconium dioxide (ZrO_2) [60], silicon dioxide (SiO_2) [60,63,68], silicon tetrahydride SiH_4 [63] or polymers of diethoxysiloxane [67].

3.3. Silica-Coated, New-Generation TiO_2

Silicon dioxide represents a useful inert layer, based on its insulating properties [66]. Several methods for coating TiO_2 with SiO_2 have been designed. One such method, the solvothermal process [63], is supported by El-Toni et al. The titania nanoparticles are coated with silica via a facile, mild, and time-efficient process that enables control over the shell's thickness and structure [63]. The cover of silica insulates titanium dioxide's catalytic surface underneath, therefore inhibiting undesired redox reactions in the presence of UV light [63]. Because the inert coat is transparent, UV light will still reach TiO_2 [69] and be scattered. The oxide's role as a sunscreen is maintained, while safety has improved [60,63].

Another methodology implemented for silica coating is the sol–gel technique [68]. Jaroenworarluck et al. utilised the same source of silica and reagents as El-Toni et al., but lower temperatures, slower mixing of chemicals, and longer reaction times [68]. The coated titania exhibited enhanced light-absorbing properties compared to the industrially available, uncoated material [68]. However, the suppression of photoactivity was not established [68]. Furthermore, although both techniques were experimented at the nanoscale [63,68], the coated TiO_2 resulting from the solvothermal process displayed a homogenous silica layer [63], compared to the coat formed via the sol–gel technique [68]. Thus, El-Toni et al. confirmed that higher temperatures and control over silica content influence shell layer uniformity, which is beneficial in the photoinhibition of nano- TiO_2 [63]. The field remains promising for future improvements because it is challenging to inactivate titanium dioxide's catalytic activity completely [60,63,67].

4. TiO_2 Impact in the Food Industry

4.1. Smart Packaging Composite

Titania's high refractive index and reactivity towards UV light [1] are remarkable properties not only in the sunscreen industry, but also in food processing [15,16,31,70–72]. The oxide's nanoform has been developed for use in packaging light-sensitive food products [15], to ensure food durability. In this regard, merging TiO_2 nanoparticles and biopolymers has given rise to "Active Food Packaging (AFP)" [72–74]. AFP displays increased mechanical strength and antimicrobial properties [16,70–75] compared to other "green" wrapping materials, such as paper, which is more sensitive to tearing, attracts moisture, and allows microbes to proliferate [70].

The biolayer is usually chitosan [70], or a derivative [16] that has an intrinsic biocidal activity [76]. The biolayer is permeable to water and oxygen, which are adsorbed on the package's surface, diffusing through it [16,72]. In the presence of UV light, TiO_2 transforms the diffused molecules into reactive oxygen species [16,70,72,74]. These species target, oxidise, and inactivate microbes that would otherwise proliferate in food products [16,71,72]. ROS were also found to contribute towards the photodegradation of ethylene [16,77], a highly active growth regulator for fruits and vegetables [77,78]. As such, fresh food discolouration [78] and accelerated food softening [77,78] are also minimized. The AFP mechanism of action has been summarized in Figure 6.

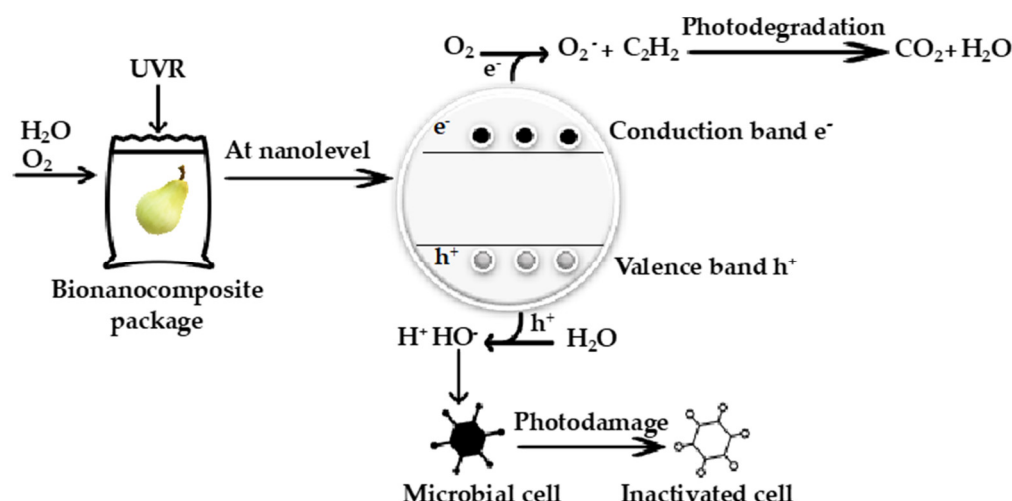


Figure 6. AFP mechanism of action. Adapted from reference [16].

One impediment observed experimentally by Siripatrawan and Kaewklin is that, during AFP manufacturing, TiO_2 nanoparticles cluster with increasing concentrations [16]. The formation of aggregates decreases titania's catalytic efficiency, due to a decrease in its effective surface area [3,5,16,20,79]. This impediment, also supported experimentally by Li et al., affects the rate of redox processes required for microbial inactivation [16,79]. To choose optimal AFP candidates, multiple synthetic trials are necessary, in order to compare the materials' catalytic efficiencies [16,79].

Overall, merging TiO_2 and biomaterials prolongs food storage, while preventing health hazards associated with contaminated food [71–73,80]. In addition, chitosan's biodegradable polysaccharide structure [70] lowers the environmental footprint specific of conventional, artificial packaging [15,72,73].

Another advantage of AFP is that it functions as an antimicrobial without releasing any chemicals onto the food supply [72,73,75,80]. In contrast, direct contact between biocides and consumer goods has led to health concerns. The diffusion of antimicrobials from the food surface into the bulk can alter the product's taste [73] or structure [80]. Spoilage, regardless of degree, is not accepted by the European Food Safety Authority (EFSA) regulations for food safety [81]. Moreover, reactive chemical release is counterproductive, because food contamination is an issue to be resolved by antimicrobial agents [81], not caused by their action. In comparison, AFP is part of an expanding area of research. Smart packaging is more efficient than previous preventive methods against food contamination because it irreversibly damages fungal and bacterial agents through a localised pathway [80]. Table 1 summarises the bactericidal and antifungal activity of smart composites against a wide range of food contaminants.

Table 1. Composition of different active food packagings and their antimicrobial activity.

Nanoparticle	Biocomposite	Antibacterial Activity	Antifungal Activity
TiO_2	Chitosan	<i>Escherichia coli</i> [16], <i>Salmonella typhimurium</i> [16], <i>Staphylococcus aureus</i> [16], <i>Pseudomonas aeruginosa</i> [16]	<i>Aspergillus oryzae</i> [16], <i>Penicillium roqueforti</i> [16]
TiO_2	Wheat gluten-cellulose	<i>S. aureus</i> [70], <i>E. coli</i> [70]	<i>Saccharomyces cerevisiae</i> [70]
TiO_2	Chitosan	<i>E. coli</i> [82]	N/A
$\text{TiO}_2 + \text{Ag}$	Chitosan	<i>E. coli</i> [83]	N/A
$\text{TiO}_2 + \text{Ag}$	Chitosan—fish gelatin	<i>E. coli</i> [84], <i>S. aureus</i> [83]	<i>Botrytis cinerea</i> [84]
TiO_2	Chitosan—graphene oxide	<i>Bacillus subtilis</i> [85]	<i>Aspergillus niger</i> [85]
$\text{TiO}_2 + \text{Ag}$	Chitosan	<i>E. coli</i> [86], <i>S. aureus</i> [86]	<i>Candida albicans</i> [86]

TiO ₂	Natural herbal and fruit extracts	<i>E. coli</i> [87], <i>Salmonella paratyphi</i> [87], <i>B. subtilis</i> [87], <i>Vibrio cholerae</i> [87], <i>Listeria monocytogenes</i> [87], <i>Streptococcus faecalis</i> [87], <i>S. aureus</i> [87], <i>P. aeruginosa</i> [87]	<i>Saccharomyces diastaticus</i> [87], <i>A. niger</i> [87]
TiO ₂	Chitosan-starch	<i>E. coli</i> [88], <i>S. aureus</i> [88]	N/A
TiO ₂	Chitosan-lemon grass oil	<i>S. aureus</i> [89], <i>Enterobacteriaceae</i> [89], psychrotrophic [89] and lactic acid bacteria [89]	N/A
TiO ₂	Chitosan	N/A	Yeasts and moulds [90]

TiO₂ = titanium dioxide, Ag = silver.

4.2. Food-Grade TiO₂

Consumer trends and dietary habits are diverse and changing at a fast pace [91]. These are mainly influenced by modern lifestyle choices and by societal demands for a diversity of new flavours [91]. Consequently, safe preservatives are necessary for tackling the waste of fresh foods and beverages, while enhancing their physical properties [91].

In the food processing sector, titanium dioxide comes under the name E171 in the European Union and INS171 in the United States [92]. The name “titanium white” [6,93] is also significant, because the metal oxide powder increases the opacity, brightness, and whiteness of many foods [93–96]. As such, it is authorised for use in creamer formulations, [94–97] to thicken their texture, and in fish [96,97], instant drinks [94], and confectionary products [93,95], to enhance their flavour. It was also found to improve the taste of many coloured foods incorporated into a daily diet [93,96,97]. Despite its presence in many foods, E171 is an additive with no nutritional value [97].

4.3. Emerging Controversies on E171

In the late 1960s, the European Union and the United States Food and Drug Administration marked food-grade titanium dioxide as safe [92]. However, in the last decade, there has been a concerning rise in publications on food-grade TiO₂. Its practical uses in the food sector are often associated with warnings on the health impact of nanosized E171 [93–95]. The plethora of publications in this field may have been propelled by the lacunes in European Food Safety Authority’s (EFSA) re-evaluation study of E171 safety, as demanded by the European Commission [98]. The panel’s target was to accurately rate the safety of titanium dioxide in food products, following oral administration [98]. In 2016, the conclusion reached by the committee is that although ingestion of E171 does not lead to carcinogenesis, its nanotoxicity on the reproductive system is inconclusive [98]. At the time, no further steps were taken by EFSA to elucidate the link between E171 and the reproductive toxicity, and an acceptable daily intake of the food additive was not established [98].

Another influencing factor for the high number of publications in this area is the analytical determination of an accurate risk assessment for nano-TiO₂ [93,99,100]. Currently, there is no documented, legally approved approach for testing particle size distribution (micro- and nano-) in E171 or in consumer goods exposed to E171 [93–95]. Concerningly, small-size nanoparticles, showing high reactivity [20,21], were found to elude detection even with modern analytical tools [93]. Ideally, E171 would only contain microscale titania to achieve the desired effects in food processing [99]. However, scientific studies highlight the presence of nanoparticles in E171 in alarming concentrations, which poses a health hazard [93,94,96,99,100]. Already in France, since January 2020, E171 is temporarily banned by decree [99]. In May 2021, relevant scientific proof of titania’s genotoxic behaviour, when accumulated in the body, was taken into consideration by the EFSA committee [100]. The panel re-emphasised the impossibility to quantify an acceptable daily intake of the preservative [100]. However, once E171 has been exposed as a potential genotoxin, a safety threshold for administration cannot be determined, because the chemical should not be administered in the first place [100]. The European Commission is now responsible for re-evaluating the legislation around E171 use [100].

Suffice to say that EFSA's decision on the topic is the most valuable regarding regulatory changes on nanomaterials in food industries [98,100]. Yet, the French government has already acted on concerns in the scientific community regarding E171 toxicity, by imposing a legal ban on its use a year before EFSA's assessment [99,100]. The nanotoxicity of E171, in all its complexity, was being investigated five years ago, as it is now. A prompt response on limiting its use is vital for ensuring the population's safety.

5. Titania Toxicity

The downside of nanoreactivity is herein described, by reviewing the methodologies employed in toxicity studies concerned with titanium dioxide. The latter's damaging potential after absorption and distribution in the body, as well as controversies around the mineral's toxicity, is the core of this section.

5.1. Post-Inhalation Toxicity

Late-20th century publications on titanium dioxide's toxicity exposed a correlation between inhaling fine particles of this mineral and the emergence of pleural diseases, especially in workers exposed to it in manufacturing plants [101,102]. At present, titanium dioxide is classified by the International Agency for Research on Cancer (IARC) as a 2B-type carcinogen, equivalent to "possibly carcinogenic to humans" [103]. This classification also implies that further studies are required to fully ascertain its cytotoxic and genotoxic potential [103].

According to clinical research, many experiments have been performed on rodents to strengthen the link between the inhalation of TiO_2 and the development of lung tumours [104], lung inflammation [105], breakdown of alveolar membranes as a result of the body's immune response [106], as well as lung tissue scarring [107]. However, there are many trials that implement questionable protocols [105,106,108]. Warheit published an extensive review on protocol discrepancies [108]. The main findings support that carcinogenesis in murine lungs is influenced by the metal oxide's particle size, crystallinity, and overload in the lungs, thus not directly or solely related to its reactivity towards pulmonary cells [108].

In toxicity assays, methodologies of relevance must be able to simulate realistic conditions, instead of introducing extreme parameters that are bound to reveal a detrimental impact. For example, in the protocol designed by Chen et al. [106], nano- TiO_2 suspensions were instilled into tracheal incisions of rodents until detrimental pulmonary changes were recorded [106]. This differs significantly from human exposure conditions. In a quality-accredited industrial laboratory, tests are performed on the metal oxide powder under rigorous health and safety regulations. These are in place in order to protect manufacturers or analytical scientists, who are trained to abide by testing procedures. To assist this, the largest study on human lung exposure to TiO_2 concludes that neither the occupational history of manufacturers nor their exposure to TiO_2 during employment is related to developing lung carcinoma [109].

To date, there are insufficient data on the duration of human lung exposure to nano- TiO_2 and the ensuing effects to make a relevant scientific correlation [101,102]. Therefore, extrapolating results from rodent studies [104–108], with little control over parameter rigour, does not represent an accurate evaluation of nano- TiO_2 toxicity in the human respiratory system.

5.2. Post-Ingestion Toxicity

For almost a decade, oral ingestion of nano-titania has represented a concerning health issue for the population [110,111]. Studies on rodents and human volunteers indicate that once titania passes the intestinal barrier, it reaches the bloodstream and is then transported to vital organs [97,110,111]. If oral consumption of the oxide's nanoform exceeds the recommended limit, it can accumulate in tissues and induce irreversible damage

[112]. The harmful effects observed on rodents include stomach and intestine inflammation [113], liver cell necrosis [112], lesions to cardiovascular tissue [112], enhancement of anxiety disorder [110], progression of colon tumours [114], and nephrotoxicity in kidneys [112]. There are many experimental variables that can influence the outcome of ingestion toxicity assays, but the main ones to consider are the concentration and reactivity of TiO₂ nanoparticles.

5.2.1. Concentration Discrepancies

The concentration parameter is crucial to the outcome of toxicity assays and therefore must be chosen carefully for in vivo experiments.

The in vivo study carried out by Wang et al. exposed rodents to a high, fixed dose of 5 g/kg_{bw} of TiO₂ per day over the course of fourteen days, showing deleterious effects on hepatocytes, nephrons, and on myocardial tissue [112]. However, inconsistencies to real life are notable when comparing the experimental protocol [112] to European regulations [98]. Realistically, the human oral daily intake of titania nanoform results from ingesting E171 and was estimated by EFSA to be between 0.5 and 14.8 mg/kg_{bw} [98]. This estimate is approximately 1000-fold smaller than the quantity employed by Wang et al. in the rodent study. Another parameter influencing the experiment is the method of administration, which registered accidental flaws. Rodents were exposed to titanium dioxide suspensions by being forcefully fed a single daily dose, which led to mortality by mistake in some of the subjects [112]. These undesired variables, though mainly kept under control, make the study weakly reliable for human risk assessments of oral exposure to nano-TiO₂, given that human digestion is a more complex, gradual, and timely process [115].

In contrast, the in vivo toxicity experiment conducted by Medina-Reyes et al. exposed rodents to 5 mg/kg_{bw} of nano-TiO₂, which can simulate the human oral intake of this mineral for different age groups [98,110]. The rodents' health levels were carefully monitored over sixteen weeks instead of two [110,112]. Another important factor considered in this study is the dieting behaviour of the rodents [110]. The latter were organised into high-fat- and regular-fat-diet categories, which enabled the researchers to compare the effects of nano-TiO₂ ingestion on both categories, by analysing injuries to the gastro-intestinal tract, liver, and reproductive system [110]. The administration pathway also differs from the study carried out by Wang et al. [112]. Instead of forcefully feeding the rodents with a high fixed dose of the metal oxide slurry [112], E171 was introduced in the rats' drinking water, alongside their diet [110]. This gradual exposure allowed for a realistic simulation of human digestion steps and substantially improved the test results' reliability.

5.2.2. Human Toxicity Models in Different Concentrations

Novel in vitro techniques in the field of toxicity assessments focus on simulating human exposure to E171 with better accuracy than murine models [116,117]. Notably, Dorier et al. exposed a model of human intestinal cells to food-grade titanium dioxide and pure nanoparticles in an environment relevant to human physiology [116,117]. The studies investigated the link between the nanoparticles' reactivity, the formation of reactive oxygen species (ROS), and intracellular oxidative stress, which can lesion deoxyribonucleic acid (DNA) strands and induce carcinogenesis [15,116,117]. The results have revealed that, regardless of the nano-TiO₂ concentration, human intestinal cells are protected from the nanoparticles' reactivity even when the cells are at their most sensitive phase of development, i.e., before differentiation to meet specific functions in the body [116,117]. E171 thus only displayed moderate genotoxicity due to ROS formation [116,117]. The technique of continuous exposure has shown better accuracy compared to an acute, time-limited exposure [117] to correlate in vitro and in vivo studies [116].

5.3. Toxicity after Dermal Exposure

Stratum corneum cells reside in the uppermost layer of the skin, covering the entire human body [12,118]. These cells represent a significant route for absorption of nano-TiO₂ [12,118], which is incorporated into the photoprotective medical make-up [119] and sun cream formulations [12,58–60]. In this context, both *in vitro* and *in vivo* studies question the harmfulness of cosmetics containing titanium dioxide [12,120–123]. The findings are concerning in the scientific community because sunscreens are designed with the aim to protect the skin from carcinoma [58,59] and not be the cause of mutagenesis.

First, controversies have appeared regarding the ability of nanosized titanium dioxide to penetrate both the epidermis and vascularised dermis, when applying sun creams topically [12,28,59,118,119,121–124]. As indicated previously, when dispersed in the circulatory system [59,110,111], the nanomaterial can accumulate in organ tissues, leading to deleterious side-effects [110–114,116,117].

Secondly, when applied on the skin, UV-illuminated titanium dioxide can easily contact moisture in the atmosphere [97,120]. As a result, e^-_{CB} and h^{+}_{VB} [2,8,9] catalyse the formation of reactive oxygen species from water and oxygen [12,18,28,123]. These include hydroxyl radicals [8,43,44] (Equation 4) and hydroxyl anions (Equation 3). As seen before, their formation is significant in environmental and food decontamination [16,32–35,70,71,74,75]. However, the same ROS were exposed as a mutagen for dermal cells [26], by causing DNA strand breaks and oxidative stress [65,120,123].

Although coating the material with inert shells may delay titanium dioxide's catalytic activity, an irreversible inhibition of its reactivity is difficult to achieve [63,67,121]. Furthermore, the crystallinity of TiO₂ nanoparticles in sun creams can also influence the catalytic effect [64,68,121]. While rutile scatters light most efficiently [1], anatase absorbs photons more strongly [64,121]. A better regulation over the polymorphs incorporated into sun cream formulations seems necessary in order to inhibit the metal oxide's catalytic power.

6. Titania Toxicity Controversy

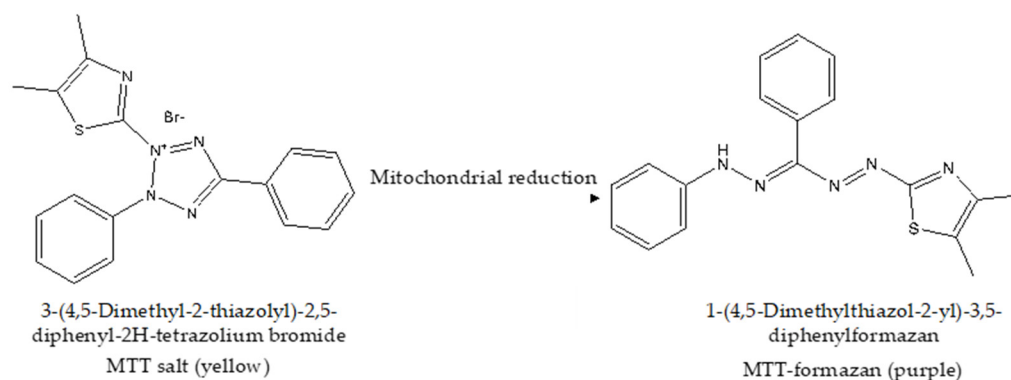
Owing to the great variety of applications of titanium dioxide, especially at the nanoscale, it is necessary to investigate its genotoxic and cytotoxic potential. At present, consistent results between toxicity assays are challenging to achieve [125–129]. Naturally, different experimental parameters and methodologies focus on distinct types of cellular damage [127–137]. Table 2 summarises the most encountered *in vitro* toxicity assays for titanium dioxide nanoparticles.

Table 2. *In vitro* toxicity assays for TiO₂.

<i>In Vitro</i> Assay	Detection Target after Exposure to Nano-TiO ₂
DCF (dichlorofluorescein) [128–133]	Cellular oxidative stress [128–133]
MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) [12,117,129,133,134]	Cell viability through metabolic activity, once in contact with the MTT dye [12,117,129,133,134]
LDH (lactate dehydrogenase) [129]	Apoptosis [129]
IL-8 (interleukin-8) with ELISA kit [129,135]	Inflammatory immune response in pulmonary cells [129,135]
Comet [127,130]	DNA strand breaks and intracellular oxidative stress [127,130]
Micronucleus [127]	ROS-induced chromosome breakage and genetic abnormalities [127]
NR (neutral red) uptake [12,133,136]	Cell viability through functional lysosomal activity [133,136]
WST-1 [137]	Cell viability, recorded as mitochondrial dehydrogenase activity in the presence of the WST-1 reagent [137]

6.3. The MTT Assay

When the yellow MTT dye comes into contact with cells, their metabolic activity reduces this salt to the purple MTT-formazan (Scheme 2) [133,134,140].



Scheme 2. MTT reduction by cellular metabolism. Adapted from reference [140].

The latter is then quantified via colourimetry, a technique that relates the MTT-formazan absorbance to its concentration with Beer–Lambert’s Law [141,142]. As such, the concentration of purple dye will indicate the population of viable cells whose metabolism was unharmed after exposure to nano-TiO₂ [117,129,133]. Often, the MTT assay is also prone to inaccuracies in titania toxicity reports [129,133]. For example, Kroll et al. ascertained that at high concentrations of titanium dioxide nanoparticles, the absorbance of the MTT formazan dye increases significantly [129]. In an experiment, this would lead to false-positive results, by detecting a higher cell viability post-exposure to the catalyst [129,133]. The outcome conforms experimentally with the findings of Guadagnini et al. [133].

6.4. The Neutral Red (NR) Assay

This method monitors the ability of healthy cells to incorporate neutral red dye into their lysosomes [136]. The supravital stain is extracted and then colourimetry detects the concentration of cells that remained intact post-exposure to the metal oxide [140]. Although this assay measures a different endpoint, it is more cost-effective and sensitive than the MTT assay [136,140]. However, light distortion was observed in the NR assay carried out by Guadagnini et al. [133], owing to titanium dioxide’s dual behaviour towards light [1]. The results expose an increase in the dye’s absorbance in the presence of the metal oxide and absence of cells [133]. This can impact the end point and, during a cytotoxic assay, an erroneous, higher cell viability would be recorded.

6.5. Meta-Analysis as a Preventive Action

Recently, novel approaches are seen through meta-analyses of experimental results. Many toxicity assays published so far for nano-TiO₂ have been analysed in order to obtain a trend in the most influential factors during experiments [125,126]. These include the size range of the nanoparticles [125,126], coating materials [60,63,68], concentration [125,126,129], and crystal phase [143], all of which influence photoreactivity. Ling et al. conducted an up-to-date comprehensive assessment of 1592 publications related to the in vitro nanotoxicity of titania [125]. Filtering was performed by excluding studies that did not offer *i*) the aforementioned factors of influence, *ii*) duplicate results of assays, in agreement with legislation requirements, and *iii*) clear, implemented protocols [125]. As such, screening allowed for the selection of 59 studies of high-quality score [125], which approximates to 3.7% of the total number of publications at the start of the meta-analysis. A similar screening system for toxicity assays on titanium dioxide was applied by Charles et al., which allowed for the selection of 36 relevant studies out of 100 [126]. The numbers

suggest that an in-depth classification of relevant experimental conditions is vital. It can serve as a reference point for future scientists, prior to performing an assay for assessing nano-titania's genotoxic and cytotoxic effects, with the aim to obtain reliable answers.

Altogether, the uncontrolled parameters in assays prompt the need for a clear, invariable definition of titania's toxicity. Implementing a proper methodology for toxicity assays of light-sensitive materials is of the utmost importance. A consensus on hazard assessments of nano-TiO₂ must be reached after extensive scientific collaborations. Unfortunately, what the accuracy percentages reveal is that scientific competition and bias may take precedence over communal efforts to design adequate, errorproof protocols for toxicity assays of light-sensitive metal oxides, such as TiO₂.

7. Conclusions and Outlook

This review has highlighted the impact, both positive and detrimental, of titanium dioxide's structure and catalytic properties in the pharmaceutical, dermato-cosmetic, environmental safety, and food industries. Each sector can be improved through extensive research efforts.

In view of medicinal breakthroughs, it becomes clear that a balance is required between using nano-titanium dioxide as an anticancer agent and its cytotoxic potential on healthy cells. As a decontaminant, the metal oxide is highly efficient, but special care is needed when dispersing the catalytic slurries into the environment, to ensure the proliferation of ecosystems is not impacted long-term by the residual accumulation of this mineral. In the food packaging sector, AFP is structurally versatile and displays great potential to combat white pollution and food waste. In the food processing sector, however, the controversial use of E171 has reached a point of international disputes. As such, the authorities in charge of elucidating its mutagenic potential must also be responsible for raising societal awareness of its incorporation into alimentation.

Another challenge is posed by achieving control over the oxide's reactivity. Given its multitude of uses linked to human exposure routes, it becomes vital to limit its catalytic activity, but preserve its light-scattering capacity. Scientific advances have been made towards coating titanium dioxide nanoparticles with inert shells. Experimental parameters are still in need of improvement, to accomplish more than a delay in titania reactivity, and possibly target an irreversible, complete catalytic inhibition.

At present, it is also difficult to assess what titania toxicity entails. Its dual behaviour towards light can mask the real experimental outcomes of toxicity assays that are based on the optical detection of cell viability parameters. This fact is concerning because many toxicity assays require protocol adjustments for light-sensitive catalysts. In this respect, modern meta-analytical tools provide useful insights, by screening databases of experimental reports. Such analyses could guide future scientists on what experimental conditions need special calibrations for titanium dioxide and how to achieve them in the laboratory. The standpoint of the author is that joint scientific efforts are required to ascertain the mechanisms of titanium dioxide toxicity, as this influences all the abovementioned uses and breakthroughs.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The author declares no conflict of interest.

Acknowledgments: The author wishes to express their sincere gratitude to Stefan A.F. Bon, for feedback and guidance on the manuscript of this review.

References

- Braun, J.H.; Baidins, A.; Marganski, R.E. TiO₂ pigment technology: A review. *Prog. Org. Coatings* **1992**, *20*, 105–138, [https://doi.org/10.1016/0033-0655\(92\)80001-d](https://doi.org/10.1016/0033-0655(92)80001-d).
- Shand, M.; Anderson, A.J. Aqueous phase photocatalytic nitrate destruction using titania based materials: Routes to enhanced performance and prospects for visible light activation. *Catal. Sci. Technol.*, **2013**, *3*, 879–899.
- Gopinath, K.P.; Madhav, N.V.; Krishnan, A.; Malolan, R.; Rangarajan, G. Present applications of titanium dioxide for the photocatalytic removal of pollutants from water: A review. *J. Environ. Manag.* **2020**, *270*, 110906, <https://doi.org/10.1016/j.jenvman.2020.110906>.
- Cihlar, J.; Navarro, L.K.T.; Kasperek, V.; Michalicka, J.; Kasty, J.; Castkova, K.; Celko, L. Influence of LA/Ti molar ratio on the complex synthesis of anatase/brookite nanoparticles and their hydrogen production. *Int. J. Hydrogen Energy* **2021**, *46*, 8578–8593, <https://doi.org/10.1016/j.ijhydene.2020.12.080>.
- Jing, L.; Zhou, W.; Tian, G.; Fu, H. Surface tuning for oxide-based nanomaterials as efficient photocatalysts. *Chem. Soc. Rev.* **2013**, *42*, 9509–9549, <https://doi.org/10.1039/c3cs60176e>.
- Bichowsky, F.V. Titanium White—A new method for its preparation. *Ind. Eng. Chem.* **1929**, *21*, 1061–1063, <https://doi.org/10.1021/ie50239a021>.
- Li, W.; Elzatahry, A.; Aldhayan, D.; Zhao, D. Core-shell structured titanium dioxide nanomaterials for solar energy utilization. *Chem. Soc. Rev.*, **2018**, *47*, 8203–8237.
- Herrmann, J.-M. Heterogeneous photocatalysis: fundamentals and applications to the removal of various types of aqueous pollutants. *Catal. Today* **1999**, *53*, 115–129, [https://doi.org/10.1016/S0920-5861\(99\)00107-8](https://doi.org/10.1016/S0920-5861(99)00107-8).
- Ireland, J.C.; Klostermann, P.; Rice, E.W.; Clark, R.M. Inactivation of *Escherichia coli* by titanium dioxide photocatalytic oxidation. *Appl. Environ. Microbiol.* **1993**, *59*, 1668–1670, <https://doi.org/10.1128/aem.59.5.1668-1670.1993>.
- Fröschl, T.; Hörmann, U.; Kubiak, P.; Kučerová, G.; Pfanzelt, M.; Weiss, C.K.; Behm, R.J.; Hüsing, N.; Kaiser, U.; Landfester, K.; et al. High surface area crystalline titanium dioxide: potential and limits in electrochemical energy storage and catalysis. *Chem. Soc. Rev.* **2012**, *41*, 5313–5360, <https://doi.org/10.1039/c2cs35013k>.
- Linsebigler, A.L.; Lu, G.; Yates, J.T. Photocatalysis on TiO₂ surfaces: Principles, mechanisms, and selected results. *Chem. Rev.* **1995**, *95*, 735–758, <https://doi.org/10.1021/cr00035a013>.
- Shukla, R.; Sharma, V.; Pandey, A.K.; Singh, S.; Sultana, S.; Dhawan, A. ROS-mediated genotoxicity induced by titanium dioxide nanoparticles in human epidermal cells. *Toxicol. Vitro* **2011**, *25*, 231–241, <https://doi.org/10.1016/j.tiv.2010.11.008>.
- Lingaraju, K.; Basavaraj, R.; Jayanna, K.; S.Bhavana; Devaraja, S.; Swamy, H.K.; Nagaraju, G.; Nagabhushana, H.; Naika, H.R. Biocompatible fabrication of TiO₂ nanoparticles: Antimicrobial, anticoagulant, antiplatelet, direct hemolytic and cytotoxicity properties. *Inorg. Chem. Commun.* **2021**, *127*, 108505, <https://doi.org/10.1016/j.inoche.2021.108505>.
- Çeşmeli, S.; Avci, C.B. Application of titanium dioxide (TiO₂) nanoparticles in cancer therapies. *J. Drug Target.* **2019**, *27*, 762–766, <https://doi.org/10.1080/1061186x.2018.1527338>.
- Wang, P.; Xiong, Z.; Xiong, H.; Cai, J. Synergistic effects of modified TiO₂/multifunctionalized graphene oxide nanosheets as functional hybrid nanofiller in enhancing the interface compatibility of PLA/starch nanocomposites. *J. Appl. Polym. Sci.* **2020**, *137*, <https://doi.org/10.1002/app.49094>.
- Siripatrawan, U.; Kaewklin, P. Fabrication and characterization of chitosan-titanium dioxide nanocomposite film as ethylene scavenging and antimicrobial active food packaging. *Food Hydrocoll.* **2018**, *84*, 125–134, <https://doi.org/10.1016/j.foodhyd.2018.04.049>.
- Golasik, M.; Herman, M.; Piekoszewski, W. Toxicological aspects of soluble titanium—A review of in vitro and in vivo studies. *Metallomics* **2016**, *8*, 1227–1242, <https://doi.org/10.1039/c6mt00110f>.
- Wang, Y.; Sauvat, A.; Lacroux, C.; Lebeau, J.; Grall, R.; Hullo, M.; Nesslany, F.; Chevillard, S. TiO₂ nanomaterials non-controlled contamination could be hazardous for normal cells located in the field of radiotherapy. *Int. J. Mol. Sci.* **2020**, *21*, 940, <https://doi.org/10.3390/ijms21030940>.
- Kiss, B.; Br, T.; Czifra, G.; Tóth, B.I.; Kertész, Z.; Szikszai, Z.; Kiss, Á.Z.; Juhász, I.; Zouboulis, C.C.; Hunyadi, J. Investigation of micronized titanium dioxide penetration in human skin xenografts and its effect on cellular functions of human skin-derived cells. *Exp. Dermatol.* **2008**, *17*, 659–667, <https://doi.org/10.1111/j.1600-0625.2007.00683.x>.
- Balachandran, K.; Mageswari, S.; Preethi, A. Photocatalytic decomposition of A549-lung cancer cells by TiO₂ nanoparticles. *Mater. Today Proc.* **2021**, *37*, 1071–1074, [doi:10.1016/j.matpr.2020.06.297](https://doi.org/10.1016/j.matpr.2020.06.297).
- Chen, X.; Mao, S.S. Titanium dioxide nanomaterials: synthesis, properties, modifications, and applications. *Chem. Rev.*, **2007**, *107*, 2891–2959.
- Irshad, M.A.; Nawaz, R.; Rehman, M.Z.U.; Adrees, M.; Rizwan, M.; Ali, S.; Ahmad, S.; Tasleem, S. Synthesis, characterization and advanced sustainable applications of titanium dioxide nanoparticles: A review. *Ecotoxicol. Environ. Saf.* **2021**, *212*, 111978, <https://doi.org/10.1016/j.ecoenv.2021.111978>.
- Thakur, B.; Kumar, A.; Kumar, D. Green synthesis of titanium dioxide nanoparticles using *Azadirachta indica* leaf extract and evaluation of their antibacterial activity. *South Afr. J. Bot.* **2019**, *124*, 223–227, <https://doi.org/10.1016/j.sajb.2019.05.024>.

24. Rajendhiran, R.; Deivasigamani, V.; Palanisamy, J.; Masan, S.; Pitchaiya, S. Terminalia catappa and carissa carandas assisted synthesis of TiO₂ nanoparticles—A green synthesis approach. *Mater. Today Proc.* **2021**, *45*, 2232–2238.
25. Abisharan, J.M.; Devikala, S.; Kumar, R.D.; Arthanareeswari M.; Kamaraj, P. Green synthesis of TiO₂ nanoparticles using Cucurbita pepo seeds extract. *Mater. Today: Proc.*, **2019**, *14*, 302–307.
26. de la Calle, I.; Menta, M.; Klein, M.; Séby, F. Screening of TiO₂ and Au nanoparticles in cosmetics and determination of elemental impurities by multiple techniques (DLS, SP-ICP-MS, ICP-MS and ICP-OES). *Talanta* **2017**, *171*, 291–306, <https://doi.org/10.1016/j.talanta.2017.05.002>.
27. Altsitsiadis, E.; Undheim, T.; de Vries, E.; Hinrichs, B.; Stockfleth, E.; Trakatelli, M.; on behalf of the EPIDERM Group Health literacy, sunscreen and sunbed use: an uneasy association. *Br. J. Dermatol.* **2012**, *167*, 14–21, <https://doi.org/10.1111/j.1365-2133.2012.11082.x>.
28. Barnard, A.S. One-to-one comparison of sunscreen efficacy, aesthetics and potential nanotoxicity. *Nat. Nanotechnol.* **2010**, *5*, 271–274, <https://doi.org/10.1038/nnano.2010.25>.
29. Veierød, M.B.; Weiderpass, E.; Thörn, M.; Hansson, J.; Lund, E.; Armstrong, B.; Adami, H.-O. A prospective study of pigmentation, sun exposure, and risk of cutaneous malignant melanoma in women. *JNCI: J. Natl. Cancer Inst.* **2003**, *95*, 1530–1538, <https://doi.org/10.1093/jnci/djg075>.
30. Rodrigues, N.D.; Stavros, V.G. From fundamental science to product: A bottom-up approach to sunscreen development. *Sci. Prog.* **2018**, *101*, 8–31, <https://doi.org/10.3184/003685018x15166183479666>.
31. Chen, Z.; Han, S.; Zhou, S.; Feng, H.; Liu, Y.; Jia, G. Review of health safety aspects of titanium dioxide nanoparticles in food application. *NanoImpact* **2020**, *18*, 100224, <https://doi.org/10.1016/j.impact.2020.100224>.
32. Li, J.; Singh, V.V.; Sattayasamitsathit, S.; Orozco, J.; Kaufmann, K.; Dong, R.; Gao, W.; Jurado-Sanchez, B.; Fedorak, Y.; Wang, J. Water-driven micromotors for rapid photocatalytic degradation of biological and chemical warfare agents. *ACS Nano* **2014**, *8*, 11118–11125, <https://doi.org/10.1021/nn505029k>.
33. Kozlova, E.A.; Smirniotis, P.G.; Vorontsov, A.V. Comparative study on photocatalytic oxidation of four organophosphorus simulants of chemical warfare agents in aqueous suspension of titanium dioxide. *J. Photochem. Photobiol., A*, **2004**, *162*, 503–511.
34. Chauhan, S.; D'Cruz, R.; Faruqi, S.; Singh, K.; Varma, S.; Singh, M.; Karthik, V. Chemical warfare agents. *Environ. Toxicol. Pharmacol.* **2008**, *26*, 113–122, <https://doi.org/10.1016/j.etap.2008.03.003>.
35. Alexpandi, R.; Abirami, G.; Balaji, M.; Jayakumar, R.; Ponraj, J.G.; Cai, Y.; Pandian, S.K.; Ravi, A.V. Sunlight-active phytol-ZnO@TiO₂ nanocomposite for photocatalytic water remediation and bacterial-fouling control in aquaculture: A comprehensive study on safety-level assessment. *Water research*, **2022**, *212*, 118081.
36. Shen, Z.; Zhong, J.-Y.; Han, X.-Y.; Wang, L.-Y.; Cui, Y.; Chen, L.-K.; Zheng, Y.-C. Decontamination of chemical warfare agents on sensitive equipment materials using Zr 4+ and Ge 4+ co-doped TiO₂ and hydrofluoroether suspension. *Chem. Eng. J.* **2016**, *302*, 111–119, <https://doi.org/10.1016/j.cej.2016.04.153>.
37. Shen, Z.; Zhong, J.-Y.; Yang, J.-C.; Cui, Y.; Zheng, H.; Wang, L.-Y.; Wang, J.-L. Decontamination of chemical warfare agents by Zn²⁺ and Ge⁴⁺ co-doped TiO₂ nanocrystals at sub-zero temperatures: A solid-state NMR and GC study. *Chem. Phys. Lett.* **2018**, *707*, 31–39, <https://doi.org/10.1016/j.cplett.2018.07.033>.
38. Legrini, O.; Oliveros, E.; Braun, A.M. Photochemical processes for water treatment. *Chem. Rev.* **1993**, *93*, 671–698, <https://doi.org/10.1021/cr00018a003>.
39. Silva, M.R.; Lourenço, M.A.; Tobaldi, D.M.; Silva, C.; Seabra, M.P.; Ferreira, P. Carbon-modified titanium oxide materials for photocatalytic water and air decontamination. *Chem. Eng. J.* **2020**, *387*, 124099, <https://doi.org/10.1016/j.cej.2020.124099>.
40. De Matteis, V.; Cascione, M.; Rinaldi, R. *Titanium Dioxide (TiO₂) and Its Applications*, eds. F. Parrino and L. Palmisano, Elsevier, Amsterdam, The Netherlands, pp. 373–393.
41. Leshuk, T.; Krishnakumar, H.; Livera, D.D.O.; Gu, F. Floating photocatalysts for passive solar degradation of naphthenic acids in oil sands process-affected water. *Water* **2018**, *10*, 202, <https://doi.org/10.3390/w10020202>.
42. Ollis, D.F.; Pelizzetti, E.; Serpone, N. Photocatalyzed destruction of water contaminants. *Environ. Sci. Technol.* **1991**, *25*, 1522–1529, <https://doi.org/10.1021/es00021a001>.
43. Sjogren, J.C.; Sierka, R.A. Inactivation of phage MS2 by iron-aided titanium dioxide photocatalysis. *Appl. Environ. Microbiol.* **1994**, *60*, 344–347, <https://doi.org/10.1128/aem.60.1.344-347.1994>.
44. Rajeshwar, K. Photoelectrochemistry and the environment. *J. Appl. Electrochem.* **1995**, *25*, 1067–1082, <https://doi.org/10.1007/bf00242533>.
45. Turchi, C. Photocatalytic degradation of organic water contaminants: Mechanisms involving hydroxyl radical attack. *J. Catal.* **1990**, *122*, 178–192, [https://doi.org/10.1016/0021-9517\(90\)90269-p](https://doi.org/10.1016/0021-9517(90)90269-p).
46. Hassaan, M.A.; El Nemr, A. Health and environmental impacts of dyes, *Am. J. Environ. Sci.*, **2017**, *1*, 64–67.
47. Treschev, S.Y.; Chou, P.-W.; Tseng, Y.-H.; Wang, J.-B.; Perevedentseva, E.V.; Cheng, C.-L. Photoactivities of the visible-light-activated mixed-phase carbon-containing titanium dioxide: The effect of carbon incorporation. *Appl. Catal. B: Environ.* **2008**, *79*, 8–16, <https://doi.org/10.1016/j.apcatb.2007.09.046>.
48. Holick, M.F. Vitamin D and bone health. *J. Nutr.* **1996**, *126*, 1159S–1164S, https://doi.org/10.1093/jn/126.suppl_4.1159s.
49. Garland, C.F.; Garland, F.C.; Gorham, E.D.; Lipkin, M.; Newmark, H.; Mohr, S.B.; Holick, M.F. The Role of Vitamin D in Cancer Prevention. *Am. J. Public Health* **2006**, *96*, 252–261, doi:10.2105/ajph.2004.045260.

50. Hansen, C.M.; Mäenpää, P.H. EB 1089, a novel vitamin D Analog with strong antiproliferative and differentiation-inducing effects on target cells. *Biochem. Pharmacol.* **1997**, *54*, 1173–1179, [https://doi.org/10.1016/s0006-2952\(97\)00181-0](https://doi.org/10.1016/s0006-2952(97)00181-0).
51. Hansen, C.; Hamberg, K.; Binderup, E. Seocalcitol (EB 1089) A Vitamin D Analogue of Anti-cancer Potential. Background, Design, Synthesis, Pre-clinical and Clinical Evaluation. *Curr. Pharm. Des.* **2000**, *6*, 803–828, <https://doi.org/10.2174/1381612003400371>.
52. Schwartz, G.G.; Wang, M.H.; Zang, M.; Singh, R.K.; Siegal, G.P. 1 α ,25-Dihydroxyvitamin D (calcitriol) inhibits the invasiveness of human prostate cancer cells. *Cancer Epidemiology Biomarkers Prev.* **1997**, *6*.
53. Koike, M.; Koshizuka, K.; Kawabata, H.; Yang, R.; E Taub, H.; Said, J.; Uskokovic, M.; Tsuruoka, N.; Koeffler, H.P. 20-Cyclopropyl-cholecalciferol vitamin D3 analogs: A unique class of potent inhibitors of proliferation of human prostate, breast and myeloid leukemia cell lines. *Anticancer Res.* **1999**, *19*.
54. Holick, M.F. Vitamin D Deficiency. *N. Engl. J. Med.* **2007**, *357*, 266–281, doi:10.1056/nejmra070553.
55. Arthey, S.; Clarke, V.A. Suntanning and sun protection: A review of the psychological literature. *Soc. Sci. Med.* **1995**, *40*, 265–274, [https://doi.org/10.1016/0277-9536\(94\)e0063-x](https://doi.org/10.1016/0277-9536(94)e0063-x).
56. Ziegler, A.; Jonason, A.S.; Leffell, D.J.; Simon, J.A.; Sharma, H.W.; Kimmelman, J.; Remington, L.; Jacks, T.; Brash, D.E. Sunburn and p53 in the onset of skin cancer. *Nature* **1994**, *372*, 773–776, <https://doi.org/10.1038/372773a0>.
57. Forestier, S. Rationale for sunscreen development. *J. Am. Acad. Dermatol.* **2008**, *58*, S133–S138, <https://doi.org/10.1016/j.jaad.2007.05.047>.
58. Baker, L.A.; Grosvenor, L.C.; Ashfold, N.R.; Stavros, V.G. Ultrafast photophysical studies of a multicomponent sun-screen: Oxybenzone–titanium dioxide mixtures. *Chem. Phys. Lett.* **2016**, *664*, 39–43.
59. Burnett, M.E.; Wang, S.Q. Current sunscreen controversies: a critical review. *Photodermatol. Photoimmunol. Photomed.* **2011**, *27*, 58–67, <https://doi.org/10.1111/j.1600-0781.2011.00557.x>.
60. Furusawa, T.; Honda, K.; Ukaji, E.; Sato, M.; Suzuki, N. The microwave effect on the properties of silica-coated TiO₂ fine particles prepared using sol–gel method. *Mater. Res. Bull.* **2008**, *43*, 946–957, <https://doi.org/10.1016/j.materresbull.2007.04.031>.
61. Sohn, M.; Amorós-Galicia, L.; Krus, S.; Martin, K.; Herzog, B. Effect of emollients on UV filter absorbance and sunscreen efficiency. *J. Photochem. Photobiol., B*, **2020**, *205*, 111818.
62. Schauder, S. Dermatological tolerance of UV absorbers, perfume components, and preservatives in sunscreen products. *Bundesgesundheitsblatt-Gesundheitsforschung/Gesundheitsschutz*, **2001**, *44*, 471–479.
63. El-Toni, A.M.; Yin, S.; Sato, T.; Ghannam, T.; Al-Hoshan, M.; Al-Salhi, M. Investigation of photocatalytic activity and UV-shielding properties for silica coated titania nanoparticles by solvothermal coating. *J. Alloy. Compd.* **2010**, *508*, L1–L4, <https://doi.org/10.1016/j.jallcom.2010.08.031>.
64. Osterwalder, U.; Sohn, M.; Herzog, B. Global state of sunscreens. *Photodermatol., Photoimmunol. Photomed.*, **2014**, *30*, 62–80, <https://doi.org/10.1111/phpp.12112>.
65. Serpone, N.; Salinaro, A.; Emeline, A. Deleterious effects of sunscreen titanium dioxide nanoparticles on DNA: efforts to limit DNA damage by particle surface modification. **2001**, *4258*, 86–99, <https://doi.org/10.1117/12.430765>.
66. Nekrashevich, S.S.; Gritsenko, V.A. Electronic structure of silicon dioxide (a review). *Phys. Solid State* **2014**, *56*, 207–222, <https://doi.org/10.1134/s106378341402022x>.
67. Siddiquey, I. A.; Furusawa, T.; Sato, M.; Honda, K.; Suzuki, N. Control of the photocatalytic activity of TiO₂ nanoparticles by silica coating with polydiethoxysiloxane. *Dyes Pigm.* **2008**, *76*, 754–759. <https://doi.org/10.1016/j.dyepig.2007.01.020>.
68. Jaroenworarluck, A.; Sunsaneeyametha, W.; Kosachan, N.; Stevens, R. Characteristics of silica-coated TiO₂ and its UV absorption for sunscreen cosmetic applications. *Surf. Interface Anal.* **2006**, *38*, 473–477.
69. Liang, C.; Gou, Y.; Wu, L.; Zhou, J.-G.; Kang, Z.; Shen, B.; Wang, Z. Nature of Electromagnetic-Transparent SiO₂ Shell in Hybrid Nanostructure Enhancing Electromagnetic Attenuation. *J. Phys. Chem. C* **2016**, *120*, 12967–12973, <https://doi.org/10.1021/acs.jpcc.6b04721>.
70. El-Wakil, N.A.; Hassan, E.A.; Abou-Zeid, R.E.; Dufresne, A. Development of wheat gluten/nanocellulose/titanium dioxide nanocomposites for active food packaging. *Carbohydr. Polym.*, **2015**, *124*, 337–346.
71. Cao, C.; Wang, Y.; Zheng, S.; Zhang, J.; Li, W.; Li, B.; Guo, R.; Yu, J. Poly(butylene adipate-co-terephthalate)/titanium dioxide/silver composite biofilms for food packaging application. *LWT--Food Sci. Technol.*, **2020**, *132*, 109874.
72. Mesgari, M.; Aalami, A.H.; Sahebkar, A. Antimicrobial activities of chitosan/titanium dioxide composites as a biological nanolayer for food preservation: A review. *Int. J. Biol. Macromol.* **2021**, *176*, 530–539, <https://doi.org/10.1016/j.ijbiomac.2021.02.099>.
73. Huang, T.; Qian, Y.; Wei, J.; Zhou, C. Polymeric Antimicrobial Food Packaging and Its Applications. *Polymers* **2019**, *11*, 560, doi:10.3390/polym11030560.
74. Park, S.; Jeon, Y.; Han, T.; Kim, S.; Gwon, Y.; Kim, J. Nanoscale manufacturing as an enabling strategy for the design of smart food packaging systems. *Food Packag. Shelf Life* **2020**, *26*, 100570, <https://doi.org/10.1016/j.fpsl.2020.100570>.
75. Sung, S.-Y.; Sin, L.T.; Tee, T.-T.; Bee, S.-T.; Rahmat, A.R.; Rahman, W.; Tan, A.-C.; Vikhrman, M. Antimicrobial agents for food packaging applications. *Trends Food Sci. Technol.* **2013**, *33*, 110–123, <https://doi.org/10.1016/j.tifs.2013.08.001>.

76. Cazón, P.; Velazquez, G.; Ramírez, J.A.; Vázquez, M. Polysaccharide-based films and coatings for food packaging: A review. *Food Hydrocoll.* **2017**, *68*, 136–148, doi:10.1016/j.foodhyd.2016.09.009.
77. de Chiara, M.L.V.; Pal, S.; Licciulli, A.; Amodio, M.L.; Colelli, G. Photocatalytic degradation of ethylene on mesoporous TiO₂/SiO₂ nanocomposites: Effects on the ripening of mature green tomatoes. *Biosystems Engineering*, **2015**, *132*, 61–70.
78. Saltveit, M.E. Effect of ethylene on quality of fresh fruits and vegetables. *Postharvest Biol. Technol.* **1999**, *15*, 279–292, doi:10.1016/s0925-5214(98)00091-x.
79. Li, G.; Lv, L.; Fan, H.; Ma, J.; Li, Y.; Wan, Y.; Zhao, X.S. Effect of the agglomeration of TiO₂ nanoparticles on their photocatalytic performance in the aqueous phase. *J. Colloid Interface Sci.*, **2010**, *348*, 342–347.
80. Quintavalla, S.; Vicini, L. Antimicrobial food packaging in meat industry. *Meat Sci.* **2002**, *62*, 373–380, [https://doi.org/10.1016/s0309-1740\(02\)00121-3](https://doi.org/10.1016/s0309-1740(02)00121-3).
81. European Food Safety Authority. Available online: <https://www.efsa.europa.eu/en/topics/topic/food-contact-materials> (accessed on 16 April 2021).
82. Tang, Y.; Hu, X.; Zhang, X.; Guo, D.; Zhang, J.; Kong, F. Chitosan/titanium dioxide nanocomposite coatings: Rheological behavior and surface application to cellulosic paper. *Carbohydr. Polym.* **2016**, *151*, 752–759, <https://doi.org/10.1016/j.carbpol.2016.06.023>.
83. Lin, B.; Luo, Y.; Teng, Z.; Zhang, B.; Zhou, B.; Wang, Q. Development of silver/titanium dioxide/chitosan adipate nanocomposite as an antibacterial coating for fruit storage. *LWT—Food Sci. Technol.*, **2015**, *63*, 1206–1213.
84. Lin, D.; Yang, Y.; Wang, J.; Yan, W.; Wu, Z.; Chen, H.; Zhang, Q.; Wu, D.; Qin, W.; Tu, Z. Preparation and characterization of TiO₂-Ag loaded fish gelatin-chitosan antibacterial composite film for food packaging. *Int. J. Biol. Macromol.* **2020**, *154*, 123–133, <https://doi.org/10.1016/j.ijbiomac.2020.03.070>.
85. Xu, W.; Xie, W.; Huang, X.; Chen, X.; Huang, N.; Wang, X.; Liu, J. The graphene oxide and chitosan biopolymer loads TiO₂ for antibacterial and preservative research. *Food Chem.* **2017**, *221*, 267–277, <https://doi.org/10.1016/j.foodchem.2016.10.054>.
86. Xiao, G.; Zhang, X.; Zhang, W.; Zhang, S.; Su, H.; Tan, T. Visible-light-mediated synergistic photocatalytic antimicrobial effects and mechanism of Ag-nanoparticles@chitosan–TiO₂ organic–inorganic composites for water disinfection. *Appl. Catal., B*, **2015**, *170–171*, 255–262.
87. Br. Pat., Nanoparticulate titanium dioxide nanomaterial modified with functional groups and with citric extracts adsorbed on the surface, for the removal of a wide range of microorganisms. EP 3012225A1, 27 April 2016. Available online: <https://patentimages.storage.googleapis.com/47/ec/07/027b1717b2d7f5/EP3012225A1.pdf> (accessed on 16 April 2021).
88. Cheng, L.; Hao, Y.; Song, X.; Chang, Q. Effects of chitosan and nano titanium dioxide on the mechanical, physicochemical and antibacterial properties of corn starch films. *J. Macromol. Sci. Part B* **2021**, 1–15, <https://doi.org/10.1080/00222348.2021.1889123>.
89. Hosseinzadeh, S.; Partovi, R.; Talebi, F.; Babaei, A. Chitosan/TiO₂ nanoparticle/ Cymbopogon citratus essential oil film as food packaging material: Physico-mechanical properties and its effects on microbial, chemical, and organoleptic quality of minced meat during refrigeration. *J. Food Process. Preserv.* **2020**, *44*, <https://doi.org/10.1111/jfpp.14536>.
90. Sami, R.; Elhakem, A.; Alharbi, M.; Benajiba, N.; AlMatrafi, M.; Jing, J.; Helal, M. Effect of titanium dioxide nanocomposite material and antimicrobial agents on mushrooms shelf-life preservation. *Processes* **2020**, *8*, 1632, <https://doi.org/10.3390/pr8121632>.
91. Zink, D.L. The impact of consumer demands and trends on food processing. *Emerg. Infect. Dis.* **1997**, *3*, 467–469, <https://doi.org/10.3201/eid0304.970408>.
92. Berardinelli, A.; Parisi, F. TiO₂ in the food industry and cosmetics. In *Titanium Dioxide (TiO₂) and Its Applications*, ed. F. Parrino and L. Palmisano, Elsevier: Amsterdam, The Netherlands, 2021.
93. Peters, R.J.B.; van Bommel, G.; Herrera-Rivera, Z.; Helsper, H.P.F.G.; Marvin, H.J.P.; Weigel, S.; Tromp, P.C.; Oomen, A.G.; Rietveld, A.G.; Bouwmeester, H. Characterization of titanium dioxide nanoparticles in food products: analytical methods to define nanoparticles. *J. Agric. Food Chem.* **2014**, *62*, 6285–6293, <https://doi.org/10.1021/jf5011885>.
94. Li, B.; Chua, S.L.; Yu, D.; Chan, S.H.; Li, A. Separation and size characterization of highly polydisperse titanium dioxide nanoparticles (E171) in powdered beverages by using Asymmetric Flow Field-Flow Fractionation hyphenated with Multi-Angle Light Scattering and Inductively Coupled Plasma Mass Spectrometry. *J. Chromatogr. A* **2021**, *1643*, 462059, <https://doi.org/10.1016/j.chroma.2021.462059>.
95. Weir, A.; Westerhoff, P.; Fabricius, L.; Hristovski, K.; von Goetz, N. Titanium dioxide nanoparticles in food and personal care products. *Environ. Sci. Technol.* **2012**, *46*, 2242–2250, <https://doi.org/10.1021/es204168d>.
96. Winkler, H.C.; Notter, T.; Meyer, U.; Naegeli, H. Critical review of the safety assessment of titanium dioxide additives in food. *J. Nanobiotechnol.*, **2018**, *16*, 51.
97. Ropers, M.-H.; Terrisse, H.; Mercier-Bonin, M.; Humbert, B. Titanium dioxide as food additive. In *Application of Titanium dioxide*, ed. M. Janus, InTech, Croatia, 2017, pp. 3–4, doi:10.5772/intechopen.68883.
98. EFSA ANS Panel, Re-evaluation of titanium dioxide (E 171) as a food additive. *EFSA J.*, **2016**, *14*, e04545.
99. Boutillier, S.; Fourmentin, S.; Laperche, B. Food additives and the future of health: An analysis of the ongoing controversy on titanium dioxide. *Futures* **2020**, *122*, 102598, <https://doi.org/10.1016/j.futures.2020.102598>.

100. EFSA Panel on Food Additives and Flavourings; Younes, M.; Aquilina, G.; Castle, L.; Engel, K.-H.; Fowler, P.; Frutos Fernandez, M.J.; Fürst, P.; Gundert-Remy, U.; Gürtler, R. et al. Safety assessment of titanium dioxide (E171) as a food additive. *EFSA J.*, **2021**, *19*, e06585, .
101. Moran, C.A.; Mullick, F.G.; Ishak, K.G.; Johnson, F.B.; Hummer, W.B. Identification of titanium in human tissues: probable role in pathologic processes. *Hum. Pathol.*, **1991**, *22*, 450–454.
102. Garabrant, D.H.; Fine, L.J.; Oliver, C.; Bernstein, L.; Peters, J.M. Abnormalities of pulmonary function and pleural disease among titanium metal production workers.. *Scand. J. Work. Environ. Heal.* **1987**, *13*, 47–51, <https://doi.org/10.5271/sjweh.2087>.
103. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans, *IARC Monogr. Eval. Carcinog. Risks. Hum.*, **2010**, *93*, 1–413.
104. Baan, R.; Straif, K.; Grosse, Y.; Lauby-Secretan, B.; El Ghissassi, F.; Coglian, V.; WHO International Agency for Research on Cancer Monograph Working Group. Carcinogenicity of carbon black, titanium dioxide, and talc. *Lancet Oncol.* **2006**, *7*, 295–296, [https://doi.org/10.1016/s1470-2045\(06\)70651-9](https://doi.org/10.1016/s1470-2045(06)70651-9).
105. Bermudez, E.; Mangum, J.B.; Asgharian, B.; Wong, B.A.; Reverdy, E.E.; Janszen, D.B.; Hext, P.M.; Warheit, D.B.; Everitt, J.I. Long-term pulmonary responses of three laboratory rodent species to subchronic inhalation of pigmentary titanium dioxide particles. *Toxicol. Sci.* **2002**, *70*, 86–97, <https://doi.org/10.1093/toxsci/70.1.86>.
106. Chen, H.-W.; Su, S.; Chien, C.; Lin, W.; Yu, S.-L.; Chou, C.; Chen, J.J.; Yang, P. Titanium dioxide nanoparticles induce emphysema-like lung injury in mice. *FASEB J.* **2006**, *20*, 2393–2395, <https://doi.org/10.1096/fj.06-6485fje>.
107. Ma, Y.; Guo, Y.; Ye, H.; Huang, K.; Lv, Z.; Ke, Y. Different effects of titanium dioxide nanoparticles instillation in young and adult mice on DNA methylation related with lung inflammation and fibrosis. *Ecotoxicol. Environ. Saf.* **2019**, *176*, 1–10, <https://doi.org/10.1016/j.ecoenv.2019.03.055>.
108. Warheit, D.B. How to measure hazards/risks following exposures to nanoscale or pigment-grade titanium dioxide particles. *Toxicol. Lett.* **2013**, *220*, 193–204, <https://doi.org/10.1016/j.toxlet.2013.04.002>.
109. Boffetta, P.; Soutar, A.; Cherrie, J.; Granath, F.; Andersen, A.; Anttila, A.; Blettner, M.; Gaborieau, V.; Klug, S.J.; Langard, S.; et al. Mortality among workers employed in the titanium dioxide production industry in Europe. *Cancer Causes Control* **2004**, *15*, 697–706, <https://doi.org/10.1023/b:caco.0000036188.23970.22>.
110. Medina-Reyes, E.I.; Delgado-Buenrostro, N.L.; Díaz-Urbina, D.; Rodríguez-Ibarra, C.; Déciga-Alcaraz, A.; González, M.I.; Reyes, J.L.; Villamar-Duque, T.E.; Flores-Sánchez, M.L.; Hernández-Pando, R.; et al. Food-grade titanium dioxide (E171) induces anxiety, adenomas in colon and goblet cells hyperplasia in a regular diet model and microvesicular steatosis in a high fat diet model. *Food Chem. Toxicol.* **2020**, *146*, 111786, <https://doi.org/10.1016/j.fct.2020.111786>.
111. Pele, L.C.; Thoree, V.; A Bruggraber, S.F.; Koller, D.; Thompson, R.P.H.; Lomer, M.; Powell, J.J. Pharmaceutical/food grade titanium dioxide particles are absorbed into the bloodstream of human volunteers. *Part. Fibre Toxicol.* **2015**, *12*, 1–6, <https://doi.org/10.1186/s12989-015-0101-9>.
112. Wang, J.; Zhou, G.; Chen, C.; Yu, H.; Wang, T.; Ma, Y.; Jia, G.; Gao, Y.; Li, B.; Sun, J.; Li, Y.; Jiao, F.; Zhao, Y.; Chai, Z. Acute toxicity and biodistribution of different sized titanium dioxide particles in mice after oral administration. *Toxicol. Lett.*, **2007**, *168*, 176–185.
113. Talamini, L.; Gimondi, S.; Violatto, M.B.; Fiordaliso, F.; Pedica, F.; Tran, N.L.; Sitia, G.; Aureli, F.; Raggi, A.; Nelissen, I.; et al. Repeated administration of the food additive E171 to mice results in accumulation in intestine and liver and promotes an inflammatory status. *Nanotoxicology* **2019**, *13*, 1087–1101, <https://doi.org/10.1080/17435390.2019.1640910>.
114. Urrutia-Ortega, I.M.; Garduño-Balderas, L.G.; Delgado Buenrostro, N.L.; Freyre-Fonseca, V.; Flores-Flores, J.O.; González Robles, R.; Pedraza-Chaverri, J.; Hernández-Pando, R.; Rodríguez Sosa, R.; León-Cabrera, S. et al. Food-grade titanium dioxide exposure exacerbates tumor formation in colitis associated cancer model. *Food Chem. Toxicol.*, **2016**, *93*, 20–31.
115. Gim-Krumm, M.; Donoso, P.; Zuñiga, R.N.; Estay, H.; Troncoso, E. A comprehensive study of glucose transfer in the human small intestine using an in vitro intestinal digestion system (i-IDS) based on a dialysis membrane process. *J. Membr. Sci.* **2018**, *564*, 700–711, <https://doi.org/10.1016/j.memsci.2018.07.080>.
116. Dorier, M.; Béal, D.; Marie-Desvergne, C.; Dubosson, M.; Barreau, F.; Houdeau, E.; Herlin-Boime, N.; Carriere, M. Continuous in vitro exposure of intestinal epithelial cells to E171 food additive causes oxidative stress, inducing oxidation of DNA bases but no endoplasmic reticulum stress. *Nanotoxicology*, **2017**, *11*, 751–761.
117. Dorier, M.; Tisseyre, C.; Dussert, F.; Béal, D.; Arnal, M.-E.; Douki, T.; Valdiglesias, V.; Laffon, B.; Fraga, S.; Brandão, F.; et al. Toxicological impact of acute exposure to E171 food additive and TiO2 nanoparticles on a co-culture of Caco-2 and HT29-MTX intestinal cells. *Mutat. Res. Toxicol. Environ. Mutagen.* **2018**, *845*, 402980, <https://doi.org/10.1016/j.mrgentox.2018.11.004>.
118. Hoet, P.H.M.; Brüske-Hohlfeld, I.; Salata, O.V. Nanoparticles—known and unknown health risks. *J. Nanobiotechnol.*, **2004**, *2*, 12.
119. Nonni, J. Medical makeup: the correction of hyperpigmentation disorders. **2012**, *139*, S170–S176, [https://doi.org/10.1016/s0151-9638\(12\)70131-x](https://doi.org/10.1016/s0151-9638(12)70131-x).
120. Dunford, R.; Salinaro, A.; Cai, L.; Serpone, N.; Horikoshi, S.; Hidaka, H.; Knowland, J. Chemical oxidation and DNA damage catalysed by inorganic sunscreen ingredients. *FEBS Lett.* **1997**, *418*, 87–90, [https://doi.org/10.1016/s0014-5793\(97\)01356-2](https://doi.org/10.1016/s0014-5793(97)01356-2).
121. Skocaj, M.; Filipic, M.; Petkovic, J.; Novak, S. Titanium dioxide in our everyday life; is it safe? *Radiol. Oncol.* **2011**, *45*, 227–247, <https://doi.org/10.2478/v10019-011-0037-0>.

122. Sadrieh, N.; Wokovich, A.M.; Gopee, N.V.; Zheng, J.; Haines, D.; Parmiter, D.; Siitonen, P.H.; Cozart, C.R.; Patri, A.R.K.; McNeil, S.E.; Howard, P.C.; Doub, W.H.; Buhse, L.F. Lack of Significant dermal penetration of titanium dioxide from sunscreen formulations containing nano- and submicron-size TiO₂ particles. *Toxicol. Sci.*, **2010**, *115*, 156–166.
123. Hidaka, H.; Horikoshi, S.; Serpone, N.; Knowland, J. In vitro photochemical damage to AND, ARN and their bases by an inorganic sunscreen agent on exposure UVA and UVB radiation. *J. Photochem. Photobiol. A: Chem.* **1997**, *111*, 205–213, [https://doi.org/10.1016/s1010-6030\(97\)00229-3](https://doi.org/10.1016/s1010-6030(97)00229-3).
124. Tan, M.-H.; A Commens, C.; Burnett, L.; Snitch, P.J. A pilot study on the percutaneous absorption of microfine titanium dioxide from sunscreens. *Australas. J. Dermatol.* **1996**, *37*, 185–187, <https://doi.org/10.1111/j.1440-0960.1996.tb01050.x>.
125. Ling, C.; An, H.; Li, L.; Wang, J.; Lu, T.; Wang, H.; Hu, Y.; Song, G.; Liu, S. Genotoxicity evaluation of titanium dioxide nanoparticles in vitro: a systematic review of the literature and meta-analysis. *Biol. Trace Elem. Res.*, **2021**, *199*, 2057–2076.
126. Charles, S.; Jomini, S.; Fessard, V.; Bigorgne-Vizade, E.; Rousselle, C.; Michel, C. Assessment of the *in vitro* genotoxicity of TiO₂ nanoparticles in a regulatory context. *Nanotoxicology* **2018**, *12*, 357–374, <https://doi.org/10.1080/17435390.2018.1451567>.
127. Karlsson, H.L.; Di Bucchianico, S.; Collins, A.; Dusinska, M. Can the comet assay be used reliably to detect nanoparticle-induced genotoxicity?. *Environ. Mol. Mutagen.* **2014**, *56*, 82–96, <https://doi.org/10.1002/em.21933>.
128. Doak, S.; Griffiths, S.; Manshian, B.; Singh, N.; Williams, P.; Brown, A.; Jenkins, G. Confounding experimental considerations in nanogenotoxicology. *Mutagenesis* **2009**, *24*, 285–293, <https://doi.org/10.1093/mutage/geb010>.
129. Kroll, A.; Pillukat, M.H.; Hahn, D.; Schnekenburger, J. Interference of engineered nanoparticles with in vitro toxicity assays. *Arch. Toxicol.* **2012**, *86*, 1123–1136, <https://doi.org/10.1007/s00204-012-0837-z>.
130. Kain, J.; Karlsson, H.L.; Moller, L. DNA damage induced by micro- and nanoparticles—interaction with FPG influences the detection of DNA oxidation in the comet assay. *Mutagenesis* **2012**, *27*, 491–500, <https://doi.org/10.1093/mutage/ges010>.
131. Wang, H.; Joseph, J.A. Quantifying cellular oxidative stress by dichlorofluorescein assay using microplate reader. *Free Radic. Biol. Med.* **1999**, *27*, 612–616, doi:10.1016/s0891-5849(99)00107-0.
132. Aranda, A.; Sequedo, L.; Tolosa, L.; Quintas, G.; Burello, E.; Castell, J.; Gombau, L. Dichloro-dihydro-fluorescein diacetate (DCFH-DA) assay: A quantitative method for oxidative stress assessment of nanoparticle-treated cells. *Toxicol. Vitro.* **2013**, *27*, 954–963, <https://doi.org/10.1016/j.tiv.2013.01.016>.
133. Guadagnini, R.; Halamoda Kenzaoui, B.; Walker, L.; Pojana, G.; Magdolenova, Z.; Bilanicova, D.; Saunders, M.; Juillerat-Jeanerret, L.; Marcomini, A.; Huk, A.; et al. Toxicity screenings of nanomaterials: challenges due to interference with assay processes and components of classicin vitro tests. *Nanotoxicology* **2015**, *9*, 13–24, <https://doi.org/10.3109/17435390.2013.829590>.
134. Liu, Y.; Peterson, D.A.; Kimura, H.; Schubert, D. Mechanism of Cellular 3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide (MTT) Reduction. *J. Neurochem.* **2002**, *69*, 581–593, <https://doi.org/10.1046/j.1471-4159.1997.69020581.x>.
135. Bickel, M. The role of interleukin-8 in inflammation and mechanisms of regulation. **1993**, *64*.
136. Repetto, G.; del Peso, A.; Zurita, J.L. Neutral red uptake assay for the estimation of cell viability/cytotoxicity. *Nat. Protoc.* **2008**, *3*, 1125–1131. <https://doi.org/10.1038/nprot.2008.75>.
137. Thirunavukkarasu, G.K.; Bacova, J.; Monfort, O.; Dworniczek, E.; Paluch, E.; Hanif, M.B.; Rauf, S.; Motlochova, M.; Capek, J.; Hensel, K.; et al. Critical comparison of aerogel TiO₂ and P25 nanopowders: Cytotoxic properties, photocatalytic activity and photoinduced antimicrobial/antibiofilm performance. *Appl. Surf. Sci.* **2021**, *579*, 152145, <https://doi.org/10.1016/j.apusc.2021.152145>.
138. Gea, M.; Bonetta, S.; Iannarelli, L.; Giovannozzi, A.M.; Maurino, V.; Bonetta, S.; Hodoroaba, V.-D.; Armato, C.; Rossi, A.M.; Schilirò, T. Shape-engineered titanium dioxide nanoparticles (TiO₂-NPs): cytotoxicity and genotoxicity in bronchial epithelial cells. *Food Chem. Toxicol.* **2019**, *127*, 89–100, <https://doi.org/10.1016/j.fct.2019.02.043>.
139. Di Bucchianico, S.; Cappellini, F.; Le Bihanic, F.; Zhang, Y.; Dreij, K.; Karlsson, H.L. Genotoxicity of TiO₂ nanoparticles assessed by mini-gel comet assay and micronucleus scoring with flow cytometry. *Mutagenesis*, **2017**, *32*, 127–137.
140. Borenfreund, E.; Babich, H.; Martin-Alguacil, N. Comparisons of two in vitro cytotoxicity assays—The neutral red (NR) and tetrazolium MTT tests. *Toxicol. in Vitro*, **1988**, *2*, 1–6.
141. Swinehart, D.F. The Beer-Lambert law. *J. Chem. Educ.*, **1962**, *39*, 333.
142. Pearce, J.M. Digital designs and scientific hardware. In *Open-Source Lab*, ed. J. M. Pearce, Elsevier, Boston, MA, USA, 2014, pp. 165–252.
143. Wang, J.; Fan, Y. Lung injury induced by tio₂ nanoparticles depends on their structural features: Size, shape, crystal phases, and surface coating. *Int. J. Mol. Sci.* **2014**, *15*, 22258–22278, <https://doi.org/10.3390/ijms151222258>.