



Article Taxonomy and Phylogenetic Relationships of *Clavulinopsis* (Clavariaceae, Agaricales): Description of Six New Species and One Newly Recorded Species from China

Jun Yan ¹^(D), Jing Wen ¹, Gui-Wu Li ¹, Shao-Wu Wu ² and Ping Zhang ^{1,*}

- ¹ College of Life Sciences, Hunan Normal University, Changsha 410006, China; alexis830@163.com (J.Y.); 18163799060@163.com (J.W.); liguiwu201909@163.com (G.-W.L.)
- ² Bureau of Forestry, Tongdao Dong Autonomous County, Huaihua 418500, China
- * Correspondence: zhangping0000@163.net

Abstract: Specimens of *Clavulinopsis* (Clavariaceae, Agaricales) collected in China were studied using morphological and molecular methods. Six species—*C. aspersa, C. bicolor, C. bispora, C. erubescens, C. incarnata,* and *C. tropicalis*—are described as new to science, and *C. trigonospora* is a newly recorded species in China. Phylogenetic analysis was conducted based on a combined dataset of internal transcribed spacer and nuclear ribosomal RNA large subunit sequences. The phylogenetic reconstruction revealed that the six new species each formed an independent lineage, and the samples of *C. trigonospora* from China were nested with accessions of *C. trigonospora* collected from Italy. The morphology of the seven Chinese species is described in detail, and is illustrated with line drawings and photographs. A key to the known *Clavulinopsis* species in China is provided.

Keywords: clavarioid fungi; Clavariaceae; phylogenetic analysis; morphology; taxonomy



Citation: Yan, J.; Wen, J.; Li, G.-W.; Wu, S.-W.; Zhang, P. Taxonomy and Phylogenetic Relationships of *Clavulinopsis* (Clavariaceae, Agaricales): Description of Six New Species and One Newly Recorded Species from China. *J. Fungi* **2023**, *9*, 656. https://doi.org/10.3390/ jof9060656

Academic Editors: Xinlei Fan, Jadson Diogo Pereira Bezerra and Sajeewa Maharachchikumbura

Received: 15 April 2023 Revised: 25 May 2023 Accepted: 8 June 2023 Published: 12 June 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

1. Introduction

The genus Clavulinopsis Overeem comprises 34 species of coral fungi distributed worldwide [1] and classified in the Clavariaceae (Agaricales, Basidiomycota). Among the 165 taxon records for the genus currently listed in the Index Fungorum database (http://www.indexfungorum.org; accessed on 15 April 2023), approximately 70 species names are legitimately published. However, the majority of taxa lack molecular evidence and some even lack adequate micromorphological data. For example, Clavulinopsis coliformis (Boud.) Corner was transferred to Clavulinopsis by E.J.H. Corner in 1950, but E.J.H. Corner considered that C. coliformis might only be an unusual state of a well-known species, such as C. corniculata (Schaeff.) Corner or C. fusiformis (Sowerby) Corner [2]. Similarly, the taxonomic placement of other *Clavulinopsis* species, such as *C. pusilla* (Coker) Corner, C. subflava (Britzelm.) Corner, C. sulphurascens (Schwein.) Corner, and C. tenella (Boud.) Corner, is dubious [2]. In addition, the relationships among *Clavaria* Vaill. ex L., Clavulinopsis, and Ramariopsis (Donk) Corner, which are classified in the same family, have historically been controversial. Considering macromorphological evidence, the basidiome of the species placed in *Clavaria* and *Clavulinopsis* are simple or branched, clavarioid, and variously colored, whereas those of *Ramariopsis* species are rarely simple [2,3]. Based on micromorphological data, R.H. Petersen noted that Clavaria and Clavulinopsis are not separable at the genus rank because clamp connections are present on the basidia and hyphae in Clavulinopsis, but this is the sole character used to distinguish the genus from Clavaria. With regard to Ramariopsis, R.H. Petersen suggested that the genus must be segregated from *Clavaria* and *Clavulinopsis* on account of its branched fruit bodies and echinulate spores [4].

With the increase in taxonomic means, the boundaries between these genera are gradually being resolved. D.N. Pegler and T.W.K. Young [5] used scanning electron microscopy to examine the basidiospore structure in *Clavulinopsis* and *Ramariopsis* and observed that, among the species examined, most *Clavulinopsis* species were smooth-spored except for *Clavulinopsis helvola* (Pers.) Corner, whereas no species of *Ramariopsis* was smooth-spored. Furthermore, the mode of formation of basidiospore ornamentation was entirely different between *Ramariopsis* species and *Clavulinopsis* species. Birkebak et al. [6] conducted a molecular phylogenetic analysis to clarify the phylogenetic resolution of *Clavaria, Clavulinopsis*, and *Ramariopsis*. Thus, the status of *Clavulinopsis* as a distinct genus has been clarified, and knowledge of the similarities and differences in *Clavulinopsis* from related genera is much improved.

Reports on the taxonomic diversity in *Clavulinopsis* have been extremely limited in the past decade, with only four new species formally described (*C. aurantiaca* Araujo-Neta, G.A. Silva & Gibertoni, *C. dimorphica* A.N.M. Furtado & M.A. Neves, *C. imperata* A.N.M. Furtado & M.A. Neves, and *C. trigonospora* Franchi & M. Marchetti) [7–9]. Moreover, the sequence data generated for the *Clavulinopsis* species remain limited in quantity and coverage. In the present study, six new species and one newly recorded species of *Clavulinopsis* in China are described based on morphological examination and molecular phylogenetic analysis. Thirty-four newly generated sequences (seventeen ITS and seventeen nrLSU) for these seven *Clavulinopsis* species have been deposited in GenBank.

2. Materials and Methods

2.1. Specimens

Seventeen specimens of *Clavulinopsis* were collected from Hainan, Hunan, Jilin, Shaanxi, Sichuan, and Yunnan provinces in China from 2014 to 2022. The fresh specimens were dried using heat or indicating silica gel. The dried vouchers were deposited in the Mycological Herbarium of Hunan Normal University (MHHNU), Changsha, China. Detailed information on the 17 specimens and other samples used in the phylogenetic analysis are provided in Table 1, including GenBank accession numbers and locality information.

2.2. DNA Extraction, Amplification, and Sequencing

Total genomic DNA was extracted from the dried samples using the Ezup Column Fungi Genomic DNA Purification Kit (Sangon Biotech, Shanghai, China) in accordance with the manufacturer's instructions. The primer pair ITS4/ITS5 [10] was used to amplify the internal transcribed spacer (ITS) region, and the universal primers LR0R, LR3, and LR5 [11,12] were used to amplify the nuclear ribosomal large subunit (nrLSU) region. Each PCR amplification was performed with an Eppendorf Mastercycler thermal cycler (Eppendorf Inc., Hamburg, Germany) in a 25 μ L reaction mixture, which contained 1× PCR buffer, 1.5 mM MgCl₂, 0.2 mM dNTPs, 0.4 μ M of each primer, 1.25 U Taq polymerase (Sangon Biotech, Shanghai, China), and 1 μ L DNA template. The thermal cycling was performed as follows: initial denaturation at 94 °C for 4 min, then 34 cycles of 94 °C for 40 s, annealing at 55 °C for 40 s, extension at 72 °C for 1 min, and a final extension at 72 °C for 8 min [13]. All amplified PCR products were electrophoresed on 1% agarose gel, and the purified PCR products were sequenced by Sangon Biotech (Shanghai, China). The specified primers were also used for sequencing reactions. All sequences newly generated in this study were submitted to GenBank.

2.3. Morphological Studies

The macromorphological characters of species were based on field notes and habitat photographs. Color codes used in descriptions follow Kornerup and Wanscher [14], and color terms are from Ridgway [15]. The micromorphological characters were observed with a light microscope. Tissue sections of dried materials were first rehydrated with 5% KOH solution or distilled water, and then stained with 1% Congo Red when necessary. The notation [n/m/p] means that *n* basidiospores were measured from *m* basidiomes of *p* specimens. The basidiospore dimensions are described using the notation $(\mathbf{a}-)\mathbf{b}-\mathbf{c}(-\mathbf{d})$. The range $\mathbf{b}-\mathbf{c}$ contains a minimum of 90% of the measured values, and \mathbf{a} and \mathbf{d} are extreme

values, provided in parentheses. The *Q* value is the length/width ratio of each basidiospore, and the **Q** value is the average $Q \pm$ standard deviation.

2.4. Alignment and Phylogenetic Analyses

Sequence data for the legitimate species of the genus *Clavulinopsis* and two *Mucronella* species were downloaded from GenBank. The downloaded and newly generated sequences constituted the dataset for the present analysis. The ITS and nrLSU sequences were, respectively, aligned using the default settings for gap openings and gap extension penalties with MAFFT v7.471 [16], and then manually adjusted where necessary in BIOEDIT v7.2.5 [17]. The concatenated ITS–nrLSU data matrix containing 79 sequences (39 ITS and 40 nrLSU) was assembled with SEQUENCEMATRIX 1.7.8 [18]. A maximum likelihood (ML) analysis was conducted using RAXML v8.0.20 [19] with 1000 bootstrap replicates and the GTR + Gamma evolutionary model. Bayesian inference (BI) was performed with MRBAYES v3.2.7 [20] and run for 1,000,000 generations with the GTR + I + G optimal evolutionary model selected with MRMODELTEST v2.4 [21] for each partition, using four Markov Chain Monte Carlo (MCMC) chains to calculate posterior probabilities. The tree files were merged and edited with FIGTREE v1.4.2 [22].

Table 1. Voucher information and GenBank accession numbers of taxa used in this study.

Identification	Specimen No.	GenBank No. (ITS)	GenBank No. (28S)	Location	References
Clavulinopsis amoena	PBM3381	_	HQ877702	Australia: Tasmania	Birkebak et al. [6]; Hyde et al. [8]
C. appalachiensis	S.D. Russell iNaturalist # 91596164	OM809324	_	USA: Indiana	Unpublished
C. appalachiensis	TENN074980	MT196965	_	USA: Tennessee	Unpublished
C. aspersa	MHHNU10153	OQ703777	OQ703794	China: Hunan	Present study
C. aspersa	MHHNU10342	OQ703778	OQ703795	China: Hunan	Present study
C. aspersa	MHHNU11103	OQ703779	OQ703796	China: Hunan	Present study
C. aurantiaca	URM <bra>: 84212</bra>	_	KX227749	Brazil: Pernambuco	Hyde et al. [8]
C. aurantiaca	URM <bra>: 84216</bra>	KC348464	NG058946	Brazil: Pernambuco	Hyde et al. [8]
C. aff. auranti- ocinnabarina	JMB08171004	—	HQ877704	USA: Tennessee	Birkebak et al. [6]; Hyde et al. [8]
C. aff. auranti- ocinnabarina	JMB08240901	—	HQ877703	USA: Tennessee	Birkebak et al. [6]; Hyde et al. [8]
C. bicolor	MHHNU10381	OQ703780	OQ703797	China: Hainan	Present study
C. bispora	MHHNU11181	OQ703781	OQ703798	China: Jilin	Present study
C. bispora	MHHNU11188	OQ703782	OQ703799	China: Jilin	Present study
C. corallinorosacea	PBM3380	KP257144	HQ877707	Australia: Tasmania	Birkebak et al. [6,23]; Hyde et al. [8]
C. corniculata	TENN064106	KP257145	HQ877713	USA: Tennessee	Birkebak et al. [6,23]; Hyde et al. [8]
C. corniculata f. bispora	AMB 18573	MT055953	—	Italy	Franchi and M. Marchetti [9]
C. erubescens	MHHNU8040	OQ703783	OQ703800	China: Hunan	Present study
C. erubescens	MHHNU10290	OQ703784	OQ703801	China: Shaanxi	Present study
C. fusiformis	HKAS122627	ON794403	_	China	Wang et al. [24]
C. fusiformis	PBM 2804	—	EF535273	USA: Massachusetts	Birkebak et al. [6]; Hyde et al. [8]
C. fusiformis	TENN064110	_	HQ877717	USA: Tennessee	Birkebak et al. [6]; Hyde et al. [8]

_

Identification	Specimen No.	GenBank No. (ITS)	GenBank No. (28S)	Location	References
C. fusiformis	2728		KM248914	_	Unpublished
C. gracillima	MO 215748	KY706170		Canada: Windsor	Hay et al. [25]
C. gracillima	TENN065662	—	HQ877708	USA: Tennessee	Unpublished
C. helvola	EL 111/04	EU118617	EU118617	Sweden	Birkebak et al. [6]; Hyde et al. [8]
C. helvola	Lueck6	KP965770	KP965788	Germany: Lueckendorf	Karich et al. [26]
C. incarnata C. incarnata C. incarnata C. incarnata	MHHNU9314 MHHNU9813 MHHNU11330 MHHNU11331	OQ703785 OQ703786 OQ703787 OQ703788	OQ703802 OQ703803 OQ703804 OQ703805	China: Hunan China: Yunnan China: Yunnan China: Yunnan	Present study Present study Present study Present study
C. laeticolor	EL 8/00	EU118618	EU118618	Finland	Birkebak et al. [6]; Hyde et al. [8]
C. luteoalba	BRACR16669	—	JQ415959	Denmark: Copenhagen	Hyde et al. [8]
C. luteoalba	BSI13 147a	OP538704	—	Switzerland	Unpublished
C. miyabeana	ZP-2118	MK427059	_	China: Hunan	Chen and Zhang, P. [27]
<i>C</i> . sp.	MCCNNU 00948	MT587808	MT587810	China	Unpublished
<i>C.</i> sp.	MCCNNU 00952	MT587809	MT587811	China	Unpublished
C. sulcata	PBM3379	—	HQ877709	Australia: Tasmania	Birkebak et al. [6]; Hyde et al. [8]
C. sulcata	PDD78241	—	DQ284904	New Zealand	Dentinger, B.T.M. and McLaughlin, D.J. [28]
C. trigonospora	AMB: 18557	NR176720	NG088120	Italy: Capezzano Pianore	Franchi and M. Marchetti [9]
C. trigonospora	AMB: 18587	—	MT055968	Italy: Capezzano Pianore	Franchi and M. Marchetti [9]
C. trigonospora	MHHNU9186	OQ703789	OQ703806	China: Sichuan	Present study
C. trigonospora	MHHNU9200	OQ703790	OQ703807	China: Sichuan	Present study
C. trigonospora	MHHNU10198	OQ703791	OQ703808	China: Gansu	Present study
C. tropicalis	MHHNU10721	OQ703792	OQ703809	China: Hainan	Present study
C. tropicalis	MHHNU10722	OQ703793	OQ703810	China: Hainan	Present study
C. umbrinella	HFRG_EJ191214_3_FRD 17588114	^{3I} OQ133539	OQ133591	United Kingdom: Hampshire	Unpublished
Ramariopsis laeticolor	CR12764	—	GU299509	Slovakia	Unpublished
R. laeticolor	UBC F23885	KJ146701	—	Canada: British Columbia Area	Unpublished
Mucronella flava	IO.16.84	MT232354	MT232307	Sweden	Olariaga et al. [29]; Yan et al. [30]
<i>Mucronella</i> sp.	PDD95742	HQ533013	—	New Zealand	Yan et al. [30,31]

Table 1. Cont.

Note: Newly generated sequences are shown in bold.

3. Results

3.1. Taxonomy

Clavulinopsis aspersa P. Zhang & Jun Yan, sp. nov.: Figures 1 and 2. MycoBank: 848958

Diagnosis: Characterized by its solitary or scattered habit, yellowish to yellow basidiomata, and smooth, thin-walled, broadly ellipsoid to ellipsoid basidiospores.

Etymology: *aspersa* (Lat.) refers to the scattered growth habit of this species.

Type: China. Hunan Province: Rucheng County, Jiulongjiang National Forest Park, 25°26′44.01″ N, 113°47′09.58″ E, alt. 500 m, in broadleaved forest, 22 June 2020, Ping Zhang (MHHNU10342, holotype).

Description: Basidiomata fragile, simple, 15–50 mm tall, 1–4 mm wide, solitary or scattered to gregarious, rarely caespitose-connate at the base. Fertile part claviform or subcylindric to fusiform, sometimes slightly curved or flexuous, occasionally longitudinal depressions or grooves when old, yellowish [1A3–4, Matius Yellow, Picric Yellow, Pale Greenish Yellow] to yellow [2A7–8, Apricot Yellow, Light Cadmium] with age. Apex rounded, concolorous, becoming yellow [2A7–8, Light Cadmium] to dark yellow [4A7–8, Aniline Yellow]. Sterile part narrow or indistinct, concolorous or slightly paler, sometimes semitransparent, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous. Taste, odor, and macrochemical reactions were not recorded.

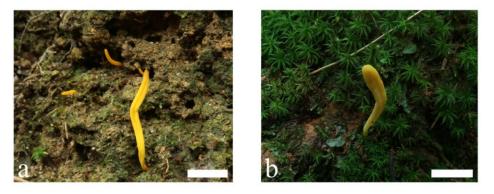


Figure 1. Basidiomata of *Clavulinopsis aspersa* (a) MHHNU10342; (b) MHHNU11103. Scale bars = 2 cm.

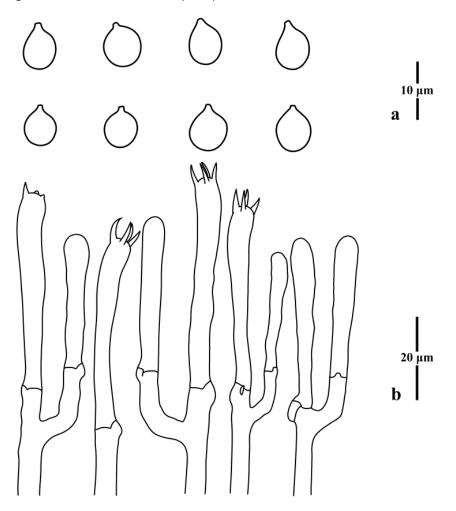


Figure 2. Microscopic features of Clavulinopsis aspersa (MHHNU10342). (a) Basidiospores; (b) basidia.

Basidiospores [100/4/3] (6.0) 6.5–8.0 (8.5) × 5.0–6.0 µm [Q = 1.17-1.40 (1.50), **Q** = 1.26 ± 0.08], thin-walled, hyaline, smooth, inamyloid, broadly ellipsoid to ellipsoid with a distinct apiculus. Basidia (36) 42–56 × 6.0–8.0 µm, thin-walled, hyaline, clavate to subcylindrical, with a clamp connection, four tapered sterigmata, 3.5–7.0 µm long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 1.5–5.0 µm wide; hyphae distant from subhymenium 8.0–10.0 µm wide.

Habitat and distribution: Solitary or scattered to gregarious, on soil rich in humus, in broadleaved forest, known only in subtropical areas of China; June to July.

Additional specimens examined: China. Hunan Province: Guzhang County, Laoyapo, 28°41′13.69″ N, 110°05′20.16″ E, alt. 953 m, in broadleaved forest, 22 July 2019, Ping Zhang (MHHNU10153); Tongdao County, Fengshuwan Forest Park, 26°09′45.66″ N, 109°46′31.52″ E, alt. 400 m, in broadleaved forest, 6 July 2022, Ping Zhang (MHHNU11103).

Comments: In the genus Clavulinopsis, approximately 10 species are similar to C. aspersa in the basidioma color. Among these species, C. dimorphica and C. fleischeriana (Henn.) Corner, with many-branched basidiomata, are immediately distinguishable [2,7]. Clavulinopsis helvola and C. trigonospora are particularly distinctive in Clavulinopsis because the former species has a spiny spore and the latter has a subtriangular spore [2,9]. According to published data, the Q of spores of C. antillarum (Pat.) Courtec. are 1.0–1.18 [32], and the average Q of spores of C. *imperata* are 1.08 [7], which indicates that C. *aspersa* has significantly narrower spores (Q = 1.17-1.40). The irregularly thick-walled hyphae of C. imperata are unique within Clavulinopsis [7]. Based on spore dimensions, C. luteoalba (Rea) Corner (5.0–8.0 \times 2.5–4.5 µm or 5.2–8.0 \times 2.8–4.4 µm) can be distinguished from C. aspersa [2,9]. Compared with C. aspersa, C. amoena (Zoll. & Moritzi) Corner, C. fusiformis, and C. laeticolor (Berk. & M.A. Curtis) R.H. Petersen often grow more densely and are very variable in form, size, and color. Moreover, C. amoena is found in tropical areas from sea level to 1300 m and has subglobose spores (4.0–7.0 \times 4.0–6.5 µm); C. fusiformis has a taller basidioma (5–14 cm) and slightly thick-walled spores; and C. laeticolor has slightly thick-walled hyphae and spores [2].

Clavulinopsis bicolor P. Zhang & Jun Yan, sp. nov.: Figures 3 and 4.





Figure 3. Basidiomata of *Clavulinopsis bicolor* (a,b) MHHNU10381. Scale bars = 2 cm.

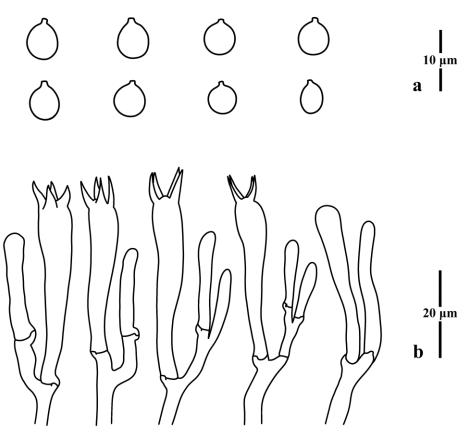


Figure 4. Microscopic features of Clavulinopsis bicolor (MHHNU10381). (a) Basidiospores; (b) basidia.

MycoBank: 848959

Diagnosis: Characterized by a simple, bicolored basidiomata and smooth, thin-walled, globose or subglobose basidiospores.

Etymology: *bicolor* (Lat.) refers to the two different colors of the basidiomata, namely, yellow to brown at the base and white or creamy white to pale green-white above.

Type: China. Hainan Province: Baoting County, Qixianling, 18°42′04.60″ N, 109°41′57.61″ E, alt. 350 m, in tropical broadleaved forest, 26 June 2020, Sainan Li (MHHNU10381, holotype).

Description: Basidiomata fragile, simple, 20–40 mm tall, 1–2 mm wide, scattered to gregarious; the base is divided. Fertile part subcylindric to fusiform, sometimes slightly twisted and with longitudinal depressions, white or creamy white to pale green-white [1A1, 26A2, White, Pale Glaucous-Green, Pale Blue-Green]. When dried, the color changes to pale yellow-white [4A2, Sea-Form Yellow]. Apex obtuse–acute, concolorous, becoming yellow [4A2–3, Martius Yellow]. Sterile part narrow, very distinct, 5–7 mm tall, yellow to brown [2A4–7, Yellowish Citrine, Dark Olive-Buff], without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous. Taste, odor, and macrochemical reactions were not recorded.

Basidiospores [60/2/1] (4.5) 4.7–5.5 (5.7) × (4.0) 4.4–5.0 μ m [Q = 1.04–1.15 (1.25), **Q** = 1.09 \pm 0.05], thin-walled, hyaline, smooth, inamyloid, globose or subglobose, with a distinct apiculus. Basidia (32) 37–46 × (5.0) 5.0–7.0 μ m, thin-walled, hyaline, clavate to subcylindrical, with a clamp connection, four tapered sterigmata, 4.0–8.5 μ m long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 1.0–6.0 μ m wide; hyphae distant from subhymenium 8.0–10.0 μ m wide.

Habitat and distribution: Scattered to gregarious, on soil rich in humus, in tropical broadleaved forest, known only in a tropical area of China; June.

Comments: *C. bicolor* is most difficult to distinguish from *C. appalachiensis* (Coker) Corner and *C. rufipes* (G.F. Atk.) Corner because all have a brown stem and light-colored fertile parts. Based on morphological records, *C. appalachiensis* not only shows a certain difference in color from *C. bicolor*, but also has a larger basidioma ($3.0-9.0 \times 0.15-0.5$ cm), a longer stem (1–4 cm), and broader basidia ($7.0-8.5 \mu$ m) than *C. bicolor*. Compared with *C. appalachiensis*, *C. rufipes* is more similar to *C. bicolor* in macromorphology except that it occasionally has sparingly branched basidiomata. However, *C. rufipes* have pip-shaped to ovoid spores, and shorter basidia ($23-30 \mu$ m) than *C. bicolor* [2].

Clavulinopsis bispora P. Zhang & Jun Yan, sp. nov.: Figures 5 and 6.



Figure 5. Basidiomata of *Clavulinopsis bispora* (a) MHHNU11188; (b) MHHNU11181. Scale bars = 2 cm.

MycoBank: 848960

Diagnosis: Differs from other taxa in the genus by having very small clamp connections and two-spored basidia.

Etymology: bispora (Lat.) refers to the two-spored basidia.

Type: China. Jilin Province: Tonghua, Baijifeng National Forest Park, 41°33′57.68″ N, 126°04′47.00″ E, alt. 680 m, in broadleaved forest, 6 Auguest 2022, Ping Zhang (MHHNU11188, holotype).

Description: Basidiomata fragile, simple, 30–50 mm tall, 1–3 mm wide, gregarious or caespitose. Fertile part subcylindric to claviform, sometimes conspicuously twisted and with a distinct longitudinal depression, yellowish [1A5–6, Lemon Yellow, Empire Yellow] to orange-yellow [1A7–8, Aniline Yellow, Ochraceous Orange] with age. Apex rounded, concolorous, becoming slightly darker when mature. Sterile part narrow, indistinct, concolorous or slightly differing in color from the upper part, occasionally semitranslucent, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous or slightly paler. Taste, odor, and macrochemical reactions were not recorded.

Basidiospores [100/4/2] 6.0–8.0 (8.5) × (4.5) 5.0–6.0 µm [Q = (1.08) 1.17–1.60 (1.70), **Q** = 1.37 ± 0.16], thin-walled, hyaline, smooth, inamyloid, most ellipsoid to broadly ellipsoid, several long ellipsoid or subglobose with a distinct apiculus. Basidia (42) 46–55 × 5.0–7.0 µm, thin-walled, hyaline, clavate to subcylindrical, with a very small clamp connection, two tapered sterigmata, 5.0–8.0 µm long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 2.0–5.0 µm wide; hyphae distant from subhymenium ~12 µm wide.

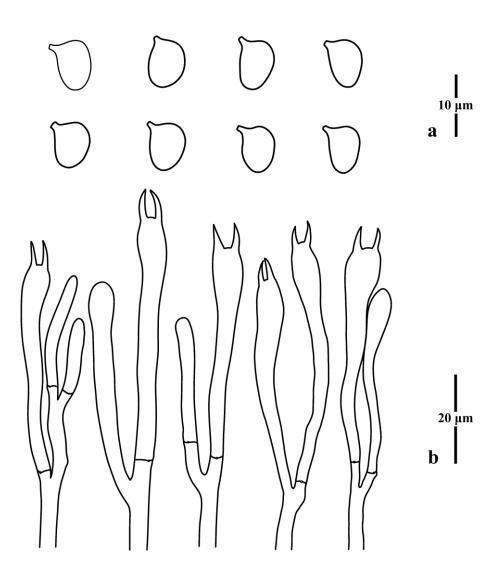


Figure 6. Microscopic features of *Clavulinopsis bispora* (MHHNU11188). (a) Basidiospores; (b) basidia.

Habitat and distribution: Gregarious or caespitose, on soil rich in humus, in broadleaved forests, known only in northeast China; August.

Additional specimen examined: China. Jilin Province: Ji'an County, Wunvfeng National Forest Park, 41°16′19.37″ N, 126°08′14.46″ E, alt. 700 m, in broadleaved forest, 5 Auguest 2022, Ping Zhang (MHHNU11181).

Comments: The characteristics of small clamp connections and two-spored basidia are extremely rare in *Clavulinopsis*, and long sterigmata is also not a typical feature in the genus. According to previous records, *C. calocera* (G.W. Martin) Corner, *C. inflatissima* Corner, *C. lingula* Corner, and *C. sibutiana* (Har. & Pat.) Corner have small clamps [2,3]. However, *C. calocera* was transferred to the genus *Lepidostroma* Mägd. & S. Winkl. [33]. *C. inflatissima* and *C. lingula* have large spores (9.0–11.5 × 8.0–9.0 µm and 10.0–12.0 × 6.0 µm, respectively) and four-spored basidia [3], and *C. sibutiana* is close to *C. lingula* and has been considered a variant of *C. lingula* [3]; thus, these species are easily distinguished from *C. bispora*. With regard to other taxa with two-spored basidia, *C. corniculata* f. *bispora* may produce a profusely branched basidioma, the hyphae and basidia lack clamps [2,9], and *C. luticola* is more specialized on account of its short basidia (8–12 µm long) and flask-shaped cystidia [2].

Clavulinopsis erubescens P. Zhang & Jun Yan, sp. nov.: Figures 7 and 8.





Figure 7. Basidiomata of *Clavulinopsis erubescens* (a) MHHNU8040; (b) MHHNU10290. Scale bars = 2 cm.

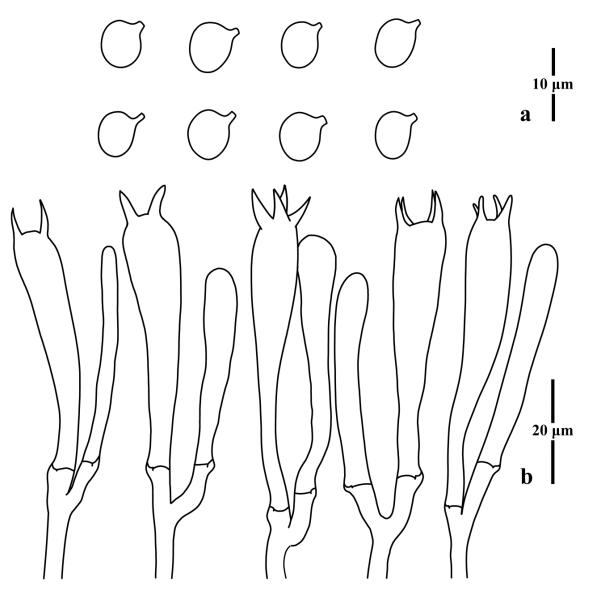


Figure 8. Microscopic features of *Clavulinopsis erubescens* (MHHNU8040). (a) Basidiospores; (b) basidia.

MycoBank: 848961

Diagnosis: Distinguished from other taxa in the genus by its simple red basidiomata, ellipsoid to broadly ellipsoid basidiospores, and two-spored or four-spored basidia.

Etymology: *erubescens* (Lat.) alludes to the basidiomata becoming red with age.

Type: China. Hunan Province: Yuanling County, Fenghuangshan Forest Park, 28°27'10.41" N, 110°25'27.32" E, alt. 150 m, in broadleaved forest, 25 June 2014, Ping Zhang (MHHNU8040, holotype).

Description: Basidiomata fragile, simple, 10–60 mm tall, 1–4 mm wide, gregarious to caespitose. Fertile part subcylindric to fusiform, occasionally slightly curved or flexuous and with a distinct longitudinal depression, red-orange [7A7–8, 7B7–8, Coral Red, Peach Red, Grenadine Red] to red [10A7–8, 10B7–8, Scarlet Red, Rose Doree, Corinthian Red]. Apex rounded or obtuse–acute when mature, concolorous or slightly paler, becoming darker with age. Sterile part narrow, indistinct, concolorous or paler than the upper part, occasionally curved, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous. Taste, odor, and macrochemical reactions were not recorded.

Basidiospores [100/4/2] (5.5) 5.8–7.0 × (4.3) 4.5–5.2 µm [Q = 1.20-1.40 (1.44), $\mathbf{Q} = 1.29 \pm 0.07$], thin-walled, hyaline, smooth, inamyloid, ellipsoid to broadly ellipsoid with a distinct apiculus. Basidia (37) 40–54 × 6–8 µm, thin-walled, hyaline, clavate to subcylindrical, clamped, two or four tapered sterigmata, 4.0–6.0 µm long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 1.0–5.0 µm wide; hyphae distant from subhymenium ~8 µm wide.

Habitat and distribution: Gregarious to caespitose, on soil rich in humus, in broadleaved forests, only known in Hunan and Shaanxi provinces, China; June to October.

Additional specimen examined: China. Shaanxi Province: Lueyang County, Baishigou Village, 33°22′28.74″ N, 106°12′05.14″ E, alt. 900 m, in broadleaved forest, 4 October 2019, Ping Zhang (MHHNU10290).

Comments: Within the genus *Clavulinopsis*, *C. corallinorosacea*, *C. depokensis* (Overeem) Corner, *C. miyabeana* (S. Ito) S. Ito, and *C. sulcata* are similar to *C. erubescens* in having a simple red basidioma. However, compared with *C. erubescens*, *C. corallinorosacea* has a taller, occasionally branched basidioma and narrower basidiospores; *C. depokensis* has four-spored basidia and is specialized among *Clavulinopsis* species in occurring on dead leaves and twigs in woods rather than on soil. Additionally, *C. sulcata* has a variably colored basidioma and globose or subglobose basidiospores [2]. Consistent with the morphological differences, the present phylogenetic analysis confirmed that *C. erubescens* has only a distant affinity with, and is placed in a separate lineage to, *C. corallinorosacea*, *C. miyabeana*, and *C. sulcata*.

Clavulinopsis incarnata P. Zhang & Jun Yan, sp. nov.: Figures 9 and 10.

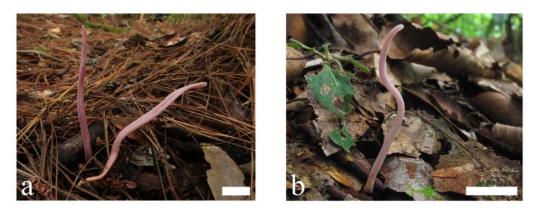


Figure 9. Basidiomata of *Clavulinopsis incarnata* (a) MHHNU11330; (b) MHHNU9314. Scale bars = 2 cm.

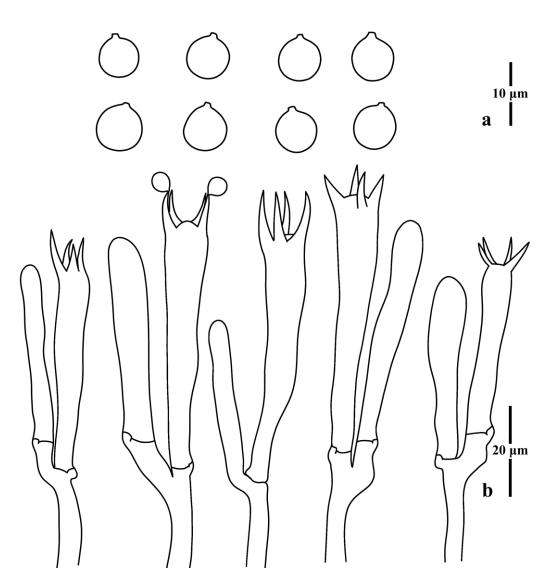


Figure 10. Microscopic features of *Clavulinopsis incarnata* (MHHNU11330). (a) Basidiospores; (b) basidia.

MycoBank: 848962

Diagnosis: Differs from other taxa in the genus in having a pinkish basidioma, fourspored basidia, globose to subglobose basidiospores, and very long sterigmata.

Etymology: incarnata (Lat.) refers to the pinkish basidiomata of this species.

Type: China. Yunnan Province: Binchuan County, Jizu Moutain, 25°57′28.20″ N, 100°23′17.37″ E, alt. 2300 m, in mixed coniferous–broadleaved forest, 29 Auguest 2022, Ping Zhang (MHHNU11330, holotype).

Description: Basidiomata fragile, simple, 30–120 mm tall, 1–7 mm wide, solitary or scattered to gregarious. Fertile part subcylindric to fusiform, sometimes conspicuously twisted and with a distinct longitudinal depression, pinkish [11A2–3, 12A4–5, Hermosa Pink, Eosine Pink Thulite Pink]. Apex rounded or obtuse–acute, obviously paler than the lower part, becoming pale yellow or brown with age. Sterile part narrow, indistinct, concolorous or subtly different in color, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous or slightly paler. Taste, odor, and macrochemical reactions were not recorded.

Basidiospores [100/4/4] 6.0–7.5 (8.0) × 6.0–7.0 (8.0) µm [Q = 1.00-1.09 (1.15), $\mathbf{Q} = 1.05 \pm 0.04$], thin-walled, hyaline, smooth, inamyloid, globose to subglobose with a distinct apiculus. Basidia (36) 43–61 × 8.0–11.0 µm, thin-walled, hyaline, clavate to sub-

cylindrical, clamped, four tapered sterigmata, 6.5–13.0 μ m long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 1.5–5.0 μ m wide; hyphae distant from subhymenium 8.0–12.5 μ m wide.

Habitat and distribution: Solitary or scattered to gregarious, on soil rich in humus, in broadleaved forest or mixed coniferous–broadleaved forest, known only in subtropical areas of China; August to September.

Additional specimens examined: China. Hunan Province: Sangzhi County, Badagong Moutain, 29°40′49.64″ N, 109°48′24.81″ E, alt. 1300 m, in broadleaved forest, 14 September 2017, Ping Zhang (MHHNU9314); Yunnan Province: Malipo County, 23°21′41.98″ N, 105°09′44.17″ E, alt. 1580 m, in broadleaved forest, 3 Auguest 2019, Ping Zhang (MHHNU9813); Binchuan County, Jizu Moutain, 25°57′28.20″ N, 100°23′17.37″ E, alt. 2300 m, in mixed coniferous–broadleaved forest, 29 Auguest 2022, Ping Zhang (MHHNU11331).

Comments: Based on the simple or sparsely branched, pinkish basidiomata, this species may be mistaken for Clavaria incarnata Weinm., Clavaria rosea Fr., Clavulina amethystinoides (Peck) Corner, and Clavulina purpurascens P. Zhang. However, as it possesses the generic characters of four-spored basidia without postpartal septation and clamped hyphae, it can be determined that *Clavulinopsis incarnata* is assignable to *Clavulinopsis* [2,3,13]. The genus Clavulinopsis includes several species with a pinkish basidioma. Compared with C. incarnata, C. alcicornis (Zoll. & Moritzi) Corner, C. carneola Corner, C. lignicola (R.H. Petersen) Corner, C. lingula, and C. moricolor Corner all have a shorter basidioma. In addition, C. alcicornis mainly has three-spored basidia and C. carneola has four-spored or six-spored basidia; C. lignicola generally has a bifurcate basidioma and smaller spores $(2.8-3.2 \times 2.1-2.3 \mu m)$ similar to those of C. moricolor $(3.3-4.2 \times 2.5-3.0 \mu m)$; and C. lingula has unique clamp connections [2,3]. C. corallinorosacea (Cleland) Corner and C. sulcata Overeem are more similar to *C. incarnata* in basidiomata size, but the basidiomata of these two species often grows more densely. In addition, the former species has a narrower, fusiform or amygdaliform basidiospore, and the basidiomata of the latter species is thinly white and villous at the whitish base [2,3].

Clavulinopsis trigonospora Franchi & M. Marchetti, Index Fungorum 457:1, 2020: Figures 11 and 12.





Figure 11. Basidiomata of *Clavulinopsis trigonospora* (a) MHHNU9186; (b) MHHNU9200. Scale bars = 2 cm.

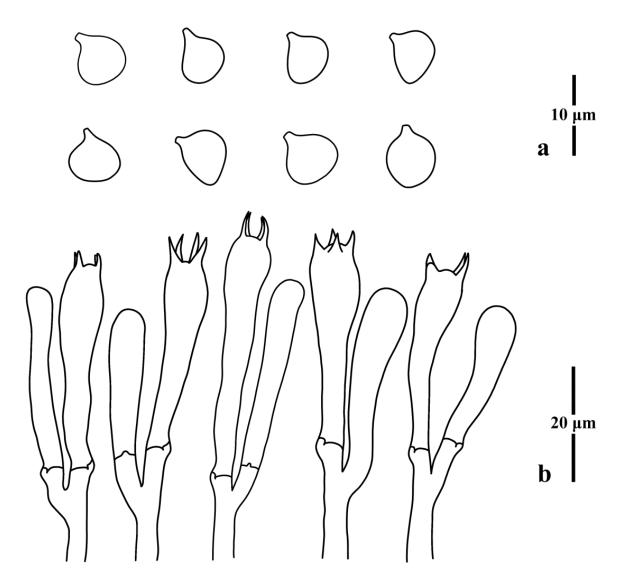


Figure 12. Microscopic features of *Clavulinopsis trigonospora* (MHHNU9200). (**a**) Basidiospores; (**b**) basidia.

Diagnosis: Distinguished from other taxa in the genus by its smooth, thin-walled, subtriangular, quadrilateral to subglobose basidiospores.

Description: Basidiomata fragile, simple, 6–90 mm tall, 1–4 mm wide, solitary or scattered to gregarious. Fertile part subcylindric, clavate to fusiform, sometimes curved or flexuous and with a longitudinal depression, yellowish to yellow [2A6–8, 30A5–6, Empire Yellow, Lemon Yellow, Clear Dull Green Yellow]. Apex rounded, concolorous, becoming darker than the lower part with age. Sterile part indistinct, concolorous or semitransparent, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous. Taste, odor, and macrochemical reactions were not recorded.

Basidiospores [100/4/3] 5.8–7.5 (8.0) × 4.5–6.5 µm $[Q = 1.21-1.51, Q = 1.29 \pm 0.08]$, thin-walled, hyaline, smooth, inamyloid, subtriangular to ellipsoid with a distinct apiculus. Basidia (30) 32–43 × 6–9 (10) µm, thin-walled, hyaline, clavate to subcylindrical, clamped, four tapered sterigmata, 5.0–7.0 µm long. Basidioles, incrustations, or crystals absent. Subhymenium clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 2.0–4.0 µm wide; hyphae distant from subhymenium 6.0–10.0 µm wide.

Habitat and distribution: Solitary or scattered to gregarious, on soil rich in humus, in broadleaved forest or mixed coniferous–broadleaved forest, known in Italy and China; August to November.

Specimens examined: China. Sichuan Province: Kangding County, Jiefang Second Village, 30°28'47.48" N, 102°15'46.38" E, alt. 2860 m, in mixed coniferous–broadleaved forest, 20 Auguest 2017, Ping Zhang (MHHNU9186, MHHNU9200); Gansu Province: Yuzhong County, Xinglongshan National Nature Reserve, 35°47'55.36" N, 104°04'20.27" E, alt. 2450 m, in broadleaved forest, 7 Auguest 2019, Ping Zhang (MHHNU10198).

Comments: Based on the records of Franchi and M. Marchetti [9], *C. trigonospora* has a simple, bright yellow to light yellow-orange, small basidioma (25–45 mm tall, 1–2 mm wide), and its most distinctive character is subtriangular spores. Compared with the type specimen, our specimens collected in China show a larger range in size, but all possess subtriangular spores. The distribution of this species in China was confirmed by phylogenetic analysis of molecular data, and thus the present account is the first record of this species outside of Italy.

Clavulinopsis tropicalis P. Zhang & Jun Yan, sp. nov.: Figures 13 and 14.





Figure 13. Basidiomata of *Clavulinopsis tropicalis* (a) MHHNU10721; (b) MHHNU10722. Scale bars = 2 cm.

MycoBank: 848963

Diagnosis: The species has a red basidioma and ellipsoid to broadly ellipsoid basidiospores, and differs from *C. erubescens* in having only four-spored basidia and growing in the tropics.

Etymology: tropicalis (Lat.) refers to the climate in which the species was discovered.

Type: China. Hainan Province: Baoting County, Qixianling, 18°42'08.44" N, 109°41'36.19" E, alt. 300 m, in tropical broadleaved forest, 31 July 2021, Ping Zhang (MHHNU10722, holotype).

Description: Basidiomata fragile, simple, 15–35 mm tall, 1–3 mm wide, solitary or gregarious to caespitose. Fertile part subcylindric to fusiform, occasionally slightly curved or flexuous and with a distinct longitudinal depression when mature, red to dark red [10B7–8, 10C7–8, Acajou Red, Pompeian Red] or pinkish [11A5–7, Spinel Pink, Spinel Red, Corinthian Red]. Apex rounded, concolorous or slightly paler, becoming darker with age. Sterile part narrow, indistinct, concolorous or paler than the upper part, occasionally curved, without tomentum or mycelial patch at the base. Context fragile, hymenium concolorous. Taste, odor, and macrochemical reactions were not recorded. Basidiospores [100/4/2] 5.0–7.0 (8.5) × 4.0–5.5 µm [Q = 1.18–1.50, $\mathbf{Q} = 1.29 \pm 0.12$] thin-walled, hyaline, smooth, inamyloid, broadly ellipsoid to ellipsoid with a distinct apiculus. Basidia (35) 43–50 × 6–9 µm, thin-walled, hyaline, clavate to subcylindrical, clamped, four tapered sterigmata, 4.0–7.0 µm long. Basidioles, incrustations, or crystals absent. Subhymenium

clearly delimited from the context, composed of densely interwoven hyphae. Hyphae of the context cylindrical to inflated, thin-walled, hyaline, parallel, without secondary septa, with clamp connections. Hyphae near subhymenium 1.5–3.5 μ m wide; hyphae distant from subhymenium 8.0–10.5 μ m wide.

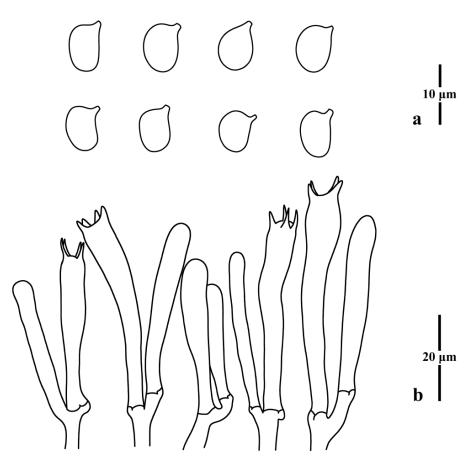


Figure 14. Microscopic features of *Clavulinopsis tropicalis* (MHHNU10722). (a) Basidiospores; (b) basidia.

Habitat and distribution: Solitary or gregarious to caespitose, on soil rich in humus, in tropical broadleaved forest, known only in tropical areas of China; July.

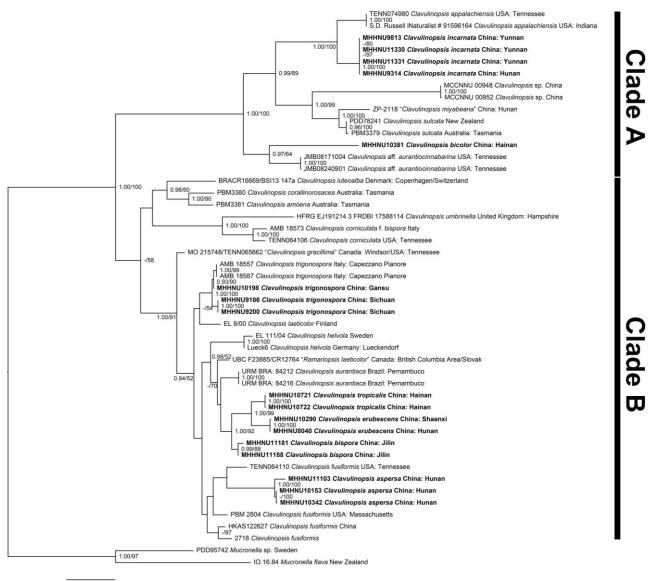
Additional specimen examined: China. Hainan Province: Baoting County, Qixianling, 18°42′08.44″ N, 109°41′36.19″ E, alt. 300 m, in tropical broadleaved forest, 31 July 2021, Ping Zhang (MHHNU10721).

Comments: Compared with *C. erubescens*, *C. tropicalis* is distributed in the tropics and only has four-spored basidia. Based on the present multi-loci phylogenetic analysis, although *C. erubescens* and *C. tropicalis* are placed on sister branches, there was a genetic distance of 0.0400 between the two taxa.

3.2. Molecular Phylogenetic Analysis

In total, 79 sequences (39 ITS and 40 nrLSU) from 50 samples (Table 1) were assembled into a multi-gene dataset for the molecular phylogenetic analysis, of which 34 sequences (17 ITS and 17 nrLSU) were newly generated in the present study. The combined ITS–nrLSU dataset consisted of 1612 sites and represented 23 species of *Clavulinopsis* and two species of *Mucronella* as the outgroup. The ML and BI (not shown) phylogenies (Figure 15) were extremely similar in topology and included two resolved clades (A and B) among the species of *Clavulinopsis*. The monophyly of clade A was strongly supported (PP = 1.00, BP = 100%) and included *C. appalachiensis* (PP = 1.00, BP = 100%), *C. incarnata* (PP = 1.00, BP = 100%), one unnamed *Clavulinopsis* species (PP = 1.00, BP = 100%), *C. miyabeana*,

C. sulcata (PP = 0.96, BP = 100%), *C. bicolor*, and *C. aff. aurantiocinnabarina* (PP = 1.00, BP = 100%). Clade B (BP= 58%) comprised 16 species: *C. luteoalba*, *C. corallinorosacea*, *C. amoena*, *C. umbrinella*, *C. corniculata* (PP = 1.00, BP = 100%), *C. gracillima* (dubious name), *C. trigonospora* (PP = 1.00, BP = 100%), *C. laeticolor*, *C. helvola* (PP = 1.00, BP = 100%), *R. laeticolor* (dubious name), *C. aurantiaca* (PP = 1.00, BP = 100%), *C. tropicalis* (PP = 1.00, BP = 100%), *C. erubescens* (PP = 1.00, BP = 100%), *C. bispora* (PP = 1.00, BP = 94%), *C. aspersa* (PP = 1.00, BP = 100%), and *C. fusiformis*. The six new species each formed an independent lineage, and the newly collected samples of *C. trigonospora* from China were nested within *C. trigonospora* accessions collected from Italy.



0.06

Figure 15. Phylogenetic relationships of *Clavulinopsis* species inferred from the combined dataset (ITS and nrLSU) using Bayesian posterior probabilities \geq 0.90; ML Bootstrap values \geq 50% are reported on the branches; the sign "-" means under the reported level. Six new species and one newly recorded species are shown in boldface text.

4. Discussion

In this study, six *Clavulinopsis* species new to science and one *Clavulinopsis* species newly recorded in China were identified. The specimens of *Clavulinopsis trigonospora* from Sichuan and Gansu, as a newly recorded species in China, conformed with the type speci-

men from Italy in molecular phylogenetic placement and morphological characteristics, and represent the first discovery of this species outside of Italy. Morphology and molecular data verified that the other six species collected were new species. The six new *Clavulinopsis* species are herein formally named and described in detail, and are illustrated with line drawings and photographs.

In the process of conducting the phylogenetic analysis, we found that available sequences of *Clavulinopsis* species are still extremely limited in GenBank and many species are represented by only one or two accessions with sequences. After adding the newly generated sequences in this study, we obtained a more stable topology for *Clavulinopsis*, with two clades resolved, in comparison with the previous works by Birkebak et al. [6] and Hyde et al. [8]. However, the common synapomorphic traits for species within clade A and clade B currently remain unclear and, in any case, samples of additional taxa are needed for reconstruction of a taxonomically complete phylogeny.

Prior to the present study, no new *Clavulinopsis* species had been described in China. Most *Clavulinopsis* specimens previously collected in China were assigned to species originating in Europe, Oceania, or the Americas. Based on our field investigations and molecular analyses, it is apparent that the main difficulties hindering the classification of *Clavulinopsis* are the currently limited availability of molecular data and the paucity of reliable, taxonomically important morphological characters. The present study has provided additional molecular data, provided a key to the known *Clavulinopsis* species in China (Table A1) and enhanced knowledge of the biological diversity in *Clavulinopsis*. However, the diversity of the genus in China is much richer than expected and additional research is needed to further explore this diversity in the future.

Author Contributions: Conceptualization: P.Z.; methodology: J.Y.; performing the experiment: J.Y., J.W. and G.-W.L.; resources: P.Z., J.Y., G.-W.L., and S.-W.W.; writing—original draft preparation: J.Y.; writing—review and editing: P.Z.; supervision: P.Z.; project administration: P.Z.; funding acquisition: P.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This study was financially supported by the National Natural Science Foundation of China (No. 31750001) and the Postgraduate Scientific Research Innovation Project of Hunan Province (No. CX20210429).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The sequence data generated in this study are deposited in NCBI GenBank.

Acknowledgments: We thank Robert McKenzie for editing the English text of a draft of this manuscript.

Conflicts of Interest: The authors declare no competing interests.

Appendix A

Table A1. Key to *Clavulinopsis* species in China.

1. Basidiomata branched	C. corniculata
1. Basidiomata unbranched	2
2. Basidiospores spinous	C. helvola
2. Basidiospores smooth	3
3. Basidiospores subtriangular to ellipsoid	C. trigonospora

Table A1. Cont.

3. Basidiospores globose, subglobose or ellipsoid	4
4. Basidia and hyphae with a very small clamp connection	C. bispora
4. Basidia and hyphae with a normal size clamp connection	5
5. Basidiomata bicolored	C. bicolor
5. Basidiomata monochromatic	6
6. Basidiomata red or pinkish	7
6. Basidiomata white, yellowish or orange	11
7. Basidiomata pinkish	C. incarnata
7. Basidiomata red	8
8. Basidia two or four tapered sterigmata	C. erubescens
8. Basidia invariably four tapered sterigmata	9
9. Basidiospores globose to subglobose	C. sulcata
9. Basidiospores ellipsoid, fusiform or amygdaliform	10
10. Basidiomata 15–35 mm tall, basidiospores broadly ellipsoid to ellipsoid	C. tropicalis
10. Basidiomata mostly 60–90 mm tall, basidiospores rather fusiform or amygdaliform	C. corallinorosacea
11. Basidiomata white or cream color	12
11. Basidiomata yellowish or orange	13
12. Basidiospores broadly pip-shaped, the wall slightly thickened	C. brevipes
12. Basidiospores subglobose, thin-walled	C. spiralis
13. Basidiomata orange, orange-red or salmon-orange	14
13. Basidiomata yellowish	15
14. Basidiomata 15–30 mm tall, basidiospores globose, rarely subglobose	C. aurantiaca
14. Basidiomata 15–70mm tall, basidiospores subglobose	C. aurantiocinnabarina
15. Distributed in the tropics	C. amoena
15. Distributed in the temperate and subtropics	16
16. Basidiospore hilar appendix not prominent (<1 μm)	C. luteoalba
16. Basidiospore hilar appendix very prominent (>1 μm)	17
17. Tramal hyphae somewhat thick-walled (up to 1 μm)	C. laeticolor
17. Tramal hyphae thin-walled	18
18. Basidiomata 50–140 mm tall, densely fasciculate	C. fusiformis
18. Basidiomata 15–50 mm tall, solitary or scattered to gregarious	C. aspersa

References

- 1. Wijayawardene, N.N.; Hyde, K.D.; Al-Ani, L.K.T.; Tedersoo, L.; Haelewaters, D.; Rajeshkumar, K.C.; Zhao, R.L.; Aptroot, A.; Leontyev, D.V.; Saxena, R.K.; et al. Outline of Fungi and fungi-like taxa. *Mycosphere* **2020**, *11*, 1160–1456. [CrossRef]
- 2. Corner, E.J.H. A Monograph of Clavaria and Allied Genera; Oxford University Press: London, UK, 1950; pp. 1–740.
- 3. Corner, E.J.H. Supplement to "A monograph of *Clavaria* and allied genera". *Beih. Zur Nova Hedwig.* 1970, 33, 1–299.
- 4. Petersen, R.H. Notes on Clavarioid Fungi. XV. Reorganization of *Clavaria, Clavulinopsis* and *Ramariopsis*. *Mycologia* **1978**, 70, 660–671. [CrossRef]
- Pegler, D.N.; Young, T.W.K. Basidiospore structure in *Ramariopsis* (Clavariaceae). *Trans. Br. Mycol. Soc.* 1985, 84, 207–436. [CrossRef]
- 6. Birkebak, J.M.; Mayor, J.R.; Ryberg, K.M.; Matheny, P.B. A systematic, morphological and ecological overview of the *Clavariaceae* (Agaricales). *Mycologia* **2013**, *105*, 896–911. [CrossRef] [PubMed]
- Furtado, A.N.M.; Daniëls, P.P.; Neves, M.A. New species and new records of *Clavariaceae* (Agaricales) from Brazil. *Phytotaxa* 2016, 253, 1–26. [CrossRef]

- Hyde, K.D.; Hongsanan, S.; Jeewon, R.; Bhat, D.J.; McKenzie, E.H.; Jones, E.G.; Phookamsak, R.; Ariyawansa, H.A.; Boonmee, S.; Zhao, Q.; et al. Fungal diversity notes 367–490: Taxonomic and phylogenetic contributions to fungal taxa. *Fungal Divers.* 2016, 80, 1–270. [CrossRef]
- Franchi, P.; Marchetti, M. I Funghi Clavarioidi in Italia; A. M. B. Fondazione Centro studi Micologici: Vicenza, Italy, 2021; Volume 1, pp. 1–664.
- 10. White, T.J.; Bruns, T.; Lee, S.; Taylor, J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR Protoc. Guide Methods Appl.* **1990**, *18*, 315–322. [CrossRef]
- 11. Vilgalys, R.; Hester, M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J. Bacteriol.* **1990**, 172, 4238–4246. [CrossRef]
- 12. Rehner, S.A.; Samuels, G.J. Taxonomy and phylogeny of Gliocladium analysed from nuclear large subunit ribosomal DNA sequences. *Mycol. Res.* **1994**, *98*, 625–634. [CrossRef]
- 13. Wu, C.L.; He, Y.; Yan, J.; Zhang, P. Two new species of *Clavulina* (Cantharellales) from southwestern China based on morphological and molecular evidence. *Mycol. Prog.* **2019**, *18*, 1071–1078. [CrossRef]
- 14. Kornerup, A.; Wanscher, J.H. *Methuen Handbook of Colour*, 3rd ed.; Fyre Methuen: London, UK, 1978; pp. 1–252.
- 15. Ridgway, R. Color Standards and Color Nomenclature; Published by the author: Washington, DC, USA, 1912; pp. 1–252.
- Katoh, K.; Standley, D.M. A simple method to control over-alignment in the MAFFT multiple sequence alignment program. Bioinformatics 2016, 32, 1933–1942. [CrossRef] [PubMed]
- 17. Hall, T.A. BioEdit: A user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. *Nucleic Acids Symp. Ser.* **1999**, *41*, 95–98.
- 18. Vaidya, G.; Lohman, D.J.; Meier, R. Sequencematrix: Concatenation software for the fast assembly of multi-gene datasets with character set and codon information. *Cladistics* **2011**, *27*, 171–180. [CrossRef]
- 19. Stamatakis, A. RAxML-VI-HPC: Maximum likelihoodbased phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* **2006**, *22*, 2688–2690. [CrossRef]
- Ronquist, F.; Huelsenbeck, J.P. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 2003, 19, 1572–1574. [CrossRef]
- 21. Nylander, J.A.A. MrModeltest v2. Program Distributed by the Author; Evolutionary Biology Centre: Uppsala, Sweden, 2004.
- 22. Rambaut, A. *FigTree v1. 4*; University of Edinburgh: Edinburgh, UK, 2012. Available online: http://tree.bio.ed.ac.uk/software/figtree (accessed on 5 April 2023).
- 23. Birkebak, J.M.; Adamčík, S.; Looney, B.P.; Matheny, P.B. Multilocus phylogenetic reconstruction of the Clavariaceae (Agaricales) reveals polyphyly of agaricoid members. *Mycologia* **2016**, *108*, 860–868. [CrossRef]
- 24. Wang, R.; Herrera, M.; Xu, W.J.; Moreno, J.P.; Colinas, C. Ethnomycological study on wild mushrooms in Pu'er Prefecture, Southwest Yunnan, China. *J. Ethnobiol. Ethnomed.* **2022**, *18*, 55. [CrossRef]
- 25. Hay, C.R.J.; Thorn, R.G.; Jocobs, C.R. Taxonomic survey of *Agaricomycetes* (Fungi: Basidiomycota) in Ontario tallgrass prairies determined by fruiting body and soil rDNA sampling. *Can. Field-Nat.* **2018**, *132*, 4. [CrossRef]
- Karich, A.; Kellner, H.; Schmidt, M.; Ullrich, R. Ein bemerkenswertes Mykotop im Zittauer Gebirge mit Microglossum rufescens als Erstnachweis für Deutschland. *Boletus* 2015, 36, 151–163.
- 27. Chen, Z.H.; Zhang, P. Atlas of Macrofungi in Hunan; Hunan Normal University Press: Changsha, China, 2019; pp. 1–425.
- Dentinger, B.T.M.; McLaughlin, D.J. Reconstructing the Clavariaceae using nuclear large subunit rDNA sequences and a new genus segregated from *Clavaria*. *Mycologia* 2006, 98, 746–762. [CrossRef] [PubMed]
- 29. Olariaga, I.; Huhtinen, S.; Læssøe, T.; Petersen, J.H.; Hansen, K. Phylogenetic origins and family classification of typhuloid fungi, with emphasis on *Ceratellopsis, Macrotyphula* and *Typhula* (Basidiomycota). *Stud. Mycol.* **2020**, *96*, 155–184. [CrossRef] [PubMed]
- 30. Yan, J.; Li, G.W.; Liu, W.H.; Chen, Z.H.; Zhang, P. Updated taxonomy of Chinese *Clavaria* subg. *Syncoryne* (Clavariaceae, Agaricales): Description of two new species and one newly recorded species. *Mycol. Prog.* **2022**, *21*, 67. [CrossRef]
- Yan, J.; Wang, X.Y.; Wang, X.H.; Chen, Z.H.; Zhang, P. Two new species of *Clavaria* (Agaricales, Basidiomycota) from Central China. *Phytotaxa* 2020, 477, 71–80. [CrossRef]
- 32. Petersen, R.H. The Clavarioid Fungi of New Zealand; DSIR Science Information Publishing: Wellington, New Zealand, 1988; pp. 1–170.
- 33. Oberwinkler, F. Fungus-alga interactions in basidiolichens. Beih. Zur Nova Hedwig. 1984, 79, 739–774.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.