

Proceeding Paper

The Growth Curve Method to Rapidly Derive the Antibacterial Potential of Polyoxovanadates [†]

Dorinda Marques-da-Silva ^{1,2}, Sib Sankar Mal ³, Manuel Aureliano ^{4,5} and Ricardo Lagoa ^{1,6,*}

¹ ESTG-Polytechnic Institute of Leiria, Morro do Lena-Alto do Vieiro, 2411-901 Leiria, Portugal; dorinda.silva@ipleiria.pt

² LSRE-LCM-ESTG, Polytechnic Institute of Leiria, 2411-901 Leiria, Portugal

³ Department of Chemistry, National Institute of Technology Karnataka, Mangalore 575025, Karnataka, India; malss@nitk.edu.in

⁴ Faculdade de Ciências e Tecnologia (FCT), University of Algarve, 8005-139 Faro, Portugal; maalves@ualg.pt

⁵ Centro de Ciências do Mar (CCMar), University of Algarve, 8005-139 Faro, Portugal

⁶ Applied Molecular Biosciences Unit, NOVA University of Lisbon, 2819-516 Caparica, Portugal

* Correspondence: ricardo.lagoa@ipleiria.pt

[†] Presented at the Biosystems in Toxicology and Pharmacology—Current challenges, 8–9 September 2022; Available online: <https://bitap.sciforum.net/>.

Abstract: In previous studies (Marques-da-Silva et al., 2019), we measured the minimum inhibitory concentrations (MICs) of three polyoxovanadates, namely V10, MnV11, and MnV13, against *Escherichia coli*. MICs were obtained following the standard method, which requires a 16–20 h culture and might neglect the effects of the compounds' metabolism during incubation. In this work, we studied the action of those compounds against *Enterococcus faecalis* by monitoring the bacterial growth kinetics, and we observed that the inhibition was evident from the beginning of the exponential phase. Notably, data collected from just a 7 h culture was enough to identify the compounds with stronger antibacterial activity according to standard MICs.

Keywords: polyoxometalates; antibacterial agents; *Enterococcus faecalis*; manganese polyoxovanadates; drug discovery; rapid methods in microbiology



Citation: Marques-da-Silva, D.; Mal, S.S.; Aureliano, M.; Lagoa, R. The Growth Curve Method to Rapidly Derive the Antibacterial Potential of Polyoxovanadates. *Med. Sci. Forum* **2022**, *11*, 2. <https://doi.org/10.3390/BiTaP-12790>

Academic Editor: Paula Videira

Published: 10 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Polyoxometalates are a well-known group of anionic polynuclear metal oxides (containing V^V, Ta^V, Nb^V, W^{VI}, and Mo^{VI}, usually in their highest oxidation state) with distinct and chemically changeable cluster structures. These compounds present important therapeutic potential as anticancer, antiviral, and antibacterial agents [1].

In previous work [2], we measured the minimum inhibitory concentrations (MICs) of three polyoxovanadates (POVs), namely Na₆V₁₀O₂₈ (abbreviated V10), K₅Mn^{IV}V₁₁O₃₂·10 H₂O (MnV11), and K₇Mn^{IV}V₁₃O₃₈·18 H₂O (MnV13) against the Gram-negative bacteria *Escherichia coli*. The MICs were obtained following the well-accepted serial two-fold dilution method. However, this method requires a 16–20 h culture and might neglect the effects of the compounds' metabolism/speciation changes during incubation.

There is a clear need for rapid methods to investigate potential antibacterial agents, antibiotic susceptibility, and novel effective combinations against different bacteria [3,4]. Sun et al. (2016) described a method based on OD600 and ATP measurements in a miniaturized set-up to identify new drugs and drug combinations against multidrug-resistant bacteria [3]. Recently, Chakansin et al. (2022) presented a dye-based assay to rapidly screen the antibacterial activity of nanomaterials whose turbidity interferes with conventional methods [4]. Although rapid and simple, it was observed that the dye could be degraded in the presence of nanomaterials with photocatalytic properties at high concentrations.

The application of dye-based assays to test POVs should also be regarded carefully because some of these compounds can catalyze the degradation of dyes [5]. In addition, many POVs are colored, and the color changes with speciation.

Several (bio)chemical modifications of metallodrugs, such as POVs, can affect their bioactivity and putative interactions with proteins [6,7]. Furthermore, the metabolic conversion of POVs by bacteria was reported before [2]. POVs' speciation and bioactivity can therefore change during incubation with microorganisms [6], and endpoint (single-time) assays, such as the conventional MIC determination method, can fail to detect the influence of compounds' speciation.

Taking into account these limitations, we further studied the antibacterial action of the POVs against the Gram-positive *Enterococcus faecalis* in this work by monitoring the growth curve. The method directly following bacterial multiplication is cheap and does not require detection reagents. Moreover, it can be automated and miniaturized for the high-throughput screening of numerous compounds against different bacteria [3]. In addition, the MICs of the POVs against *E. faecalis* were determined for the first time.

2. Materials and Methods

2.1. Polyoxovanadates, Bacteria and Culture Medium

The synthesis of the POVs and preparation of the stock solutions were performed as described in our previous work [2]. The antibacterial studies were carried out with *E. faecalis* (ATCC 29212) in LB broth.

2.2. Minimum Inhibitory Concentrations Determination

MICs were measured by the serial two-fold dilution method, using dilutions of each POV in concentrations 1024, 2048, and 4096 $\mu\text{g}\cdot\text{mL}^{-1}$. After inoculation with the bacteria and incubation for 18 h, the occurrence or absence of bacterial growth was visually examined (turbidity or presence of a cell deposit).

2.3. Bacterial Growth Monitoring

Growth curves of *E. faecalis* in LB were obtained by monitoring the optical density of the cultures at 600 nm (OD600), for up to 9 h incubation. The POV compounds were tested at 0.5 mM concentration, based on the antibacterial data obtained before with *E. coli* [2].

3. Results and Discussion

The antibacterial action of V1, V10, MnV11, and MnV13 against *E. faecalis* was investigated by monitoring the growth of the bacteria at different incubation times and by the conventional MIC determination method.

For the growth curve method, the kinetics of bacterial growth was followed from the OD600 measured in a laboratory spectrophotometer. Figure 1 illustrates the effect of a POV with low or insignificant antibacterial action, V1, and a clearly inhibitory compound, V10. The comparison between the growth curves in the presence of POVs and the corresponding control cultures indicates that the compounds did not affect the lag time of the culture. However, the inhibitory effects of V10, MnV11, and MnV13 on the growth of *E. faecalis* were evident from the beginning of the exponential phase. The monomeric vanadate species (V1) showed only a modest inhibition of bacterial growth.

In addition, the MICs of the POVs against *E. faecalis* were determined from 18 h cultures in the presence of a series of concentrations up to 4096 $\mu\text{g}\cdot\text{mL}^{-1}$. The MIC values obtained were 2048 $\mu\text{g}\cdot\text{mL}^{-1}$ for V10 and MnV11, 4096 $\mu\text{g}\cdot\text{mL}^{-1}$ for MnV13, and >4096 $\mu\text{g}\cdot\text{mL}^{-1}$ for V1. These results are in line with the efficacy of the same compounds against *E. coli* measured previously [2], suggesting that POVs yield broad antibacterial action.

There was also a good correlation between the effect of each compound in the *E. faecalis* growth curve and the corresponding MIC values (Figure 2). Notably, the inhibition data obtained after 7 h culture was sufficient to identify the POVs with stronger antibacterial activity, V10, MnV11, and MnV13, in agreement with the results of the MIC determination

at 18 h. The inhibition of bacterial growth was calculated from the increase in OD600 of cultures in the presence of the tested compound, which was normalized to the increase observed in the control culture that started with the same bacterial inoculum, but without the compounds. At 7 h, V10, MnV11, and MnV13 inhibited bacterial growth by 51%, 84%, and 73%, respectively. At this incubation time, V1 only inhibited 11% of *E. faecalis* growth, an indication of its low antibacterial activity.

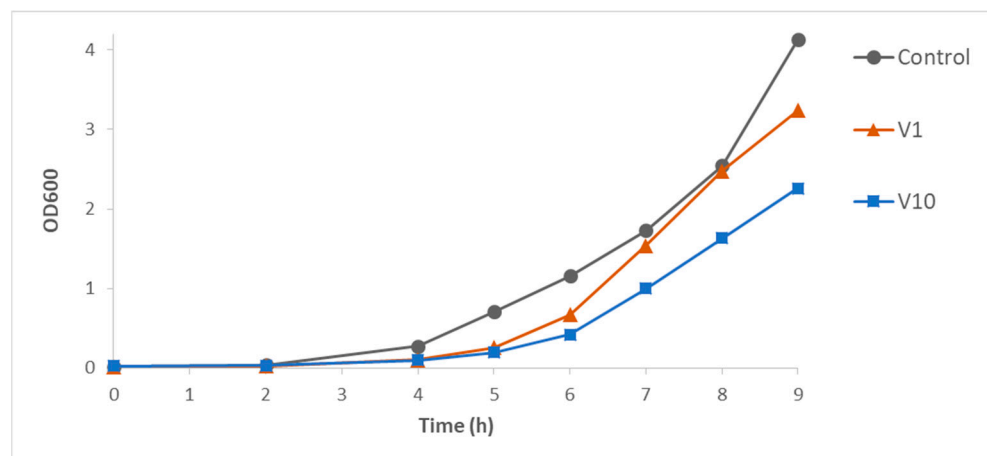


Figure 1. Time evolution of *Enterococcus faecalis* growth in the absence and in the presence of V1 or V10 compounds at 0.5 mM concentration.

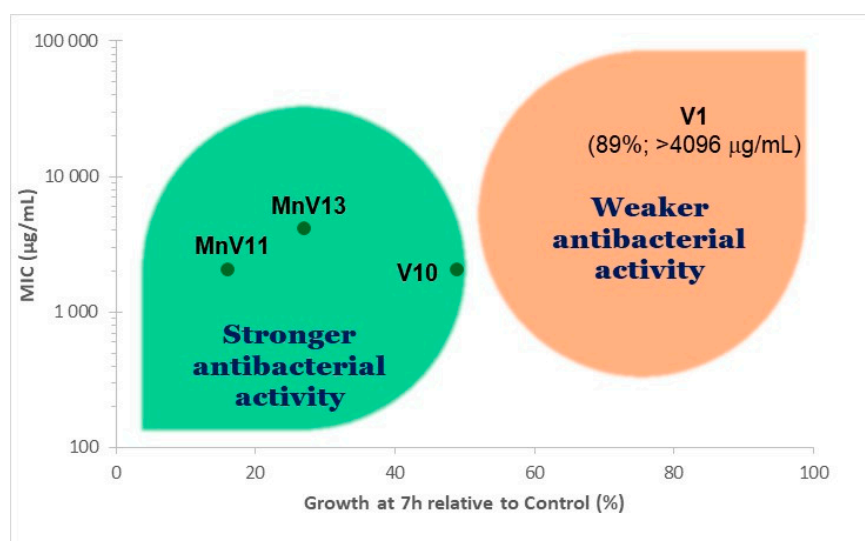


Figure 2. Relationship between the effect of the tested polyoxovanadates on *Enterococcus faecalis* growth measured at 7 h incubation and the corresponding minimum inhibitory concentrations (MIC) determined by the serial two-fold dilution method at 18 h.

4. Conclusions

The present study expands our understanding of the antibacterial action of vanadates and manganese polyoxovanadates. The POVs were tested against the Gram-positive bacteria *E. faecalis* by monitoring the growth of the microorganism at different incubation times and by the conventional MIC determination method.

The MICs herein obtained for the POVs against *E. faecalis*, and previously against *E. coli* [2], point to MnV11 as the compound with a higher potential for antibacterial applications, with activity towards both Gram-negative and Gram-positive bacteria.

The growth curve method implemented in this work to assess the antibacterial activity of POVs returned coherent results that validate the method for the fast screening of compounds. The method can provide preliminary indications of high- and low-activity compounds within a few hours.

As a future perspective, the study points out that the automated monitoring of the growth curve can meet the need for rapid methods to screen antibacterial POVs against different bacteria.

Author Contributions: Investigation, D.M.-d.-S. and S.S.M.; writing—original draft preparation, D.M.-d.-S. and R.L.; writing—review and editing, M.A. and R.L. All authors have read and agreed to the published version of the manuscript.

Funding: The authors' research received Portuguese national funds from FCT—Foundation for Science and Technology through unit projects UIDB/50020/2020, UIDP/50020/2020 and LA/P/0045/2020 (D.M.S.); UIDB/04326/2020 and LA/P/0101/2020 (M.A.); and UIDP/04378/2020, UIDB/04378/2020 and LA/P/0140/2020 (R.L.). S.S.M. thanks the Indian Council of Scientific & Industrial Research (CSIR) for providing financial support (01(2906)/17/EMR-II).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Aureliano, M.; Marques-da-Silva, D.; Serrano, A.; Martins, J.; Faleiro, L.; Fonseca, C.; Fraqueza, G.; Lagoa, R. Polyoxometalates with anticancer, antibacterial and antiviral activities. In *Polyoxometalates: Advances, Properties, and Applications*, 1st ed.; Jenny Stanford Publishing: United Square, Singapore, 2022; *in press*.
2. Marques-Da-Silva, D.; Fraqueza, G.; Lagoa, R.; Vannathan, A.A.; Mal, S.S.; Aureliano, M. Polyoxovanadate inhibition of: *Escherichia coli* growth shows a reverse correlation with Ca²⁺-ATPase inhibition. *New J. Chem.* **2019**, *43*, 17577–17587. [[CrossRef](#)]
3. Sun, W.; Weingarten, R.A.; Xu, M.; Southall, N.; Dai, S.; Shinn, P.; Sanderson, P.E.; Williamson, P.R.; Frank, K.M.; Zheng, W. Rapid antimicrobial susceptibility test for identification of new therapeutics and drug combinations against multidrug-resistant bacteria. *Emerg. Microbes Infect.* **2016**, *5*, e116. [[CrossRef](#)] [[PubMed](#)]
4. Chakansin, C.; Yostaworakul, J.; Warin, C.; Kulthong, K.; Boonrungsiman, S. Resazurin rapid screening for antibacterial activities of organic and inorganic nanoparticles: Potential, limitations and precautions. *Anal. Biochem.* **2022**, *637*, 114449. [[CrossRef](#)] [[PubMed](#)]
5. Missina, J.M.; Leme, L.B.; Postal, K.; Santana, F.S.; Hughes, D.L.; De Sá, E.L.; Ribeiro, R.R.; Nunes, G.G. Accessing decavanadate chemistry with tris(hydroxymethyl)aminomethane, and evaluation of methylene blue bleaching. *Polyhedron* **2020**, *180*, 114414. [[CrossRef](#)]
6. Aureliano, M.; Gumerova, N.I.; Sciortino, G.; Garribba, E.; Rompel, A.; Crans, D.C. Polyoxovanadates with emerging biomedical activities. *Coord. Chem. Rev.* **2021**, *447*, 214143. [[CrossRef](#)]
7. Aureliano, M.; Gumerova, N.I.; Sciortino, G.; Garribba, E.; McLauchlan, C.C.; Rompel, A.; Crans, D.C. Polyoxidovanadates' interactions with proteins: An overview. *Coord. Chem. Rev.* **2022**, *454*, 214344. [[CrossRef](#)]