

# Novel Purification Process for Amyloid Beta Peptide(1-40)

Kenji Usui <sup>\*,†</sup>, Shin-ichiro Yokota <sup>†</sup>, Kazuya Iwata and Yoshio Hamada <sup>\*</sup>

Faculty of Frontiers of Innovative Research in Science and Technology (FIRST), Konan University, Chuo-ku, Kobe 650-0047, Japan; m1861013@a.konan-u.ac.jp (S.-i.Y.); s1791005@s.konan-u.ac.jp (K.I.)

<sup>\*</sup> Correspondence: kusui@konan-u.ac.jp (K.U.); pynden@gmail.com (Y.H.)

<sup>†</sup> These authors contributed equally to this work.

Received: 12 March 2020; Accepted: 8 April 2020; Published: date

## Table of Contents

Page 1: Contents

Page 2: Materials and methods, and references

Page 3: Figure S1 for the HPLC chart of A $\beta$ (1-40) cleaved from HMBA resin after the reduction process without mercaptoacetic acid

Page 4: Figure S2 for the HPLC chart of A $\beta$ (1-40) by conventional peptide synthesis using Wang resin

## S1 Materials and Methods

### S1.1. Fmoc Peptide Synthesis of A $\beta$ (1-40) Using Wang Resin

A $\beta$ (1-40) was synthesized using Wang-PEG Resin (Watanabe Chemical Industries, Ltd., Hiroshima, Japan) by Fmoc solid-phase synthesis [S1] using the (2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate) (HBTU)–1-hydroxy benzotriazole monohydrate (HOBT) method (10 eq., double coupling). The DIPCI (N,N'-Diisopropylcarbodiimide)–DMAP (N,N-dimethyl-4-aminopyridine) method was used for the first residue (valine). The side-chain-protecting groups used were t-butyloxy carbonyl (Boc) for Lys; 2,2,4,6,7-pentamethyldihydrobenzofuran-5-sulfonyl (Pbf) for Arg; t-butyl (tBu) for Ser, Tyr, Asp, and Glu; and trityl (Trt) for His, Asn, and Gln.

### S1.2. Deprotection and Cleavage of A $\beta$ (1-40) Using Wang Resin

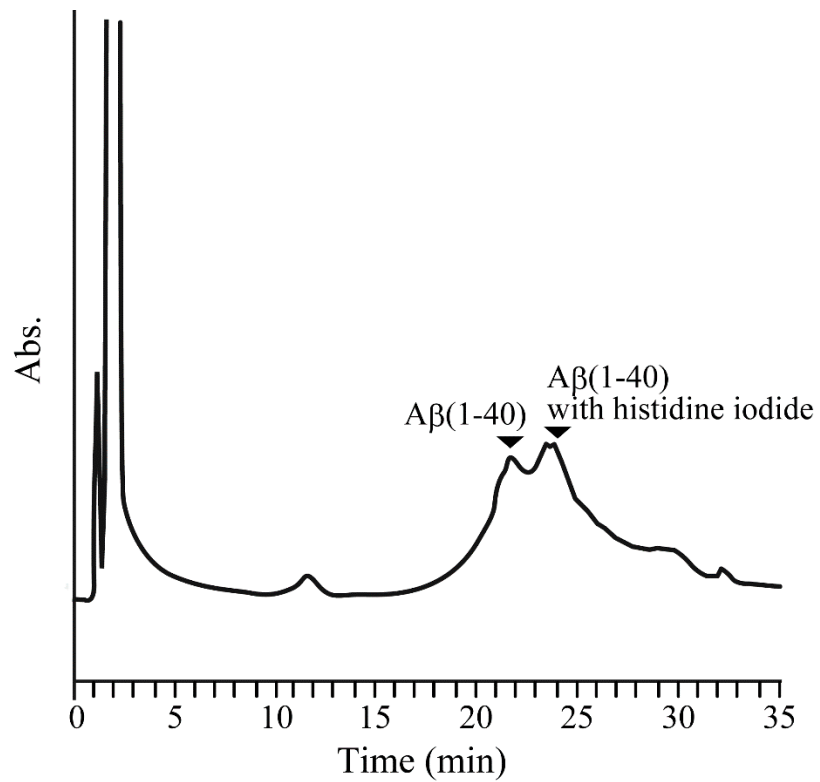
The side-chain-protecting groups on the resins were removed by incubating the peptide-resin for 2 h in deprotection and cleavage solution (trifluoroacetic acid (TFA, Watanabe Chemical Industries)/triisopropylsilane (Wako Pure Chemical Industries, Tokyo, Japan)/water (90/5/5, v/v)). The peptides were precipitated by the addition of cold diethyl ether and collected by centrifugation.

### S1.3. Monomerization and HPLC Analysis of A $\beta$ (1-40) Cleaved from Wang Resin

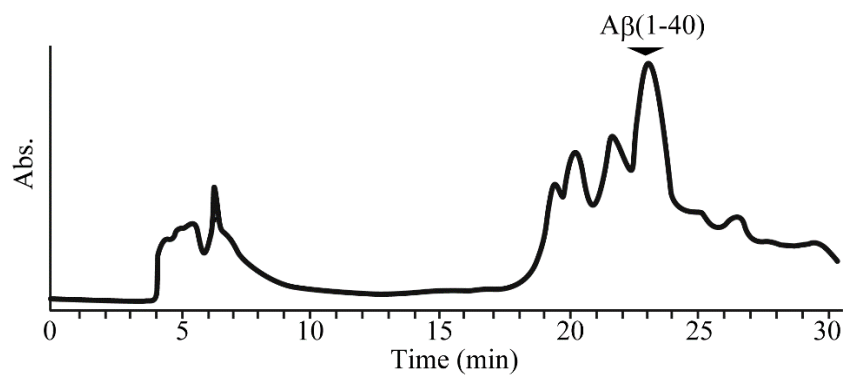
Before the HPLC, the crude peptide cleaved from Wang resin was dissolved in GdnHCl containing phosphate buffer (50 mM sodium phosphate, 8 M GuHCl, pH 7.5). In addition, the sample was incubated at 37 °C for 1 day in order to monomerize the sample completely. Then, the HPLC was performed on the GL7410 pump and GL7450 detector system (GL Sciences) using 220 nm absorbance. The peptide was analyzed on a Shodex Asahipak ODS-50 column (10.0 × 250 mm, Showa Denko K.K., Tokyo, Japan) using an isocratic condition with 100% of A solvent over 5 min and then a linear gradient from 0% to 30% of B solvent (90% acetonitrile, 10% Milli-Q, and 0.1% NH<sub>4</sub>OH) over 30 min at a flow rate of 1.0 mL/min.

## References

S1. Chan W. C.; White, P. D. Fmoc solid-phase peptide synthesis; Oxford University Press: New York, 2000.



**Figure S1.** HPLC chart of A $\beta$ (1-40) cleaved from HMBA resin after the reduction process without mercaptoacetic acid. The peaks were characterized by MALDI-TOF MS: A $\beta$ (1-40) with histidine iodide, m/z 4453.2 ((M+H)<sup>+</sup> calcd. 4456.5); A $\beta$ (1-40), m/z 4329.4 ((M+H)<sup>+</sup> calcd. 4330.6).



**Figure S2.** HPLC chart of A $\beta$ (1-40) by conventional peptide synthesis using Wang resin. The peak was characterized by MALDI-TOF MS.