Recent Advances in Nanomaterial-Based Sensing for Food Safety Analysis

Xiujuan Qiao 1,2,3,†, Jingyi He 1,2,†, Ruixi Yang 1,2, Yanhui Li 1,2, Gengjia Chen 1,2, Sanxiong Xiao 4, Bo Huang 1,2, Yahong Yuan 1,2, Qinglin Sheng 1,2,* and Tianli Yue 1,2,*

1 College of Food Science and Technology, Northwest University, Xi’an 710069, China
2 Laboratory of Nutritional and Healthy Food-Individuation Manufacturing Engineering/Research Center of Food Safety Risk Assessment and Control, Xi’an 710069, China
3 College of Chemistry and Molecular Engineering, Qingdao University of Science and Technology, Qingdao 266042, China
4 Xi’an Product Quality Supervision & Inspection Institute, Xi’an 710069, China
* Correspondence: qlsheng@nwu.edu.cn (Q.S.); yuetl@nwu.edu.cn (T.Y.)
† These authors contributed equally to this work.

Abstract: The increasing public attention on unceasing food safety incidents prompts the requirements of analytical techniques with high sensitivity, reliability, and reproducibility to timely prevent food safety incidents occurring. Food analysis is critically important for the health of both animals and human beings. Due to their unique physical and chemical properties, nanomaterials provide more opportunities for food quality and safety control. To date, nanomaterials have been widely used in the construction of sensors and biosensors to achieve more accurate, fast, and selective food safety detection. Here, various nanomaterial-based sensors for food analysis are outlined, including optical and electrochemical sensors. The discussion mainly involves the basic sensing principles, current strategies, and novel designs. Additionally, given the trend towards portable devices, various smartphone sensor-based point-of-care (POC) devices for home care testing are discussed.

Keywords: nanomaterials; sensors; sensor principle; food safety

1. Introduction

With the environmental pollution caused by social and technological development, the food safety problem has become increasingly serious. In addition, illegal businesses’ blind pursuit of interests has forced food safety problems to occur frequently, which is closely related to people’s health [1]. For example, clenbuterol, a banned chemical, was used by farmers in pigs to promote their leanness [2]. In 2008, melamine illegally added to infant formula caused more than 290,000 infants to suffer from pathological urinary tract stones [3,4]. All these events have made the public aware of the importance and urgency of addressing food safety issues.

Food safety issues continue to occur, prompting us to seek more effective food analysis techniques [5,6]. However, food analysis is a complicated process and a big challenge for human beings. For example, many potentially dangerous ingredients may occur in different kinds of food and at every stage in handling food [7]. These dangerous contaminants include heavy metals, various toxins, residual drugs, pesticides, viruses, illegal additives, and bacteria [8–10]. In addition, the complications will be increased with food industry globalization [11]. In view of the requirement of consumers for health, food safety issues have prompted the public to construct fast, cost-effective, specific, and sensitive analytical methods and analytical techniques for food analysis.

Sensitivity and specificity are the most important features of sensor sensing applications, and their sensing performance can be enhanced by proper modification of the sensing surface [12]. One of the most important ways to increase the sensing performance...
is the application of nanomaterials. Nanomaterials, with a size from 1 nm to 100 nm, can be easily developed into different forms for different detection needs, such as 0 dimension (0D) of nanoparticles/nanoclusters [13–17], I dimension (1D) of nanowire/nanorod [18–20], II dimension (2D) of nanosheet [21,22], and III dimension (3D) of nanonet/nanoflower/nanobulk [23–25]. Due to their unique light, electrical, and mechanical properties, their large surface area, good biocompatibility, catalytic activity, and rich bonding sites, nanomaterials have been widely used in various fields [26–31], especially for the production of sensing surface elements, that can not only increase the sensitivity but also provide lower limits of detection (LOD) [32–34].

There are many kinds of nanomaterial-based sensors and biosensors, which could be classified as optical and electrochemical sensors according to their transduction signals. Despite the fact that some reviews on the applications of nanomaterials for use in food safety sensing have been reported [35–38], a comprehensive overview of various nanomaterial-based sensors for food safety detection needs to be updated. Therefore, in this manuscript, we review the latest advances in novel sensors based on nanomaterials to detect food contamination over the past few years, including toxins, heavy metals, pesticides, drug residues, pathogens, and other hazardous substances (Figure 1). Various sensors in this manuscript could be divided into diverse optical methods and multiple electrochemical methods. In addition, the current smartphone-based portable devices have also been summarized here for the determination of contaminants in household food.

**Figure 1.** Novel sensors and biosensors based on nanomaterials for food detection.

### 2. Current Approaches for Food Safety Sensing

Nowadays, nanomaterial-based sensors and biosensors are being explored to reduce the occurrence of food safety problems. Generally, a biosensor contains several parts: sensing elements, recognition elements, and transducer elements. Sensing elements, involving nanomaterials of 0D, 1D, 2D, and 3D nanomaterials. The high conductivity, large surface area and unique physicochemical properties of nanomaterials mean that they are characterized by good sensitivity in the detection of food contaminants [39–41].
Apart from the sensitivity, the selectivity is also one of the most prominent features for the construction of biosensors. The integration of a recognition element with sensing element can provide a useful tool to improve the selectivity for monitoring food pollutants. The recognition elements usually involve aptamers [42], antibodies [43,44], DNA [45], peptides [46], and cells [47], which can selectively and specifically respond to analysis targets [48]. The transducer element, which plays crucial roles in the performance of sensor devices [49], can convert a biological response into detectable signals [50]. According to different kinds of transducer signals, current approaches based on nanomaterials for food contaminants’ detection could be classified into optical methods or electrochemical methods. Optical methods contain colorimetry, chemiluminescence (CL), fluorescence, and so on. Electrochemical methods contain impedimetric sensors, potentiometric sensors, and amperometric sensors (Figure 2).

Non-covalent and covalent methods are used for the biofunctionalization of nanomaterials [51,52]. Non-covalent bonding forces include electrostatic interaction (Figure 3A), π-π stacking (Figure 3B), van der Waals forces, etc. The non-covalent method is to simply link biomolecules with nanomaterials. Compared with non-covalent interaction, covalent binding has advantages in terms of reproducibility and the stability of the nanomaterials’ surface functionalization and physisorption. Covalent links could be formed by biomolecular links (Figure 3C), classic amide coupling reactions (Figure 3D), cross-linking, or click chemistry. These strategies preserve all specific properties of both the biomolecule and nanomaterial.
3. Optical Sensors and Biosensors

In optical sensors, quantum dots, gold/silver nanoparticles, upconversion nanoparticles, metal oxide nanomaterials, and organic fluorescent molecular-based nanomaterials have been widely used to improve sensing performance [53,54]. Depending on the signal output format, this section focuses on colorimetric, fluorescent, and surface plasmon resonance (SPR) sensors for food inspection.

3.1. Colorimetric Sensors and Biosensors

Colorimetric assays have attracted more attention because they are the simplest and easiest sensing strategies among various optical methods. Regarding colorimetric assay, color changes can be easily detected by visual observation without any complex and expensive equipment [55]. Compared to traditional detection methods that require high-pressure input and the presence of inert gases, nanomaterials provide a feasible in-situ detection solution [56]. Among them, gold nanoparticles (Au NPs), the most preferred candidate materials, are widely applied to fabricate sensors of colorimetric assays for food contaminants due to their unique optical features, simple synthesis method, high stability, and easy modification [57,58].

The typical mechanism of the fabrication of colorimetric sensors based on Au NPs mainly relies on Au NPs’ distance changes. Au NPs dispersed in solution usually appear red with the maximum absorption wavelength at ~520 nm. However, when Au NPs polymerize, the solution color changes to dark blue or purple (surface plasmon band from visible region to the near-infrared region). Directly prepared gold nanoparticles cannot meet the needs of sensing detection, and their surfaces need to be modified [59]. As shown in Figure 4, the aggregation can be both induced by targets on unmodified Au NPs and functional Au NPs. Additionally, the aggregation of Au NPs can also be induced by salt. Conversely, when the aggregated Au NPs are redispersed, it causes the solution color to change from purple or blue to red. The redispersion of Au NPs can be mediated by specific target molecule interaction (Figure 5). Surface microenvironment and external environment changes lead to the agglomeration and redispersion of Au NPs, which promotes their application in the field of colorimetric assays.

Recently, researchers have developed a solid-phase sandwich-type colorimetric immunosensor for the rapid detection of Staphylococcus A (SEA) in food. With the help of covalent affinity with protein A, SEA antibodies are modified on the slides to form a test region. The same antibody is conjugated to the gold nanoparticles by physical adsorption as nanoimmune probes. When slides are continuously exposed to SEA and AuNP-antibody bioconjugates, distinct red spots appear in the detection area due to the aggregation of gold nanoparticles. The limit of detection of SEA in milk by biosensors is 1.5 ng mL\(^{-1}\) [59]. The sensor does not require any signal amplification strategy, and the detection of various targets can be achieved by changing the immune probe. The colorimetric properties of gold nanoparticles also contribute to the detection of food freshness. Li et al. [60] constructed a detection system composed of polyethylene glycol (PEG)-modified AuNPs and dopamine to achieve convenient and effective colorimetric detection of food freshness (Figure 6). The system exhibits a significant burgundy to black color change at amine concentrations of 1–100 µg mL\(^{-1}\) with a detection limit of 2.8 µg mL\(^{-1}\).
Figure 4. Several schemes of colorimetric sensors based on Au NPs' aggregation.

Redispersion strategies

1. Target-specific molecule interaction mediated redispersion strategy

2. DNA-induced interaction mediated redispersion strategy

Figure 5. Several schemes of colorimetric sensors based on Au NPs' redispersion.
Recently, researchers have developed a solid-phase sandwich-type colorimetric immunoassay biosensor for the rapid and accurate quantification of morphine fluorescence methods in food. Compared with subjective colorimetric method, fluorescence phenomena are more attractive due to their low backgrounds, high sensitivity, strong objectivity, low detection limit, and high repeatability in food detection. Thus, developing sensitive, low cost and eco-friendly fluorescence materials with desirable fluorescence properties is urgently essential for food analysis. Since carbonate quantum dots (CDs) were firstly discovered by Xu et al. in 2004 [62], the fluorescence emission property of CDs makes them attractive as sensing probes for food analysis [63,64]. For example, Yue et al. [65] synthesized copper-modified fluorescent carbon dots (Cu-CDs) with stable double emission by hydrothermal methods, which were successfully applied to the detection of methylthlobutazine (TM) in fruits according to the linear relationship between the degree of fluorescence quenching of Cu-CDs and the concentration of the target. Yin et al. [66] prepared a novel N,Cl co-doped carbon dots (N,Cl-CDs) method based on deep eutectic solvent (DES) to achieve the rapid and accurate quantification of morphine fluorescence methods in food.

In addition to its excellent fluorescence properties, CDs can be prepared from any starting material containing carbon, include most food by-products, such as fruits, livestock, and vegetables. Based on this property, agricultural products containing a large amount of natural carbon to form CDs have great potential in bio-environmental sensing platforms [67–70], and also provide new solutions to the serious problem of food waste in global production and consumption processes [71–73]. The sensors for food analysis based on CDs produced from agriculture products are summarized as follows in Table 1.

### 3.2. Fluorescence Sensors and Biosensors

![Figure 6. Schematic diagram of the principle of food freshness detection based on AuNPs’ colorimetry [61].](image)

**Table 1.** Several sensors for food analysis based on CDs produced from agriculture products.

<table>
<thead>
<tr>
<th>Carbon Precursor</th>
<th>Methods</th>
<th>Target</th>
<th>Limit of Detection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wang et al. [74]</td>
<td>Papaya flesh</td>
<td>Thermal/200 °C/5 h</td>
<td>Escherichia coil</td>
</tr>
<tr>
<td>Das et al. [75]</td>
<td>Gram shells</td>
<td>Pyrolysis/315 °C/3 h</td>
<td>Escherichia coil</td>
</tr>
<tr>
<td>Hu et al. [76]</td>
<td>Orange peel</td>
<td>Microwave/900 W/1 min</td>
<td>Vanillin</td>
</tr>
<tr>
<td>Liu et al. [77]</td>
<td>Tomato puree</td>
<td>Microwave/10 min</td>
<td></td>
</tr>
<tr>
<td>Bao et al. [78]</td>
<td>Eleocharis dulcis</td>
<td>Hydrothermal/120 °C/5 h</td>
<td>Fe^{3+}</td>
</tr>
<tr>
<td>Bandi et al. [79]</td>
<td>Onion waste</td>
<td>Hydrothermal/120 °C/2 h</td>
<td>Fe^{3+}</td>
</tr>
<tr>
<td>Shen et al. [80]</td>
<td>Sweet potatoes</td>
<td>Hydrothermal/180 °C/18 h</td>
<td>Fe^{3+}</td>
</tr>
<tr>
<td>Zhao et al. [81]</td>
<td>Corn bract</td>
<td>Reflux/100 °C/24 h</td>
<td>Hg^{2+}</td>
</tr>
<tr>
<td>Bano et al. [82]</td>
<td>Tamarindus indica leaves</td>
<td>Hydrothermal/210 °C/5 h</td>
<td>Hg^{2+}</td>
</tr>
<tr>
<td>Tyagi et al. [83]</td>
<td>Lemon peels</td>
<td>Hydrothermal/200 °C/12 h</td>
<td>Cr^{6+}</td>
</tr>
<tr>
<td>Kumar et al. [84]</td>
<td>Tulsi leaves</td>
<td>Hydrothermal/180 °C/4 h</td>
<td>Pb^{2+}</td>
</tr>
<tr>
<td>Bhatt et al. [85]</td>
<td>Tulsi leaves</td>
<td>Hydrothermal/200 °C/4 h</td>
<td>Cr^{6+}</td>
</tr>
<tr>
<td>Wen et al. [86]</td>
<td>Pigskin</td>
<td>Hydrothermal/250 °C/2 h</td>
<td>Cd^{2+}</td>
</tr>
<tr>
<td>Liu et al. [87]</td>
<td>Chocolate</td>
<td>Hydrothermal/200 °C/8 h</td>
<td>Pb^{2+}</td>
</tr>
</tbody>
</table>

Other nanomaterials with non-fluorescent properties can achieve fluorescence effect-based food detection by combining with fluorescent molecules. The chromophore of most
conventional fluorescent probes produces strong fluorescence in its separated state, but the presence of aggregation-caused quenching (ACQ) results in molecules not radiating energy at high concentrations or in a concentrated state. In fact, the presence of the ACQ phenomenon limits the application of fluorescent probes to some extent [88–90]. Since the discovery of aggregation-induced emission effect (AIE) by Tang Group in 2001 [91], the use of AIE materials as fluorescent probes has gradually become a research hotspot [92–96]. This is because the AIE effect can overcome the shortcomings of the ACQ effect mentioned above, and thus is able to be used for high concentrations of fluorescence [97–101]. AIE emission can be clearly explained by the key role of molecular motions [102, 103].

Generally, detection strategies for AIE luminescence (AIEgens) include: (1) electrostatic interaction and hydrogen bond interaction, (2) solubility change of AIEgens, (3) disruption of AIE luminescence quenching, (4) preparation of nano-sized AIE particles, and (5) target-induced disaggregation of AIEgens. Based on these sensing mechanisms, a number of AIEgens have been exploited as sensing probes for food detection. Jia et al. [104] proposed a novel label-free fluorescent aptamer that was modified on graphene oxide (GO) to detect AFB1 in food samples by observing the fluorescence of quaternized tetraphenylethene salt and AFB1 aptamer aggregates (TPE-Z/AFB1) (Figure 7A). The fluorescence of the TPE-Z aptamer aggregates almost has non-fluorescence emission in the Tris-HCl buffer solution. In the presence of AFB1, the release of TPE-Z/AFB1 aptamers from the GO surface results in fluorescence recovery. The sensor has a detection limit of 0.25 ng mL−1. Wang et al. [105] constructed a tetraphenylethylene derivative functionalized mesoporous silica nanoparticle for the detection of furazolidone (FZD) and furacillin (NF) antibiotics in water.

In addition, the self-polymerization and disassembly of AIE nanomaterials can also be used for food inspection. As shown in Figure 7B, Mehta et al. [106] developed fluorescent peptidyl probes (2,3) with benzimidazolyl-cyanovinylidene (1) fluorophores based on their AIE effect for the detection of Ag+ and Ag NPs in water samples with detection limits of 0.64 ppb and 1.1 ppb, respectively. As for Figure 6C, Han et al. [107] reported a copper nanocluster (Cu NCs) with 2,3,5,6-tetrafluorothiophenol as a reducing agent and protective agent for the detection of histamine in food. When histamine is present, the nanoaggregates of Cu NCs are destroyed and spherical particles appear, leading to the quenching of their AIE effect.
3.3. Surface Plasmon Resonance (SPR) Sensors and Biosensors

Surface plasmon resonance (SPR) technology is a special optical method that detects changes in the refractive index caused by the interaction between molecules and ligands in a sample on the sensor surface [108]. When a molecule fixed on the surface of the sensor binds to a target in solution, it causes a change in the refractive index, resulting in a surface plasmon at the dielectric interface. Compared with conventional optical techniques, SPR technique exhibits some advantages, for example, the direct monitoring of the refractive index changes for food safety without need of specific properties (absorption, fluorescence, or scattering bands) and without the requirement of radioactive or fluorescent markers [109] of the analyte. Generally, SPR biosensors consist of a light source, optical system, sensing system, and detection system. The optical system emits the incident light; the change in resonance angle/wavelength can be converted by the sensing system; the magnitude signal of the resonance angle or wavelength is detected by system measures. SPR biosensors’ design principles always include fiber-optic surface plasmon resonance (FOSPR) [110], localized surface plasmon resonance (LSPR) [111], surface plasmon resonance imaging (SPRI), and transmission surface plasmon resonance (TSPR) [112]. The schematic illustrations with basic composition and advantages about the four types of SPR biosensors’ platforms have been provided in Figure 8.

Figure 8. Schematic illustrations of SPR biosensor-based platforms with advantages for food analysis.

Nanoparticle-based SPR technology has been widely used for the detection of foodborne contaminants. Vaisocherová-Lísalová et al. [113] constructed a multichannel SPR biosensor modified with an antibody-functionalized poly(carboxybetaine acrylamide)
(pCBAA) coating, which enhanced the sensor response with the help of Au NPs for food-borne *E. Coli O157:H7* and *Salmonella* sp., which were detected with limits of 57 cfu mL\(^{-1}\) and 17 cfu mL\(^{-1}\), respectively. Écija-Arenas et al. [114] modified a single layer of graphene on the transducer gold surface by chemical vapor deposition (CVD), and then modified 1-pyrenebutyric acid (PBA) to graphene by \(\pi\) stacking to ensure the covalent binding of subsequent aptamers. The specific capture of kanamycin by aptamers resulted in a change in SPR signaling with a detection limit of 285 nmol L\(^{-1}\).

4. Electrochemical Sensors and Biosensors

The electrochemical-based detection method for food safety is one of the most popular methods among detection strategies [115]. Due to their cost-effectiveness, simplicity, inherent sensitivity, high sensing speed, and compatibility with portable devices, electrochemical-based sensors have become the most rapidly growing sensor class [116]. Traditional electrodes based on mercury [117,118] have gradually been replaced by other suitable nanomaterials’ modified electrodes, such as biocompatible and excellent conductive carbon nanomaterials [119–123] or metal nanoparticles [124–126], nanozymes with high stability and tunable catalytic activity [127], and metal-organic frameworks (MOFs) with more active sites and high porosity [128].

Electrochemical sensors can be divided into potentiometric sensors, electrochemical sensors, and impedance sensors, according to the different conduction behavior of impedance, current, or potential signals caused by the interaction between the target and the recognition element on the sensor surface. This section focuses on the application of the above three types of nanomaterial-based electrochemical sensors in food safety detection.

4.1. Impedance Sensors and Biosensors

Electrochemical impedance spectroscopy (EIS) is a method based on electrical impedance detection at the electrode/electrolyte interface that can effectively detect food contaminants [129]. When the target in the electrolyte binds to the recognition element on the electrode surface, a specific concentration of the detected target is achieved by observing the impedance change at the electrode/electrolyte interface. This value is obtained by applying a small sinusoidal voltage at a specific frequency. For example, Mejri-Omrani et al. [130] reported an electrochemical aptasensor for directly detecting Ochratoxin A (OTA) by EIS technique. Polypyrrole (PPy) covalently bonded with polyamidoamine dendrimers as sensing elements, were coated on the surface of a gold electrode to reach PAMAM G4. Label-free OTA aptamers were thereafter covalently bonded to the PAMAM G4 for building an electrochemical aptasensor. When OTA and aptamers bind, the conformation of PAMAM G4 changes, leading to a difference in the electrical signal. The results showed that the method was able to detect concentrations of OTA in foods below the European regulations allowed, reaching 2 ng L\(^{-1}\). Miao et al. [131] constructed an electrochemical impedance sensor based on magnetic molecularly imprinted polymer for the detection of the insecticide dichlorodiphenyltrichloroethane (DDT) in food. The sensor uses a molecularly imprinted polymer synthesized from magnetic Fe\(_3\)O\(_4\) and polydopamine (PDA@Fe\(_3\)O\(_4\) MIP MNPs) as the electrode signal material, which is adsorbed by the identification unit of the PDA layer when DDT is present, resulting in a change in the electrochemical impedance signal of the material. The results show that the sensor has a good linear relationship with DDT concentration in the range of \(1 \times 10^{-11}\) to \(1 \times 10^{-3}\) mol L\(^{-1}\), and the detection limit reaches \(6 \times 10^{-12}\) mol L\(^{-1}\) (Figure 9A).
b) Electrochemical impedance sensor scheme for detecting DDT by EIS, including synthesis of PDA@Fe\(_3\)O\(_4\) MIP MNPs (a) and sensor construction (b); (B) four microelectrode array modes (A) and platform assembly mode (B) of sensing modules in a label-free miniaturized platform protocol for the detection of L. monocytogenes by EIS [132].

In addition, EIS is also used in the field of portable, miniaturized platforms for sensing foodborne bacteria of L. monocytogenes. These antibody-functionalized EIS microelectrodes are not only specific for the identification of Listeria monocytogenes, but their microfluidic devices can also be used for detection in liquid samples, such as milk. This highly efficient microfluidic device provides a platform for the detection of Listeria monocytogenes with a limit of detection (LOD) of 5.5 cfu mL\(^{-1}\) (Figure 9B) [132].

4.2. Voltammetry Sensors and Biosensors

Cyclic voltammetry (CV), linear sweep voltammetry (LSV), differential pulse voltammetry (DPV), and square wave voltammetry (SWV) are the most commonly used techniques in building sensors for food safety analysis. CV is one of the most widely used voltammetric techniques, which is based on the qualitative and quantitative analysis of targets by determining electrochemical information in the presence of redox intermediates or reversible reactions. In CV experiments, the potential superimposed on the working electrode is positively and linearly related to time. When the set ramp potential is reached, the applied potential returns to the initial potential, and a CV scan signal is obtained by recording the generated current. For other voltammetry sensors, DPV technique and SWV is generally applied for quantitative analysis in biosensors, especially for aptasensors, due to its merits of high sensitivity, short analysis time, and simultaneous multi-component detection. In addition, these techniques can be used to explore the kinetics, thermodynamics, and mechanisms of chemical reactions and analytical measurements. DPV differs from ESI technology in that DPV detects by applying a series of regular voltage pulses to the current immediately after a linear sweep and recording the resulting current difference as a function of the applied potential. In SWV, an excitation signal is obtained by applying a symmetrical square wave pulse of the staircase waveform to the working electrode.

Summerson and Prasco [133] report an inexpensive and convenient electrochemical sensor for the real-time monitoring of ochratoxin A (OTA) (Figure 10). When the target binds specifically to the aptamer, the conformational change of the aptamer causes MB to approach the electrode surface and enhance the electron transport ability of the electrode. SWV records
the signals generated by different concentrations of OTA into the detection system. This is a simple design without any signal amplification strategy for food safety analysis.

Figure 10. The scheme of electrochemical aptasensors for monitoring OTA. The sensing interface consists of methylene blue-modified DNA aptamers (A), and sensor current changes are observed at higher (B) and lower (C) square wave frequencies [133].

To achieve higher detection goals, designers often introduce enzymes. For example, Ezhilan et al. [134] reported that a sensitive acetylcholinesterase cyclic voltammetric biosensor based on Pt/ZnO/acetylcholinesterase/chitosan enables the detection of melamine and urea in adulterated milk. The acetylthiocholine acted as an electrochemical active substance so that the subsequent redox signal was recorded in CV by acetylcholinesterase catalysis. Urea and melamine bind to the serine hydroxyl group of acetylcholinesterase as competitive inhibitors, thus affecting the catalytic activity of acetylcholine. This biosensor detection mechanism is based on linear regression models for the calculation of binary mixtures of urea and melamine in adulterated cow milk. The proposed bioelectrode for the detection of urea and melamine has a limit of detection of 1 pM and 3 pM, respectively.

For signal amplification, Wang et al. [135] constructed an exonuclease III (Exo III)-driven double-amplified electrochemical aptamer sensor for the detection of chloramphenicol (CAP). First, the researchers synthesized Zr-MOF complexes of PtPd@Ni-Co hollow nanoboxes (PtPd@Ni-Co HNBs) and poly (diallyldimethylammonium chloride)-functionalized graphene (PDDA-Gr) as electrode modification materials to increase electrode surface area and conductivity. The captured DNA and assistant DNA were then modified on the surface of Zr-MOF to promote signal amplification. When CAP is present, it causes the release of a large amount of trigger DNA (Tr DNA) in Cycle I, then Tr DNA and Exo III initiating Cycle II, causing the exposed capture DNA to further bind to the signal probe (MB/HP-Uio-66/Signal DNA) and resulting in an electrochemical ‘signal on’. Under optimal conditions, the aptamer sensor has a good linear range, from 10 fM to 10 nM with a chloramphenicol detection limit of 0.985 fM. By changing the aptamer sequence, this sensing strategy can be applied to the detection of a variety of different targets (Figure 11).
Xu and Hou et al. [136] reported a highly sensitive aptamer sensor based on two electroactive species for the detection of malathion. When malathion is present, its specific binding to the aptamer causes the thiamine (Tn)-labeled aptamer to detach from the electrode surface while inducing the ferrocene (Fc)-labeled capture probe to form a hairpin structure, so detection can be achieved by signal shutdown of Tn and signal conduction of Fc. At the same time, the signal cycle amplification is achieved with the help of exonuclease I (Exo I). In this work, under the detection conditions, the proposed aptasensor exhibited excellent stability, specificity, and repeatability with a wide linear range from 0.5 to 600 ng L\(^{-1}\) (Figure 12).

**Figure 11.** Schematic diagram of exonuclease III-driven dual-amplification electrochemical aptamer sensor for chloramphenicol detection [135].

**Figure 12.** The scheme of dual-signal aptasensor for detection of malathion [136].
As described above, enzymes play a vital role in food testing and have been used to detect chemical or biological contaminants such as small molecules, heavy metal ions, and proteins [137]. However, the disadvantages of natural enzymes are that they are costly, time-consuming, vulnerable to extreme pH and temperature, and so on, which brings great challenges to their application. In this regard, nanozymes, the newly emerging artificial enzyme mimics, could be accepted to surpass these obstacles [138]. Nanozymes, having both nanomaterial features and natural enzyme-like properties, are attractive substitutes to natural enzymes [139]. Currently, nanozymes are widely considered to be the next generation of artificial enzyme stars due to their attractive nanoscale advantages of low cost, long-term storage, high stability to severe environments, and ease of mass production [140,141]. The materials used to synthesize nanoenzymes mainly include metals, metal oxides, MOFs, covalent organic frameworks (COF), polymers, etc. Various enzyme-like materials include nanomaterials, polymers, compounds, micelles, metalloproteins, and coordination complexes. These nanozymes are usually applied to mimic many kinds of enzymes, for example: oxidase (OXD), catalase (CAT) [142], superoxide dismutase (SOD) [143], peroxidase (POD) [144], glucose oxidase (GOX) [145], sphosphatase, and so on (Figure 13).

Nanozymes still have excellent test performance in complex food matrix environments. Wu et al. [146] developed a two-dimensional MnO$_2$ nanosheet-mediated electrochemical sensor with oxidase-like and peroxidase properties for the detection of organophosphate pesticides. Hu et al. [147] developed a functional 2D MOF nanoenzyme for the detection of Staphylococcus aureus. The 2D MOF material has peroxidase activity, which can effectively catalyze o-phenylenediamine to 2,2'-diaminoazobenzene to detect the concentration of Staphylococcus aureus. Under optimal conditions, the sensor has a detection limit of 6 cfu mL$^{-1}$ (Figure 14).

![Figure 13. Various nanomaterials and the enzymes nanomaterials mimicked.](image)

![Figure 14. Schematic diagram of 2D MOF nanoenzymes for the detection of Staphylococcus aureus [147].](image)
4.3. Potentiometric Sensors and Biosensors

Potentiometry (PM) is used in quantitative electroanalytical analysis of the target analyte by measuring the electrochemical potential of charged species between two electrodes (reference electrode and working electrode) in electrolytic tanks. Typically, PM testing is used when there is no or very little current in the system. This PM technique can achieve outstanding sensitivity for detection sensors. For example, Arvand et al. [148] report a potential sensor based on a mesoporous aluminosilicate-modified poly(vinyl chloride) film for the determination of Al$^{3+}$ in food and drug products. However, a highly accurate and stable reference electrode is required in this system, which will affect the application of PM in analytical detection [149].

In addition to the above examples, Table 2 (below) lists other nanomaterial-based (photo)electrochemical sensing food detection schemes.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Modified Electrode</th>
<th>Food Samples</th>
<th>Detection Techniques</th>
<th>LOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rapini et al.</td>
<td>Acetamiprid PANI/AuNPs/GPSEs</td>
<td>Fruit juice</td>
<td>DPV</td>
<td>86 nM</td>
</tr>
<tr>
<td>Jiao et al.</td>
<td>Chlorpyrifos FC@MWCNTs/OMC/GCE</td>
<td>Fresh pakchoi, lettuce and leek</td>
<td>CV</td>
<td>0.33 ng mL$^{-1}$</td>
</tr>
<tr>
<td>Liu et al.</td>
<td>Acetamiprid CdTe-MWCNTs/rGONRs/ITO</td>
<td>Apples and tomatoes</td>
<td>PEC</td>
<td>0.2 pM</td>
</tr>
<tr>
<td>Qiao et al.</td>
<td>BPA</td>
<td>Milk and drinking water</td>
<td>PEC</td>
<td>0.5 nM</td>
</tr>
<tr>
<td>Chen et al. [152]</td>
<td>β-agonists: RAC, CLB, PHL, SAL and PRO</td>
<td>AP-Ago/AuE</td>
<td>DPV</td>
<td>40 (RAC), 0.35 (CLB), 1.0 (PHL), 0.53 (SAL) and 1.73 (PRO) pg mL$^{-1}$</td>
</tr>
<tr>
<td>Li et al. [151]</td>
<td>Acetamiprid Co-doped ZnO/ITO</td>
<td>Cucumber</td>
<td>PEC</td>
<td>0.18 nM</td>
</tr>
<tr>
<td>Prabhakar et al.</td>
<td>Malathion CHIT-IO/FTO</td>
<td>Lettuce leaves</td>
<td>DPV</td>
<td>0.001 ng/mL</td>
</tr>
<tr>
<td>Song et al.</td>
<td>KMY</td>
<td>Milk</td>
<td>DPV</td>
<td>0.005 μg L$^{-1}$</td>
</tr>
<tr>
<td>Chen et al.</td>
<td>KMY and OTC</td>
<td>Dynabead</td>
<td>SWV</td>
<td>0.15 and 0.18 pM</td>
</tr>
<tr>
<td>Chen et al.</td>
<td>CAP</td>
<td>cDNA/GCE</td>
<td>SWV</td>
<td>0.33 pg mL$^{-1}$</td>
</tr>
<tr>
<td>Wang et al.</td>
<td>KMY</td>
<td>AuE</td>
<td>DPV</td>
<td>1.3 f M</td>
</tr>
<tr>
<td>Yan et al. [161]</td>
<td>CAP and OTC</td>
<td>AuNPs/GCE</td>
<td>SWV</td>
<td>0.15 and 0.1 ng mL$^{-1}$</td>
</tr>
<tr>
<td>Ge et al. [162]</td>
<td>TTC</td>
<td>BiPO$_4$/3DNGH/ITO</td>
<td>Milk</td>
<td>0.033 nM</td>
</tr>
<tr>
<td>Danesh et al.</td>
<td>SMY</td>
<td>Apt-CHIT/SPGE</td>
<td>Milk</td>
<td>11.4 nM</td>
</tr>
<tr>
<td>Yin et al. [164]</td>
<td>SMY</td>
<td>NP-PtTi/AuMWCNTsFe$_2$O$_4$/GCE</td>
<td>Milk</td>
<td>7.8 pg mL$^{-1}$</td>
</tr>
<tr>
<td>Wang et al. [165]</td>
<td>AMP</td>
<td>ITO electrode</td>
<td>Milk</td>
<td>4.0 pM</td>
</tr>
<tr>
<td>Zhao et al. [166]</td>
<td>Penicillin PEDOT-AuNPs/GRFe$_2$O$_3$NPs/GCE</td>
<td>Milk</td>
<td>DPV</td>
<td>0.057 ng mL$^{-1}$</td>
</tr>
<tr>
<td>Li et al. [167]</td>
<td>OTA</td>
<td>CDs-BP</td>
<td>EIS</td>
<td>0.03 fg mL$^{-1}$</td>
</tr>
</tbody>
</table>
Table 2. Cont.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Modified Electrode</th>
<th>Food Samples</th>
<th>Detection Techniques</th>
<th>LOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hao et al. [168]</td>
<td>OTA and FB₁</td>
<td>BFE</td>
<td>Maize</td>
<td>SWV 5 (OTA); 20 (FB₁) pg mL⁻¹</td>
</tr>
<tr>
<td>Zheng et al. [169]</td>
<td>AFB₁</td>
<td>TS-AuNPsc-cDNA/AuE</td>
<td>Animal feed and food samples</td>
<td>SWV 0.6 × 10⁻⁴ ppt</td>
</tr>
<tr>
<td>Goud et al. [170]</td>
<td>AFB₁</td>
<td>SPCE</td>
<td>Beer and wine sample</td>
<td>EIS 0.12 (seqA) and 0.25 (seqB) ng mL⁻¹</td>
</tr>
<tr>
<td>Istamboulié et al. [171]</td>
<td>AFM₁</td>
<td>SPCE</td>
<td>Milk</td>
<td>EIS 1.15 ng L⁻¹</td>
</tr>
<tr>
<td>Mishra et al. [172]</td>
<td>OTA</td>
<td>Apt/SPCE</td>
<td>Cocoa beans</td>
<td>DPV 0.07 ng mL⁻¹</td>
</tr>
<tr>
<td>Bagheryan et al. [173]</td>
<td>S. typhimurium</td>
<td>Diazonium-grafting layer modified SPCE</td>
<td>Apple juice sample</td>
<td>EIS 6 cfu mL⁻¹</td>
</tr>
<tr>
<td>Jia et al. [174]</td>
<td>Salmonella</td>
<td>rGO-MWCNT/GCE</td>
<td>Chicken sample</td>
<td>EIS 25 cfu mL⁻¹</td>
</tr>
<tr>
<td>Sheikhzadeh er al. [175]</td>
<td>S. typhimurium</td>
<td>Poly[pyrrole-co-3-carboxylpyrrole]copolymer/Au</td>
<td>Apple juice sample</td>
<td>EIS 3 cfu mL⁻¹</td>
</tr>
</tbody>
</table>

5. Mobile Sensors and Biosensors

Most of the current conventional sensing techniques for food safety are still dependent on laboratories, which are costly, time-consuming, complicated, labor-intensive, etc. Thus, the conventional sensing techniques make these sensing platforms unsuitable for point-of-care (POC) and consumer-oriented detection. These disadvantages could not timely prevent harmful substances from harming the human body and the environment. Therefore, there is an intense desire to develop portable sensing devices as homecare test devices [176]. To this end, the detection application of smartphone-based biosensors in food safety has become a hot spot for researchers [177–180].

With the rapid development of nanomaterials and the widespread use of smartphones for everyone, the combination of food safety monitoring services and smartphones will have the potential to offer portable sensing devices as homecare test devices. For example, Man et al. [181] developed a Au NPs-based microfluidic colorimetric immunoassay device for the detection of alternariol monomethyl ether (AME) with a detection limit of 200 pg mL⁻¹ and a recovery rate of 90.63% to 93.9% for smartphone imaging (Figure 15A). Cheng et al. [182] report a two-dimensional (2D) Pt-Ni(OH)₂ nanosheet-based amplified bidirectional lateral flow immunoassay device for the simultaneous detection of acetochlor and cymethrin, with detection limits of 0.63 ng/mL and 0.24 ng/mL, respectively (Figure 15B). In order to apply POC to home life, its devices need to be highly reliable, low cost, and user-friendly. Based on the above requirements, the rapid development of smartphones, which contain various embedded sensors, could enable successful application in food safety monitoring. For example, smartphones’ built-in sensors of audio jacks and cameras and external sensors (e.g., probe, microfluidic devices and circuit) have been widely used in POC devices [183,184]. Smartphone-based POC sensors aim to accomplish more timely, more efficient, and easier testing in resource-limited settings.
5.1. Smartphone-Based Optical Biosensors

Because of the advantages of smartphones, they are commonly applied in building optical POC biosensing systems, including colorimetry [185,186], surface plasmon resonance [187], fluorescence [188,189], image analysis, etc. The combination of smartphones and biosensing devices can promote precision and immediacy in food safety analysis.

Zeinhom et al. [190] have established a novel colorimetric biosensor for the detection of S. Enteritidis in cheese, milk, and water samples through visual and quantitative techniques. This testing system is based on magnetic beads labeled S. Enteritidis antibodies as a capture platform, horseradish peroxidase (HRP) as a signal amplification, where HRP enzyme and capture antibodies are both located on inorganic nanoflowers. The visual signal produced by nanotechnology at low concentrations is easily recorded by smartphones. The detection principle of the capture platform form and antibody-enzyme-inorganic nanoflower signal amplification strategy for S. Enteritidis is shown in Figure 16. This successful application of smartphones in the detection of S. Enteritidis in cheese and milk indicates the possibility of its application in various food products.
Processes 2022, 10, 2576 18 of 29

Figure 16. The novel smartphone-based colorimetric biosensor for detection of S. Enteritidis [190].

Compared with colorimetric technics, fluorescence technics show more attractive features of high precision, sensitivity, and accuracy. The same group have developed a smartphone-based fluorescence biosensors for *E. coli* O157:H7 detection. This POC biosensor, based on smartphones, includes a long-pass thin-film interference filter, a laser-diode-based photosource, a 3D printer, and insert lenses. A commercial blue laser (eBay) was employed as the excitation light source, powered by two batteries. This process was excited at 405 nm with output power of ~5 mW, and the detection fluorescence emission was collected by a signal-collecting external lens. The proposed biosensor provides a low noise to background imagine system, with a detection limit of 1 cfu mL$^{-1}$ and 10 cfu mL$^{-1}$ for yoghurt and egg, respectively (Figure 17A) [191]. Typically, actual food samples contain a number of different substances with similar biochemical/chemical structures and properties. Therefore, the smartphone-based multiplexing detection equipment has been applied to food [192–195]. Xing et al. [196] report a microfluidic biosensor based on the principle of fluorescence resonance energy transfer (FRET) between GO and fluorescent molecules, combined with smartphones, for the simultaneous detection of multiplex foodborne bacterial ssDNA (Figure 17B).

Figure 17. (A) Schematic diagram (a) and profile (b) of a smartphone-based fluorescence biosensors for *E. coli* O157:H7 detection [191]; (B) smart biosensor for simultaneous detection of multiplex foodborne bacterial ssDNA [196].
5.2. Smartphone-Based Electrochemical Biosensors

Nowadays, different kinds of electrochemical techniques, such as impedimetric [197], potentiometric [198], and amperometric [199] techniques based on smartphones have been researched. A number of POC or miniaturized electrochemical devices have been developed. Generally, electrochemical assay based on smartphone needs to meet the requirements of an external potentiostat for electrochemical signal acquisition and processing, disposable and low-cost electrodes, data transmission mode, and extra batteries for power supply.

In view of the above requirements, screen printing technology can be put into consideration to develop POC and low-cost electrochemical biosensors, because it is possible to easily prepare different kinds of electrodes on paper, plastics, and other flexible substrates [200]. Ji et al. [201] developed an electrochemical detection sensor based on a combination of screen-printed electrodes and a smartphone. This system was composed of a simple GO modification electrode, an energy transformation module to provide stimuli signals, a low-cost potentiostat for CV measurements, a bluetooth module for transmitting electrochemical data, and a smartphone as the controller and display. The redox couples CV signal could perform on smartphone by roughly controlling different scan rates with test errors less than 3.8% when compared to detection results on commercial electrochemical workstations (Figure 18A).

In addition, Mishra et al. [202] have developed a wearable electrochemical platform with electrodes imprinted on the surface of nitrile experimental gloves, which have the advantages of low cost and being disposable. Characterization was performed at room temperature by using a Palmsens handheld potentiostat (EmStat3 Blue with 10.0 × 6.0 × 3.4 cm³ dimensions, Palmsens, Houten, Netherlands) powered by a rechargeable Li-Po battery. This enzyme-based, disposable ‘glove lab’ biosensing system transmits data to Android smartphone devices in real time with a compact electronic interface for electrochemical sensing and wireless Bluetooth. The electrode system and the wireless electronic interface in the form of a long snake are modified on the surface of the glove with three layers of stress-resistant elastic ink. The first layer is a silver layer of Ag/AgCl particles combined with elastic Ecoflex material, the second layer is an elastic styrene-isoprene copolymer-modified carbon ink, and the third layer is a transparent flexible insulator. This organophosphorus hydrolase (OPH)-based biosensor provides a new idea for detecting organophosphate (OP) in food, and the cost, convenience, and speed advantages of the sensor also open the door to more applications of flexible wearable sensors in hand multiplexed sensing (Figure 18B).

In order to further reduce external attachments, Near Field Communication (NFC) technology is gradually entering the field of view. NFC is able to provide wireless power and data transmission through inductive coupling, thus solving the problem of using additional battery accessories in smartphone electrochemical sensors. NFC technology shows great potential in the development of new POC sensing systems due to its characteristics, such as low cost and possessing no battery to develop. Currently, NFC-based smartphone-integrated POC biosensors have already been used for biochemical detection [203,204] and food safety sensing [205]. Figure 18C shows an example of switchable NFC tags to detect food spoilage with a smartphone [205]. This protocol reports a gas sensor based on iron(III) p-toluenesulfonate-doped polyaniline (PAni) nanostructures for the detection of biogenic amines in meat.
In the future, mobile diagnostics will show great potential by combining technologies such as portable smartphones, cloud computing, and new sensing designs. If these sensors are used in the food industry, they will be beneficial for food quality control. In the future, mobile diagnostics will show great potential by combining technologies such as portable smartphones, cloud computing, and new sensing designs.

6. Conclusion and Outlook

Safety issues in all aspects of food production have always been a global concern, and in order to protect people from the threat of food contaminants, more convenient detection methods are needed. Compared with traditional methods, nanomaterial-based sensors are widely used in food safety testing because of their low cost, high efficiency, and reliability. In terms of electrode modification, nanomaterials can improve detection sensitivity due to their large specific surface area, good electrical conductivity and good catalytic activity. In terms of selectivity, nanomaterials provide excellent selectivity for sensing analysis by binding to aptamers, antibodies, cells, etc. This review summarizes various nanomaterial-based optical and electrochemical sensing analysis techniques for food contaminants, mainly mentioning basic sensing principles, examples of current sensing strategies, and novel designs based on nanomaterials. Through the design of nanomaterials and the construction of sensors, the performance of sensors has been significantly improved, providing new topics for food safety testing.

In addition, given the trend of portable devices and widespread use of mobile phones, the new generation of smartphone-based point-of-care devices has significant advantages (cost-effectiveness, ease of operation, minimal equipment, and ease of data management) when assembling various sensors for home care testing, which is the direction of the future. If these sensors are used in the food industry, they will be beneficial for food quality control. In the future, mobile diagnostics will show great potential by combining technologies such as portable smartphones, cloud computing, and new sensing designs.
Author Contributions: Writing—original draft preparation, X.Q. and J.H.; investigation and discussion, R.Y., Y.L. and G.C.; revision and discussion, S.X., B.H. and Y.Y.; revision, discussion, and project administration, T.Y. and Q.S. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by National Key R&D Program of China (2019YFC1606703) and the Natural Science Foundation of Shaanxi Province in China (2020JM-429).

Conflicts of Interest: The authors declare no conflict of interest.

References


33. Holzinger, M.; Goff, A.L.; Cosnier, S. Synergetic effects of combined nano_materials for biosensing applications. Sensors 2017, 17, 1010. [CrossRef]


42. Kalita, J.J.; Sharma, P.; Bora, U. Recent developments in application of nucleic acid aptamer in food safety. Food Control 2022, 145, 109406. [CrossRef]


Processes 2022, 10, 2576


61. Li, H.; Gan, J.C.; Yang, Q.; Fu, L.L.; Wang, Y.B. Colorimetric detection of food freshness based on amine-responsive dopamine polymerization on gold nanoparticles. Talanta 2021, 234, 122706. [CrossRef] [PubMed]


77. Liu, W.J.; Li, C.; Sun, X.B.; Pan, W.; Yu, G.G.; Wang, J.P. Highly crystalline carbon dots from fresh tomato: UV emission and quantum confinement. Nanotechnology 2017, 28, 485705. [CrossRef]

78. Bao, R.Q.; Chen, Z.Y.; Zhao, Z.W.; Sun, X.; Zhang, J.Y.; Hou, L.R.; Yuan, C.Z. Green and facile synthesis of nitrogen and phosphorus co-doped carbon quantum dots towards fluorescent ink and sensing applications. Nanomaterials 2018, 8, 386. [CrossRef]


133. Somerson, J.; Plaxco, K. Electrochemical aptamer-based sensors for rapid point-of-use monitoring of the mycotoxin ochratoxin a directly in a food stream. Molecules 2018, 23, 912. [CrossRef] [PubMed]


164. Wang, X.Z.; Dong, S.S.; Gai, P.P.; Duan, R.; Li, F. Highly sensitive homogeneous electrochemical aptasensor for antibiotic residues detection based on dual recycling amplification strategy. *Biosens. Bioelectron.* 2016, **15**, 49–54. [CrossRef]

165. Zhao, J.; Guo, W.J.; Pei, M.S.; Ding, F. GR-Fe3O4NPs and PEDOT-AuNPs composite based electrochemical electrochemical aptasensor for the sensitive detection of penicillin. *Anal. Methods* 2016, **8**, 4391–4397. [CrossRef]

166. Li, K.X.; Qiao, X.J.; Zhao, H.Y.; He, Y.P.; Sheng, Q.L.; Yue, T.L. Ultrasensitive and label-free electrochemical aptasensor based on carbon dots-black phosphorus nanohybrid for the detection of Ochratoxins A. *Microchem. J.* 2021, **168**, 106378. [CrossRef]


177. Rateni, G.; Dario, P.; Cavallo, F. Smartphone-based food diagnostic technologies: A review. *Sensors* 2017, **17**, 1453. [CrossRef]


196. Xing, G.W.; Li, N.; Lin, H.F.; Shang, Y.T.; Pu, Q.S.; Lin, J.M. Microfluidic biosensor for one-step detection of multiplex foodborne bacteria ssDNA simultaneously by smartphone. *Talanta* 2022, 253, 123980. [CrossRef]


