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Electrochemical Characterization of a Polymer Inclusion Membrane Made of Cellulose Triacetate and Aliquat 336 and Its Application to Sulfonamides Separation

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Abstract: An electrochemical characterization of a polymer inclusion membrane (PIM) fabricated with the ionic liquid (IL) Aliquat 336 (26%) and the polymer cellulose triacetate (CTA) (76%) is presented. Considering the use of PIMs in separation systems to remove pollutants from water, the characterization was performed with NaCl solutions by measuring membrane potential, electrochemical impedance spectroscopy, and salt diffusion and results were compared with those obtained from dry membranes. Results showed a significant reduction in the membrane diffusive permeability and electrical conductivity as well as the transport number of cation Na⁺ across the PIM when compared with solution values, which could be mainly related to the dense character of the membrane. Membrane application in the separation of different sulfonamides (sulfathiazole, sulfapyridine, sulfamethazine, and sulfamethoxazole) from water, with 1 M NaCl solution as striping phase, was also considered. These results indicated that the different chemical characteristics of the compounds, as well as the compact structure of the PIM, limited the transport of the organic molecules though it.

Keywords: polymer inclusion membrane; Aliquat 336; membrane potential; impedance spectroscopy; sulfonamides

1. Introduction

As it is well known, ionic liquids (ILs) are salts composed of an organic cation and an inorganic/organic anion with very low vapor pressure, low solubility, good electrical conductivity, and thermal stability [1], which allow their use in a large number of different applications such as solvents for organic reactions and catalysis, energy, electro-deposition, green chemistry, separation processes or CO₂ capture, among others [2–9]. Moreover, ILs can be used as extractants in the preparation of polymer inclusion membranes (PIMs). PIMs basically consist of a polymeric matrix, normally made of cellulose triacetate (CTA) or poly(vinyl chloride) (PVC) [10,11] and an extractant (named carrier), which is responsible for transport across the membrane. Usually the addition of a plasticizer or some modifiers is needed to obtain adequate elastic characteristics of the membranes [12]. However, the use of ILs as carriers favors the flexibility and mechanical strength of PIMs avoiding the need of the addition of other components, and their high viscosity helps to retain them in the polymer gel-like network. Different commercial ILs such as Cyphos 101 (trihexyl(tetradecyl)phosphonium chloride), Cyphos 102 (trihexyl(tetradecyl)phosphonium bromide),

In previous works, different physicochemical characteristics of PIMs (contact angle, Young modulus, conductivity, or dielectric constant) were analyzed for membranes containing different Aliquat 336 (AlqCl) percentages in both CTA or PVC polymers [18], as well as for PIMs where the counter-anion of Alq was modified (AlqCl, AlqNO₃ or AlqSCN) [19]. However, information on other electrochemical characteristic parameters, such as ion transport numbers or ionic diffusion coefficients [20,21], determined from measurements carried out with PIMs in contact with saline solutions seems to be also of interest for their applications.

Thus, in this study an electrochemical characterization of an Aliquat 336/CTA PIM (26% AlqCl/74% CTA) is presented performed by membrane potential and impedance spectroscopy measurements using a typical model electrolyte (NaCl) at different solution concentrations. A comparison of other physicochemical parameters depending on the PIM state (dry or wet) is also indicated to have a more adequate picture of the membrane behavior in usual working conditions for water treatment.

Separation abilities of the characterized PIM are studied using a mixture of four common antibiotics, belonging to the sulfonamides family, which can be found in natural waters due to their wide use and incomplete elimination through sewage treatment.

2. Results and Discussion

Taking into account that PIMs are aimed to transport and separate compounds of interest in aqueous solution, it is important to determine their physicochemical characteristics in a wet media and to compare them with respect to dry membranes. It is important to bear in mind that PIMs can lose part of the IL incorporated in the polymeric network when membranes are immersed in distilled water or aqueous electrolyte solutions [22,23]. This mass loss is associated with both the nature of the IL and the polymer. In particular, Kagaya et al. [23] found a relative mass loss of around 20% for an Aliquat 336/PVC membrane after 48 h immersion in distilled water. However, the mass loss diminished to 2% when immersed in a NaCl solution.

In this study, chemical surface differences between a wet PIM (sample 26% AlqCl/74% CTA-w, w indicating 'wet state', that is, after 48 h immersed in distilled water) and a dry PIM (sample 26% Alq336/74% CTA-d, d indicating 'dry state') was determined by analyzing X-ray photoelectron spectroscopy (XPS) spectra. XPS is a well-known technique able to give information on chemical changes on membrane surfaces since it provides elemental identification, relative composition, and chemical state of the elements present in a surface region [24]. Consequently, differences in the atomic concentration percentages (A.C. %) determined for the elements found on the surfaces of wet and dry membrane samples should be an indication of Aliquat 336 modification/lost. Table 1 shows the A.C. % of the elements present on the surfaces of the PIMs (C, O, N, Cl, and Si) in dry and wet states (samples 26% AlqCl/74% CTA-d and 26% AlqCl/74% CTA-w), which were determined by analyzing the XPS spectra. Moreover, A.C. % of the elements found on the surface of a CTA film in dry and wet states (CTA-d and CTA-w samples, respectively) are also indicated in Table 1 for comparison reasons. As expected, both CTA-films do not show any Cl content since this element is associated to Aliquat 336. However, Si, a non-characteristic membrane/film element, is detected in all samples. This fact can be attributed to a manufacture contamination, and its higher percentage in wet samples might be related to element surface reorganization associated to the hydrophobic character of silica [25,26]. In fact, surface reorganization of more hydrophobic organic matter and N impurities could be one of the reasons for differences between dry and wet samples. In this context, it should also be indicated that the total N (%) obtained for dry and wet PIM samples also seems to include contamination contribution, while the specific Aliquat 336-N contribution (binding energy at 402.2 eV [27]) is indicated in brackets. In that case, the N/Cl ratio is similar to the theoretical one

 $((N/Cl)^{T} = 1)$. Consequently, the comparison of the results obtained for wet and dry PIM samples shows a reduction in the percentages of Cl and N associated to Aliquat 336 in the wet sample (around 25%). This fact seems to indicate a partial loss of the IL from the membrane surface, but the effect of element surface reorganization depending on their higher/lower hydrophobic character might also be involved.

Table 1. Atomic concentration percentages of the elements found * on the surfaces of wet (X-w) and dry (X-d) PIM or CTA film.

PIM	C (%)	O (%)	N (%)	Cl (%)	Si (%)
26% AlqCl/74% CTA-w	72.6	23.7	1.6 (0.38)	0.31	1.6
26% AlqCl/74% CTA-d ^a	66.7	29.6	0.89 (0.50)	0.44	0.3
CTA-w	73.7	24.1	0.65	n.d.	1.6
CTA-d	66.6	32.3	n.d.	n.d.	0.9

* Other elements with A.C. % < 0.3 are not indicated; ^a values from reference [18]. n.d. = not detected.

Wet PIMs also exhibit different elastic behavior than dry ones, as can clearly be observed in Figure 1, where a comparison of the strain-stress/elongation curves for wet and dry PIM samples is presented. Wet PIM sample shows a more plastic character, with a significant increase in the elongation at break (around eight times higher) and lower Young modulus (40% reduction), which is determined from the slope of the linear relationship obtained at low strain-stress values (insert in Figure 1). These changes are attributed to the plasticizer character of water, as it was previously obtained for regenerated cellulose membranes [28]. This fact should be considered in the selection of the Aliquat 336 content used for PIM manufacture, since it can play an important role on the mechanical behavior (high plasticity or brittle) of the membranes, according to the significant reduction in Young modulus associated to Aliquat 336 content in dry AlqCl/CTA membranes [18].



Figure 1. Strain-stress vs. elongation curves for wet (□) and dry (■) samples of the 26% AlqCl/74% CTA PIM.

Ion transport across a membrane is usually characterized by the ionic diffusion coefficients (D_i) or the ionic transport numbers (t_i), which are related by the following expression: $t_i = z_i D_i / \Sigma i z_i D_i$, or $t_i = D_i / (D_+ + D_-)$ in the case of 1:1 single electrolyte solutions [29], being z_i the valence of ion *i*. Ion transport numbers represent the fraction of the total current (I_T) transported for one ion ($t_i = I_i / I_T$)

and, consequently, in the case of single 1:1 electrolytes they should fulfil that $t_+ + t_- = 1$. Ion transport numbers are usually determined from membrane potential ($\Delta \Phi_{mbr}$) values by [20,30]

$$\Delta \Phi_{\rm mbr} = (RT/F)(1 - 2t_{-})\ln(a_{1}/a_{2}) = (RT/F)(1 - 2t_{-})\ln(c_{1}/c_{2}) \tag{1}$$

where *R* and F are the gas and Faraday constants, T represents the temperature of the system, while a_1 and a_2 are the activities of the NaCl solutions at both membrane sides, which can be replaced by solution concentration (c_1 and c_2) in the case of diluted solutions. Figure 2 shows the variation of membrane potentials with the solution concentration for the 26% AlqCl/74% CTA membrane, as well as the calculated solution diffusion potential values ($\Delta \Phi_{dif}^{o} = (RT/F)(2t_+^{o} - 1)\ln(c_1/c_2)$), where t_+^{o} represents the Na⁺ transport number in solution, that is, when there is not any membrane separating the electrolyte solutions (dot-dashed line) [29]. For comparison reasons, membrane potentials for a commercial positively charged ion-exchange membrane (AR204-SZRA-412 by Ionics) are also drawn in Figure 2.



Figure 2. Membrane potential as a function of NaCl solution concentration: (◊) 26% AlqCl/74% CTA membrane, (▲) ion-exchanger AR204-SZRA-412 membrane. Solution diffusion potential: dash dot line.

As it can be observed in Figure 2, $\Delta \Phi_{mbr}$ values for 26% AlqCl/74% CTA and AR204-SZRA-412 membranes are rather similar, being those corresponding to the ion-exchange sample slightly more negative. Moreover, in both cases, $\Delta \Phi_{mbr}$ values differ significantly from those corresponding to the NaCl solution, which is an indication of the control of both membranes on ions transport. Ion transport numbers across the PIM were determined by means of Equation (1), and the following average values were obtained: $t_{-} = 0.91 \pm 0.03$ and $t_{+} = 0.090 \pm 0.003$. Consequently, the diffusive transport of cations across the studied PIM seems to be practically excluded. In fact, taken into account the relationship between ion transport numbers and diffusion coefficients [29], we obtain: $t_{-}/t_{+} = D_{-}/D_{+} = 10.1$; that means, Cl⁻ seems to move10 times higher than Na⁺ in the PIM. Ion transport number values are rather similar to those reported by other authors for PIMs obtained for different ILs [20].

Electrochemical impedance spectroscopy (EIS) measurements also provide valuable information on electrical characteristics of membranes. Figure 3 shows the Nyquist plot ($-Z_{img}$ vs. Z_{real} , Figure 3a) and the Bode plot ($-Z_{img}$ vs. frequency, Figure 3b) obtained for the 26% Alq336/74% CTA membrane separating two 0.03 M NaCl solutions. Two separated contributions associated to the membrane (frequency ranging between 10 kHz and 1000 kHz) and the electrolyte solution placed between the membrane and the electrodes (frequency > 1000 kHz) can be observed in Figure 3, which correspond to a depressed semicircle (membrane) and a semicircle (electrolyte solution). This kind of impedance curve is typical of composite systems, in particular it is similar to that exhibited by low permeability systems such as dense non-swollen membranes [31] or nanofiltration/reverse osmosis membranes [32–34], and it is related with the significant difference in the electrical characteristics of the two parts forming the composite system: the membrane and the electrolyte solution, respectively. The equivalent circuit for the whole electrolyte(e)/membrane(m) system consists in the addition (series association) of two sub-circuits: a parallel association of a resistance (R) and a capacitor (C) for the electrolyte solution placed between the electrodes and the membrane (R_eC_e), plus a parallel association of a resistance and a generalized capacitor (R_mQ_m) for the membrane contribution due to a distribution of relaxation times [35] (see Materials and Methods section).



Figure 3. Impedance plots for the 26% AlqCl/74% CTA membrane separating two NaCl solutions at 0.03 M concentration. (**a**) Nyquist plot ($-Z_{img}$ vs. Z_{real}); (**b**) Bode plot ($-Z_{img}$ vs. frequency).

The fit of the impedance curves obtained for the NaCl concentrations studied (see electrochemical impedance spectroscopy (EIS) measurements in the Experimental section) allows us the determination of the membrane and electrolyte electrical resistance (R_m and R_s , respectively) at each NaCl solution, and Figure 4a shows the dependence R_m and R_s with electrolyte concentration. Moreover, taking into account the relation between membrane electrical resistance and electrical conductivity (σ_m) [36]

$$\sigma_{\rm m} = \Delta x_{\rm m} / R_{\rm m} S_{\rm m} \tag{2}$$

where Δx_m and S_m represent the membrane thickness and area respectively (assuming the transport takes place through the whole membrane area), variation of membrane electrical conductivity with NaCl concentration can also be obtained. Figure 4b shows the σ_m - σ_s relationship (membrane conductivity *vs.* solution conductivity), where the significant barrier effect of the 26% AlqCl/74% CTA PIM on the ion transport can be observed, which could be related to the dense structure and low ionic diffusion across this kind of membranes [20,31].



Figure 4. (a) Variation of 26% AlqCl/74% CTA membrane (\Diamond) and NaCl solution (\times) electrical resistance with electrolyte concentration. (b) Membrane conductivity versus solution (NaCl) conductivity.

Diffusive permeability across a membrane, P_s, is another parameter usually determined for membrane characterization, obtained from salt diffusion measurements. According to Fick's first law,

the solute flux crossing the membrane at steady-state, J_s , and the concentration difference which causes such flux, $\Delta c = (c_d - c_r)$, are related by [37]

$$J_s = dn/dt = P_s \Delta c = P_s (c_d - c_r)$$
(3)

Equation (3) can also be expressed as

$$dc_r/(c_d - c_r) = (S_m/V_0)P_s dt$$
(4)

Vo and S_m are the volume of the receiving cell and the membrane area, respectively. Taking into account the mass continuity: $c_d^{o} + c_r^{o} = c_d^{t} + c_r^{t} = constant$, where c_d^{o} and c_r^{o} indicate the initial concentrations and c_d^{t} and c_r^{t} at a time t, we obtain the expression

$$\ln([1 - (2c_r^{t}/c_d^{t})] = -2[S_m/(Vo \Delta x_m)] P_s t$$
(5)

where Vo represents the cell volume (that means at t = 0). Figure 5 shows time evolution of solution concentration separated by the PIM at a NaCl donor solution concentration of 0.01 M. Diffusive NaCl permeability across the PIM was determined from the slope of the straight line shown in Figure 5, according to Equation (5), and the value $P_s = 3.8 \times 10^{-8}$ m/s was obtained. This value is similar to that obtained for low permeability or reverse osmosis/nanofiltration membranes [31,32] and, consequently, it seems to be associated to the dense structure of the membrane, which controls the transport when no carried mediated effect is involved.



Figure 5. Time evolution of solution concentrations difference ratio at both sides of the 26% AlqCl/74% CTA PIM for NaCl donor concentration $c_d^o = 0.01 \text{ M}$. Δc^t : solution concentration difference at time t; ΔC^o : initial solution concentration difference (t = 0).

The low permeability and barrier effect to NaCl transport exhibited by the PIM determined by different kinds of measurements is of interest for its application in natural water systems (which usually have also NaCl in solution and/or other salt) since it supports the elimination of their contribution with respect to carried-mediated transport.

Application of the PIM to the Separation of Sulfonamides

The transport kinetics of four sulfonamides across the PIM is a simultaneous combination of extraction and back-extraction reactions occurring in non-equilibrium conditions at the same time.

The fundamentals of sulfonamides (S) extraction at basic pH by Aliquat 336 incorporated in a PIM can be described by the generic stoichiometric equation (Equation (6)) [15]

$$[AlqCl-]_{mem} + Ant-_{aq} \leftrightarrows [AlqAnt-]_{mem} + Cl-_{aq}$$
(6)

where mem and aq refer to the membrane and aqueous phases, respectively.

As it is indicated in Equation (6), sulfonamides are extracted via formation of an ionic pair with the triocylmethyl ammonium cation while chloride is released in the aqueous solution. Thus, the release of the extracted sulfonamides is possible in a stripping phase containing high amounts of chloride anion.

The transient concentration profiles of sulfathiazole (STZ), sulfapyridine (SPY), sulfamethazine (SMZ), and sulfamethoxazole (SMX) in the feed and stripping solutions as a function of time are presented in Figure 6. The results show an important decrease of the concentration of both STZ (with a thiazole functional group) and SMX (which is the most acidic antibiotic in this study) in the feed solution the first 8 h of experiment. However, the transport of these species to the stripping phase is much slower. Hence, it can be observed that 60% of the initial STZ and 50% of SMX are remain in the membrane after 24 h of experiment, pointing out that the transport of these species through the dense membrane is the limiting step for their separation. SPY and SMZ exhibit less affinity towards Aliquat 336 due to their less acidic characteristics and their transport is not significant.



Figure 6. Transient concentration profiles in sulfonamides transport experiments using the 26% AlqCl/74% CTA PIM. (a) STZ; (b) SMX; (c) SPY; (d) SMZ. Triangles: feed solution; squares: stripping solution. (Feed solution: 5 mg L⁻¹ of sulfonamides at pH 9, stripping solution: 1 M NaCl).

3. Materials and Methods

3.1. Membrane Preparation

PIMs were prepared dissolving 200 mg of cellulose triacetate (CTA) in 20 mL of chloroform and stirred during 5 h. Afterwards, 1.3 mL of a 0.5 M Aliquat 336 IL (trioctyl methylammonium chloride or AliquatCl) solution in chloroform was added to the polymer solution and let stirred 2 h more. Then, the solution was poured into a flat bottom glass dish and the solvent was allowed to evaporate for 24 h at room temperature. The resulting film was carefully peeled off from the bottom of the glass dish. Prepared PIMs had a composition of 26% of AliquatCl and 74% of CTA and thickness of $30 \pm 3 \mu m$.

The IL Aliquat 336 and the polymer CTA were purchased from Fluka Chemie (Steinheim, Germany), while other chemicals are from Panreac (Barcelona, Spain).

XPS Measurements

Membrane surface chemical characterization was performed by analyzing XPS spectra obtained with a Physical Electronics spectrometer PHI 5700 (Physical Electronics, Chanhanssen, MN, USA), using X-ray Mg K α radiation (300 W, 15 kV, 1253.6 eV) as the excitation source. High-resolution spectra were recorded at a given take-off angle of 45° by a concentric hemispherical (±0.1 eV binding energy accuracy) and analyzed following the procedure and experimental conditions already explained in previous papers [24,38]. Measurements were performed with a membrane in wet state (26% AlqCl/74% CTA-w sample), which means, after their immersion in distilled water for 48 h, sample were gently surface dried with paper, and stored for one week to ensure the loss of the adsorbed water; XPS analysis of dry PIM sample (26% AlqCl/74% CTA-d) was already reported [18]. Moreover, a cellulose triacetate film (without IL) in dry and wet states (CTA-d and CTA-w, respectively) was also analyzed by XPS for comparison.

3.2. Elastic Measurements

Elastic characterization was carried out using a force digital gauge (ES20 model, Mark-10 Corporation, New York, NY, USA), with a maximum tension of 100 N, length accuracy of ± 0.01 mm, and at a strength rate of 10 mm/s, connected to a computer. Elastic measurements were performed with the membrane in air (sample 26% AlqCl/74% CTA-d) and placed in a distilled water tank (sample 26% AlqCl/74% CTA-w).

3.3. Electrochemical Characterization: Cell Potentials, Electrochemical Impedance Spectroscopy, and Salt Diffusion Measurements

Electrochemical characterization of the 26% AlqCl/74% CTAPIM was carried out in a dead-end test cell similar to that described in [39], with the solution stirred at 540 rpm to minimize solution concentration-polarization at the membrane surfaces and at room temperature (25 ± 2) °C.

Concentration or cell potential (ΔE) measurements were performed using two reversible Ag/AgCl electrodes connected to a digital voltmeter (Yokohama 7552, 1 G Ω input resistance) and changing the concentrations of the NaCl solutions at both membrane sides by keeping constant the concentration ratio $c_1/c_2 = 2$, from c_2 ranging between 0.005 M and 0.2 M. The membranes were maintained overnight in contact with the lowest concentration solution, but it was renovated previously to start the measurements. Membrane potential ($\Delta \Phi_{mbr}$) or equilibrium electrical potential difference between two solutions of the same electrolyte but different concentrations (c_1 and c_2) separated by the membrane were obtained from measured cell potential values by subtracting the electrode potential ($\Delta \Phi_{elect} = (RT/F)\ln(c_1/c_2)$, where *R* and F represent the gas and Faraday constants, and T is the system temperature [37], that is, $\Delta \Phi_{mbr} = \Delta E - \Delta \Phi_{elect}$.

Electrochemical impedance spectroscopy (EIS) measurements were performed with the membrane placed in the dead-end test cell between two NaCl solutions of the same concentration (concentration ranging between 0.01 M and 0.05 M), with Pt electrodes connected to a frequency response analyzer

(FRA, Solartron 1260, England), that is: NaCl solution (c)/electrode//membrane//electrode/NaCl solution (c) system [4]. Measurements were recorded for 100 data points for frequency (f) ranging between 1 Hz and 10⁷ Hz, at a maximum voltage of 0.01 V [4,31,33]. The impedance of a system, Z, is a complex number, $Z = Z_{real} + j Z_{img}$, derived from experimental data obtained from a broad range of frequencies, which can be separated into real (Z_{real}) and imaginary (Z_{img}) parts by algebra rules. For simple and homogeneous systems, Z_{real} and Z_{img} are related to the electrical resistance (R) and the capacitance (C) by the following expressions

$$Z_{\text{real}} = (R/[1 + (\omega RC)^2]); Z_{\text{img}} = -(\omega R^2 C/[1 + (\omega RC)^2])$$
(7)

where ω represents the angular frequency ($\omega = 2\pi f$). The analysis of the impedance data is usually carried out by plotting –Zimg versus Zreal (Nyquist plot), where a semi-circle corresponds to the parallel association of a resistance and a capacitance or (RC) circuit [35]. This description is only valid for homogeneous systems, which present a unique relaxation time τ (with $\tau_{max} = 2\pi RC$), but in the case of non-homogeneous systems the Nyquist plot is a depressed semi-circle due to a distribution of relaxation times, then a non-generalized capacitor (usually represented as Q) is considered [35]. Moreover, for composite systems consisting of layers with different electrical characteristics, a series association of semi-circles/depressed semi-circles (one for each layer) is commonly obtained [31–34].

For NaCl diffusion, the membrane separated a donor solution ($c_d = 0.01$ M) from a receiving diluted solution (c_r), which initially was distilled water (that means, $c_r^o = 0$) [39]. Time variation of donor and receiving solution concentrations was determined by measuring conductivity changes by means of two conductivity cells, each one submerged in a half-cell, connected to two digital conductivity meters (Crison GLP 31, Barcelona, Spain).

3.4. Transport Experiments

Transport across the PIM of four different antibiotics—sulfathiazole (STZ), sulfapyridine (SPY), sulfamethazine (SMZ), and sulfamethoxazole (SMX)—all purchased from Sigma-Aldrich (Steinheim, Germany) was analyzed. Antibiotics stock solutions (250 mg L⁻¹) were prepared in methanol. Experiments were carried out in a permeation cell similar to the one described in [15] in which the mechanically stirred (800 rpm) feed and stripping solutions (150 mL each) were separated by the PIM with an exposed membrane surface area of 11.5 cm². All experiments were conducted at an ambient temperature of 22 ± 1 °C.

Working solution containing a mixture of 5 mg L^{-1} each compound was prepared by appropriate dilution of the stock solutions with deionized water (Milli-Q Plus system, Millipore Iberica S.A., Madrid, Spain) and the pH was adjusted to 9 with NaOH. Sodium chloride (Carlo Erba, Cornaredo, Italy) was used to prepare the 1 M NaCl stripping solution. Samples were taken from the feed and stripping solutions at regular time intervals during the 24 h period of the experiments and were analyzed by an HPLC system (1200 Series Agilent Technologies, Santa Clara, CA, USA) with a diode array detector (1290 Infinity DAD Agilent Technologies, Santa Clara, CA, USA) following the procedure described in [15].

4. Conclusions

Electrochemical characterization of a polymer inclusion membrane prepared with the ionic liquid Aliquat 336 (26%) and the polymer cellulose triacetate (74%) was performed with a model electrolyte (NaCl) by measuring membrane potential, electrochemical impedance spectroscopy, and salt diffusion. These results enable the estimation of characteristic electrochemical membrane parameters such us the membrane conductivity, and its dependence with NaCl solution concentration, the diffusive permeability, and the ion (Na⁺ and Cl⁻) transport numbers, which clearly show the barrier effect of this membrane to the transport of NaCl. Moreover, membrane contact with water or aqueous solutions seems to affect both chemical surface characteristics and elastic parameters. The separation of sulfonamides using this membrane is affected by its dense character, which limits the transport of the organic molecules through it.

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