



Remiern

# **Eco-Friendly Processing of Wool and Sustainable Valorization of This Natural Bioresource**

Crisan Popescu 1 and Michaela Dina Stanescu 2,\*

- <sup>1</sup> KAO Germany GmbH, Pfungstaedter Str. 98-100, 64297 Darmstadt, Germany; crisan717@yahoo.co.uk
- <sup>2</sup> Department of Natural and Technical Sciences, Aurel Vlaicu University, 77 Revolutiei Blvd., 310130 Arad, Romania
- \* Correspondence: stanescu@uav.ro

**Abstract:** The environmental invasion of plastic waste leads to, among other things, a reassessment of natural fibers. Environmental pollution has shown the importance of the degradability, among other properties, of the raw materials used by the textile industry or other industrial fields. Wool seems to be a better raw material than the polymers that generate large quantities of micro- and nano-plastics, polluting the soil, water, and air. However, the usual processing of raw wool involves a number of chemically very polluting treatments. Thus, sustainable procedures for making wool processing environmentally friendly have been considered, leading to the reappraisal of wool as a suitable raw material. Besides their applications for textile products (including *smart textiles*), new directions for the valorization of this natural material have been developed. According to the recent literature, wool may be successfully used as a thermal and phonic insulator, fertilizer, or component for industrial devices, or in medical applications, etc. In addition, the wool protein  $\alpha$ -keratin may be extracted and used for new biomaterials with many practical applications in various fields. This review makes a survey of the recent data in the literature concerning wool production, processing, and applications, emphasizing the environmental aspects and pointing to solutions generating sustainable development.

**Keywords:** sustainable development in sheep breeding; wool a natural raw material; sustainable enzymatic wool processing; wool sustainable valorization

Citation: Popescu, C.; Stanescu, M.D. Eco-Friendly Processing of Wool and Sustainable Valorization of This Natural Bioresource. *Sustainability* **2024**, *16*, 4661. https://doi.org/10.3390/ su16114661

Academic Editor: Maria L. Auad

Received: 12 April 2024 Revised: 21 May 2024 Accepted: 27 May 2024 Published: 30 May 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).

#### 1. Introduction

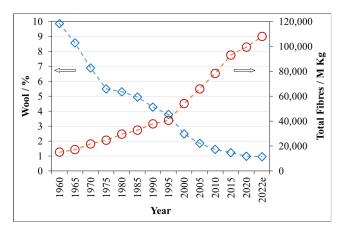
The pollution generated by micro- and nano-plastics, spread all around us, and disturbing the flora and fauna, has changed attitudes concerning the utilization of plastic materials [1–4]. Thus, in the textile field, interest has turned back to the natural fibers of vegetal (bast, cotton, etc.) and animal (silk and wool) origin [1].

Wool has accompanied mankind since the dawn of the latter. It features in legends ("golden fleece"), the Bible, and historical facts (The Woolsack in the Parliament of the UK). Over the centuries, wool has been an important fiber for textile producers. The products have ranged in quality and price from the relatively inexpensive to the ultra-luxurious scarlet woolens aimed at aristocratic markets [5].

Wool is sheep hair, and it is a raw material with many qualities, like elasticity, moisture management, good heat insulation due to air retention, gentle luster, etc. [6]. Wool products are comfortable to wear and are considered by consumers to be a renewable and environmentally friendly material. The study of Collie and coworkers [7] evidenced the high level of wool-product biodegradation in seawater (more than 20%), compared with cotton (10%) and synthetic fibers (less than 1%) during the same period. Wearing wool brings many benefits such as reduced body odor, improved sleep quality, protection against fire, and reduced eczema symptoms [8–10]. Despite all these excellent

Sustainability **2024**, 16, 4661 2 of 36

properties, wool is today only about 1% of the total amount of consumed fibers [11], as illustrated in Figure 1.



**Figure 1.** World fiber consumption evolution (blue-wool; red-total fibres) [11] (2022e: estimated value for 2022).

This situation is created by the complexity of wool processing, generating high water and carbon footprints [12]. The implementation of a circular economy policy requires serious investments, the pay-off taking time [13]. Indicators for comparing the Life Cycle Assessments (LCAs) of natural and synthetic fibers [14] and the products from such fibers [15,16] have been apprised.

The environmental impact depends also on the type of wool processed [17]. New indicators have been introduced for measuring wool, like *ZQ*, involving animal welfare, environmental sustainability, fiber quality, and the traceability of products, as well as social responsibility, together with *ZQRX* (regenerative index), measuring how well the producers compensate carbon, restore watercourses, and protect native species [18]. The *eco-label*, established in 2014 by the EU Commission [19], is another criterion for comparing the environmental impacts of different textile products [20]. Such labels stimulate producers and consumers to produce and, respectively, to buy *ecofriendly* products [21].

These environmental problems have appeared alongside the evolution of sheep farming due to food (land) and water consumption and pollution harms. The greenhouse effect is largely due to methane releases [22]. Wiedemann and coworkers [23] investigated the environmental effects of farming for wool supply and the production of woolen sweaters. Energy consumption is determinant as to wool processing and greenhouse emissions at the farming stage. But those environmental costs have to be considered alongside the benefits of other sheep products like milk and cheese, as well as meat. [24,25].

The economic analysis revealed that sheep breeding is not very efficient, but the social and cultural aspects, the traditions, the community cohesion, also have to be put in balance [26]. Further on, the trail of wool along the value chain, from farms to spinning mills and finished product, is important [27].

As is generally known, clothes are, together with food and shelter, basic needs for everyone. The fabrication of textiles for clothing involves the production and processing of fibers, followed by the finishing operations. All of these steps require water and chemicals consumption, generating pollution [28]. Natural fibers, such as wool, consume, for their processing, more water than do synthetic fibers. The water consumption for wool fabric is 200–300 kg/kg, less than for cotton (250–350 kg/kg), but more than the amount used for synthetic polyester (100–200 kg/kg) [29].

The evaluation of wool's value also has to consider that, in addition to its uses in textile material manufacture, it may be valorized in construction, agriculture, cosmetics, etc.

Sustainability **2024**, 16, 4661 3 of 36

The processing of wool fibers into textiles plays an important role in the pollution of the environment, and several solutions have been applied in efforts to reduce this. This paper aims mainly to survey the enzymatic processes available for wool preparation and finishing as a clean alternative to the traditional procedures, as well as some non-traditional and sustainable procedures for the valorization of wool.

# 2. Sustainable Development in Sheep Breeding

Farming systems may be cost-effective, diminishing the pressure to choose between biodiversity preservation and economic development [30]. Modern technology may also help animal husbandry farms to become cost-effective [31–33]. The level of technology adoption determines the increases in the ecological aspects of the pastoral system [34].

Several papers provide information regarding the efficient management of sheep breeding. There are many factors influencing its efficiency, like the relations with stakeholders [35], socio-technical networks in the field [27], animal genetics [36], animal welfare [33], flock management [37], and pasture productivity [38].

The implementation of a correct EU Common Agriculture Policy for supporting the pastoral economy is also of importance. Determining the correct value chain of the products and having a right appreciation of the profession, which can attract young people by introducing sustainable financial facilities, are compulsory [39]. Integration of crops and sheep breeding is of importance for land-use efficiency, improvements in productivity, and decrease in the environmental burden [40]. Also, special computer programs like Smart Sheep Breeder applied in India [41] have been developed for improving decision making in sheep breeding.

#### 3. Wool Structure and Properties

Wool, like other animal hairs, is a natural composite system, one which has a complex dual structure at all levels. The whole fiber appears like a ring/core arrangement, with the cortex wrapped by the cuticle. The cortex contains ortho- and para-cortical cells and the cell membrane complex. The cortical cells are further composed of macrofibrils and intermacrofibrillar material. The macrofibrils consist of keratin intermediate filaments (KIF) and an intermicrofibrillar matrix made of keratin-associated proteins (KAP) and cytoplasmatic, as well as nuclear, remnants. As such, the cortex matches, at all levels, the 'filament in matrix' model. By considering the lateral interactions between the KIFs and the surrounding matrix as an interphase, the more complex view of wool uses the three-phase model, which provides better explanations of fiber mechanics and reactions with chemicals [42–44].

Chemically, wool is a protein fiber made of  $\alpha$ -keratin, a fibrous protein. The elemental analysis of wool shows carbon, hydrogen, oxygen, nitrogen, and sulfur. The high sulfur percentage (around 5 wt%) results from the cystine content of the fiber. Total hydrolysis of the peptide bonds in proteins yields the 20 common natural  $\alpha$ -amino acids found in wool and their mol percentages, which are approximated by the values indicated in parentheses, namely, alanine (5%), arginine (7.2%), asparagine and aspartic acid (6%), cystine (11.2%), glutamine and glutamic acid (12.1%), glycine (8.1%), histidine (0.7%), iso-leucine (2.8%), leucine (6.9%), lysine (2.3%), methionine (0.5%), phenylalanine (2.5%), proline (7.5%), serine (10.2%), threonine (6.5%), tryptophan (1.2%), tyrosine (4.2%), and valine (5.1%) [45].

The peptide arrangement in wool fiber has been a topic of study since the early use of X-rays for investigating protein structures [46,47]. The X-ray diffraction pattern of wool shows a meridian reflection at 0.51 nm and an equatorial reflection at 0.98 nm. Interpreting these results, the  $\alpha$ -helical structure was proposed to give an account of the secondary structure of the keratin fiber [48]. The organization of the  $\alpha$ -helices in keratin intermediate filaments, KIF, which form the macrofibrils, the ordered (crystalline) part of the wool fiber, is driven entropically and follows a lateral and end-to-end association (polymerization) rule, as illustrated in Scheme 1 [49].

Sustainability **2024**, 16, 4661 4 of 36



**Scheme 1.** Keratin  $\alpha$ -helices organized to form macrofibrils [49].

Wool fibers, like any other animal fiber, have a slightly elliptic cross-section and are protected by the scales of cuticle arranged on their surface like tiles on a roof. These scales cause wool to have different fiber–fiber friction coefficients when measured along or against the scale orientation. This property of a differential friction coefficient is unique among textile fibers and is the reason for the felting ability of wool fibers [45].

Wool fibers also contain lipids. The wool grease is a complex mixture produced by sebaceous glands in sheep skin. In addition to this external grease, there are internal lipids trapped in the mass of wool proteins [50].

## 4. Wool Availability

Keratin fibers are available almost everywhere in the world, and wool produced by sheep is by far the most widely used keratin fiber. The total amount of wool produced yearly fluctuates with market demands but decreased over the last 70 years to around 1100 M Kg [11], as indicated by Figure 2.

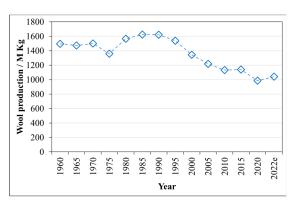


Figure 2. Wool, yearly production [11] (2022e: estimated value for 2022).

Economically the sheep is a green factory without wastes: consuming only water and grass as raw materials, a sheep produces wool, milk, and meat, and even the excrement is useful for fertilizing pastures. Because the sheep graze, not pulling the grass as the goats do, the pasture regenerates quickly, within weeks after the passage of sheep. Very roughly, a sheep produces annually 1 kg greasy wool (which means some 0.6 kg clean fibers) from 1 ha of average pasture [51].

#### 5. Classical Processing of Wool Fibers

Bringing wool to the textile circuit is a labor-intensive process. The first steps of shearing and collecting the greasy wool, followed by skirting (selecting the parts of the fleece) and classing wool according to the fiber diameter and strength, are manual operations. The collected greasy (raw) wool contains various amounts of impurities collected by the wool during the sheep's grazing, namely sand, dirt, and vegetable matters, to which one adds the grease and suint produced by sheep during its daily biological activity. Depending on the type of wool and the pasture quality, clean wool fibers range from 30 to 75% of the total greasy fleece collected from the sheep.

The numerous chemical and mechanical processes through which the wool fibers are turned into an end-product can be grouped into the following operations:

• Operations of cleaning the raw wool;

Sustainability **2024**, 16, 4661 5 of 36

- Operations which transform the fibers into fabric;
- Operations of cleaning yarns and fabrics;
- Operations of stabilizing the dimensions of fabrics;
- Operations of surface treatment and infiltration of fibers and fabrics;
- Operations for imparting a finished aspect to fabrics.

The details of the operations differ as to the ways the fabrics were obtained: by weaving or by knitting, respectively. In all of these cases, various chemical products are used for assisting in, or for producing, the required effects. Most operations make extensive use of surfactants for assisting processes, namely, for washing/scouring, for improving the quality of bleaching, of dyeing, or for finishing (i.e., softening and stiffening) the materials. This results in the further production of pollutants, which are either associated with effluents, or emitted into the atmosphere.

## 5.1. Operations of Cleaning the Raw Wool

For the delivery of clean fibers to the industry, the greasy wool goes through scouring, which is the operation of washing the raw fibers with about 1%wt surfactant in a continuous 5–6 bowl line in order to remove the grease, suint, sand, and dirt from the fibers. The waters discharged from the first three bowls used to wash fibers with more than 10% grease (usually from fine wool, Merino type) are further used for separating lanolin, an important ingredient in many personal and health care creams, from the effluent, by means of an Alfa-Laval process [52]. Despite lanolin separation, the wastewater from wool scouring is heavily polluted with organic matters, and much effort is dedicated to processes to clean and reuse them, aiming to reduce water consumption. The recent development of bio-degradable surfactants helps in the initiative to build a more environmentally friendly wool scouring process. Some 5–7 L of water are required to obtain 1 kg of clean wool under severely controlled parameters, but 20 L is more common.

The vegetable matters, if more than 2–3%, have also to be removed before the material is subjected to further processing. This operation, known as "carbonization", makes use of the good resistance of wool to strong acids (particularly sulfuric acid) and of the hydrolysis of cellulose by the same environment. The fibers with vegetable matters (e.g., grass and burrs) are soaked in a 20% sulfuric acid solution, dried and baked, and then crushed to separate the carbonized cellulosic matters from the rest. The fibers are then neutralized and dried. The process produces acid-polluted waters and carbon dust in the air, besides weakening the wool fibers, for which reason alternatives, like the use of enzymes, are sought.

## 5.2. Operations which Transform Fibers into Fabric

This group gathers several mechanical operations which help in the transforming of the fibers into a yarn, namely, carding, combing, and spinning, and further process a yarn into a knitted or woven fabric by knitting or weaving, respectively.

The making of the yarn requires the grouping of fibers into a roving of fairly even density, which is eventually spun. The fibers are kept together in the yarn by fiber–fiber friction, and the twisting movement during spinning ensures a compact packing. Providing that the fibers in the yarn are parallel or randomly arranged by the previous mechanical operations, the yarn is named worsted—its formation consists of a carding operation used to produce a web of fibers, one which is further combed to arrange the fibers in parallel before spinning them into a fine and smooth yarn—or woolen-carded yarn, for the latter of which the combing operation is not used, and the yarn is, thus, bulky. Any of these yarns can then further be used for producing a 3D structure (fabric) by knitting or weaving.

Alternatively, the non-woven fabrics are produced by directly transforming fibers from the carding web into a fabric by felting, without the formation of a yarn. The oper-

Sustainability **2024**, 16, 4661 6 of 36

ation makes use of the fiber-to-fiber friction of wool in forming a felt. Another way of making an end-product is by punching tufts of wool fibers into a woven support and using resins to bond the ensemble to produce a tufted product, usually a carpet. This felting process is also used for obtaining handcraft objects by using needle felting and lasers for cutting [53,54].

All of these operations use oils (1...2 wt%) for easing the mechanical action and reducing the breakage of the fibers during the process.

## 5.3. Cleaning Operations of Yarns and Fabrics

The *washing/scouring* and the bleaching are the operations used for cleaning the yarns and the fabrics. Scouring makes use of surfactants and warm water (around 40 °C) for washing out the oils and other chemical auxiliaries used in assisting the previous mechanical operations, namely, the spinning, knitting, and weaving. The *bleaching* process is carried out as a last step after washing the materials, and the chemical agents used for it depend on the types of fibers to be treated. For most of the wool fibers, the common and environmentally friendly bleaching agent is hydrogen peroxide of a 6% concentration. Some yellow wools may require a harsher bleaching, in which cases sodium dithionite or sodium formaldehyde sulfoxylate are used, with a resulting polluting impact made by the effluents.

## 5.4. Stabilizing Dimensions

Wool fabrics are dimensionally stabilized by using at least one of the following setting operations: crabbing and wet and dry decatising. The chemistry behind the process is based on the breaking and reformation of the interchain cystine bonds in the desired places by using a mechanical stretch of the fabric in a hot, wet environment, followed by a cold, wet shock and the release of the mechanical stress. The temperature difference of the shock is usually around 80–100 degrees Celsius. The time required for stretching in the hot, wet environment (water or steam) depends on the weight and density of the fabric, going from 2 to 5 min in saturated steam of 2 bar (120 °C), and from 30 to 50 min in hot water of 98 °C.

An alternative operation is chemical setting, which can cope with a larger amount of cystine bonds than any of the above processes, thus producing long-lasting effects. It makes use of thio-glycols under a stretching operation, followed by hydrogen peroxide reformation of the di-sulfide bonds, like in permanent waving of hair. The process does not require a temperature shock, but the chemical reagents required make it, however, too expensive relative to the usual end-products.

## 5.5. Surface Treatment and Coating and Infiltrating Operations

Surface treatments and coating and infiltrating operations cover the *coloring* and most of the wet-finishing processes. The *coloring* process renders most of the commercial value of a fabric. It is achieved either by coating (*pigment printing*) or by infiltrating (*printing* and *dyeing*) the fibers. While pigment printing binds by the action of a resin coating the coloring pigments, which have no affinity for wool, the other printing systems and the dyeing make use of molecules which diffuse and eventually bind on active sites of the fiber or aggregate inside the fiber to form difficult-to-remove clusters. The dyes used for wool fibers are the same as those from practically any dyeing class; it is only the disperse dyes which are of low interest for wool dyeing, due to their poor hydrophilicity and affinity, and this class remains dedicated exclusively to chemical fibers. The coloring process, no matter which class of dyes is used, pollutes the water with dye molecules that were not successfully bonded by the fiber, as the yield of dyeing cannot reach 100%. A coating operation is usable not only for pigment printing, but also for covering the fibers with certain chemical products which may impart new properties to the end-product, like, for example, an improved easy-care finish, as discussed further on.

Sustainability **2024**, 16, 4661 7 of 36

The wet-finishing operations comprise softening, milling and easy-care treatment. The softening process is achieved by using cationic or silicon-based surfactants (softeners) to improve the handleability of the fabric, particularly as to the smooth touch. The milling process makes use of the unique ability of wool to felt, because of the scales on the fiber surface which enable the directional frictional effect. The process is carried out by mechanical action applied to the fabric in a wet environment of controlled pH, with the help of ethoxylated surfactants, and produces the specific wool-like look of the material. The ability of wool to felt has a downside: the machine-washing and tumbler-drying of wool clothes at home change (shrink) the product dramatically. As a result, home easy-care operations are not possible for wool products, unless they receive a specific easy-care finish which can arrest the felting property of wool fibers. The easy-care treatment aims to erode the scales on the fiber's surface and/or arrest the fiber-to-fiber movement using chemical resins. The eroding of the scales is achieved by Allwörden's reaction, which involves a controlled cold chlorination followed by the neutralization of residual chlorine with sodium sulfite [55]. Resin is further applied in order to improve the results. The process is a heavily polluting one, as it releases large amounts of halogenated organic compounds (AOX) in the effluent, as well as gaseous chlorine to the atmosphere, for which reasons new methods are under investigation. Treatments with enzymes, with ozone, or with plasma, followed by resin application, are some of the available alternatives to chlorination. It is of interest to note that researchers identified that the wool fiber's ability to felt is a heritable trait, one dependent on fiber diameter and curvature [56]. Their results suggest that it would be possible to breed for naturally shrink-resistant wool, producing, on the back of the sheep, wool for easy-care products which would not need any special further treatment [57].

#### 5.6. Treatments for a Finished Aspect

The treatments for a finished aspect are, generally, mechanical operations which belong to the dry-finishing routine, and comprise raising, shearing, pressing (ironing), and steaming. The dry-finishing operations make use of vegetal or metal-made brushes for raising; helicoidal blades for shearing; hot cylinders, or plates, for pressing; and steamers for steaming. These operations are used to give the final aspect to the fabric's surface, and their main polluting product is the wool powder, formed mainly at shearing and collected by the vacuum cleaners of the finishing machines.

## 6. Pollution in Wool Processing

As previously described, conventional wool processing uses many polluting chemicals, consumes large quantities of water, and generates a large amount of wastewater. In agreement with the sustainable development goals [58], efficient processes need to be applied, with reduced consumption of chemicals and energy, safer solvents, and reduced amounts of by-products [59].

The main polluting steps of wool processing are shown in Scheme 2 [60].



Scheme 2. The main polluting steps of wool processing [60].

In some of these steps, the polluting chemicals may be replaced by enzymatic treatments [61]. The replacement of conventional procedures with enzymatic ones leads thus to decreases in the chemicals and energy required for the process and lowers the amount of byproducts, due to the specificity of the enzyme.

Sustainability **2024**, 16, 4661 8 of 36

## 7. Biotechnology in Wool Processing

Biotechnology replaces polluting chemicals with enzymes and represents an opportunity for future textile developments from the environmental, economic, and even public health points of view [62].

Enzymes are Proteases with catalytic activities. Their classification is based on the type of reaction catalyzed. For the textile industry, the types of enzymes mostly used are Oxidoreductases (class 1) and Hydrolases (class 3) [63]. A number of properties, like high catalytic activity, specificity for reaction, substrate regio- and stereo-selectivity, ability to work in mild conditions, and biodegradability, recommend the use of specific enzymes for sustainable processes. The representative enzymes applied in wool processing are presented below.

## Enzymes for Wool Processing

The most used enzymes in wool manufacture are *Hydrolyses*, enzymes that fragment different substrates by reaction with water. From among these enzymes, representative examples are detailed in the following.

*Proteases* are enzymes which break the chains of proteins. Proteins are biopolymers having as synthons amino-acid molecules. The structure of the protein is complex, being characterized by primary, secondary ( $\alpha$ -helix and  $\beta$ -sheet), tertiary, and in some cases, quaternary structures (see Figure 3) [64].

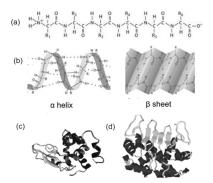


Figure 3. Protein structure: (a) primary, (b) secondary, (c) tertiary, and (d) quaternary [64].

The *Proteases* may be classified according to their active center structures as *serine*, *cysteine*-, *threonine*-, *aspartic*-, *glutamic*-, or *metallo-Proteases*, as well as *aspara-gine-Peptidelyases* [65]. The most widespread enzymes are serine-*Proteases*, which have a triad in the reaction center consisting of the following amino acids: *serine* (Ser), *histidine* (His), and *aspartic acid* (Asp) (see Figure 4). This splits the amide moiety of the protein by transforming it into a carboxylic acid (RCOOH) and an amine (R'NH<sub>2</sub>).

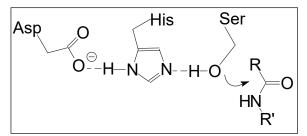


Figure 4. Catalytic triad of a serine-Protease.

To this class belong *Trypsin*, *Chymotrypsin*, *Elastase*, and *Subtilisin*. The cysteine-*Proteases* act as a dyad comprising a *cysteine* as nucleophile and a *hystidine* as base deprotonating the *cysteine*. *Papain* and *Bromelain* belong to this class. The threo-

Sustainability **2024**, 16, 4661 9 of 36

nine-Proteases act with only the threonine, which has both OH and NH2 groups involved in the catalytic process. Aspartic- and glutamic- Proteases have acid-based proteolytic mechanisms. Pepsin and Renin are included in aspartic-Proteases, and Scytalidopepsin B in the glutamic-Proteases. In metallo-Proteases, a metal, usually Zn and sometimes Co, is involved in the catalysis. Matrix Metallo Proteases are examples of such enzymes.

*Proteases* are involved in the processes of wool finishing, mainly with the antifelting treatments. A specific group of *Proteases* are the *Keratinases*. Keratin, the wool protein, is not easily degraded, due to the disulfide bonds [66].

*Keratinases* hydrolyze the wool protein chain. These enzymes, according to the reaction center, are *serine* or *metallo-Proteases*. For transforming keratin, beside *Keratinases*, an Oxidoreductase, namely, *Disulfidereductase* (EC 1.8.1.8), is needed [67]. In fact, the process of keratin degradation starts by the breaking of the –S-S- bonds, as catalyzed by the redox enzyme, resulting in –SH groups. This transformation changes the stereochemistry of the chain, making possible the access of the *Keratinase*, which may hydrolyze the amino acid chain either internally (*endo*) or at the N/C terminal (*exo*) [68].

*Cutinases* (EC 3.1.1.74) are *Esterases*, reacting with esters and also presenting the catalytic triad *Ser-His-Asp* [69]. These enzymes act on the wool cuticle's hydrolyzing lipids and protein bonds and thus reduce the hydrophobicity of wool [70].

Cellulases are Hydrolyses which fragment cellulose, a 1–4  $\beta$  glucose polymer. The bond cleavage is performed by an acid–base catalysis involving two dicarboxylic amino acids working in tandem [71]. Cellulose is found in the vegetal impurities accompanying raw wool. The classical procedure used for eliminating them is by means of treatment with concentrated sulfuric acid, as previously described. Such a process is very polluting and its replacement with an enzymatic treatment is desirable. The enzymatic removal of vegetative impurities on raw wool is known as bio-scouring. There are three types of Cellulases: endo- and exo-glucanase, which hydrolyze the cellulose chain inside or at the ends, and  $\beta$ -glycosidase, which hydrolyzes cellobiose, the glucose dimer. These types may be used together or, for more efficiency, mixed with Pectinases [72].

*Lipases* of microbial origin (EC 3.1.1.3) are also *Hydrolyses*, ones specific to lipids, catalyzing the hydrolysis of long-chain triacylglycerols. These enzymes may be used for eliminating grease from wool. Microbial *Lipases* belong to the *serine Hydrolyses* group [73].

Laccases (EC 1.10.3.2) are Oxidoreductases which improve the dyeing process by the polymerization of different phenols on the textile fibers, generating the chromophore compounds. The polymer remains fixed on the material, avoiding the loss of color with time [74]. Laccases are metallo-enzymes with four copper atoms in the reaction center which transport the electron during the redox process [75]. The huge number of applications of Laccase (dyeing and finishing of wool, discoloration of wastewaters) leads to the optimization of preparation procedures for obtaining a mass production of this enzyme [76].

The use of other enzymes, like *Collagenases* [77] or *Transaminases* [78], has also been mentioned.

Transglutaminases (EC 2.3.1.13) belong to the Transferases group, comprising enzymes moving the acyl group based on *glutamine* to different amines including the diamino acids from wool fibers. The treatment repairs previous damage instances by cross-linking, and improves dye fixation [79]. The discovery of microbial *Transglutaminases* enlarged their industrial applications due to facility in separation (extracellular enzymes) and activity in large intervals of pH (5–8) and temperature (40–70 °C) [80]. Besides the improvement in yarn resistance, the treatment increases wool softness and reduces pilling and felting [81].

The inconveniences associated with the applications of enzymes consist in their lower stability and sensitivity to the reaction conditions, and their relatively high cost. One solution for improving enzymatic treatments is enzyme immobilization [82]. The immobilized enzyme is more stable and can easily be separated and recycled. The pro-

Sustainability **2024**, 16, 4661 10 of 36

cedures for immobilization are performed by physical or chemical methods (see Table 1) [83].

	Table 1.	. Immobilization	procedures	for enzy	ymes	[83]	
--	----------	------------------	------------	----------	------	------	--

Methods	Advantages	Disadvantages		
	Physical			
	No modifications of the enzyme structu	re		
Adsorption	Simple, Regeneration of carrier,	Desorption; Enzyme subject to microbial attack		
	Reduced cost			
Entrapment/Encapsulation No modifications of the enzyme structure Diffusion problems; Leakage				
Chemical				
Covalent bond	Highon stability	Possible modification of the enzyme structure,		
Cooutent bonu	Higher stability	leading to reduced activity; Costly process		
Cupacliuking	Mara stable than free engume	Possible modification of the enzyme structure,		
Crosslinking	More stable than free enzyme	leading to reduced activity		

The possibility of separating and reusing the immobilized enzymes, which would reduce the process cost, endorses their application in numerous fields. The choice of carrier is of great importance.

The carrier's properties under consideration are the following: high stability, biocompatibility, insolubility, possibility of reuse, and lower cost [73]. A list of carriers compatible with plant *Proteases* is presented by Troncoso and coworkers. Natural and synthetic polymers, as well as inorganic oxides or magnetic particles, are described together with the plant enzymes immobilized on these carriers [84]. Immobilization of an alkaline *Keratinase* on chitosan-based carriers improved the thermal and operational stability of the enzyme [85].

To facilitate immobilized enzyme separation, magnetic carriers may be used. Carboxymethyldextran, combined with ions of iron, was employed for the magnetic nanoparticle, the enzyme being fixed covalently with glutaraldehyde or pentaethylenehexamine [86].

Enzymes are obtained from plants, animals, and microbes, the last source occupying almost 90% of the overall enzyme market [87]. Due to the wide area of industrial applications, the preparation of enzymes has lately been improved, with progress in extraction and purification methods, as well as in formulation, resulting in a larger quantity of enzyme production [88].

## 8. Processing Wool by Sustainable Methods

The replacement of conventional procedures with enzymatic ones leads to decreased chemicals and energy usage for the process and lowers the amounts of by-products due to the specificity of the enzyme.

#### 8.1. Bio-Scouring

The conventional scouring process described previously may be performed with enzymes like *Cellulases, Pectinases, Proteases, Cutinases, Lipases,* and *Collagenases,* which are good for selectively removing vegetable residue and grease from the raw wool.

A mixture of hydrolytic enzymes (*Bactosol WO*, Clariant) was successfully used to remove the wool grease [89]. Comparison of the chemically and enzymatically treated wool revealed a better result for the procedure when using the enzyme mixture as scouring agent [89].

A thermophilic lipase, produced by the bacterial strain *Bacillus aerius*, was employed successfully to process raw wool. It produced good results for scouring and dyeing wool, both in the same bath, thus reducing energy and water consumption [90].

Sustainability **2024**, 16, 4661 11 of 36

An immobilized *Lipase* was used to eliminate the wool surface lipids without damaging the interior of the fiber. An enzyme, produced by an extremophilic organism, was employed for this purpose, and the immobilization stabilized the enzyme and made its separation and reuse possible after each treatment. In this way, an enzyme covalently immobilized on a polyethylene imine–sericin hybrid may be used up to five times [91]. The same carrier has been used for immobilization of a *Protease* from *Bacillus safensis* FO-36bMZ836779, with treatment studies showing good results as to improvement in wool shrink-proofing [92].

By using *Proteases*, the scouring process is improved and the cuticular layer modified, increasing the dyeability of wool fibers. Elimination of lipids may be performed with *Keratinases*. The immobilization of the enzyme made it reusable and consequently reduced the cost of the process [85].

The treatment with *Cutinases* followed by *Proteases* is more efficient in wool scouring [93].

The combination of the enzymatic treatment with silver nanoparticles improved the results for *Cellulases* but not for *Lipases* [94].

Along with enzymes as scouring agents, ultrasonic irradiation also reduces the environmental impact of the process without damaging the quality of the wool fibers [95,96]. The development of the ultrasonic scouring procedure creates a cleaner and greener process, making the use of wool fiber more sustainable [97,98].

Summing up, the use of enzymes during wool scouring is a viable alternative to the traditional scouring, providing that the mechanical action is retained. The results for bio-scouring establish that, in addition to wax, vegetable matter and undesirable fibers can also be removed, and at a lower temperature than classical scouring, saving energy.

The water of the washing process may be reused after treatment with cationic polyamide to separate the solid waste, from which lanolin is to be extracted, with an ethanol–methanol mixture [99].

Another procedure for wool cleaning without water is based on the use of supercritical (sc) CO<sub>2</sub>. The white index is increased, and the Gram-positive and Gram-negative bacteria deactivated by treatment with scCO<sub>2</sub> [100].

Wool scouring should preserve as much as possible the properties of the fibers, such as luster, tensile strength, humidity, hygroscopicity, etc. Enzyme scouring, being a milder process than the classical one, best protects the wool fiber's properties. An after-scouring treatment of wool using *Transglutaminase* improves even more the wool's luster, softness, and tensile strength [101].

#### 8.2. Carbonization

The classical way, as described previously, to remove vegetable matters contained in raw wool is by a treatment with sulfuric acid. The procedure is very polluting, and many attempts have been made to replace or reduce the quantity of the sulfuric acid. Treatments with enzymes like *Cellulases*, *Pectinases*, *Hemicellulases*, and *Xylanases* have been investigated in this respect. So far, no single enzymatic treatment for wool carbonization has been suggested [93].

Cellulases may be used to make the procedure eco-friendly. There were a few papers published on this topic some years ago showing the cellulolytic treatment as having a reduced efficiency [102,103]. A combination of enzymatic and chemical treatments was proposed by Sedelnik [104]. The advantage was associated with the reduction of the quantity of sulfuric acid, which means a lower degree of pollution. A mixture of Glucanases (EC 3.2.1.4), acid Pectinase (EC 3.2.1.15), and Xylanase (3.2.1.8) proved more efficient, being successfully used for wool treatment without sulfuric acid [105]. No recent publications concerning biocarbonization were found.

Sustainability **2024**, 16, 4661 12 of 36

## 8.3. Bleaching

This process eliminates the last impurities and improves the whiteness of the wool. The most used agent is hydrogen peroxide. The procedure is applied under various conditions, either acidic, or alkaline. By means of this treatment, it is claimed that wool obtains a smoother surface than that of the untreated fibers [50].

The addition of enzymes is recommended in order to reduce the bleaching time. For instance, combining bleaching with a *Protease* treatment improves the resultant wool's properties [106].

The hydrogen peroxide may also be produced in situ by using *Glucose oxidase*, which catalyzes the transformation of glucose to gluconic acid and hydrogen peroxide [107] Promising results have been obtained by using this procedure for wool bleaching [108].

## 8.4. Antifelting Enzyme Treatments

One particular property of wool is its ability to felt, due to the scales on the fiber surface, which lead to interlocking of the fibers and shrinking of the material. Shrink-proofing treatments are required in order to make wool materials machine-washable. As has been described above, the treatments either etch the scales and/or cover the fiber surface with a polymer. Such treatments are very polluting due to the use of chlorine.

Hassan and Carr [109] compare several methods used for anti-felting treatments including oxidative, enzymatic, and plasma procedures. The authors stated that the chlorine-Hercoset procedure is the most effective treatment but underlined the negative environmental impact of it. The enzymatic treatment is less efficient and difficult to control, the *Proteases* having the potential to damage the wool fibers.

A new *Protease*, a fungal enzyme AtP produced by *Aspergillus tubingensis*, proved to be successful in reducing felting and eliminating lice eggshells from wool [110].

A mixture of proteolytic enzymes was produced by *Bacillus* sp. 51, containing serine *Proteases* and *Metallopeptidases* in combination with *Oxidoreductases* capable of breaking the –S-S- bonds. These enzymes reduce the felting of wool top with a very small weight loss and no fiber damage [111].

A combination of enzymes produced by *Bacillus* sp. 51 or *Bacillus patagoniensis* PATO5<sup>T</sup>, when used with a biosurfactant in the anti-felting treatments, reduced the enzyme diffusion into wool by acting mainly on the fiber's surface [112].

*Proteinase K,* which is produced by a fungus (*Tritirachium album Limber*), and easily hydrolyses keratin, was also efficient when used in in shrink-proofing and anti-pilling treatments of wool [113].

A recycled keratin extract treatment proved to be efficient as an anti-felting solution, improving also the softness, whiteness, and dyeability of the treated wool. [114].

Shrink-proofing treatments have been performed with *Proteases* extracted from a plant (*Cynara cardunculus* L.). The treatment ensures a good shrink resistance for wool yarn and fabric [115].

Treatments with *Lacase* or a *Protease*, followed by coating with poly-carbohydrates (chitosan, wheat, gum arabic, or starch), have been performed. The best results for shrink resistance have been obtained by the combination *Protease*—chitosan [116]. The treated wool has a reduced area shrinkage and unaffected tensile and bending properties.

Another enzyme used in anti-felting treatments is *Savinase* (EC 3.4.21.14), which is produced by *Bacillus lentus*. *Savinase* 16 L combined with an organic phosphine [P[(CH<sub>2</sub>)<sub>n</sub> OH]<sub>3</sub>, n = 1 to 10)] as activator, and employed in a multiple short time padding process which may be applied successfully at an industrial level [117].

To avoid damage to the wool fiber by the *Protease, Savinase* was covalently immobilized on a complex carrier based on poly (ethylene glycol) bis (carboxymethyl) ether and L-cysteine: HSCH<sub>2</sub> NH (COOH) COCH<sub>2</sub>[OCH<sub>2</sub>CH<sub>2</sub>]<sub>n</sub>OCONH(COOH)CH<sub>2</sub>SH. A good anti-felting result was obtained by this treatment [118].

Sustainability 2024, 16, 4661 13 of 36

> Proteolytic extracellular enzyme produced by a thermophilic bacterium (Bacillus safensis FO-36bMZ836779) was extracted and used for a wool treatment, both free and immobilized on an activated agar carrier. The treated wool became machine-washable without significant deterioration of the fibers [92].

> The proteolytic enzyme treatment, besides its anti-felting effect, improves the whiteness and dyeability of the wool [119].

> Mixed treatments have also been tried, some of them being successful. For example, the Protease treatment was combined with different pretreatments like corona discharge or plasma, as well as with other enzymes such as Cutinases, Lipases, and Transaminases, and the results in terms of shrink-proofing and whiteness degree were encouraging [120].

> Also, the combination of a commercial Keratinase with chitosan improves the dimensional stability of wool, as well as its wettability and dyeability [121].

## 8.5. Reduced Pollution in the Dyeing Operation

According to an evaluation of the footprint of the processing of woolen textiles, the dyeing phase is the most toxic for the environment [60]. Textile dyeing wastewaters represent 75% of all wastewater containing dyes [122]. Thus, the application of sustainable, ecologically friendly solutions seems appropriate, and several recent studies have addressed this topic.

One issue relevant to diminishing the environmental impact of dyeing is the use of natural dyes. The classification of natural dyes based on their sources is given in Figure 5 [123].

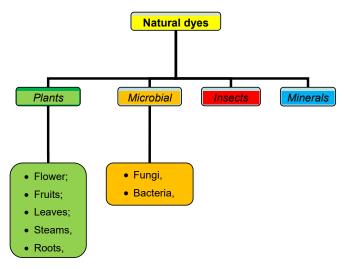


Figure 5. Sources for natural dyes [123].

Table 2. Dyeing wool with natural dyes.

Numerous studies have been dedicated to examining this aspect and, as a result, there are various solutions which have been suggested for dyeing various colors (see Table 2).

Source	Color	Procedure

Source	Color	Procedure	Properties	Lit.
Marigold flower as powder	Yellow	Mordants: Alum, Iron or Tin Chloride	Alum and Tin mordant bright color, Iron dark; Light fastness: 5; Wash fastness: 4.5–5	[124]
Onion skin	Dark yellow	Plasma, Alum as mordant, Dyeing by spraying	Wash fastness: 4.0; Reduced energy and water consumption	[122]

Sustainability **2024**, 16, 4661 14 of 36

Nutshell	Brown	Mordants: Alum, Iron sulfate and Chitosan	Wash fastness: 4–5; UV-protection for skin	[125]
Little hogweed	Brown	Mordant: Banana tree leaf	Wash fastness: 4–5; UV and Antibacterial protection; Di- minished toxicity (bio-mordant)	[126]
North Indian rosewood	Dark red	Mordants: Aloe Vera and Amla	Wash fastness: 4–5; Antibacterial protection; Diminished toxicity (bio-mordant)	[127]
Camphor tree	Brown	-	Wash fastness: 4–5; UV and Antibacterial protection	[128]
leaves	Pink Pink–Red Red	Mordants: Arjun, Gallnut, Pomegranate, Citric acid, and Chlorophyl	Wash fastness: 4–5; Light fastness 4–5 (with higher quantity of mordant); Diminished toxicity (bio-mordant)	[129]
Black rice	Pink (pH 3) Yellow (pH 10)	Mordants: Copper sulfate and Aluminum sulfate	UV-protection to skin	[130]
Kesudo herb Cinnamon herb Goldenrod herb	Yellow–Red Red Brown		Wash fastness: 4–5; Light fastness poor; Diminished toxicity (no mordant) Covers Sustainable Development Goals: 3, 13, and 14	[131]
Cinnamon bark	Yellow	Mordants: Henna leaves, Rose petals, Pomegranate peels, and Turmeric rhizome	Wash fastness: 4–5; Light fastness: 4–5; Rub fastness: 4–5; Diminished toxicity (bio-mordant)	[132]
Common basilisk	Yellow to Yellow–Brown	3	Wash fastness: 4–5; Rub fastness: 4–5; Light fastness: 5–8, depending on mordant type and quantity	[133]
	Red	Chitosan-polypropylene imine dendrimer	Wash fastness: 3–4; Light fastness: 7	[134]
Cochineal beetle	Red-Orange	Mordants: Henna leaves and Pomegranate peels; Microwave	Wash fastness: 3–4; Light fastness: 3–5; Diminished toxicity (bio-mordant)	[135]
	Red-Orange	Plasma; Dendrimer (Polypropylene imine)	Improved dyeability	[136]
Madder	Red (Al, Sn, Ni, Co, Zn); Grey (Fe, Cr); Brown (Cu)		yWash fastness: 4 (Cu, Cr, Al+Cu, Fe+Cr, Fe+Zn, Sn+Cr, Co+Ni, Co+Cr); 4.5 (Al+Cr, Cu+Cr); Light fastness: 8–7	[137]
Madder Gardenia (blue and yellow)	Combination of dyes: Red, Violet Yellow, Green, Brown	-	Wash fastness: 5; Uniformity of dyeing; Diminished toxicity (no mordant)	[138]
Madder Cochineal beetle	Red	Mordants: Chitosan and a den- drimer (methyl acrylate + eth- ylene diamine)	Antibacterial and antioxidant properties	[139]
Madder Reseda	Red Beige	Mordant: Pomegranate peel; US: dye extraction	Wash fastness: 4–5; Light fastness: 4–5; Diminished toxicity (bio-mordant)	[140]
Madder Weld	Red Beige	Mordant: Oak	Wash fastness: 4–5; Rub fastness: 4–5; Light fastness moderate;	[141]

Sustainability **2024**, 16, 4661 15 of 36

Purple cluster ge-			Diminished toxicity (bio-mordant) Wash fastness: 4–5;	
ranium	Yellow-Red	Mordant: Tin chloride	Light fastness: 4–5,	[142]
Tariiuiii			Wash fastness: 4–5;	
Roasted peanut		M:		[140]
skin	Red	Microwave	Light fastness: 5–6;	[143]
			Diminished toxicity (no mordant)	
Henna + Acacia	Dark red	Chitosan;	Wash fastness: 4–5; Light fastness: 4–5;	[144]
nilotica pod (An)		Mordants: Tannins from <i>An</i>	Antibacterial activity	
		Mordant: Alum; Nano Ag,TiO2, Al2O3	Antibacterial activity	[145]
Walnut dyes	Brown		Wash fastness: 4; Light fastness: 5;	
, rumius al jes	210 ((1)	Mordants: Acacia bark, and	Rub fastness: 4–5;	[146]
		Turmeric	Diminished toxicity (bio-mordant)	[110]
Pomogranato	Beige	Mordants: AgNO 7nO and	Wash fastness: 4–5;	
Pomegranate Walnut Green		Mordants: AgNO3, ZnO and Cu2O		[147]
wainut Green	Brown		Light fastness: 5–7	
Saffron flower	Brown to Green	Mordants: Alum, Cu and Fe sul-		[148]
0.16 . 11 6		fates and Tin chloride	Acidic and alkaline perspiration: 4–5	
Sulfonated kraft lignin	Yellow	Hydrogen peroxide	Wash fastness: 4	[149]
M	Green	Mordants: Alum, Aluminum	Wash fastness: 4–5; UV protecting;	[150]
Mugwort leaves	Brown	chloride, and Ferrous sulfate	Antibacterial	[150]
T. 1.	D 1	0 1.0 6.1	Wash fastness: 4–5;	
Talaromyces atr.	Red	Genetic modification of the pig-	Acidic and alkaline perspiration: 4–5;	[151]
TRP-NRC	Brown	ment source	Diminished toxicity (no mordant)	[]
			Wash fastness: 5: Rub fastness: 4–5:	
Red grape pomace	Red	Mordants: Tannic acid, Mimosa	Light fastness: 4–5;	[152]
rica grupe pormice	1100	extr.; US	Diminished toxicity (bio-mordant)	[10_]
		Mordants: Turmeric rhizomes,	, , , , , , , , , , , , , , , , , , ,	
Harmal seeds	Light brown		Acidic and alkaline perspiration: 4–5;	[153]
Tarmar seeds	Light blown	Pomegranate peels	Diminished toxicity (bio-mordant)	[155]
		Mordants: Pomegranate peels,	Diffinished toxicity (bio-mordant)	
Tea leaves' tannins	Brown	Acacia bark and Turmeric rhi-	Good fastness;	[154]
rea leaves tailinis	brown		Diminished toxicity (bio-mordant)	[154]
D	D	zomes		
Rose geranium	Brown	-	Diminished toxicity (no mordant)	[155]
waste	Beige			
Lac insect		Mordants: Turmeric rhizomes	Wash fastness: 3–4;	
(Lacaic acid)	Deep purple	and Acacia bark T	Light fastness: 4–5;	[156]
			Diminished toxicity (bio-mordant)	
Melanoid from			Wash fastness: 4–5; Antibacterial;	
Lycium barbarum	Brown	Pomegranate peels as mordant	Acidic and alkaline perspiration: 3–5;	[157]
			Diminished toxicity (bio-mordant)	
Alkanet		Mordante: Acacia Turmoria	Wash fastness: 4;	
	Red	Mordants: Acacia, Turmeric, and	Light fastness: 4–5;	[158]
(Alkanna tinctoria)		Pomegranate; US	Diminished toxicity (bio-mordant)	
T 1			Wash fastness: 4;	
Trachyspermum 	Yellow-Red	Mordant: Date seeds	Light fastness: 5–6;	[159]
coptiocum			Diminished toxicity (bio-mordant)	
			Wash fastness: 4–5;	
Reseda	Yellow	Mordant: Wild olive	Light fastness: 4–5;	[160]
_100000	1 0110 44		Diminished toxicity (bio-mordant)	[-00]
			Eministrea toxicity (Dio-Intordant)	

Sustainability **2024**, 16, 4661 16 of 36

Wild tumeric Yellow Mordants: Acacia, Pomegranate, Usah fastness: 4–5;

And Pistachio Diminished toxicity (bio-mordant)

For fixation of the natural dyes the dyeing needs to be performed on pre- or post-mordant-treated wool. The mordant is either a metal salt or an organic compound containing OH functional groups. The dye fixation is performed with the metal ion in the metal salt case and with the functional groups in the bio-mordant case. The fixation of the dye is due to the bonds created between the wool surface and the dye (see Figure 6).

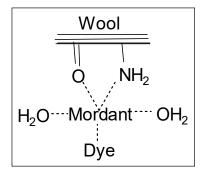


Figure 6. Dye fixation to wool through a mordant.

Also, ionic bonding with the COOH group from the dye is possible in the cases of dendrimers (Den) used as mordant (see Figure 7).

$$\begin{array}{ccc} & \overset{\oplus}{\operatorname{NH_3}} & \overset{\ominus}{\operatorname{OCODye}} \\ \operatorname{WoolCONH--- Dendrimer} -- \operatorname{NH_3}^{\overset{\oplus}{\operatorname{OCODye}}} & \overset{\ominus}{\operatorname{OCODye}} \\ & & & & & \\ \operatorname{NH_3} & & \operatorname{OCODye} \end{array}$$

Figure 7. Dye fixation to wool through a dendrimer with amino terminal groups as mordant.

Usually, the natural dyes have oxygen for coordination with the mordant, either as OH or as C=O groups (see Figure 8).

Figure 8. Chemical structure of components from natural dyes.

The color hue of natural dye is shifted after complexing with mordants, and different mordants result in different color changes, causing problems in controlling the color stability. Less toxicity and better fixation have been realized by using mixtures of plant extracts [138]. The use of organic mordants is considered ecologically sound and has solved the problem of dye fixation and that of the fastness of the dyed wool.

Sustainability **2024**, 16, 4661 17 of 36

Another aspect of the making of sustainable dyeing processes is the reduction of dyeing energy. Experiments have been performed studying dyeing with natural dye by using green energy, and a reduction of 24% of the power supply was realized [162].

Reduction of dye pollution was also achieved by the synthesis of the dye on the wool fiber through polymerization by oxidation of small organic compounds. Those include a dyeing process in the presence of a *Peroxidase* (EC 1.11.1.7), using various organic compounds like 1,4-dihydroxybenzene, 2,7-dihydroxynaphthalene, catechin hydrate, 2,5-diaminobenzenesulfonic acid, 3-amino-4-hydroxybenzenesulfonic acid, 1,4-diaminobenzene, 2- and 4-aminophenol. The dyeing proved to be efficient under mild conditions without damage to fibers, and with good fastness to washing and light. Moreover, depending on the pH used, different colors and shades may be obtained [163].

One of the catalysts often used for dyeing by means of the polymerization of small molecules is *Laccase*. Some examples are presented further on.

Wool was dyed by Laccase polymerization of aromatic phenols or amines. Brown shades were obtained with 1,4-dihydroxybenzene or 2,5-diaminobenzenesulfonic acid, and beige-to-green with 2,7-dihydroxynaphthalene. Very good wash fastness was obtained for all the dyed materials [164]. 1,3-Dihydroxybenzene treated with Laccase together with 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) dyed wool, giving a material with good fastness properties. The dyeing bath may be reused (six cycles); the cost of the dyeing process is thus reduced [165]. Starting from pyrrole, by treatments with Laccase, wool was dyed. The resulting polypyrrole was fixed to the wool fiber by covalent and hydrogen bonds. The color depends on pyrrole quantity, going from green to black. The dyed wool has good wash and light fastness and is conductive, and therefore suitable for technical textile applications [166]. A pale-yellow color was obtained by treatment of wool with ferulic acid in the presence of *Laccase*. The dyed wool has a wash fastness of 4–5 and improved antioxidant and deodorizing activities. The light fastness showed poor values (3), the dyed wool being not fit for applications demanding high light-fastness values [167]. Dyeing wool by 2,5-diaminobenzenesulfonic acid polymerization catalyzed by Laccase was the basis of a series of experiments which used different pH values. The color obtained depends on the pH value, ranging from dark purple (pH 1.8) to yellowish-brown (pH 10). The structure of the polymer is complex (linear polymer of 2,5-diaminobenzenesulfonic acid including phenazine moieties) [168]. Gallic acid and catechin are also used for wool dyeing by Laccase catalyzed reactions. The resulting polymer is fixed in the wool fiber by covalent bonds. Better properties for the dyed wool are obtained when polyethyleneimine (PEI) is added in the dyeing bath. It seems that PEI, by binding through hydrogen bonds to wool fibers, confers a shrink resistance [169]. Caffeic acid polymerized in the presence of polyethyleneglycol gave a brown shade to wool. The material has better wettability and electroactivity [170]. Syringic acid was used as monomer, in the presence of Laccase and dyed wool pretreated with 1-ethyl-3-(3-dimethyl aminopropyl) carbodiimide hydrochloride. A deeper yellowish-brown color and better fixation of the polymer on the wool resulted [171].

The *Oxidoreductases* are also efficient in improving dyeing with natural dyes. One of the problems regarding natural dyes is the relatively reduced washing fastness compared with synthetic dyes. Bai and coworkers [172] have applied *Laccase* catalytic polymerization for dyeing wool to gallnut, grape seed, and turmeric extracts, obtaining a material with good washing fastness. Green tea extract with *Laccase* is the basis of an efficient wool dyeing procedure by the polymerization of phenolic compounds from the extract. The dyed wool has antibacterial, antioxidant, and UV-protective properties [173].

Enzymes may be used for improving dye fixation by modifying the fiber's surface. Madder as dye with bentonite as mordant has been applied to wool pretreated with microbial-*Transglutaminase*. The enzyme catalyzes cross-linking lysine-glutamine on the wool fiber's surface, helping the fixation of the dye. An increase in the washing fastness properties was observed for the wool pretreated with enzymes [174].

Sustainability **2024**, 16, 4661 18 of 36

Another enzymatic pretreatment of wool fiber was applied in dyeing with bloodroot seeds (beige) and madder roots (red) extracts. The pretreatment enzyme was a mixture of *Amylases, Lipases*, and *Proteases* produced by animal pancreas. The madder root extract contains salicylic, ellagic, and benzoic acids and quercetin, with OH, C=O and COOH functional groups that are expected to attach on wool [175].

Other enzymes used for the modification of the surface of wool fabric are kerationlytic *Proteases* produced by *Streptomyces harbinensis* and *Streptomyces carpaticus*. This treatment, coupled with functionalization, helps to provide UV protection, antibacterial activity and washing fastness [176].

Enzymatic pretreatment also improves dyeing with synthetic dyes [177]. Protease and sodium alginate treatment improved wool dyeability with reactive dyes [178].

Enzymatic treatment may help the surface patterning in wool/polyester blends, patterns in relief resulting due to wool degradation by the *Proteases* [179].

Besides the choices of dyes and pretreatments, water consumption represents a problem in the dyeing process. Recent publications analyzed this aspect and proposed some ecologically sound solutions. For instance, the water from bleaching operation may be further used for dyeing, with consequent improvements in the dyeability [180]. The reduction of water for the dyeing has to be accompanied by the addition of auxiliaries in order to prevent dye aggregation, which would lead to an uneven dyeing [181].

Some studies have revealed that wastewaters may be successfully reused, thus reducing water consumption [182].

Magnetically treated water presents another opportunity for dyeing wool, reducing the dye consumption, and improving dyeing quality [183].

Engineered water nanostructures were efficient in improving the pilling-resistance of knitted wool fabric [184].

#### 8.6. Bio-Treatments for a Finished Aspect

New properties of a wool may be developed by grafting the fibers with different compounds. Enzymes play an important role in this process. Enzymatic treatment with *Laccase* to change the wool yarn shape by the grafting of tyrosine was successfully performed [185]. Wool grafted with  $\beta$ -cyclodextrin was prepared using *Laccase* as catalyst [186].

Ecologically sound solutions have also been considered for imparting anti-bug properties to textiles, and natural, biodegradable compounds are used for this finishing operation [187]. For example, the mothproofing treatment of wool with nano kaolinite showed promising results [188], and rare earth ions addition proved efficient in an anti-bacterial treatment [189].

## 9. Sustainable Aspects in the Valorization of Wool

#### 9.1. Textile Products

Wool is primarily a valued textile fiber. The fact that wool cloth is good to wear in all seasons is an important feature which promotes the use of wool for cloth manufacture based on woven or knitted fabrics. An analysis of knitted wool has revealed the variation of the pores (opening and closing) by sweating and shivering to keep the skin dry and give comfort and safety, regardless of the season [190].

Depending on the fiber diameter and length, wool can be used for both interior textiles (medium wool, 24.6...32.5 micron, and coarse wool, with diameters of more than 32.5 microns) and for apparel (fine wool, up to 24.5 microns in diameter, and medium wool), as shown in Figure 9.

Sustainability **2024**, 16, 4661 19 of 36

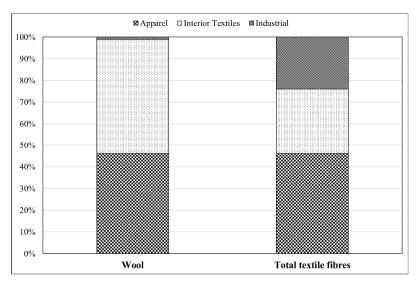


Figure 9. World fiber consumption by end-use, in 2022 [11].

The price of raw wool fiber is well above those of the other textile fibers (cotton and other cellulosic and synthetic fibers), being 4 to 10 times higher than any of them [11]. This supports wool's position as a premium fiber for textiles (apparel and interior products) and explains the small amount used for commercially cheaper industrial applications (around 1% of total wool fibers), like felts for filtering, or house insulation, which is described further on.

A strong type of wool is needed for carpet production, which comprises 45% of the world's wool production, New Zealand being the largest supplier [191]. According to Broda and coworkers [192], the coarse wool carpets produced by the tufted technique seem to be very good sound absorbents. The works of Kobiela-Mendrek and coworkers support the same assumption [193].

Knitting is another way of processing wool. The improvement of knitting machines, digitalization, and robotization ensure a sustainable development implementation in this wool-exploitation area [194].

Blends with synthetic fibers have better properties and longer lives. Wool and propylene yarns have a reduced pilling tendency and are thus suitable for winter socks [195].

As discussed above, the processing of raw wool for obtaining woven or knitted materials involves a number of polluting steps. A group of researchers from Norway proposed the reduction of pollution by local production development, which eliminates the pollution of transport operations and develops the transparency of the value chain and the concern for the environment [196].

While wool is primarily used for textiles, for which the diameter of the fiber dictates the end-use, more recent research has put into evidence new fields of interest for which the diameter of the fiber does not play any role. The environmental cost of wool processing has imposed the discovery of new applications in order to justify the expenditure. Thus, a performed *Strengths*, *Weaknesses*, *Opportunities*, *Threats* (SWOT) analysis revealed that wool may be used successfully, instead of plastic materials, in many products other than clothes [197].

## 9.2. Other Usages of Wool

The sustainable development principle has stimulated the occurrence of many technically valuable applications for wool. Some examples of these new wool-applications are presented.

Sustainability **2024**, 16, 4661 20 of 36

#### 9.2.1. Smart Textiles

One such application is the production of *smart textiles* serving as wearable electronic sensors (e-sensors). Thus, wool socks have been produced which can monitor the number of footsteps [198]. Knitted wool coated with silver nanowire may be used as electronic textiles, which can used as e-sensors for body motions [199]. Efficient motion e-sensors have been prepared by coating knitted wool with a suspension of graphene nanoplates, in acetone, by sonication. The prepared sensors are highly stretchable and flexible, being able to build up and to discharge electric charges, property applicable for their functionality as super-capacitors [200].

Studies considering the deposition of conductive components on wool single-jersey have been performed by Wilson and coworkers [201]. The fabric was treated with graphene ink, reduced grapheme oxide, and three encapsulants. A good interaction between graphene and wool was evidenced, with deposits found on the wool scales, while the encapsulants filled the interstitial spaces. The authors underlined the fact that the deposition of conductive components on the textile depends on the structure of the textile material, no general procedure being applicable.

The successful involvement of wool as raw material for obtaining *smart textiles* for super-capacitors is mentioned by Grube and coworkers [202]. Merino wool, used with nylon and coated with conductive polymers like poly(3,4-ethylenedioxythiophene) and poly(styrenesulfonate), enables the fabrication of good-quality *smart textiles* for sensors [203].

Highly conductive wool fabrics have been prepared by a multi-step process by coating wool with an ink based on graphene nano-platelets and particles of carbon black. Low voltage heating devices have been fabricated from such products [204].

A substrate based on wool with silver nanoflower deposition was employed for a successful surface-enhanced Raman scattering measurement of traces of the pesticide Carbendazim [205].

## 9.2.2. Agriculture Applications

Wool is a perfect source of carbon and nitrogen. Thus, sheep's wool may be used as substrate for the cultivation of different plants. For example, the use of sheep's wool in cucumber cultivation reduces the water consumption as well as the contributions to the greenhouse effect [206].

Pellets from sheep's wool are efficient bio-fertilizers which can be used for organic vegetable production. Experiments performed with tomato and spinach cultivation showed wool pellets and commercial fertilizers to result in similar production levels [207].

Coarse wool was applied as fertilizer to a bean cultivar (*Phaseolus vulgaris* L.), decreasing the *carbon footprint* by 10% [208].

Mulch mats obtained from wool prevent soil erosion, gradually release moisture into soil, and promote the growth of planted vegetation, stopping the growth of weeds [209]. Wool was productively used as vegetation mats for eight perennial plants. The content levels of carbon and nitrogen, as well as the water storage capacity, recommend wool as a vegetation mat for landscapes [210].

Wool is also a source of plant biostimulants, as a substrate for the cultivation of keratinolytic fungi such as *Paecilomyces lilacinus* 112 [211].

## 9.2.3. Building Sector

One sector requiring carbon policy actions is the building sector. Due to climate change, the decarbonization of this sector is compulsory. New biodegradable materials have to be considered, given their sustainability. Wool is such a material. Its properties like breathability, moisture management, and low heat and sound conduction, together with natural flame-retardancy [9], make wool a useful material for house insulation. For

Sustainability **2024**, 16, 4661 21 of 36

the efficient application of wool as a building insulator, a study identifying the best locations for wool collection centers is of great help, reducing transport pollution and achieving the sustainable development of the region [212].

Besides its thermal properties, wool is a good acoustic insulator. Non-woven wool products may be used in multi-layer room insulation, its sound adsorption depending on surface density, thickness, and air permeability [213]. Valorization of coarse wool fibers for insulation in the context of eco-building was presented by a team of Romanian researchers. Many benefits may be obtained by good local management [214]. Good insulators were obtained by combining wool not appropriate for textile manufacture with hemp. Compared with other insulating panels, wool-hemp panels are a sustainable solution capable of decreasing the environmental impact [215,216]. The combination of wool with agricultural waste, like sugar bagasse, provided excellent insulators. Natural fibers are a good option for replacing synthetic materials in buildings [217]. Insulating properties have been improved by doping the fibers with boron. The new material is a good thermal and sound insulator for buildings [218]. Sheep's wool soy protein bio-composite is another efficient acoustic insulator [219].

Solid bricks presenting good thermal insulation and mechanical resistance have been obtained from wool fibers and clay. Wool fibers have an important effect on this new biomaterial, leading to higher energy efficiency [220]. Wool composite with natural rubber or polyurethane acrylate-based resin makes another good thermal and acoustic insulator material [221]. Complex mixtures containing polyester, date-palm fibers, and wool have been prepared. The new material is a good-performing and low-cost insulator that may be applied in building systems, home furniture, automotive parts, etc. [222].

Other insulators were obtained from wool and cement mortar. The resulting panels have good thermal and mechanical properties [223]. The incorporation of wool fibers in magnesium phosphate cement leads to the optimization of thermal comfort and energy efficiency. The material may be applied as an external thermal insulator [224].

According to the literature, with the use of bio-based insulators in France, by 2050, a saving of 75,000 tons of fossil fuels and a reduction of the greenhouse effect to the order of 312,771 tCO2eq is expected [225].

## 9.2.4. Materials for Industrial Applications

Wool may be used as a constituent of new biomaterials comprising a large field of applications. This is justified by the number of properties of the wool fibers recommending them as components in composite materials, like durability, ability to flex, fire resistance, etc. [226]. Blends with other materials may have various applications. A combination with palm fibers, as nonwoven material, was used for automotive interiors [227]. A mixture of soy protein and wool was used in the manufacture of membranes for lithium batteries [228].

Wool fibers may be included successfully in railway brake shoe composition, imparting a low cost, and lowering the environmental impact [229].

Wool blends show good results in coating anode electrodes. According to the experimental results, such an anode is suitable for photo-catalytic fuel cells, as well as for wearable electronics [230].

A combination of the expensive clay used as subgrade material in roads with wool and banana fibers provides a new, less costly composite which maintains a high level of the road's carrying capacity [231].

Bio-composites obtained from wool fibers and wheat gluten present good mechanical and fire-retardancy performance, and are useful as fire protection materials [232].

Good results have been obtained by combining wool fibers with pineapple leaf or sisal fibers in a natural rubber matrix. The new materials may be applied for packing and other household purposes [233].

Wool's ability to retain various chemicals has been exploited for centuries, the legend of "the golden fleece" being based on the real fact of the utilization of wool in screening

Sustainability **2024**, 16, 4661 22 of 36

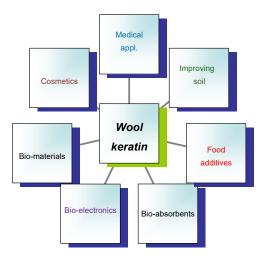
gold particles from flowing rivers. This property is now used for making wool filters for purifying waters used in the food or pharmaceutical industry, wool filters being able to absorb even the traces of heavy metals present in waters [234,235]. Raw wool is also a good metal adsorbent for metal ions like Zn and Cu, with a sorption efficiency of over 90% [236]. Wool filters reduce the virus content in water better than polymers like polypropylene and polyester or river sand [237]. Wool composite made with magnetite and polysiloxane is a good adsorbent for oil spill removal. By centrifugation, the adsorbed oil is recovered [238]. Wool has also been proved to absorb pollutants from the surrounding atmosphere. In this respect, of particular interest is wool's capacity to absorb formaldehyde [239], tobacco smoke, and various other unpleasant odors [240].

The protein chains of wool fiber sequester cations. This property was mentioned previously as being used for filtering water, but it is also the basis for dyeing wool with chrome dyes. Recently it has been shown that the sequestering of metals by amino acids of protein chains make an enzyme-like structure and may be further used for enabling wool fibers to act as catalysts, or catalyst supports in various complex reactions [241]. This field is newly under development and, with the increased demand for new biocatalysts, one may expect an increased demand for wool as well. Wool, combined with poly(2-amino thiophenol), was used as carrier for palladium nanoparticles. The resulting product may be used as an industrial catalyst [242]. A Fe-Co nanocatalyst was prepared by pyrolysis of the mixture of wool, dicyandiamide, and the corresponding metal salts [243].

# 9.2.5. Keratin Extraction and Applications

Wool is a source of protein, its main component being  $\alpha$ -keratin. Like all the keratins (e.g., hairs, nails, and feathers) wool fiber is practically insoluble in water under normal conditions, due to its highly cross-linked structure based on disulfide bonds. Temperature and certain chemicals may, however, break these bonds and render wool soluble, producing solutions of polypeptides [244,245].

Several procedures for the extraction of keratin from wool using environmentally friendly processes have been described [246,247]. Preparation methods for obtaining keratin have also been presented by Wang and Tong [99]. The comparative evaluation of five methods for keratin extraction described ionic liquid extraction as having the highest yield (95%) [248]. The necessity to continue the research for new eco-friendly and cost-effective methods for obtaining keratin films from wool and other sources has been underlined in these papers.  $\alpha$ -Keratin extract has various fields of application (see Figure 10) [249,250].



**Figure 10.** Applications of wool keratin extracts.

Sustainability **2024**, 16, 4661 23 of 36

## Medical applications

Because keratin contains several cell-binding motifs of amino acids, it may help cell proliferation, which recommends such films for medical applications, particularly for wound dressing [251–253]. The composites of wool, collagen, and ionic liquids have produced films for wound dressing, drug delivery, and sensors [254]. A very good composite for wound dressing was produced by combining wool with a zirconium-based metal-organic frameworks. The material obtained has air permeability, wash-robustness and moderate hydrophilicity [255].

Keratin based materials may also be applied in tissue engineering. The properties of keratin, like degradation rate, mechanical properties, porosity, swelling, and wettability, support this biomedical application [256].

The material derived by the electrospinning of polycaprolactone with keratin appears to be good for bone engineering [257]. Keratin mixed with bioactive glass and polycaprolactone provides a material suitable for bone repair [258].

Keratin nanofibers obtained by electrospinning are good components in polymer blends for bio-medical applications [259].

Keratin has been developed into hydrogels used as bone scaffolds, wound dressing, and wearable medical devices [260].

#### Biomaterials

Bioplastics have become of great interest in the context of green sustainable solutions. Keratin is a suitable partner for obtaining such materials, and keratin extracted from wool is a source for such new biomaterials [250]. The advantage is in the biodegradability of these materials, keratin being easily biodegradable. Keratin films, under composting conditions, degrade in 5 days [261]. The combination of keratin with various compounds leads to new improved materials. By grafting keratin with lipoic acid (see Figure 11a) different films may be produced. Lipoic acid binds to keratin by amide bonds, conferring a new geometry on the material obtained. This grafted keratin is more thermally stable and has improved physical properties [262]. Combination of keratin with polybutylene succinate (PBS) (see Figure 11b) resulted in nanofibrous materials successfully used for drug delivery and scaffolds for cell development [263].

Figure 11. Compounds for keratin grafting: (a) lipoic acid and (b) polybutylene succinate.

#### Bioelectronics

Wool keratin is employed in bioelectronics in such applications as quantum dots for the fluorescent detection of toxic metal traces (ions of chromium and iron). The multiple functional groups (CONH, COOH, NH<sub>2</sub>) of the keratin may interact with the metal by coordination or ionic bonds [264]. Keratin extracted from wool (WK) may bind on carbon nanotubes (CNT), giving dispersed inks used in high performance bioelectronics (flexible circuits or health monitoring electrodes) (see Figure 12) [265].

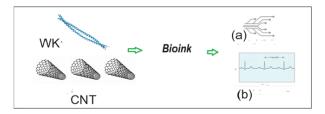


Figure 12. Quantum dots based on wool keratin (WK) with (a) electronic circuits; (b) electrocardiogram [265].

Sustainability **2024**, 16, 4661 24 of 36

New fibers are obtained by combining wool keratin with a polyanion (poly 4-styrenesulfonate), followed by centrifugation and dry-spinning of the mixture. Due to the outstanding mechanical properties, ion conductivity, and humidity awareness these fibers may be successfully used in strain sensors [266].

#### Food additives

Wool keratin may also be used as a substituent for casein in food manufacture [267]. Wool keratin food supplements may improve performance in the growth parameters of animals. Such results make keratin a sustainable protein source [268].

#### Biosorbent

The membrane obtained by combining keratin and polyamide 6 may be doped with silver, Ag, resulting in a good absorbent which can be used in air purification [269].

## Soil improvement

Good results in soil fertilization are obtained by mixing biochar obtained by the pyrolysis of lignocellulosic residues (carbon source) and hydrolyzed wool (nitrogen source). This combination gives a C/N ratio corresponding to those of efficient fertilizers [270].

#### Cosmetics

Keratin is suitable for skin application due to hydrophilicity and the capacity to form films. A film of keratin applied on the skin offers a silky sensation [271]. Keratin particles are proficient as components of cosmetics for hair care [272]. Thus, keratin is successfully used in lotions, shampoos, and conditioners, and for skincare products [273,274].

Concerning the sustainable aspects of wool valorization and the prospects of wool for replacing plastic materials, we have taken into consideration the advantages and disadvantages of such replacement (see Table 3) [197].

Advantages	Disadvantages
Biodegradability	Produced in low quantities
Good acoustic and thermal insulator	Processing for textile materials still polluting
Hygroscopic capacity	Classification of coarse wool as a by-product by the EU resulted in less restrictions on disposal (mainly landfill)
Odor-preventing capacity	More expensive than synthetics
Ensures comfortable sleep	Unsuitable management for collection and processing
Hypoallergenic	Difficulty in standardizing production
High content of carbon and nitrogen—good fertilizer	EU policy directed more to bioplastic than to natural resources like wool
Nontoxic	
Low flammability	
Potential pesticide (snake slug)	

**Table 3.** The advantages and disadvantages of using wool instead of plastic materials.

As listed in Table 3, wool has many qualities useful for successfully challenging the use of plastics, and, for the environment, it would be beneficial if wool at least partially replaced these materials.

The sustainability of wool valorization is even more underlined by the many links between the non-polluting processing of wool (e.g., enzymes and natural dyes) and the United Nations Sustainable Development Goals, SDG, as listed in Table 4.

Sustainability **2024**, 16, 4661 25 of 36

Table 4. Sustainable Development Goals (SDG) and the application of environmentally safe pro-
cedures for wool collection, processing, and disposal.

Procedure to Be Improved	Related SDGs
Sheep breeding and genetic selection	1, 2, 8, 15
Management of wool collection and distribution	9, 11
Application of new "green" technology for wool processing	3, 6, 7, 13, 14
Regulations and economic policy	15, 17

#### 10. Conclusions

Pollution generated by synthetic polymers has increased the interest in natural fibers like wool which, due to its properties, has seen growth in consumer preferences. This review surveys the state of the art in wool production, processing, and applications in the context of using less-polluting technologies that make use of enzymes.

There are several issues which need to be underlined when discussing the development of the environmentally friendly wool chain, namely:

- The necessity to develop sustainable sheep breeding for increasing profitability and obtaining an appropriate LCA of wool, starting with its production;
- New solutions for sustainable wool processing using enzymes and natural dyes are emerging;
- New non-traditional usages of wool in replacing non-biodegradable materials have been proposed and developed, opening new horizons for wool of any fiber, regardless of its diameter;
- It has to be stressed that, for any further development, the preservation of the environment has to play a role more important than the purely economic aspects;
- In maintaining sheep breeding and the processing of wool and its use, the social aspects have to be also pondered and included in the whole wool chain;
- An EU policy for encouraging the collection, processing and choice of suitable applications for coarse, processed, and waste wool.

**Author Contributions:** There were equal contributions made by both C.P. and M.D.S. as to conceptualization, data curation, and the writing of the original draft. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Conflicts of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

- 1. Nayak, R.; Nguyen, L.V.T.; Panwar, T.; Jajpura, L. Sustainable technologies and processes adapted by fashion brands. In *Sustainable Technologies for Fashion and Textiles*; Nayak, R., Ed.; Elsevier: London, UK, 2020; pp. 233–248. https://doi.org/10.1016/B978-0-08-102867-4.00011-6.
- 2. Dris, R.; Gasperi, J.; Mirande, C.; Mandin, C.; Guerrouache, M.; Langlois, V.; Tassin, B. A first overview of textile fibers, including microplastics, in indoor and outdoor environments. *Environ Pollut.* **2017**, 221, 453–458. https://doi.org/10.1016/j.envpol.2016.12.013.
- 3. Yuan, Z.; Nag, R.; Cummins, E. Human health concerns regarding microplastics in the aquatic environment—From marine to food systems. *Sci. Total Environ.* **2022**, *823*, 153730. https://doi.org/10.1016/j.scitotenv.2022.153730.
- 4. Thomas, J.; Patil, R.S.; Patil, M.; John, J. Addressing the Sustainability Conundrums and Challenges within the Polymer Value Chain. *Sustainability* **2023**, *15*, 15758. https://doi.org/10.3390/su152215758.
- 5. Popescu, C. European Sheep Industry. In *International Sheep and Wool Handbook*; Cottle, D.J., Ed.; Nottingham University Press: Nottingham, UK, 2010; pp. 153–164. https://doi.org/10.7313/UPO9781907284595.
- 6. Xueliang, X. Animal Fibers: Wool. In *Handbook of Fibrous Materials*, 1st ed.; Hu, J., Kumar, B., Lu, J., Eds.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2020; pp. 37–74. https://doi.org/10.1002/9783527342587.ch2.
- Collie, S.R.; Ranford, S.L.; Fowler, I.J.; Brorens, P.H. Microfibre pollution—what's the story for wool? In Proceedings of the AUTEX 2019—19th World Textile Conference on Textiles at the Crossroads, Ghent, Belgium, 11–15 June 2019. https://doi.org/openjournals.ugent.be/autex/article/id/63730/.

Sustainability **2024**, 16, 4661 26 of 36

8. Doyle, E.K.; Preston, J.W.V.; McGregor, B.A.; Hynd, P.I. The science behind the wool industry. The importance and value of wool production from sheep. *Anim. Front.* **2021**, *11*, 15–23. https://doi.org/10.1093/af/vfab005.

- 9. Kuffner, H.; Popescu, C. Wool fibres. In *Handbook of Natural Fibres*; Kozlowski, R., Ed.; Series in Textiles; Woodhead Publishing: Cambridge, UK, 2012; Volume 1, pp. 171–195. https://doi.org/10.1533/9780857095503.1.171.
- 10. Lehmann, M.; Tärneberg, S.; Tochtermann, T.; Chalmer, C.; Eder-Hansen, J.; Javier, J.; Seara, F.; Boger, S.; Hase, C.; von Berlepsch, V.; et al. *Pulse of the Fashion Industry*; Global Fashion Agenda and The Boston Consulting Group: Copenhagen, Denmark, 2018. Available online: https://www.peta.org.uk/wp-content/uploads/2019/03/Pulse\_of\_the\_fashion\_industry\_report\_2018-1.pdf (accessed on 11 April 2024).
- 11. International Wool Textile Organization. IWTO Market Information, Ed. 18. 2023. Available online: https://iwto.org/resources/statistics/ (accessed on 11 April 2024).
- 12. Li, X.; Zhu, L.; Dong, Y.; Chen, B.; Li, Q.; Wang, X.; Zhang, Y.; Wang, L. Water footprint assessment of wool products with a low-water footprint baseline. *Sustain. Prod. Consum.* **2022**, *34*, 310–317. https://doi.org/10.1016/j.spc.2022.09.023.
- 13. Furferi, R.; Volpe, Y.; Mantellassi, F. Circular Economy Guidelines for the Textile Industry. *Sustainability* **2022**, *14*, 11111. https://doi.org/10.3390/su141711111.
- 14. Wiedemann, S.G.; Nguyen, Q.V.; Clarke, S.J. Using LCA and Circularity Indicators to Measure the Sustainability of Textiles—Examples of Renewable and Non-Renewable Fibres. *Sustainability* **2022**, *14*, 16683. https://doi.org/10.3390/su142416683.
- 15. Sim, J.; Prabhu, V. The life cycle assessment of energy and carbon emissions on wool and nylon carpets in the United States. *J. Clean. Prod.* **2018**, *170*, 1231–1243. https://doi.org/10.1016/j.jclepro.2017.09.203.
- 16. Angelis-Dimakis, A.; Whitehouse, A.; Vyrkou, A.; Hebden, A.; Rana, S.; Goswami, P. Life cycle environmental impact and economic assessment of British Wool face masks. *Clean. Environ. Syst.* **2022**, *6*, 100084. https://doi.org/10.1016/j.cesys.2022.100084.
- 17. Bianco, I.; Picerno, G.; Blengini, G.A. Life Cycle Assessment (LCA) of Worsted and Woollen processing in wool production: ReviWool® noils and other wool co-products. *J. Clean. Prod.* **2023**, 415, 137877. https://doi.org/10.1016/j.jclepro.2023.137877.
- 18. McDonald Textiles. Available online: https://mcdonaldtextiles.com/blogs/news/what-is-zq-merino-wool (accessed on 11 April 2024).
- 19. European Commission. Commission Decision of 5 June 2014 Establishing the Ecological criteria for the Award of the EU Ecolabel for Textile Products. Official Journal of the European Union L 174/45. Available online: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32014D0350 (accessed on 11 April 2024).
- 20. Plakantonaki, S.; Kiskira, K.; Zacharopoulos, N.; Chronis, I.; Coelho, F.; Togiani, A.; Kalkanis, K.; Priniotakis, G. A Review of Sustainability Standards and Ecolabeling in the Textile Industry. *Sustainability* **2023**, *15*, 11589. https://doi.org/10.3390/su151511589.
- 21. Vezzoli, C.; Conti, G.M.; Macrì, L.; Motta, M. Designing Sustainable Clothing Systems. Franco Angeli: Milan, Italy, 2022; p. 120.
- 22. Scott, J.T.; Robertson, F.A. Environmental effects of sheep farming. In *Environmental Impacts of Pasture-Sheep Farming*; McDowell, R.W., Ed.; CABI Publishing: Boston, MA, USA, 2008, pp. 144–186. https://doi.org/10.1079/9781845934118.0144.
- 23. Wiedemann, S.G.; Biggs, L.; Nebel, B.; Bauch, K.; Laitala, K.; Klepp, I.G.; Swan, P.G.; Watson, K. Environmental impacts associated with the production, use, and end-of-life of a woollen garment. *Int. J. Life Cycle Assess.* **2020**, *25*, 1486–1499. https://doi.org/10.1007/s11367-020-01766-0.
- 24. Wiedemann, S.G.; Ledgard, S.; Henry, B.; Yan, M.; Mao, N.; Russell, S. Application of life cycle assessment to sheep production systems: Investigating co-production of wool and meat using case studies from major global producers. *Int. J. Life Cycle Assess.* 2015, 20, 463–476. https://doi.org/10.1007/s11367-015-0849-z.
- 25. Wiedemann, S.G.; Yan, M.; Henry, B.; Murphy, C. Resource use and greenhouse gas emissions from three wool production regions in Australia. *J. Clean. Prod.* **2016**, *122*, 121–132. https://doi.org/10.1016/j.jclepro.2016.02.025.
- 26. de Arriba, R.; Barac, M. Homo Economicus and the Shepherd: The Traditional Sheep Farmer Facing the Modernisation (or Intensification) of European Livestock. *East. Eur. Countrys.* **2018**, 24, 171–187. https://doi.org/10.2478/eec-2018-0009.
- 27. Gonzalez, E.B.; Easdale, M.H.; Sacchero, D.M. Socio-technical networks modulate on-farm technological innovations in wool production of North Patagonia, Argentina. *J. Rural Stud.* **2021**, *83*, 30–36. https://doi.org/10.1016/j.jrurstud.2021.02.015.
- 28. Shabbir, M.; Naim, M. Introduction to Textiles and the Environment In *Textiles and Clothing*; Shabbir, M., Ed.; Scrivener Publishing (Wiley) LLC: Beverly, MA, USA, 2019; pp. 1–9. https://doi.org/10.1002/9781119526599.ch1.
- 29. Zhu, L.; Chen, B.; Liu, J.; Chen, S.; Zhang, Y.; Wang, X.; Wang, L. Assessing baseline water footprints of natural fiber textile products in China. *J. Clean. Prod.* **2022**, *379*, 134747. https://doi.org/10.1016/j.jclepro.2022.134747.
- 30. Ribeiro, P.F.; Santos, J.L.; Santana, J.; Reino, L.; Beja, P.; Moreira, F. An applied farming systems approach to infer conservation-relevant agricultural practices for agri-environment policy design. *Land Use Policy* **2016**, *58*, 165–172. https://doi.org/10.1016/j.landusepol.2016.07.018.
- 31. Senapati, S.; Paikaray, A.; Das, B.C.; Swain, P. Application of Remote Sensing in Agriculture & Animal Husbandry, ICT Programmes in Livestock Development Problems & Prospect of ICT in Livestock Development. *Int. J. Agric. Environ. Biotechnol.* **2016**, *1*, 920–925. https://doi.org/10.22161/ijeab/1.4.42.
- 32. Constantin, P.; Nechifor, I.; Florea, A.; Cristian, C. Heritability determination for reproduction characters in the new milk population formed in the North-East part of Romania. *Sci. Pap.-Anim. Sci. Ser. Lucr. Ştiinţifice-Ser. Zooteh.* **2019**, 72, 3–6.

Sustainability **2024**, 16, 4661 27 of 36

33. Cecchini, L.; Vieceli, L.; D'Urso, A.; Magistrali, C.F.; Forte, C.; Mignacca, S.A.; Trabalza-Marinucci, M.; Chiorri, M. Farm efficiency related to animal welfare performance and management of sheep farms in marginal areas of Central Italy: A two-stage DEA model. *Ital. J. Anim. Sci.* 2021, 20, 955–969. https://doi.org/10.1080/1828051X.2021.1913076.

- Hara, S.M.; Faverín, C.; Villagra, E.S.; Easdale, M.H.; Tittonell, P. Exploring drivers and levels of technology adoption for ecological intensification of pastoral systems in north Patagonia drylands. *Agric. Ecosyst. Environ.* 2022, 324, 107704. https://doi.org/10.1016/j.agee.2021.107704.
- Ogawa, K.; Garrod, G.; Yagi, H. Sustainability strategies and stakeholder management for upland farming, Land Use Policy 2023, 131, 106707. https://doi.org/10.1016/j.landusepol.2023.106707.
- 36. Mohammed, T. A conceptual review on breeding and conservation approaches for Tikur sheep population. *Cogent Food Agric.* **2023**, *9*, 2236824. https://doi.org/10.1080/23311932.2023.2236824.
- 37. Alcock, D.J.; Harrison, M.T.; Rawnsley, R.P.; Eckard, R.J. Can animal genetics and flock management be used to reduce greenhouse gas emissions but also maintain productivity of wool-producing enterprises? *Agric. Syst.* **2015**, *132*, 25–34. https://doi.org/10.1016/j.agsy.2014.06.007.
- 38. Norton, M.R.; Garden, D.L.; Orchard, B.A.; Armstrong, P.; Brassil, T. Effects of lime, phosphorus and stocking rate on an extensively managed permanent pasture: Botanical composition and groundcover. *Crop Pasture Sci.* **2020**, *71*, 700–713. https://doi.org/10.1071/CP20135.
- 39. Farinella, D.; Nori, M.; Ragkos, A. Change in Euro-Mediterranean pastoralism: Which opportunities for rural development and generational renewal? In *Grassland Resources for Extensive Farming Systems in Marginal Lands: Major Drivers and Future Scenarios, Proceedings of the 19 Symposium of European Grassland Federation, Alghero, Italy, 7–10 May 2017*; Porqueddu, C., Franca, A., Lombardi, G., Molle, G., Peratoner, G., Hopkins, A., Eds.; Wageningen Academic Publishers: Wageningen, The Netherland, 2017; Volume 22, pp. 23–36.
- 40. Farias, G.D.; Bremm, C.; Savian, J.V.; de Souza Filho, W..; de Lima, L.C.; de Albuquerque Nunes, P.A.; Alves, L.A.; Sacido, M.; Montossi, F.; Tiecher, T.; et al. Opportunities and challenges for the integration of sheep and crops in the Rio de la Plata region of South America. *Small Rumin. Res.* **2022**, 215, 106776. https://doi.org/10.1016/j.smallrumres.2022.106776.
- 41. Hamadani, A.; Ganai, N.A. Development of a multi-use decision support system for scientific management and breeding of sheep. *Sci. Rep.* **2022**, *12*, 19360. https://doi.org/10.1038/s41598-022-24091-y.
- 42. Crewther, W.G. The effects of disaggregating agents on the stress–strain relationship for wool fibers, *Text. Res. J.* **1972**, *42*, 77–85. https://doi.org/10.1177/004051757204200202.
- 43. Feughelman, M. A model for the mechanical properties of the  $\alpha$ -keratin cortex. *Text. Res. J.* **1994**, *64*, 236–239. https://doi.org/10.1177/004051759406400408.
- 44. Hearle, J.W.S. A critical review of the structural mechanics of wool and hair fibres. *Int. J. Biol. Macromol.* **2000**, *27*, 123–138. https://doi.org/10.1016/S0141-8130(00)00116-1.
- Popescu, C.; Wortmann, F.J. Wool—Structure, Mechanical Properties and Technical Products based on Animal Fibres. In Industrial Applications of Natural Fibres; Mussig, J., Ed.; John Wiley & Sons Ltd.: Chichester, UK, 2010; pp. 255–268. https://doi.org/10.1002/9780470660324.ch12.
- 46. Astbury, W.T.; Street, A. X-ray studies of the structures of hair, wool and related fibres. I. General. *Phil. Trans. R. Soc. Lond.* **1931**, *A230*, 75–101. https://doi.org/10.1098/rsta.1932.0003.
- 47. Astbury, W.T.; Woods, H.J. X-ray studies of the structures of hair, wool and related fibres. II. The molecular structure and elastic properties of hair keratin. *Phil. Trans. R. Soc. Lond.* **1933**, *A232*, 333–394. https://doi.org/10.1098/rsta.1934.0010.
- 48. Pauling, L.; Corey, R.B.; Branson, H.R. The structure of proteins; two hydrogen-bonded helical configurations of the polypeptide chain. *Proc. Nat. Acad. Sci. USA* **1951**, *37*, 205–211. https://doi.org/10.1073/pnas.37.4.205.
- 49. Popescu, C. The Thermodynamics of Trichocyte Keratins. In *The Hair Fibre: Proteins, Structure and Development. Advances in Experimental Medicine and Biology;* Plowman, J., Harland, D., Deb-Choudhury, S., Eds.; Springer: Singapore, 2018; Volume 1054, pp. 185–203. https://doi.org/10.1007/978-981-10-8195-8\_13.
- 50. Allafi, F.A.; Hossain, M.; Shaah, M.; Lalung, J.; Ab Kadir, M.O.; Ahmad, M.I. A Review on Characterization of Sheep Wool for Wool Impurities and Existing Techniques of Cleaning: Industrial and Environmental Challenges. *J. Nat. Fibers* **2022**, *19*, 8669–8687. https://doi.org/10.1080/15440478.2021.1966569.
- 51. Möller, M.; Popescu, C. Natural Fibers. In *Sustainable Solutions for Modern Economies*; Höfer, R., Ed.; RSC Publishing: Cambridge, UK, 2009; RSC Green Chemistry No. 4., pp. 368–393. https://doi.org/10.1039/9781847552686-00368.
- 52. Möller, M.; Popescu, C. Natural Fibers. In *Polymer Science: A Comprehensive Reference*; Matyjaszewski, K., Möller, M., Eds.; Elsevier: Amsterdam, The Netherland, 2012; Volume 10, pp. 267–280. https://doi.org/10.1016/B978-0-444-53349-4.00266-1.
- 53. Zhang, L. Yellowstone Impression. In *International Textile and Apparel Association Annual Conference Proceedings*; Iowa State University Digital Press: Ames, IA, USA, 2020.
- 54. Bülbül, S.; Urper, N. Development of felting and its usage areas in tourism industry. *IRSJ Vis.* **2023**, *8*, 135–153. https://doi.org/10.55843/ivisum2382135b.
- 55. von Allwörden, K. The nature of sheep's wool and a new research method for the chemical identification of damaged wool. *Z Angew. Chem.* **1916**, *29*, 77–78. https://doi.org/10.1002/ange.19160291704.
- 56. Greeff, J.C.; Schlink, A.C. The inheritance of felting of Merino wool. Int. J. Sheep Wool Sci. 2002, 50, 6-10.
- 57. Schlink, A.C.; Greeff, J.C.; Haigh, M. You Can Breed for Easy-care Woollen Garments. Wool Technol. Sheep Breed. 2002, 50, 443–448.

Sustainability **2024**, 16, 4661 28 of 36

58. Stanescu, M.D. Goals for a sustainable development and the environmental protection, *Rom. J. Ecol. Environm.* **2022**, *4*, 56–61. https://doi.org/10.21698/rjeec.2022.205.

- 59. Gulzar, T.; Farooq, T.; Kiran, S.; Ahmad, I.; Hameed, A. Green chemistry in the wet processing of textiles. In *The Textile Institute Book Series: The Impact and Prospects of Green Chemistry for Textile Technology;* Shahid-ul-Islam; Butola, B.S., Eds; Woodhead Publishing: Cambridge, UK, 2019; pp. 1–20. https://doi.org/10.1016/B978-0-08-102491-1.00001-0.
- 60. Ji, X.; Qian, W.; Tian, Z.; Li, Y.; Wang, L. Quantification and evaluation of chemical footprint of woollen textiles. *Ind. Text.* **2021**, 72, 68–73. https://doi.org/10.35530/IT.072.01.1773.
- 61. Sen, A.; Kapila, R.; Chaudhary, S.; Nigam, A. Biotechnological Applications of Microbial Enzymes to Replace Chemicals in the Textile Industry—A Review. *J. Text. Assoc.* **2021**, *82*, 68–72.
- 62. Rahman, M.; Billah, M.M.; Hack-Polay, D.; Alam, A. The use of biotechnologies in textile processing and environmental sustainability: An emerging market context. *Technol. Forecast. Soc. Chang.* **2020**, *159*, 120204. https://doi.org/10.1016/j.techfore.2020.120204.
- 63. Islam, M.T.; Asaduzzaman, S. Environmentally-Friendly Textile Finishing. In *Textiles and Clothing: Environmental Concerns and Solutions*; Shabbir, M., Ed.; Scrivener Publishing (Wiley) LLC: Beverly, MA, USA, 2019; pp. 101–129. https://doi.org/10.1002/9781119526599.ch6.
- 64. Chou, K.-C.; Cai, Y.-D. Predicting protein quaternary structure by pseudo amino acid composition. *Proteins* **2003**, *53*, 282–289. https://doi.org/10.1002/prot.10500.
- 65. Rawlings, N.D. Protease Families, Evolution and Mechanism of Action. In *Proteases: Structure and Function*; Brix, K.; Stocker, W. Eds.; Springer, Vienna, Austria, 2013; pp. 1–36. https://doi.org/10.1007/978-3-7091-0885-7\_1.
- 66. Gopinath, S.C.; Anbu, P.; Lakshmipriya, T.; Tang, T.H.; Chen, Y.; Hashim, U.; Ruslinda, A.R.; Arshad, M.K. Biotechnological Aspects and Perspective of Microbial Keratinase Production. *Biomed. Res. Int.* **2015**, 2015, 140726. https://doi.org/10.1155/2015/140726.
- 67. Sharma, R.; Devi, S. Versatility and commercial status of microbial keratinases: A review. *Rev. Environ. Sci. Biotechnol.* **2018**, 17, 19–45. https://doi.org/10.1007/s11157-017-9454-x.
- 68. Qiu, J.; Wilkens, C.; Barrett, K.; Meyer, A.S. Microbial enzymes catalyzing keratin degradation: Classification, structure, function. *Biotechnol. Adv.* **2020**, 44, 107607. https://doi.org/10.1016/j.biotechadv.2020.107607.
- 69. Martínez, A.; Maicas, S. Cutinases: Characteristics and Insights in Industrial Production. *Catalysts* **2021**, *11*, 1194. https://doi.org/10.3390/catal11101194.
- 70. Liang, X.; Zou, H. Biotechnological Application of Cutinase: A Powerful Tool in Synthetic Biology. *Syn. Bio.* **2023**, *1*, 54–64. https://doi.org/10.3390/synbio1010004.
- 71. Stanescu, M.D. Applications of enzymes in processing cellulosic textiles—A review of the latest developments. *Cellulose Chem. Technol.* **2023**, *57*, 1–15.
- 72. Jayasekara, S.; Ratnayake, R. Microbial Cellulases: An Overview and Applications. In *Cellulose*; Pascual, A.R., Martín, M.E.E., Eds.; IntechOpen: Rijeka, Croatia, 2019; pp. 1–21. https://doi.org/10.5772/intechopen.84531.
- 73. Chandra, P.; Enespa; Singh, R.; Arora, P.K. Microbial lipases and their industrial applications: A comprehensive review. *Microb Cell Fact.* **2020**, *19*, 169. https://doi.org/10.1186/s12934-020-01428-8.
- 74. Tochetto, G.A.; Aragão, A.M.I.; de Oliveira, D.; Immich, A.P.S. Can enzymatic processes transform textile processes? A critical analysis of the industrial application, *Process Biochem.* **2022**, *123*, 27–35. https://doi.org/10.1016/j.procbio.2022.10.030.
- 75. Noreen, S.; Perveen, S.; Bilal, M.; Iqbal, H.M.N. Laccases: Catalytic and functional attributes for robust biocatalysis. In *Nanomaterials for Biocatalysis*; Castro, G.R., Nadda, A.K., Nguyen, T.A., Qi, X., Yasin, G., Eds.; Elsevier: Amsterdam, The Netherland, 2022; pp. 567–594. https://doi.org/10.1016/B978-0-12-824436-4.00018-6.
- 76. Patel, N.; Shahane, S.; Shivam; Majumdar, R.; Mishra, U. Mode of Action, Properties, Production, and Application of Laccase: A Review. *Recent Pat. Biotechnol.* **2019**, *13*, 19–32. https://doi.org/10.2174/1872208312666180821161015.
- 77. Mojsov, K. Enzymes in textile industry: A review. Int. J. Manag. IT Eng. 2014, 4, 34–44.
- 78. Kelly, S.A.; Mix, S.; Moody, T.S.; Gilmore, B.F. Transaminases for industrial biocatalysis: Novel enzyme discovery. *Appl. Microbiol. Biotechnol.* **2020**, 104, 4781–4794. https://doi.org/10.1007/s00253-020-10585-0.
- 79. Yu, J.; Su, J.; Wang, P.; Yu, Y.; Wang, Q.; Cavaco-Paulo, A. Enzymatic processing of protein-based fibers. *Appl. Microbiol. Biotechnol.* **2015**, 99, 10387–10397. https://doi.org/10.1007/s00253-015-6970-x.
- 80. Duarte, L.; Matte, C.R.; Bizarro, C.V.; Ayub, M.A.Z. Review transglutaminases: Part II-industrial applications in food, biotechnology, textiles and leather products. *World J. Microbiol. Biotechnol.* **2019**, *36*, 11. https://doi.org/10.1007/s11274-019-2792-9.
- 81. Mojsov, K. Enzymatic treatment of wool fabrics: Opportunity of the improvement on some physical and chemical properties of the fabrics. *J. Text. Inst.* **2017**, *108*, 1136–1143. https://doi.org/10.1080/00405 000.2016.12228 56.
- 82. Maghraby, Y.R.; El-Shabasy, R.M.; Ibrahim, A.H.; Azzazy, H.M.E. Enzyme Immobilization Technologies and Industrial Applications. *ACS Omega.* **2023**, *8*, 5184–5196. https://doi.org/10.1021/acsomega.2c07560.
- 83. Stanescu, M.D. Enzymes in Textile Finishing. Overview. In *Textile Engineering at the Dawn of a New Millennium: An Exciting Challenge, Proceedings of 2nd AUTEX World Textile Conference, Bruges, Belgium, 1–3 July 2002*; ResearchGate: Berlin, Germany, 2002.
- 84. Troncoso, F.D.; Sanchez, D.A.; Luján Ferreira, M. Production of Plant Proteases and New Biotechnological Applications: An Updated Review. *ChemistryOpen* **2022**, *11*, e202200017. https://doi.org/10.1002/open.202200017.

Sustainability **2024**, 16, 4661 29 of 36

85. Srivastava, B.; Singh, H.; Khatri, M.; Singh, G.; Arya, S.K. Immobilization of keratinase on chitosan grafted-β-cyclodextrin for the improvement of the enzyme properties and application of free keratinase in the textile industry. *Int. J. Biol. Macromol.* **2020**, *165A*, 1099–1110. https://doi.org/10.1016/j.ijbiomac.2020.10.009.

- 86. Gajsek, M.; Jancic, U.; Vasic, K.; Knez, Z.; Leitgeb, M. Enhanced activity of immobilized transglutaminase for cleaner production technologies. *J. Clean. Prod.* **2019**, 240, 118218. https://doi.org/10.1016/j.jclepro.2019.118218.
- 87. Ghosh, D.; Talukdar, P. Relevance of Microbial Enzymes in Textile Industries Emphasizing Metabolic Engineering Panorama. In *Microbial Fermentation and Enzyme Technology*; Thatoi, H., Das Mohapatra, P.K., Mohapatra, S., Mondal, K.C., Eds.; CRC Press: Boca Raton, FL, USA, 2020; pp. 195–226. https://doi.org/10.1201/9780429061257.
- 88. Sharma, H.; Upadhyay, S.K. Enzymes and their production strategies. In *Biomass, Biofuels, Biochemicals*; Singh, S.P., Pandey, A., Singhania, R.R., Larroche, C., Li, Z., Eds.; Elsevier: Amsterdam The Netherland, 2020; pp. 31–48. https://doi.org/10.1016/B978-0-12-819820-9.00003-X.
- 89. Vujasinović, E.; Tarbuk, A.; Pušić, T.; Dekanić, T. Bio-Innovative Pretreatment of Coarse Wool Fibers. *Processes* **2023**, *11*, 103. https://doi.org/10.3390/pr11010103.
- 90. El-Fiky, A.F.; Khalil, E.M.; Mowafi, S.; Zaki, R.A.; El-Sayed, H. A Novel Approach towards Removal of Lipid Barrier from Wool Fibers' Surface Using Thermophilic Lipase. *J. Nat. Fibers* **2022**, *19*, 9471–9485. https://doi.org/10.1080/15440478.2021.1982835.
- 91. Taleb, M.A.; Gomaa, S.K.; Wahba, M.I.; Zaki, R.A.; El-Fiky, A.F.; El-Refai, H.A.; El-Sayed, H. Bioscouring of wool fibres using immobilized thermophilic lipase. *Int. J. Biol. Macromol.* **2022**, *194*, 800–810. https://doi.org/10.1016/j.ijbiomac.2021.11.128.
- 92. Gomaa, S.K.; Zaki, R.A.; Wahba, M.I.; Taleb, M.A.; El-Rafai, H.A.; El-Fiky, A.F.; El-Sayed, H. Green method for improving performance attributes of wool fibres using immobilized proteolytic thermozyme. 3 *Biotech* **2022**, 12, 254. https://doi.org/10.1007/s13205-022-03323-y.
- 93. Shen, J. Enzymatic treatment of wool and silk fibers. In *Advances in Textile Biotechnology*, 2nd ed.; Cavaco-Paulo, A., Nierstrasz, V.A., Wang, Q., Eds.; Elsevier: Amsterdam, The Netherland, 2019; pp. 77–105. https://doi.org/10.1016/B978-0-08-102632-8.00005-0.
- 94. Eisazadeh, B.; Mirzajani, F.; Sefidbakht, Y. How is the Effect of Silver Nanoparticles and Lipase/Cellulase Enzymes on Each Other? *Iran J. Sci. Technol. Trans. Sci.* **2020**, 44, 27–35. https://doi.org/10.1007/s40995-020-00820-8.
- 95. Czaplicki, Z.; Matyjas-Zgondek, E.; Strzelecki, S. Scouring of Sheep Wool Using an Acoustic Ultrasound Wave. *Fibres Text. East. Eur.* **2021**, 29, 44–48. https://doi.org/10.5604/01.3001.0015.2721.
- 96. Pan, Y.; Hurren, C.J.; Li, Q. Effect of sonochemical scouring on the surface morphologies, mechanical properties, and dyeing abilities of wool fibres. *Ultrason. Sonochem.* **2018**, *41*, 227–233. https://doi.org/10.1016/j.ultsonch.2017.09.045.
- 97. Li, Q.; Hurren, C.J.; Wang, X. Ultrasonic assisted industrial wool scouring. *Procedia Eng.* 2017, 200, 39–44. https://doi.org/10.1016/j.proeng.2017.07.007.
- 98. Czaplicki, Z.; Matyjas-Zgondek, E.; Serweta, W. Directions of the Finishing of Fibres and Textile Development. *Fibres Text. East. Eur.* **2018**, *26*, 133–141. https://doi.org/10.5604/01.3001.0012.5167.
- 99. Wang, R.; Tong, H. Preparation Methods and Functional Characteristics of Regenerated Keratin-Based Biofilms. *Polymers* **2022**, 14, 4723. https://doi.org/10.3390/polym14214723.
- 100. Allafi, F.A.; Hossain, M.S.; Shaah, M.; Lalung, J.; Ab Kadir, M.O.; Ahmad, M.I. Optimizing Supercritical Carbon Dioxide in the Bacterial Inactivation and Cleaning of Sheep Wool Fiber by Using Response Surface Methodology. *J. Nat. Fibers* **2022**, *19*, 8399–8414. https://doi.org/10.1080/15440478.2021.1964131.
- 101. Soun, B.; Kaur, D.; Jose, S. Effect of Transglutaminase Enzyme on Physico-mechanical Properties of Rambouillet Wool Fiber. *J. Nat. Fibers* **2020**, *17*, 793–801. https://doi.org/10.1080/15440478.2018.1534187.
- 102. Heine, E.; Ruers, A.; Hocker, H. Enzymatic degradation of vegetable residues in wool. DWI Rep. 2000, 123, 475-479.
- 103. Bucur, S.; Stanescu, M.D. Enzymatic treatment of cotton: A model for wool carbonization. In Proceedings of the 10th International Wool Textile Research Conference, Aachen, Germany, 26 November–1 December 2000.
- 104. Sedelnik, N. Biotechnology to improve the quality of wool. Res. J. Text. Appar. 2003, 7, 1–10. https://doi.org/10.1108/RJTA-07-02-2003-B001.
- 105. El-Sayed, H.; Kantouch, A.; El-Gabry, L. Effect of bio-carbonization of coarse wool on its dyeability. *Indian J. Fibre Text. Res.* **2010**, 35, 330–336.
- 106. Senthilkumar, P.; Vigneswaran, C.; Kandhavadivu, P. A novel approach in single stage combined bleaching and protease enzyme treatments on wool fabrics. *Fibers Polym.* **2015**, *16*, 397–403. https://doi.org/10.1007/s12221-015-0397-v.
- 107. Khatami, S.H.; Vakili, O.; Ahmadi, N.; Soltani Fard, E.; Mousavi, P.; Khalvati, B.; Maleksabet, A.; Savardashtaki, A.; Taheri-Anganeh, M.; Movahedpour, A. Glucose oxidase: Applications, sources, and recombinant production. *Biotechnol. Appl. Biochem.* 2022, 69, 939–950. https://doi.org/10.1002/bab.2165.
- 108. Avcı, B.B.; Erkan, G. The effect of glucose oxidase enzyme on wool fibres. *Color. Technol.* **2022**, 139, 1–18. https://doi.org/10.1111/cote.12658.
- 109. Hassan, M.M.; Carr, C.M. A review of the sustainable methods in imparting shrink resistance to wool fabrics. *J. Adv. Res.* **2019**, *18*, 39–60. https://doi.org/10.1016/j.jare.2019.01.014.
- 110. Liu, W.; Yuan, F.; Wang, J.; Qin, C.; Pang, Z.; Teng, Y.; Li, F.; Liu, T. Simultaneous lice eggshell removal from wool and anti-felting with a single protease treatment. *J. Clean. Prod.* **2023**, *386*, 135828. https://doi.org/10.1016/j.jclepro.2022.135828.

Sustainability **2024**, 16, 4661 30 of 36

111. Iglesias, M.S.; Sequeiros, C.; García, S.; Olivera, N.L. Newly isolated *Bacillus* sp. G51 from Patagonian wool produces an enzyme combination suitable for felt-resist treatments of organic wool. *Bioprocess Biosyst. Eng.* **2017**, 40, 833–842. https://doi.org/10.1007/s00449-017-1748-4.

- 112. Iglesias, M.S.; Sequeiros, C.; García, S.; Olivera, N.L. Eco-friendly anti-felting treatment of wool top based on biosurfactant and enzymes, J. Clean. Prod. 2019, 220, 846–852. https://doi.org/10.1016/j.jclepro.2019.02.165.
- 113. Li, W.; Zhang, N.; Wang, Q.; Wang, P.; Yu, Y.; Zhou, M. A Sustainable and Effective Bioprocessing Approach for Improving Anti-felting, Anti-pilling and Dyeing Properties of Wool Fabric. *Fibers Polym.* **2021**, 22, 3045–3054. https://doi.org/10.1007/s12221-021-1148-x.
- 114. Du, Z.; Ji, B.; Yan, K. Recycling keratin polypeptides for anti-felting treatment of wool based on L-cysteine pretreatment. *J. Clean. Prod.* **2018**, *183*, 810–817. https://doi.org/10.1016/j.jclepro.2018.02.196.
- 115. Chemchame, Y.; Benzbir, H.; Kachachi, A.; El Bouchti, M.; Kharchafi, A. Improvement of enzymatic antifelting treatment of wool fiber and its effect on dyeing with juglone from a common walnut plant. *IOP Conf. Ser. Mater. Sci. Eng.* **2020**, 827, 012042. https://doi.org/10.1088/1757-899X/827/1/012042.
- 116. Kadam, V.; Rani, S.; Jose, S.; Shakyawar, D.B.; Shanmugam, N. Biomaterial based shrink resist treatment of wool fabric: A sustainable technology. *Sustain. Mater. Technol.* **2021**, *29*, e00298. https://doi.org/10.1016/j.susmat.2021.e00298.
- 117. Wang, L.; Yao, J.; Niu, J.; Liu, J.; Li, B.; Feng, M. Eco-Friendly and Highly Efficient Enzyme-Based Wool Shrinkproofing Finishing by Multiple Padding Techniques. *Polymers* **2018**, *10*, 1213. https://doi.org/10.3390/polym10111213.
- 118. Mei, J.; Zhang, N.; Yu, Y.; Wang, Q.; Yuan, J.; Wang, P.; Cui, L.; Fan, X. A novel "trifunctional protease" with reducibility, hydrolysis, and localization used for wool anti-felting treatment. *Appl. Microbiol. Biotechnol.* **2018**, 102, 9159–9170. https://doi.org/10.1007/s00253-018-9276-y.
- 119. Madhu, A.; Chakraborty, J.N. Developments in application of enzymes for textile processing. *J. Clean. Prod.* **2017**, *145*, 114–133. https://doi.org/10.1016/j.jclepro.2017.01.013.
- 120. Shahid, M.; Mohammad, F.; Chen, G.; Tang, R.C.; Xing, T. Enzymatic processing of natural fibres: White biotechnology for sustainable development. *Green Chem.* 2016, 18, 2256–2281. https://doi.org/10.1039/C6GC00201C.
- 121. Wang, Y.; Zhang, N.; Wang, Q.; Yu, Y.; Wang, P. Chitosan grafting via one-enzyme double catalysis: An effective approach for improving performance of wool, *Carbohydr. Polym.* **2021**, 252, 117157. https://doi.org/10.1016/j.carbpol.2020.117157.
- 122. Ahsanul, A.N.M.; Maryam, N. Zero-water discharge and rapid natural dyeing of wool by plasma-assisted spray-dyeing. *J. Clean. Prod.* **2023**, 402, 136807. https://doi.org/10.1016/j.jclepro.2023.136807.
- 123. Lara, L.; Cabral, I.; Cunha, J. Ecological Approaches to Textile Dyeing: A Review. *Sustainability* **2022**, *14*, 8353. https://doi.org/10.3390/su14148353.
- 124. Shabbir, M.; Rather, L.J.; Mohammad, F. Economically viable UV-protective and antioxidant finishing of wool fabric dyed with Tagetes erecta flower extract: Valorization of marigold. *Ind. Crops Prod.* **2018**, *119*, 277–282. https://doi.org/10.1016/j.indcrop.2018.04.016.
- 125. Dulo, B.; De Somer, T.; Phan, K.; Roosen, M.; Githaiga, J.; Raes, K.; De Meester, S. Evaluating the potential of natural dyes from nutshell wastes: Sustainable colouration and functional finishing of wool fabric. *Sustain. Mater. Technol.* **2022**, *34*, e00518. https://doi.org/10.1016/j.susmat.2022.e00518.
- 126. Zhang, W.; Wang, X.; Weng, J.; Liu, X.; Qin, S.; Li, X.; Gong, J. Eco-dyeing and functional finishing of wool fabric based on Portulaca oleracea L. as colorant and Musa basjoo as natural mordant. *Arab. J. Chem.* **2022**, *15*, 103624. https://doi.org/10.1016/j.arabjc.2021.103624.
- 127. Iqbal, K.; Afzal, H.; Siddiqui, M.O.R.; Bashir, U.; Jan, K.; Abbas, A.; Abid, H.A. Dyeing of wool fabric with natural dye extracted from Dalbergia Sissoo using natural mordants. *Sustain. Chem. Pharm.* 2023, 33, 101094. https://doi.org/10.1016/j.scp.2023.101094.
- 128. Khan, A.; Hussain, M.T.; Jiang, H.; Gul, S. Development of functional wool fabric by treatment with aqueous and alkaline extracts of *Cinnamomumcamphora* plant leaves. *J. Nat. Fibers* **2020**, *17*, 472–481. https://doi.org/10.1080/15440478.2018.1500339.
- 129. Gong, K.; Rather, L.J.; Zhou, Q.; Wang, W.; Li, Q. Natural dyeing of merino wool fibers with *Cinnamomumcamphora* leaves extract with mordants of biological origin: A greener approach of textile coloration. *J. Text. Inst.* **2020**, 111, 1038–1046. https://doi.org/10.1080/00405000.2019.1698228.
- 130. Haque, M.A.; Mia, R.; Mahmud, S.T.; Bakar, M.A.; Ahmed, T.; Farsee, M.S.; Hossain, M.I. Sustainable dyeing and functionalization of wool fabrics with black rice extract. *Resour. Environ. Sustain.* **2022**, *7*, 100045. https://doi.org/10.1016/j.resenv.2021.100045.
- 131. Thakker, A.M.; Sun, D. Ecological application of natural biomaterial on natural fibres. *Clean. Mater.* **2022**, *3*, 100038. https://doi.org/10.1016/j.clema.2021.100038.
- 132. Adeel, S.; Habib, N.; Batool, F.; Amin, N.; Ahmad, T.; Arif, S.; Hussaan, M. Environmentally friendly exploration of cinnamon bark (*Cinnamomum verum*) based yellow natural dye for green coloration of bio-mordanted wool fabric. *Environ. Prog. Sustain. Energy.* **2021**, *41*, e13794. https://doi.org/10.1002/ep.13794.
- 133. Safapour, S.; Rather, L.J. Assessment of Colorimetric and Fastness Properties of Prangos Ferulacea (*Jashir*) Dyed Wool Yarns in Conjunction with Mixed Metal Mordant Combinations via Reflectance Spectroscopy. *J. Nat. Fibers* **2023**, *20*, 2134267. https://doi.org/10.1080/15440478.2022.2134267.
- 134. Mehrparvar, L.; Safapour, S.; Sadeghi-Kiakhani, M.; Gharanjig, K. Chitosan-polypropylene imine dendrimer hybrid: A new ecological biomordant for cochineal dyeing of wool. *Environ. Chem. Lett.* **2016**, 14, 533–539. https://doi.org/10.1007/s10311-016-0559-1.

Sustainability **2024**, 16, 4661 31 of 36

135. Adeel, S.; Hussaan, M.; Rehman, F.; Habib, N.; Salman, M.; Naz, S.; Amin, N.; Akhtar, N. Microwave-assisted sustainable dyeing of wool fabric using cochineal-based carminic acid as natural colorant. *J. Nat. Fibers* **2019**, *16*, 1026–1034. https://doi.org/10.1080/15440478.2018.1448317.

- 136. Sajed, T.; Haji, A.; Mehrizi, M.K.; Boroumand, M.N. Modification of wool protein fiber with plasma and dendrimer: Effects on dyeing with cochineal. *Int. J. Biol. Macromol.* **2018**, *107 Pt A*, 642–653. https://doi.org/10.1016/j.ijbiomac.2017.09.038.
- 137. Safapour, S.; Mazhar, M.; Abedinpour, S. Broadening Color Shade Range of *Rubia tinctorum* L. Natural Colorants on Wool Fibers via Combination of Metal Mordants: Color Characteristics and Fastness Studies. *J. Nat. Fibers* **2023**, *20*, 2157923. https://doi.org/10.1080/15440478.2022.2157923.
- 138. Lin, L; Jiang, T.; Xiao, L.; Pervez, M.N.; Cai, X.; Naddeo, V.; Cai, Y. Sustainable fashion: Eco-friendly dyeing of wool fiber with novel mixtures of biodegradable natural dyes. *Sci. Rep.* **2022**, *12*, 21040. https://doi.org/10.1038/s41598-022-25495-6.
- 139. Sadeghi-Kiakhani, M.; Safapour, S.; Golpazir-Sorkheh, Y. Sustainable Antimicrobial and Antioxidant Finishing and Natural Dyeing Properties of Wool Yarn Treated with Chitosan-poly(amidoamine) Dendrimer Hybrid as a Biomordant. *J. Nat. Fibers* **2021**, *19*, 9988–10000. https://doi.org/10.1080/15440478.2021.1993475.
- 140. Hosseinnezhad, M.; Gharanjig, K.; Adeel, S.; Rouhani, S.; Imani, H.; Razani, N. The effect of ultrasound on environmentally extraction and dyeing of wool yarns. *J. Eng. Fibers Fabr.* **2022**, *17*, 15589250221104471. https://doi.org/10.1177/15589250221104471.
- 141. Hosseinnezhad, M.; Gharanjig, K.; Jafari, R.; Imani, H.; Razani, N. Cleaner colorant extraction and environmentally wool dyeing using oak as eco-friendly mordant. *Environ. Sci. Pollut. Res.* **2021**, *28*, 7249–7260. https://doi.org/10.1007/s11356-020-11041-2.
- 142. Zhao, Z.; Yan, C.; Xu, F.; Liu, J. Study on Dyeing Properties and Color Characteristics of Wool Fabrics Dyed with *Geranium caespitosum* L. Extract-A New Natural Yellow Dye. *Coatings* **2023**, *13*, 1125. https://doi.org/10.3390/coatings13061125.
- 143. Pandey, R.; Patel, S.; Pandit, P.; Nachimuthu, S.; Jose, S. Colouration of textiles using roasted peanut skin- an agro processing residue. *J. Clean. Prod.* **2018**, *172*, 1319–1326. https://doi.org/10.1016/j.jclepro.2017.10.268.
- 144. Alebeid, O.K.; Pei, L.; Elhassan, A.; Zhou, W.; Wang, J. Cleaner dyeing and antibacterial activity of wool fabric using Henna dye modified with *Acacia nilotica* pods. *Clean Technol. Environ. Policy* **2020**, 22, 2223–2230. https://doi.org/10.1007/s10098-020-01951-7.
- 145. Mehrizi, M.K.; Malakan, F.; Veysian, M.; Shahi, Z. The Effect of Different Nanoparticles on Dyed Wool Carpet with Walnut Shell Natural Dye. *Prog. Color. Color. Coat.* **2023**, *16*, 331–341. https://doi.org/10.30509/pccc.2023.167045.1191.
- 146. Arifeen, W.; Rehman, F.U.; Adeel, S.; Zuber, M.; Ahmad, M.N.; Ahmad, T. Environmental friendly extraction of walnut bark-based juglone natural colorant for dyeing studies of wool fabric. *Environ. Sci. Pollut. Res.* **2021**, *28*, 49958–49966. https://doi.org/10.1007/s11356-021-14277-8.
- 147. Sadeghi-Kiakhani, M.; Tehrani-Bagha, A.R.; Gharanjig, K.; Hashemi, E. Use of pomegranate peels and walnut green husks as the green antimicrobial agents to reduce the consumption of inorganic nanoparticles on wool yarns. *J. Clean. Prod.* **2019**, 231, 1463–1473. https://doi.org/10.1016/j.jclepro.2019.05.283.
- 148. Lachguer, K.; El Ouali, M.; Essaket, I.; El Merzougui, S.; Cherkaoui, O.; Serghini, M.A. Eco-Friendly Dyeing of Wool with Natural Dye Extracted from Moroccan *Crocus sativus* L. Flower Waste. *Fibers Polym.* **2021**, 22, 3368–3377. https://doi.org/10.1007/s12221-0256-y.
- 149. Hassan, M.M. Valorization of Sulfonated Kraft Lignin as a Natural Dye for the Sustainable Dyeing of Wool Fabrics: Effect of Peroxide Oxidation. *ACS Sustain. Chem. Eng.* **2023**, *11*, 13787–13797. https://doi.org/10.1021/acssuschemeng.3c04267.
- 150. Rather, L.J.; Zhou, Q.; Ali, A.; Mohd, Q.; Haque, R.; Li, Q. Valorization of Natural Dyes Extracted from Mugwort Leaves (*Folium artemisiae argyi*) for Wool Fabric Dyeing: Optimization of Extraction and Dyeing Processes with Simultaneous Coloration and Biofunctionalization. *ACS Sustain. Chem. Eng.* **2020**, *8*, 2822–2834. https://doi.org/10.1021/acssuschemeng.9b06928.
- 151. Salim, R.G.; Fadel, M.; Youssef, Y.A.; Taie, H.A.A.; Abosereh, N.A.; El-Sayed, G.M.; Marzouk, M. A local *Talaromyces atroroseus* TRP-NRC isolate: Isolation, genetic improvement, and biotechnological approach combined with LC/HRESI-MS characterization, skin safety, and wool fabric dyeing ability of the produced red pigment mixture. *J. Genet. Eng. Biotechnol.* **2022**, 20, 62. https://doi.org/10.1186/s43141-022-00335-2.
- 152. Baaka, N.; Haddar, W.; Ticha, M.B.; Amorim, M.T.P.; M'Henni, M.F. Sustainability issues of ultrasonic wool dyeing with grape pomace colourant. *Nat. Prod. Res.* **2017**, *31*, 1655–1662. https://doi.org/10.1080/14786419.2017.1285303.
- 153. Adeel, S.; Anjum, F.; Zuber, M.; Hussaan, M.; Amin, N.; Ozomay, M. Sustainable Extraction of Colourant from Harmal Seeds (*Peganum harmala*) for Dyeing of Bio-Mordanted Wool Fabric. *Sustainability* 2022, 14, 12226. https://doi.org/10.3390/su141912226.
- 154. Adeel, S.; Azeem, M.; Habib, N.; Hussaan, M.; Kiran, A.; Haji, A.; Haddar, W. Sustainable Application of Microwave Assisted Extracted Tea Based Tannin Natural Dye for Chemical and Bio-Mordanted Wool Fabric. *J. Nat. Fibers* **2023**, *20*, 2136322. https://doi.org/10.1080/15440478.2022.2136322.
- 155. Moussa, I.; Ghezal, I.; Sakli, F. Valorization of *Pelargonium graveolens* L'Hér. Hydrodistillation Solid Waste as Natural Dye for Wool Fabrics. *J. Nat. Fibers* **2023**, *20*, 2156966. https://doi.org/10.1080/15440478.2022.2156966.
- 156. Adeel, S.; Rehman, F.; Pervaiz, M.; Hussaan, M.; Amin, N.; Majeed, A.; Rehman, H. Microwave Assisted Green Isolation of Laccaic Acid from Lac Insect (*Kerria lacca*) for Wool Dyeing. *Prog. Color. Color. Coat.* **2021**, 14, 293–299. https://doi.org/10.30509/PCCC.2021.166734.1090.
- 157. Wang, X.; Guo, X.; Zhao, H.; Zhang, D.; Hong, G.; Wang, X.; Hou, X.; Yi, F. Optimization of the application of melanoidin microcapsules from lycium barbarum residue in wool fabric dyeing process. *J. Nat. Fibers* **2023**, 20, 2156021. https://doi.org/10.1080/15440478.2022.2156021.
- 158. Adeel, S.; Kiran, S.; Alam, M.; Farooq, T.; Amin, N.; Gulzar, T. *Alkanna tinctoria-based* sustainable alkanin natural colorant for eco-dyeing of wool. *Environ. Sci. Pollut. Res.* **2023**, *30*, 27073–27080. https://doi.org/10.1007/s11356-022-23806-y.

Sustainability **2024**, 16, 4661 32 of 36

159. Baseri, S. Sustainable dyeing of wool yarns with renewable sources. *Environ. Sci. Pollut. Res.* **2022**, *29*, 53238–53248. https://doi.org/10.1007/s11356-022-19629-6.

- 160. Hosseinnezhad, M.; Gharanjig, K.; Adeel, S.; Nahavandi, A.M. In quest for improvement of dyeing properties using agriculture waste: Utilization of Oleaster as new bio-mordant for wool yarns. *Environ. Sci. Pollut. Res.* **2023**, *30*, 122262–122273. https://doi.org/10.1007/s11356-023-30603-8.
- 161. Habib, N.; Ali, A.; Adeel, S.; ur-Rehman, F.; Aftab, M.; Inayat, A. Assessment of wild turmeric-based eco-friendly yellow natural bio-colorant for dyeing of wool fabric. *Environ. Sci. Pollut. Res.* 2023, 30, 4570–4581. https://doi.org/10.1007/s11356-022-22450-w.
- 162. Zong, S. The systematic application of plant dyeing techniques based on sustainable energy systems in ecological textile design. *Clean. Environ. Sys.* **2023**, *11*, 100143. https://doi.org/10.1016/j.cesys.2023.100143.
- 163. Netithammakorn, N.; Smith, E.; Lerpiniere, C.; Shen, J. Peroxidase-catalysed coloration of wool fabrics. *Color. Technol.* **2021**, *137*, 93–107. https://doi.org/10.1111/cote.12510.
- 164. Prajapati, C.; Smith, E.; Kane, F.; Shen, J. Laccase-catalysed Coloration of Wool and Nylon. *Color. Technol.* **2018**, 134, 423–439. https://doi.org/10.1111/cote.12350.
- 165. Antunes Barros, A.A.; da Rocha Cardoso, A.P.; Rodrigues, A.C.; dos Santos Marinho da Silva, C.J.; Zille, A. Optimizing enzymatic dyeing of wool and leather. SN Appl. Sci. 2019, 1, 1232. https://doi.org/10.1007/s42452-019-1241-6.
- 166. Bai, R.; Yu, Y.; Wang, Q.; Yuan, J.; Fan, X. Laccase-mediated in situ polymerization of pyrrole for simultaneous coloration and conduction of wool fabric. *Text. Res. J.* 2018, 88, 27–35. https://doi.org/10.1177/0040517516673336.
- 167. Sun, S.-S.; Xing, T.; Tang, R. Enzymatic dyeing and functional finishing of textile fibres with ferulic acid. *Indian J. Fibre Text Res.* **2015**, 40, 62–69.
- 168. Zhang, T.; Bai, R.; Shen, J.; Wang, Q.; Wang, P.; Yuan, J.; Fan, X. Laccase-catalyzed polymerization of diaminobenzenesulfonic acid for pH-responsive colour-changing and conductive wool fabrics. *Text. Res. J.* **2018**, *88*, 2258–2266. https://doi.org/10.1177/0040517517720497.
- 169. Yuan, M.; Wang, Q.; Shen, J.; Smith, E.; Bai, R.; Fan, X. Enzymatic coloration and finishing of wool with laccase and polyethylenimine. *Text. Res. J.* 2018, 88, 1834–1846. https://doi.org/10.1177/0040517517712096.
- 170. Bai, R.; Yu, Y.; Wang, Q.; Fan, X.; Wang, P.; Yuan, J.; Shen, J. Laccase-catalyzed poly(ethylene glycol)-templated 'zip' polymerization of caffeic acid for functionalization of wool fabrics. *J. Clean. Prod.* **2018**, 191, 48–56. https://doi.org/10.1016/j.jclepro.2018.04.213.
- 171. Zhang, T.; Bai, R.; Wang, Q.; Fan, X.; Wang, P.; Yuan, J.; Yu, Y. A novel strategy to improve the dyeing properties in lac-case-mediated coloration of wool fabric. *Color. Technol.* 2016, 133, 65–72. https://doi.org/10.1111/cote.12252.
- 172. Bai, R.; Yu, Y.; Wang, Q.; Yuan, J.; Fan, X. Effect of laccase on dyeing properties of polyphenol-based natural dye for wool fabric. *Fibers Polym.* **2016**, *17*, 1613–1620. https://doi.org/10.1007/s12221-016-5598-5.
- 173. Garg, H.; Singhal, N.; Singh, A.; Khan, M.D.; Sheikh, J. Laccase-assisted colouration of wool fabric using green tea extract for imparting antioxidant, antibacterial, and UV protection activities. *Environ. Sci. Pollut. Res.* **2023**, *30*, 84386–84396. https://doi.org/10.1007/s11356-023-28287-1.
- 174. Pour, R.A.; Bagheri, R.; Naveed, T.; Ali, N.; Rehman, F.; He, J. Surface functionalization of wool via microbial-transglutaminase and bentonite as bio-nano-mordant to achieve multi objective wool and improve dyeability with madder. *Heliyon* **2020**, *6*, e04911. https://doi.org/10.1016/j.heliyon.2020.e04911.
- 175. Salem, M.Z.M.; Ibrahim, I.H.M.; Ali, H.M.; Helmy, H.M. Assessment of the Use of Natural Extracted Dyes and Pancreatin Enzyme for Dyeing of Four Natural Textiles: HPLC Analysis of Phytochemicals. *Processes* **2020**, *8*, 59. https://doi.org/10.3390/pr8010059.
- 176. Ibrahim, N.A.; Amin, H.A.; Abdel-Aziz, M.S.; Eid, B.M. A green approach for modification and functionalization of wool fabric using bio- and nano-technologies. *Clean Technol. Environ. Policy* **2022**, 24, 3287–3302. https://doi.org/10.1007/s10098-022-02385-z.
- 177. Ferrero, F.; Mossotti, R.; Innocenti, R.; Coppa, F.; Periolatto, M. Enzyme-aided wool dyeing: Influence of internal lipids. *Fibers Polym.* **2015**, *16*, 363–369. https://doi.org/10.1007/s12221-015-0363-8.
- 178. An, F.; Fang, K.; Liu, X.; Yang, H.; Qu, G. Protease and sodium alginate combined treatment of wool fabric for enhancing inkjet printing performance of reactive dyes. *Int. J. Biol. Macromol.* **2020**, *146*, 959–964. https://doi.org/10.1016/j.ijbiomac.2019.09.220.
- 179. Prajapati, C.D.; Smith, E.; Kane, F.; Shen, J. Selective enzymatic modification of wool/polyester blended fabrics for surface patterning. *J. Clean. Prod.* **2019**, 211, 909–921. https://doi.org/10.1016/j.jclepro.2018.11.079.
- 180. Allam, O.; Elshemy, N.; El-Sayed, H. Simple and Easily Applicable Method for Reducing Freshwater Consumption in Dyeing of Wool Fabric. *J. Nat. Fibers* **2022**, *19*, 895–904. https://doi.org/10.1080/15440478.2020.1764439.
- 181. Hassan, M.M.; Bhagvandas, M. Sustainable low liquor ratio dyeing of wool with acid dyes: Effect of auxiliaries on agglomeration of dye molecules in a dyebath and dyeing uniformity. *J. Clean. Prod.* **2017**, *152*, 464–473. https://doi.org/10.1016/j.jclepro.2017.03.139.
- 182. Ozturk, E.; Cinperi, N.C.; Kitis, M. Green textile production: A chemical minimization and substitution study in a woolen fabric production. *Environ. Sci. Pollut. Res.* **2020**, *27*, 45358–45373. https://doi.org/10.1007/s11356-020-10433-8.
- 183. Czaplicki, Z.; Matyjas-Zgondek, E.; Strzelecki, S. Dyeing of Wool and Woolen Fabrics in Magnetically Treated Water. *J. Nat. Fibers* 2021, *18*, 2055–2062. https://doi.org/10.1080/15440478.2019.1711286.
- 184. Zhu, L.; Ding, X.; Wu, X. A novel method for improving the anti-pilling property of knitted wool fabric with engineered water nanostructures. *J. Mater. Res. Technol.* **2020**, *9*, 3649–3658. https://doi.org/10.1016/j.jmrt.2020.01.102.

Sustainability **2024**, 16, 4661 33 of 36

185. Li, Y.; Noro, J.; Martins, M.; Jing, S.; Silva, C.; Cavaco-Paulo, A. Changing the shape of wool yarns via laccase-mediated grafting of tyrosine. *J. Biotechnol.* **2021**, 339, 73–80. https://doi.org/10.1016/j.jbiotec.2021.08.001.

- 186. Yu, Y.; Wang, Q.; Yuan, J.; Fan, X.; Wang, P. A novel approach for grafting of β-cyclodextrin onto wool via laccase/TEMPO oxidation. *Carbohydr. Polym.* **2016**, *153*, 463–470. https://doi.org/10.1016/j.carbpol.2016.08.003.
- 187. Chatha, S.A.S.; Asgher, M.; Asgher, R.; Hussain, A.I.; Iqbal, Y.; Hussain, S.M.; Bilal, M.; Saleem, F.; Iqbal, H.M.N. Environmentally responsive and anti-bugs textile finishes—Recent trends, challenges, and future perspectives. *Sci. Total Environ.* **2019**, 690, 667–682. https://doi.org/10.1016/j.scitotenv.2019.06.520.
- 188. Jose, S.; Nachimuthu, S.; Das, S.; Kumar, A. Moth proofing of wool fabric using nano kaolinite. *J. Text. Inst.* **2018**, 109, 225–231. https://doi.org/10.1080/00405000.2017.1336857.
- 189. Akioka, S.; Hirai, S.; Iijima, K.; Hirai, A.; Alharbi, M.A.H. Rare-earth ions as antibacterial agents for woven wool fabric. *Chem. Pap.* **2022**, *76*, 3557–3567. https://doi.org/10.1007/s11696-021-01999-9.
- 190. Hu, J.; Iqbal, M.I.; Sun, F. Wool Can Be Cool: Water-Actuating Woolen Knitwear for Both Hot and Cold. *Adv. Funct. Mater.* **2020**, 30, 2005033. https://doi.org/10.1002/adfm.202005033.
- 191. Allafi, F.; Hossain, M.S.; Lalung, J.; Shaah, M.; Salehabadi, A.; Ahmad, M.I.; Shadi, A. Advancements in Applications of Natural Wool Fiber: Review. J. Nat. Fibers 2020, 19, 497–512. https://doi.org/10.1080/15440478.2020.1745128.
- 192. Broda, J.; Kobiela-Mendrek, K.; Baczek, M.; Rom, M.; Espelien, I. Sound Absorption of Tufted Carpets Produced from Coarse Wool of Mountain Sheep. J. Nat. Fibers 2023, 20, 2246103. https://doi.org/10.1080/15440478.2023.2246103.
- 193. Kobiela-Mendrek, K.; Baczek, M.; Broda, J.; Rom, M.; Espelien, I.; Klepp, I. Acoustic Performance of Sound Absorbing Materials Produced from Wool of Local Mountain Sheep. *Materials* **2022**, *15*, 3139. https://doi.org/10.3390/ma15093139.
- 194. Čuden, A.P. Knitting towards sustainability, circular economy and Industry 4.0. Appl. Res. 2023, 2, e202200087. https://doi.org/10.1002/appl.202200087.
- 195. Anas, M.S.; Abbas, A.; Awais, H.; Sarwar, M.E.; Hassan, T.U.; Abbas, H. A study on the effect of material type, structure tightness and finishing process on the physical and thermo-physiological properties of sandwich terry socks for winter wear. *J. Eng. Fibers Fabr.* **2023**, *18*, 15589250231153398. https://doi.org/10.1177/15589250231153398.
- 196. Klepp, I.G.; Haugrønning, V.; Laitala, K. Local clothing: What is that? How an environmental policy concept is understood. *Int. J. Fash. Stud.* **2022**, *9*, 29–46. https://doi.org/10.1386/infs\_00060\_1.
- 197. Berg, L.L.; Klepp, I.G.; Sigaard, A.S.; Broda, J.; Rom, M.; Kobiela-Mendrek, K. Reducing Plastic in Consumer Goods Opportunities for Coarser Wool. *Fibers* **2023**, *11*, 15. https://doi.org/10.3390/fib11020015.
- 198. Hossain, G.; Rahman, M.; Hossain, I.Z.; Khan, A. Wearable socks with single electrode triboelectric textile sensors for monitoring footsteps. *Sens. Actuator A Phys.* **2022**, *333*, 113316. https://doi.org/10.1016/j.sna.2021.113316.
- 199. Gurarslan, A.; Özdemir, B.; Bayat, I.H.; Yelten, M.B.; Kurt, G.K. Silver nanowire coated knitted wool fabrics for wearable electronic applications. *J. Eng. Fibers Fabr.* **2019**, *14*, 1558925019856222. https://doi.org/10.1177/1558925019856222.
- 200. Zhou, Y.; Myant, C.W.; Stewart, R. Multifunctional and stretchable graphene/textile composite sensor for human motion monitoring. *J. Appl. Polym. Sci.* **2022**, *139*, e52755. https://doi.org/10.1002/app.52755.
- 201. Wilson, S.A.; Laing, R.M.; Tan, E.; Wilson, C.A.; Arachchige, P.S.G.; Gordon, K.C.; Fraser-Miller, S.J. Determining deposits on knit fabrics, yarns, and fibers, from sensor-related treatments. *J. Text. Inst.* **2023**, 1–13. https://doi.org/10.1080/00405000.2023.2221427.
- 202. Grube, A.; Shamsabadi, A.A.; Firouzjaei, M.D.; Mohamed, S.I.G.P.; Hilger, L.; Elliott, M.; McKenzie, K.; Bavarian, M. Emperor's new clothes: Novel textile-based supercapacitors using sheep wool fiber as electrode substrate. *Nano Trends* **2023**, *3*, 100014. https://doi.org/10.1016/j.nwnano.2023.100014.
- 203. Ghosh, S.; Nitin, B.; Remanan, S.; Bhattacharjee, Y.; Ghorai, A.; Dey, T.; Das, T.K.; Das, N.C. A Multifunctional Smart Textile Derived from Merino Wool/Nylon Polymer Nanocomposites as Next Generation Microwave Absorber and Soft Touch Sensor. *ACS Appl. Mater. Interfaces.* 2020, 12, 17988–18001. https://doi.org/10.1021/acsami.0c02566.
- 204. Souri, H.; Bhattacharyya, D. Wool fabrics decorated with carbon-based conductive ink for low-voltage heaters, *Mater. Adv.* **2022**, 3, 3952–3960. https://doi.org/10.1039/D1MA00981H.
- 205. Nguyen, M.C.; Ngan Luong, T.Q.; Vu, T.T.; Anh, C.T.; Dao, T.C. Synthesis of wool roll-like silver nanoflowers in an ethanol/water mixture and their application to detect traces of the fungicide carbendazim by SERS technique. RSC Adv. 2022, 12, 11583–11590. https://doi.org/10.1039/d1ra09286c.
- 206. Komorowska, M.; Niemiec, M.; Sikora, J.; Gródek-Szostak, Z.; Gurgulu, H.; Chowaniak, M.; Atilgan, A.; Neuberger, P. Evaluation of Sheep Wool as a Substrate for Hydroponic Cucumber Cultivation. *Agriculture* **2023**, *13*, 554. https://doi.org/10.3390/agriculture13030554.
- 207. Bradshaw, T.; Hagen, K. Wool Pellets Are a Viable Alternative to Commercial Fertilizer for Organic Vegetable Production. *Agronomy* **2022**, *12*, 1210. https://doi.org/10.3390/agronomy12051210.
- Komorowska, M.; Niemiec, M.; Sikora, J.; Szelag-Sikora, A.; Gródek-Szostak, Z.; Findura, P.; Gurgulu, H.; Stuglik, J.; Chowaniak, M.; Atılgan, A. Closed-Loop Agricultural Production and Its Environmental Efficiency: A Case Study of Sheep Wool Production in Northwestern Kyrgyzstan. *Energies* 2022, 15, 6358. https://doi.org/10.3390/en15176358.
- 209. Dorugade, V.; Taye, M.; Qureshi, S.A.; Agazie, T.; Seyoum, B.; Badasa Abebe, B.; Komarabathina, S. Agrotextiles: Important Characteristics of Fibres and Their Applications—A Review. *J. Nat. Fibers* **2023**, 20, 2211290. https://doi.org/10.1080/15440478.2023.2211290.

Sustainability **2024**, 16, 4661 34 of 36

210. Herfort, S.; Pflanz, K.; Larsen, M.-S.; Mertschun, T.; Grüneberg, H. Influence of Sheep's Wool Vegetation Mats on the Plant Growth of Perennials. *Horticulturae* **2023**, *9*, 384. https://doi.org/10.3390/horticulturae9030384.

- 211. Constantin, M.; Raut, I.; Gurban, A.-M.; Doni, M.; Radu, N.; Alexandrescu, E.; Jecu, L. Exploring the Potential Applications of *Paecilomyces lilacinus* 112. *Appl. Sci.* **2022**, *12*, 7572. https://doi.org/10.3390/app12157572.
- Parlato, M.C.M.; Valenti, F.; Midolo, G.; Porto, S.M.C. Livestock Wastes Sustainable Use and Management: Assessment of Raw Sheep Wool Reuse and Valorization. *Energies* 2022, 15, 3008. https://doi.org/10.3390/en15093008.
- 213. Broda, J.; Bączek, M. Acoustic Properties of Multi-Layer Wool Nonwoven Structures. J. Nat. Fibers 2020, 17, 1567–1581. https://doi.org/10.1080/15440478.2019.1584078.
- 214. Ghiţuleasa, P.C.; Cărpuş, E.; Dorogan, A.; Bulacu, C.; Enciu, A. Sustainable wool insulation textile products—An opportunity for entrepreneurial initiatives in Romania *IOP Conf. Ser. Mater. Sci. Eng.* **2020**, *827*, 012016. https://doi.org/10.1088/1757-899X/827/1/012016.
- 215. Pennacchio, R.; Savio, L.; Bosia, D.; Thiebat, F.; Piccablotto, G.; Patrucco, A.; Fantucci, S. Fitness: Sheep-wool and Hemp Sustainable Insulation Panels. *Energy Procedia* **2017**, *111*, 287–297. https://doi.org/10.1016/j.egypro.2017.03.030.
- 216. Stanimirovic, M.; Vasov, M.; Mancic, M.; Rancev, B.; Medenica, M. Sustainable Vernacular Architecture: The Renovation of a Traditional House on Stara Planina Mountain in Serbia. *Buildings* **2023**, *13*, 1093. https://doi.org/10.3390/buildings13041093.
- 217. Beheshti, M.H.; Firoozi, A.; Jafarizaveh, M.; Tabrizi, A. Acoustical and Thermal Characterization of Insulating Materials Made from Wool and Sugarcane Bagasse. *J. Nat. Fibers* **2023**, *20*, 2237675. https://doi.org/10.1080/15440478.2023.2237675.
- 218. Altin, M.; Yildirim, G.S. Investigation of usability of boron doped sheep wool as insulation material and comparison with existing insulation materials. *Constr. Build. Mater.* **2022**, *331*, 127303. https://doi.org/10.1016/j.conbuildmat.2022.127303.
- 219. Urdanpilleta, M.; Leceta, I.; Guerrero, P.; de la Caba, K. Sustainable Sheep Wool/Soy Protein Biocomposites for Sound Absorption. *Polymers* **2022**, *14*, 5231. https://doi.org/10.3390/polym14235231.
- 220. Atbir, A.; Cherkaoui, M.; El Wardi, F.Z.; Khabbaz, A. Improvement of thermomechanical characteristics of multilayer biomaterial of sheep wool and clay. *Mater. Today Proc.* 2022, 58 Pt 4, 1331–1336. https://doi.org/10.1016/j.matpr.2022.02.224.
- 221. Dénes, T.-O.; Istoan, R.; Tamas-Gavrea, D.R.; Manea, D.L.; Hegyi, A.; Popa, F.; Vasile, O. Analysis of Sheep Wool-Based Composites for Building Insulation. *Polymers* 2022, 14, 2109. https://doi.org/10.3390/polym14102109.
- 222. Abdellah, M.Y.; Sadek, M.G.; Alharthi, H.; Abdel-Jaber, G.T.; Backar, A.H. Characteristic properties of date-palm fibre/sheep wool reinforced polyester composites. *J. Bioresour. Bioprod.* **2023**, *8*, 430–443. https://doi.org/10.1016/j.jobab.2023.09.003.
- 223. Cardinale, T.; Arleo, G.; Bernardo, F.; Feo, A.; De Fazio, P. Thermal and mechanical characterization of panels made by cement mortar and sheep's wool fibres. *Energy Procedia* **2017**, *140*, 159–169. https://doi.org/10.1016/j.egypro.2017.11.132.
- 224. Maldonado-Alameda, A.; Alfocea-Roig, A.; Huete-Hernández, S.; Giro-Paloma, J.; Chimenos, J.M.; Formosa, J. Magnesium phosphate cement incorporating sheep wool fibre for thermal insulation applications. *J. Build. Eng.* **2023**, *76*, 107043. https://doi.org/10.1016/j.jobe.2023.107043.
- 225. Rabbat, C.; Awad, S.; Villot, A.; Rollet, D.; Andrès, Y. Sustainability of biomass-based insulation materials in buildings: Current status in France, end-of-life projections and energy recovery potentials. *Renew. Sustain. Energy Rev.* **2022**, *156*, 111962. https://doi.org/10.1016/j.rser.2021.111962.
- 226. Beckman, I.P.; Lozano, C.; Freeman, E.; Riveros, G. Fiber Selection for Reinforced Additive Manufacturing. *Polymers* 2021, 13, 2231. https://doi.org/10.3390/polym13142231.
- 227. Azmami, O.; Sajid, L.; Boukhriss, A.; Majid, S.; El Ahmadi, Z.; Benayada, A.; Gmouh, S. Mechanical and aging performances of Palm/ Wool and Palm/ Polyester nonwovens coated by waterborne polyurethane for automotive interiors. *Ind. Crops Prod.* **2021**, 170, 113681. https://doi.org/10.1016/j.indcrop.2021.113681.
- 228. Serra, J.P.; Barbosa, J.C.; Silva, M.M.; Gonçalves, R.; Uranga, J.; Costa, C.M.; Guerrero, P.; de la Caba, K.; Lanceros-Mendez, S. Wool/soy protein isolate membranes as separators toward more sustainable lithium-ion batteries. *J. Energy Storage* **2024**, *75*, 109748. https://doi.org/10.1016/j.est.2023.109748.
- 229. Monreal-Perez, P.; Elduque, D.; López, D.; Sola, I.; Yaben, J.; Clavería, I. Full-scale dynamometer tests of composite railway brake shoes including latxa sheep wool fibers. J. Clean. Prod. 2022, 379 Pt 1, 134533. https://doi.org/10.1016/j.jclepro.2022.134533.
- 230. Caglar, A.; Sahan, B.; Sayin, A.G.; Pelen, Y.T.; Korkmaz, N.; Kivrak, H. The advanced polymer composite coated fabrics as an anode electrode and photocatalytic glucose micro fuel cell design. *J. Photochem. Photobiol. A Chem.* 2023, 444, 115005. https://doi.org/10.1016/j.jphotochem.2023.115005.
- 231. Qamar, W.; Khan, A.H.; Rehman, Z.u.; Masoud, Z. Sustainable Application of Wool-Banana Bio-Composite Waste Material in Geotechnical Engineering for Enhancement of Elastoplastic Strain and Resilience of Subgrade Expansive Clays. *Sustainability* **2022**, *14*, 13215. https://doi.org/10.3390/su142013215.
- 232. Kim, N.K.; Bruna, F.G.; Das, O.; Hedenqvist, M.S.; Bhattacharyya, D. Fire-retardancy and mechanical performance of protein-based natural fibre-biopolymer composites. *Composites* **2020**, *1 Pt C*, 100011. https://doi.org/10.1016/j.jcomc.2020.100011.
- 233. Jose, S.; Shanumon, P.S.; Paul, A.; Mathew, J.; Thomas, S. Physico-Mechanical, Thermal, Morphological, and Aging Characteristics of Green Hybrid Composites Prepared from Wool-Sisal and Wool-Palf with Natural Rubber. *Polymers* **2022**, *14*, 4882. https://doi.org/10.3390/polym14224882.
- 234. Malik, R.; Bhaskaran, D.; Meena, M.; Lata, S. Heavy metal removal using wool filters. Asian Tex. J. 2001, 10, 88–90.
- 235. Malik, R.; Bhaskaran, D.; Meena, M.; Lata, S. Heavy Metal Removal from Wastewater Using Adsorbents. In *Water Pollution and Remediation: Heavy Metals*; Inamuddin, Ahamed, M.I., Lichtfouse, E., Eds.; Environmental Chemistry for a Sustainable World Series; Springer: Cham, Switzerland, 2021; Volume 53, pp. 441–469. https://doi.org/10.1007/978-3-030-52421-0\_13.

Sustainability **2024**, 16, 4661 35 of 36

236. Simonič, M.; Flucher, V.; Luxbacher, T.; Vesel, A.; Zemljič, L.F. Adsorptive Removal of Heavy Metal Ions by Waste Wool. *J. Nat. Fibers* **2022**, *19*, 14490–14503. https://doi.org/10.1080/15440478.2022.2064401.

- 237. Pang, L.; Lin, S.; Krakowiak, J.; Yu, S.; Hewitt, J. Performance analysis of sheep wool fibres as a water filter medium for human enteric virus removal. *J. Water Process. Eng.* **2022**, 47, 102800. https://doi.org/10.1016/j.jwpe.2022.102800.
- 238. Condurache, B.-C.; Cojocaru, C.; Pascariu, P.; Samoila, P.; Harabagiu, V.C.R. *Chimie* **2022**, 25 (Suppl. S3), 245–260. https://doi.org/10.5802/crchim.168.
- 239. Thome, S. Untersuchungen zur Sorption von Innenraum-Luftschadstoffen durch Wolle (Investigations into the Sorption of Indoor Air Pollutants by Wool). Ph.D. Thesis, DWI and er RWTH Aachen eV, Aachen, Germany, 2006.
- 240. McNeil, S.J.; Zaitseva, L.I. The Development of Wool-Based Passive Filters to Improve Indoor Air Quality. *Key Eng. Mater.* **2016**, 671, 219–224. https://doi.org/10.4028/www.scientific.net/KEM.671.219.
- 241. McNeil, S.J.; Sunderland, M.R.; Leighs, S.J. The utilisation of wool as a catalyst and as a support for catalysts. *Appl. Catal. A-Gen.* **2017**, *541*, 120–140. https://doi.org/10.1016/j.apcata.2017.04.021.
- 242. Başbug, B.; Erdoğan, M.K.; Karakışla, M.; Saçak, M. Easily Applicable Heterogeneous Catalyst from Sustainable Resources: Anchoring of Pd Nanoparticles onto Chemically Poly(2-amino thiophenol)-Modified Wool Fabrics. *ACS Appl. Polym. Mater.* **2022**, *4*, 6834–6849. https://doi.org/10.1021/acsapm.2c00776.
- 243. Liang, X.; Xiao, H.; Zhang, T.; Zhang, F.; Gao, Q. A unique nanocomposite with FeCo nanoalloy anchored on S, N co-doped carbonaceous matrix for high bifunctional oxygen reduction reaction/oxygen evolution reaction electrocatalytic property in Zn-air battery. *J. Colloid Interface Sci.* **2023**, *630*, 170–181. https://doi.org/10.1016/j.jcis.2022.10.001.
- 244. Yamauchi, K.; Yamauchi, A.; Kusunoki, T.; Kohda, A.; Konishi, Y. Preparation of stable aqueous solution of keratins, and physiochemical and biodegradational properties of films. *J. Biomed. Mater. Res.* **1996**, 31, 439–444. https://doi.org/10.1002/(SICI)1097-4636(199608)31:4<439::AID-JBM1>3.0.CO;2-M.
- 245. Zhang, Q.; Liebeck, B.M.; Yan, K.; Demco, D.E.; Körner, A.; Popescu, C. Alpha-Helix Self-Assembly of Oligopeptides Originated from Beta-Sheet Keratin. *Macromol. Chem. Phys.* **2012**, *213*, 2628–2638. https://doi.org/10.1002/macp.201200446.
- 246. Fahim, N.H.; Sarkandi, A.F.; Montazer, M. Keratin extraction and its application: Extraction of wool keratin and application in diversified fields. In *The Textile Institute Book Series, The Wool Handbook, Morphology, Structure, Properties, Processing, and Applications*; Jose, S., Thomas, S., Basu, G., Eds.; Woodhead Publishing: Cambridge, UK, 2024; pp. 501–531. https://doi.org/10.1016/B978-0-323-99598-6.00008-6.
- 247. Feroz, S.; Muhammad, N.; Dias, G.; Alsaiari, M.A. Extraction of keratin from sheep wool fibres using aqueous ionic liquids assisted probe sonication technology. *J. Mol. Liq.* **2022**, *350*, 118595. https://doi.org/10.1016/j.molliq.2022.118595.
- 248. Shavandi, A.; Bekhit, A.E.D.A.; Carne, A.; Bekhit, A. Evaluation of keratin extraction from wool by chemical methods for bio-polymer application. *J. Bioact. Compat. Polym.* **2017**, *32*, 163–177. https://doi.org/10.1177/0883911516662069.
- 249. Sharma, S.; Gupta, A.; Kumar, A. Keratin: An Introduction. In *Keratin as a Protein Biopolymer*; Sharma, S., Kumar, A., Eds.; Series on Polymer and Composite Materials; Springer Nature: Cham, Switzerland, 2019; pp. 1–18. https://doi.org/10.1007/978-3-030-02901-2\_1;
- 250. Chilakamarry, C.R.; Mahmood, S.; Saffe, S.N.B.M.; Arifin, M.A.B.; Gupta, A.; Sikkandar, M.Y.; Begum, S.S.; Narasaiah, B. Extraction and application of keratin from natural resources: A review. 3 *Biotech* **2021**, 11, 220. https://doi.org/10.1007/s13205-021-02734-7.
- 251. Ye, W.; Qin, M.; Qiu, R.; Li, J. Keratin-based wound dressings: From waste to wealth. *Int. J. Biol. Macrom.* **2022**, 211, 183–197. https://doi.org/10.1016/j.ijbiomac.2022.04.216.
- 252. Kelly, R. Keratins in wound healing. In *Wound Healing Biomaterials. Functional Biomaterials*; Ågren, M.S., Ed.; Woodhead Publishing: Sawston, UK, 2016; Volume 2, pp. 353–365. https://doi.org/10.1016/B978-1-78242-456-7.00017-9.
- 253. Gaidau, C.; Stanca, M.; Niculescu, M.D.; Alexe, C.A.; Becheritu, M.; Horoias, R.; Cioineag, C.; Râpă, M.; Stanculescu, I.R. Wool Keratin Hydrolysates for Bioactive Additives Preparation. *Materials* **2021**, *14*, 4696. https://doi.org/10.3390/ma14164696.
- 254. Andonegi, M.; Correia, D.M.; Costa, C.M.; Lanceros-Mendez, S.; de la Caba, K.; Guerrero, P. Tailoring physicochemical properties of collagen-based composites with ionic liquids and wool for advanced applications. *Polymer* **2022**, 252, 124943. https://doi.org/10.1016/j.polymer.2022.124943.
- 255. Rezaee, R.; Montazer, M.; Mianehro, A.; Mahmoudirad, M. Single-step Synthesis and Characterization of Zr-MOF onto Wool Fabric: Preparation of Antibacterial Wound Dressing with High Absorption Capacity. *Fibers Polym.* **2022**, 23, 404–412. https://doi.org/10.1007/s12221-0211-y.
- 256. Ranjit, E.; Hamlet, S.; George, R.; Sharma, A.; Love, R.M. Biofunctional approaches of wool-based keratin for tissue engineering. J. Sci-Adv. Mater. Dev. 2022, 7, 100398. https://doi.org/10.1016/j.jsamd.2021.10.001.
- 257. Cho, Y.S.; Quan, M.; Kang, N.U.; Jeong, H.-J.; Hong, M.; Kim, Y.Y.; Cho, Y.-S. Strategy for enhancing mechanical properties and bone regeneration of 3D polycaprolactone kagome scaffold: Nano hydroxyapatite composite and its exposure. *Eur. Polym. J.* **2020**, *134*, 109814. https://doi.org/10.1016/eurpolymj.2020.109814.
- 258. Sun, L.; Li, S.; Yang, K.; Wang, J.; Li, Z.; Dan, N. Polycaprolactone strengthening keratin/bioactive glass composite scaffolds with double cross-linking networks for potential application in bone repair. *J. Leather Sci. Eng.* **2022**, *4*, 1. https://doi.org/10.1186/s42825-021-00077-w.
- 259. Sanchez Ramirez, D.O.; Vineis, C.; Cruz-Maya, I.; Tonetti, C.; Guarino, V.; Varesano, A. Wool Keratin Nanofibers for Bioinspired and Sustainable Use in Biomedical Field. *J. Funct. Biomater.* **2023**, *14*, 5. https://doi.org/10.3390/jfb14010005.

Sustainability **2024**, 16, 4661 36 of 36

260. Putra, N.E.; Zhou, J.; Zadpoor, A.A. Sustainable Sources of Raw Materials for Additive Manufacturing of Bone-Substituting Biomaterials. *Adv. Healthc. Mater.* **2024**, *13*, e2301837. https://doi.org/10.1002/adhm.202301837.

- 261. Fernández-d'Arlas, B. Tough and Functional Cross-linked Bioplastics from Sheep Wool Keratin. Sci. Rep. 2019, 9, 14810. https://doi.org/10.1038/s41598-019-51393-5.
- 262. Li, B.; Sun, Y.; Yao, J.; Wu, H.; Shen, Y.; Zhi, C.; Li, J. An environment-friendly chemical modification method for thiol groups on polypeptide macromolecules to improve the performance of regenerated keratin materials. *Mater. Des.* **2022**, *217*, 110611. https://doi.org/10.1016/j.matdes.2022.110611.
- 263. Guidotti, G.; Soccio, M.; Posati, T.; Sotgiu, G.; Tiboni, M.; Barbalinardo, M.; Valle, F.; Casettari, L.; Zamboni, R.; Lotti, N.; et al. Regenerated wool keratin-polybutylene succinate nanofibrous mats for drug delivery and cells culture. *Polym. Degrad. Stab.* **2020**, *179*, 109272. https://doi.org/10.1016/j.polymdegradstab.2020.109272.
- 264. Song, Y.; Qi, N.; Li, K.; Cheng, D.; Wang, D.; Li, Y. Green fluorescent nanomaterials for rapid detection of chromium and iron ions: Wool keratin-based carbon quantum dots. *RSC Adv.* 2022, *12*, 8108–8118. https://doi.org/10.1039/d2ra00529h.
- 265. Zhu, S.; Zhou, Q.; Yi, J.; Xu, Y.; Fan, C.; Lin, C.; Wu, J.; Lin, Y. Using Wool Keratin as a Structural Biomaterial and Natural Mediator to Fabricate Biocompatible and Robust Bioelectronic Platforms. *Adv. Sci.* (*Weinh.*) **2023**, *10*, e2207400. https://doi.org/10.1002/advs.202207400.
- 266. Sun, J.; Santiago, G.M.; Zhou, W.; Portale, G.; Kamperman, M. Water-Processable, Stretchable, and Ion-Conducting Coacervate Fibers from Keratin Associations with Polyelectrolytes. *ACS Sustain. Chem. Eng.* **2022**, *10*, 15968–15977. https://doi.org/10.1021/acssuschemeng.2c05411.
- 267. Dias, G.J.; Haththotuwa, T.N.; Rowlands, D.S.; Gram, M.; Bekhit, A.E.A. Wool keratin—A novel dietary protein source: Nutritional value and toxicological assessment. *Food Chem.* **2022**, *383*, 132436. https://doi.org/10.1016/j.foodchem.2022.132436.
- 268. Giteru, S.G.; Ramsey, D.H.; Hou, Y.; Cong, L.; Mohan, A.; Bekhit, A.E.A. Wool keratin as a novel alternative protein: A comprehensive review of extraction, purification, nutrition, safety, and food applications. Compr. Rev. Food Sci. Food Saf. 2023, 22, 643–687. https://doi.org/10.1111/1541-4337.13087.
- 269. Shen, B.; Zhang, D.; Wei, Y.; Zhao, Z.; Ma, X.; Zhao, X.; Wang, S.; Yang, W. Preparation of Ag Doped Keratin/PA6 Nanofiber Membrane with Enhanced Air Filtration and Antimicrobial Properties. *Polymers* **2019**, *11*, 1511. https://doi.org/10.3390/polym11091511.
- 270. Marchelli, F.; Rovero, G.; Curti, M.; Arato, E.; Bosio, B.; Moliner, C. An Integrated Approach to Convert Lignocellulosic and Wool Residues into Balanced Fertilisers. *Energies* **2021**, *14*, 497. https://doi.org/10.3390/en14020497.
- 271. Sharma, S.; Gupta, A. Sustainable Management of Keratin Waste Biomass: Applications and Future Perspectives. *Braz. Arch. Biol. Technol.* **2016**, *59*, e16150684. https://doi.org/10.1590/1678-4324-2016150684.
- 272. Tinoco, A.; Gonçalves, J.; Silva, C.; Loureiro, A.; Gomes, A.C.; Cavaco-Paulo, A.; Ribeiro, A. Keratin-based particles for protection and restoration of hair properties. *Int. J. Cosmet. Sci.* **2018**, 40, 408–419. https://doi.org/10.1111/ics.12483.
- 273. Mendoza-Muñoz, N.; Leyva-Gómez, G.; Piñón-Segundo, E.; Zambrano-Zaragoza, M.L.; Quintanar-Guerrero, D.; Del Prado Audelo, M.L.; Urbán-Morlán, Z. Trends in biopolymer science applied to cosmetics. *Int. J. Cosmet. Sci.* 2023, 45, 699–724. https://doi.org/10.1111/ics.12880.
- 274. Sionkowska, A. The potential of polymers from natural sources as components of the blends for biomedical and cosmetic applications. *Pure Appl. Chem.* **2015**, *87*, 1075–1084. https://doi.org/10.1515/pac-2015-0105.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.