Review

The Prevalence of Multidrug-Resistant Acinetobacter baumannii and Its Vaccination Status among Healthcare Providers

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Abstract: There is growing concern among healthcare providers worldwide regarding the prevalence of multidrug-resistant Acinetobacter baumannii (A. baumannii). Some of the worst hospital-acquired infections, often in intensive care units (ICUs), are caused by this bacterial pathogen. In recent years, the rise in multidrug-resistant A. baumannii has been linked to the overuse of antimicrobial drugs and the lack of adequate infection control measures. Infections caused by this bacterial pathogen are the result of prolonged hospitalization and ICU stays, and they are associated with increased morbidity and mortality. This review outlines the epidemiology, risk factors, and antimicrobial resistance associated with A. baumannii in various countries, with a special focus on the Kingdom of Saudi Arabia. In response to the growing concern regarding this drug-resistant bacteria, fundamental information about its pathology has been incorporated into the development of vaccines. Although these vaccines have been successful in animal models, their effectiveness in humans remains unproven. The review will discuss the development of A. baumannii vaccines, potential related obstacles, and efforts to find an effective strategy against this pathogen.

Keywords: multidrug-resistant Acinetobacter baumannii; prevalence; healthcare settings; vaccination strategies

1. Introduction

Acinetobacter baumannii is a widely distributed Gram-negative, non-fermentative, strictly non-motile, and oxidase-negative microbial pathogen that has been found in both natural and clinical environments [1,2]. It is also one of the six most significant pathogens in the ESKAPE group, which includes Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, A. baumannii, Pseudomonas aeruginosa, and Enterobacter species [3,4]. According to the Infectious Disease Society of America, A. baumannii is a serious global nosocomial concern [3,5]. It is a primary cause of various infections, particularly in immunosuppressed individuals or those receiving mechanical ventilation in intensive care units (ICUs) [6].
The most common infections associated with *A. baumannii* include pneumonia, bacteremia, meningitis, respiratory tract infection, and urinary tract infection [5]. Over the past decade, mortality rates due to *A. baumannii* infections have increased in many regions worldwide, ranging from 30% to 75% [7–9]. Infections caused by *A. baumannii* are associated with various risk factors, including burns, preterm birth, prolonged hospital stays, mechanical ventilation, indwelling devices, and the extensive use of antibiotic therapy [10–12].

According to the World Health Organization, multidrug-resistant *A. baumannii* is one of the most critical pathogens due to its resistance to numerous antibiotic classes, particularly carbapenems and third-generation cephalosporins [7]. Several studies conducted in the Kingdom of Saudi Arabia have indicated the increasing resistance of *A. baumannii* isolates to various antibiotics [13]. During the coronavirus 2019 (COVID-19) pandemic, the healthcare crisis caused by *A. baumannii*, particularly the carbapenem-resistant strains, reached its peak in ICUs [14,15]. Therefore, developing novel vaccines and drug targets remains imperative to combat the relentless emergence of drug resistance in *A. baumannii* against currently available antibacterial drugs [16,17]. Developing novel vaccines and therapeutics is a time-consuming process that often spans several years [18,19]. Before advancing to clinical trials, vaccine candidates must undergo testing in experimental animals to assess reactogenicity and immune response [20]. Furthermore, it is essential to evaluate the pharmacological and toxicological properties of the targeted vaccine candidates to ensure their safety and efficacy [21]. This review investigates the spread of multidrug-resistant *A. baumannii* infections in healthcare settings in Saudi Arabia and explores potential measures to prevent their transmission.

2. *A. baumannii*’s Mechanisms of Antibiotic Resistance

Healthcare facilities are currently grappling with an outbreak of multidrug-resistant *A. baumannii* [13,22]. Surveillance investigations worldwide have reported a significant increase in antimicrobial resistance by *A. baumannii* across diverse regions, including South East Asia [23], the Arabian Peninsula [24], and other parts of the world [22]. This has posed substantial healthcare and financial challenges in Saudi Arabia, where numerous cases of nosocomial multidrug-resistant *A. baumannii* have been documented [13].

*A. baumannii*, like other Gram-negative bacteria, possesses various defense mechanisms that enable it to evade the bactericidal and bacteriostatic effects of antimicrobial agents. These mechanisms include efflux pumps, which expel antibiotics from the cell through efflux mechanisms, and modifications to outer-membrane proteins (OMPs) that reduce the permeability of porins [25]. Additionally, *A. baumannii* has been found to produce metallo-β-lactamase enzymes such as imipenemase (IMP), Verona integron-mediated metallo-β-lactamase (VIM), New Delhi metallo-β-lactamase (NDM), and Seoul imipenemase (SIM). These enzymes can degrade antibiotics, rendering them ineffective [26]. However, the most crucial resistance mechanism employed by *A. baumannii* is the production of β-lactamases, particularly enzymes that hydrolyze carbapenems [14], penicillins [27], cephalosporins [27], and monobactams [14].

Studies conducted in Africa have revealed that the emergence of class D β-lactamases (OXA-23, -24, -51, and -58) and New Delhi metallo-β-lactamase (NDM-1) represents a significant mechanism of *A. baumannii* resistance. The presence of the *blaOXA-23*-like gene has been documented in several African countries, indicating its prevalence in these regions [28–31]. Furthermore, reports have highlighted the isolation of *A. baumannii* strains carrying Guiana extended-spectrum (GES) types, including GES-11, in Tunisia [32]. In addition to these mechanisms, other factors have contributed to multidrug resistance in this bacterium, such as the expression of efflux pumps (*TetA, TetB, and AdeABC*) [33] and the loss of or reduction in OMPs. In Saudi Arabia, El-Mahdy et al., (2017) conducted a study on carbapenem-resistant *A. calcoaceticus-baumannii* complex isolates from the eastern district. The findings revealed that these isolates carried the genes for carbapenem-resistant *blaOXA-23*-like and *ISAba1* phage, and nine of the isolates also possessed the *blaADC* gene [34].
3. The Epidemiology and Risk Factors Associated with *A. baumannii*

There have been numerous instances in which *A. baumannii* infections are transmitted from one medical facility to another due to the movement of infected patients. For instance, the Vietnam extended-spectrum-β-lactamase (VEB-1)-producing *A. baumannii* clone was responsible for spreading the VEB-1 strain across 55 hospitals in northern and southeastern France. Furthermore, European clonal types I and II have been observed circulating in healthcare settings in Italy, with evidence suggesting the transfer of this strain from healthcare settings in the Mediterranean area to those in southwestern Germany [35–37]. Initial reports from hospitals in New York also associated outbreaks involving the OXA-48 β-lactamase produced by *A. baumannii* with outbreaks detected within the hospitals. The movement of patients and healthcare staff between healthcare providers has facilitated the spread of multidrug-resistant *A. baumannii* [38].

In Saudi Arabia, several investigations have been conducted to identify potential risk factors for *A. baumannii* infections [13,39–42]. Several studies have indicated that *A. baumannii* endemic strains may spread to tracheal secretions in patients undergoing mechanical ventilation [43]. Patients aged 60 or older and those receiving prolonged oxygen therapy are at a higher risk of developing ventilator-associated pneumonia caused by *A. baumannii* [41,44]. The colonization of the intestinal tract by *A. baumannii* in hospitalized patients is a potential risk factor for antibiotic resistance and outbreaks of serious infections [45]. Additionally, patients with chronic conditions that compromise their immune systems, such as diabetes, malignancy, kidney disease, and persistent lung disease, are particularly susceptible [39,46,47]. An earlier study showed that out of 72 patients infected with *A. baumannii*, 36% had underlying illnesses, and 11% were diabetics [48].

4. The Impact of *Acinetobacter baumannii* among Saudi Healthcare Facilities

Saudi Arabia is geographically divided into 13 administrative regions (Figure 1), each with its own political authority. With a combined population of over 36 million as of 2023, the country accommodates millions of Muslims from around the globe during the annual Hajj pilgrimage in the cities of Makkah and Madinah. This massive congregation poses a significant risk for the transmission of infectious diseases. Consequently, Saudi Arabia serves as a central hub for the international dissemination of multidrug-resistant (MDR) strains [49–51].

![Figure 1](image_url). The Saudi Arabian map showing administrative regions and hospitals (highlighted in red).

According to the Saudi Ministry of Health, Acinetobacter infections are among the leading causes of healthcare expenditure [52]. The negative impact on outcomes and high mortality rates associated with Acinetobacter infections are due to inappropriate therapies and limited therapeutic options, especially in ICU patients [53,54]. Saudi Arabia has observed an increase in carbapenem-resistant *A. baumannii* over the past decade com-
pared to the 1990s [24]. A previous study reported that between 2005 and 2009, 26.5% of ventilator-associated pneumonia cases in Riyadh, Saudi Arabia, were attributed to Acinetobacter species [52]. In another investigation [55], the incidence of multidrug-resistant A. baumannii was 92.1%, the highest percentage recorded to date. A study published in 2015 unveiled the high resistance of A. baumannii infections in the El-Qassim region of Saudi Arabia to multiple antibiotics [56]. Similarly, an investigation conducted in Jeddah found that multidrug-resistant A. baumannii was present in 55% of patients in 2010 and 67% of patients in 2013 [42]. Table 1 provides a comprehensive overview of the incidence of multidrug-resistant A. baumannii over the past 15 years, with each investigation indicating a progressively higher prevalence. According to Table 1, A. baumannii displayed a variable degree of resistance, up to 100%, against carbapenems, penicillins, and cephalosporins during surveys conducted between 2004 and 2015 throughout Saudi Arabia.

**Table 1.** An evaluation of the resistance rates of A. baumannii to β-lactam antibiotics was carried out in hospitals across Saudi Arabia (the numbers in the table represent the percentage of A. baumannii resistant to antibiotics).

<table>
<thead>
<tr>
<th>Area</th>
<th>Region</th>
<th>Survey Year</th>
<th>Carbapenems</th>
<th>Penicillins</th>
<th>Cephalosporins</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Riyadh</td>
<td>2006</td>
<td>0</td>
<td>36</td>
<td>19</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2006–2008</td>
<td>-</td>
<td>79.1</td>
<td>92.1</td>
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<td></td>
<td></td>
<td>2009</td>
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<td>2010</td>
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<td>2011</td>
<td>100</td>
<td>64</td>
<td>-</td>
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<tr>
<td></td>
<td></td>
<td>2006–2014</td>
<td>-</td>
<td>77</td>
<td>88.5</td>
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<tr>
<td></td>
<td></td>
<td>2013–2015</td>
<td>-</td>
<td>73</td>
<td>82</td>
</tr>
<tr>
<td></td>
<td>Qassim</td>
<td>2011</td>
<td>-</td>
<td>9</td>
<td>-</td>
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<tr>
<td></td>
<td></td>
<td>2011–2015</td>
<td>-</td>
<td>-</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Makkah</td>
<td>2004–2005</td>
<td>90</td>
<td>14</td>
<td>-</td>
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<tr>
<td></td>
<td></td>
<td>2005–2006</td>
<td>95.3</td>
<td>46</td>
<td>28</td>
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<tr>
<td></td>
<td></td>
<td>2015</td>
<td>-</td>
<td>90</td>
<td>64</td>
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<td></td>
<td>Madinah</td>
<td>2014</td>
<td>96</td>
<td>-</td>
<td>-</td>
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<tr>
<td></td>
<td>Jeddah</td>
<td>1999–2000</td>
<td>81</td>
<td>15</td>
<td>-</td>
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<tr>
<td></td>
<td></td>
<td>2012</td>
<td>-</td>
<td>66</td>
<td>-</td>
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<tr>
<td></td>
<td></td>
<td>2010–2012</td>
<td>-</td>
<td>33</td>
<td>33.3</td>
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<td>2014</td>
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<td>2015</td>
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</tbody>
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Table 1. Cont.

<table>
<thead>
<tr>
<th>Area</th>
<th>Region</th>
<th>Survey Year</th>
<th>β-Lactam Antibiotics</th>
<th>Reference</th>
</tr>
</thead>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Carbapenems</td>
<td>Penicillins</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Amoxicillin-Clavulanic Acid</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>Piperacillin</td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>Piperacillin-Tazobactam</td>
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<td></td>
<td></td>
<td></td>
<td>Ceftazidime</td>
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<td></td>
<td>Cefotaxime</td>
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<td></td>
<td></td>
<td></td>
<td>Pefloxacin</td>
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<td></td>
<td>Tetracycline</td>
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<tr>
<td>South</td>
<td>Aseer</td>
<td>2011–2012</td>
<td>- 3</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2013–2014</td>
<td>0 52 50</td>
<td>81.5</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2014–2015</td>
<td>92 95.5 100</td>
<td>97.1</td>
</tr>
<tr>
<td>Najran</td>
<td></td>
<td>2012–2013</td>
<td>- 7.4 1.5</td>
<td>-</td>
</tr>
</tbody>
</table>

5. Controlling Multidrug-Resistant Acinetobacter baumannii Outbreaks in Healthcare Settings

There have been numerous instances worldwide where multidrug-resistant *A. baumannii* has been successfully controlled through the implementation of various methods [73]. The concept of a source control program encompasses antibiotic stewardship, hand hygiene, adherence to contact precautions, education, thorough environmental cleaning, and vaccination strategies [74]. To effectively manage multidrug-resistant *A. baumannii*, it is imperative to gather microbial susceptibility profile data from specific regions or hospitals and establish rapid and cost-effective testing methods [75]. Several measures need to be taken to address this issue, including providing healthcare professionals with training on conducting laboratory tests and enhancing research skills among caregivers, particularly those working in the ICU [76,77].

Despite colistin being one of the few remaining effective treatments for multidrug-resistant *A. baumannii*, its efficacy has been compromised due to the degradation of lipopolysaccharide (LPS) and the emergence of lipid A target modifications [78]. Consequently, there is an urgent need for novel therapeutic approaches to tackle these challenges. Different organizations have recommended various strategies as the primary treatment for multidrug-resistant *A. baumannii* infections [79–81]. In line with Shields et al.’s recommendation, a combination of high-dose ampicillin–sulbactam with ceftiderocol, polymyxin B, or tigecycline is recommended as the prescribed regimen in 2023 [82].

Alternative therapies, such as phage therapy, are increasingly being used to treat multidrug-resistant *A. baumannii* infections [83,84]. While phage therapy offers several advantages, certain aspects still require further clarification. These include the long-term viability of formulation and manufacturing scale, the potential resurgence of bacteriophage resistance, and the overall impact on gut microbiota [85]. In the future, virulent phages may emerge as potent bactericidal agents, potentially replacing antibiotics; however, the rapid emergence of phage resistance significantly impedes their widespread and continuous utilization [86].

6. Vaccine Development against Acinetobacter baumannii

*A. baumannii* vaccine is currently unavailable despite several vaccine candidates being tested [87]. Vaccines targeting *A. baumannii* are crucial for safeguarding individuals with weakened immune systems and patients with underlying medical conditions [88]. In the aftermath of McConnell et al.’s 2013 report [89], many laboratories are working on vaccines targeted to specific populations. Given the similarity among clinical isolates of *A. baumannii*, vaccines targeting conserved antigens may offer effective protection [90].
In the past decade, several multivalent vaccines have been considered, including live-attenuated strains, ghost bacteria, outer-membrane vesicles, and DNA (Table 2). Extensive studies have been conducted on the types and characteristics of potential vaccine candidates for *A. baumannii*. In this section of the review article, our objective is to assess each of these platforms, describe the experiments conducted with rodents to evaluate their ability to elicit immune responses, and assess their protective efficacy.

**Table 2. Types and characteristics of potential vaccine candidates for *Acinetobacter baumannii*.**

<table>
<thead>
<tr>
<th>Vaccine Type</th>
<th>Formula for a Vaccine</th>
<th>Examples</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Live-attenuated</td>
<td>Formalin-inactivated cells with aluminum-based adjuvant and alhydrogel® adjuvant 2%</td>
<td>Inactivated lipopolysaccharide-deficient whole cells</td>
<td>[91]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Inactivated <em>A. baumannii</em> Lac4 strain</td>
<td>[92]</td>
</tr>
<tr>
<td>Outer-membrane vesicles (OMVs) and complex (OMCs)</td>
<td>OMVs in phosphate buffer saline with an aluminum phosphate adjuvant</td>
<td>OMVs</td>
<td>[93]</td>
</tr>
<tr>
<td></td>
<td>OMCS in phosphate buffer saline with an aluminum phosphate adjuvant</td>
<td>OMCS</td>
<td>[93]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>LPS-free OMVs</td>
<td>[93]</td>
</tr>
<tr>
<td>DNA</td>
<td>3 µg of rickettsial outer-membrane protein A (rOmpA) in 0.1% aluminum hydroxide</td>
<td>OmpA</td>
<td>[94]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>OmpA and PAL</td>
<td>[95]</td>
</tr>
<tr>
<td>Bacterial ghosts</td>
<td><em>A. baumannii</em> Ali190 being incubated in a solution of hydrogen peroxide, sodium hydroxide, and sodium carbonate</td>
<td>LPS/surface</td>
<td>[96]</td>
</tr>
</tbody>
</table>

**6.1. Live-Attenuated Vaccines**

Live-attenuated vaccines are produced by modifying bacteria to reduce their harmfulness, allowing the immune system to combat the infectious agent without causing illness [90]. Since live-attenuated strains display antigens similar to those found in fully pathogenic strains, they provide multiple targets for both immunoglobulin and cellular immune responses [97]. However, a drawback of live-attenuated vaccines is their potential to regain their harmful properties through horizontal gene transmission or spontaneous mutation [98]. Furthermore, it remains unclear whether live-attenuated vaccinations can be administered to individuals with weakened immune systems, who are most susceptible to *A. baumannii* infection. Recently, a live-attenuated vaccine against *A. baumannii* was developed based on a D-glutamate strain [98]. By modifying the *murI1* and *murI2* genes, a significantly less virulent strain of *A. baumannii* was created, which still elicited both antibody- and cell-mediated immune responses and improved survival in immunized animals exposed to multiple pathogenic strains of *A. baumannii* [98]. Additionally, the immunity induced by an *A. baumannii* strain lacking thioredoxin (ΔtrxA) was investigated [99]. Thioredoxins are vital in regulating redox balance and oxidative stress in bacteria. It was observed that a clinical isolate of *A. baumannii* lacking thioredoxin did not cause illness in mice vaccinated with the ΔtrxA strain [99,100].

**6.2. Outer-Membrane Vesicles and Complex-Based Vaccines**

Virulent Gram-negative bacteria release outer-membrane vesicles (OMVs), which are non-infectious particles with diameters ranging from 10 to 300 nanometers [90,101]. *A. baumannii* is one of the bacteria that produce OMVs [102]. These spherical structures are typically composed of phospholipids, protein molecules, DNA, and LPS [103]. Alaniz et al. [104] suggest that OMVs are involved in delivering virulence factors to host cells, thereby initiating the infection process. McBroom and Kuehn [105] found that OMV production increases when stress levels are high and severe environments are present, such as during an infection. While the exact mechanism of OMV formation is yet to be determined, Deatherage and colleagues suggested that OMVs are generated when the density of outer-membrane–peptidoglycan connections is temporarily reduced. Furthermore, the
researchers discovered that OMPs in OMVs contain specific domains that can interact with peptidoglycan and influence their development [106]. The effectiveness of OMV-based vaccines has been demonstrated in various studies conducted in animal models, with a variety of microorganisms, including *Escherichia coli* [107,108], *Bordetella pertussis* [109], and *Burkholderia pseudomallei* [110,111]

When considering a vaccination based on OMVs, it is important to remember that LPS plays a significant role in immunogenicity, as well as the potential side effects that result from the excessive presence of endotoxin. Significant progress has been made in addressing some of these issues by isolating OMVs from the *A. baumannii* strain IB010 [93]. It has been reported that this strain cannot produce LPS due to a mutation in the *lpxD* gene [93]. A study investigating the protective effects of LPS- *A. baumannii* outer-membrane complexes (OMCs) utilized the same strain. Mice vaccinated with the LPS-deficient OMCs showed little to no toxicity, similar to that of the LPS-deficient OMVs. On day 7 of the same experiment, mice vaccinated with LPS-deficient OMCs had a 60% survival rate, while mice vaccinated with LPS-deficient OMCs supplemented with exogenous LPS had a 95% survival rate [112]. A noticeable difference was observed between the two groups compared to the untreated control group, in which no survivors were found [112]. These investigations suggest that the LPS antigen contained within an OMV-based vaccination plays a crucial role in protecting against infection, despite the potential absence of LPS, which could help prevent excessive inflammation levels.

6.3. Nucleic-Acid-Based Vaccines

The concept of nucleic acid vaccines has gained considerable attention as a potential platform for vaccinations in recent years. However, very few studies have been conducted on nucleic acid vaccines intended to protect against *A. baumannii* [113]. Recent studies have demonstrated that DNA vaccines carrying the *OmpA* gene from *A. baumannii* can protect mice in animal pneumonia models from potentially fatal bacterial infections [114,115]. Researchers have found that immunization with a plasmid-vector-mediated vaccine (*pVAXI*) carrying the *OmpA* and proteoglycan-associated lipoprotein (*PAL*) genes of *A. baumannii* is effective against a lethal pulmonary challenge with four heterologous strains, resulting in 50% survival rates. Additionally, a decrease in inflammatory cytokines and the infiltration of inflammatory cells were observed in the bronchoalveolar lavage (BAL) [114], along with a strong humoral response, mixed cellular responses from T-helper cells (Th1/Th2/Th17), and a reduction in microbial burdens. Another vaccine utilizing the alternative plasmid-encoded *OmpA* expressed through the eukaryotic expression vector pBudCE4.1 protected 16% of mice from a lung infection for up to 15 days following a challenge. This vaccine also showed modest responses in Interleukin (IL-2, IL-4, and IL-12) and interferons (IFNs) [116]. These DNA vaccination strategies are still in their early stages but show promise as potentially safe and affordable approaches for administering multivalent vaccines.

6.4. Bacterial-Ghost-Based Vaccines

Bacterial ghosts (BGs) are initially living bacteria that undergo a process in which all their cytosolic components are removed, leaving only their outer membranes intact. Due to their non-living nature, BGs possess several inherent advantages, including the inability to revert to a pathogenic phenotype when utilized [90]. BGs can also serve as self-adjuvants and can be engineered to contain specific proteins, DNA, medications, or other small molecules to enhance their efficacy. However, while BGs maintain their original outer membrane, an excessive amount of natural LPS may lead to increased inflammation [97]. In an experiment, Sprague Dawley rats were vaccinated with *A. baumannii* BGs (strain Ali190) via oral, subcutaneous, intramuscular, or intraperitoneal administration before being infected with 10⁸ CFU of the homologous *A. baumannii* strain [96]. *A. baumannii* BGs proved highly effective in protecting immunized rats against *A. baumannii* infection in all administration modes, except for oral vaccination, which resulted in a 67% survival rate [96]. However, despite the favorable outcomes of this initial study, it is crucial to investigate
whether *A. baumannii* BGs can confer immunity against a wide range of *A. baumannii* strains.

### 7. Lessons Learned and Future Perspectives

Despite efforts made, the development of vaccines against *A. baumannii* has fallen behind that of nosocomial pathogens such as *Clostridium difficile*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* [95]. No vaccine against *A. baumannii* has undergone clinical trials, indicating the challenges associated with creating a safe and effective vaccine against this pathogen.

#### 7.1. Technology of Modern Vaccines

Currently, research on anti-*A. baumannii* vaccine is still in its early stages. Most studies have focused on traditional methods and platforms, such as subunit and killed vaccines. However, there are ongoing explorations of novel antigen transporters and enhanced delivery mechanisms, including nucleic acid vaccines, virus vectors, conjugated carriers, and co-delivery [21,113,117]. It is worth noting that mRNA and DNA vaccine technologies have gained significant recognition due to the competition surrounding COVID-19 vaccines. Therefore, we can anticipate the increasing importance of nucleic-acid-based vaccines in treating *A. baumannii* and other infections in the coming years [113]. Additionally, proteomic approaches and reverse vaccination offer the benefit of utilizing bioinformatics to systematically evaluate and select potential immunogens for vaccine candidates in silico [113]. This method may be employed in the future to identify antigens and design vaccines that protect against *A. baumannii* infections.

#### 7.2. The Long-Term Effects of Cellular Immunity

A promising vaccine candidate against *A. baumannii* should demonstrate its protective effects in preclinical trials by improving the survival rates of immunized animals compared to non-immunized ones, reducing bacterial burdens within organs, and suppressing the levels of inflammatory markers in the serum [113]. While vaccination and infection have been extensively researched, less attention has been given to studying cell-mediated immunity following vaccination. To formulate effective vaccines against *A. baumannii*, researchers need to align their assessment procedures, including animal models, intervals between vaccinations and challenges, challenge strains, doses, and administration methods. This harmonization will enable more accurate comparisons of experimental vaccines developed in different laboratories [95].

#### 7.3. Cross-Protective Multivalent Vaccines

Vaccine candidates have been evaluated not only against homologous strains but also against challenges posed by other clinical isolates to demonstrate a broad level of protection. However, the degree of risk reduction varied significantly depending on the specific strain, vaccination protocol, and/or challenge dose [113]. Consequently, it is necessary to further assess the protective properties of these vaccines, especially when dealing with highly infectious strains at extremely high doses. Developing a vaccine that offers broad or diverse protection is challenging due to strain-dependent variation in selected antigens such as *OmpA* [118,119]. The complete ATCC 179878 genome revealed the presence of 28 putative alien islands, suggesting the potential for repeated infections and the difficulty of developing a vaccine [120]. *A. baumannii* has a high propensity for acquiring foreign DNA, leading to the emergence of novel genetic variants. Therefore, it is crucial to select conserved antigens and design vaccines with a wide range of components to provide adequate protection.

### 8. Limitations

This review has limitations, such as focusing solely on the prevalence of multidrug-resistant *A. baumannii* in Saudi Arabia without addressing other countries. To provide
a more comprehensive view, further studies should be conducted in other countries to compare the data. This will enable us to better understand the global distribution of multidrug-resistant *A. baumannii* and the strategies needed to control its spread. Additionally, we did not highlight any of the virulence factors associated with *A. baumannii* infection, which should be addressed. Identifying these virulence factors is essential for understanding how this bacterium causes infections and developing strategies for their prevention and treatment. Moreover, this review did not discuss the potential risks and safety issues associated with developing a vaccine against multidrug-resistant *A. baumannii*, nor did it provide any information regarding the costs or potential barriers associated with its development and implementation.

9. Conclusions

The review shows that multidrug-resistant *A. baumannii* infections are prevalent in Saudi Arabia and that combination therapies are a key strategy for overcoming drug resistance. In particular, antibiotic stewardship is essential for preventing the emergence of drug-resistant strains, and multivalent vaccines composed of outer-membrane vesicles (OMVs), bacterial ghosts (BGs), or multiple subunit vaccines hold promise in reducing these infections. The development of vaccines has undergone significant changes due to advancements in technology, particularly in the wake of COVID-19 vaccine development.


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References

2. Carvalheira, A.; Silva, J.; Teixeira, P. Acinetobacter spp. in food and drinking water—A review. *Food Microbiol.* 2021, 95, 103675. [CrossRef]


14. Kyratakis, I.; Vasilieou, E.; Pana, Z.D.; Tragianidis, A. Acinetobacter baumannii antibiotic resistance mechanisms. Pathogens 2021, 10, 373. [CrossRef]


44. Al-Saeed, A.; Balkhy, H.H.; Al-Dorzi, H.M.; Khan, R.; Rishu, A.H.; Arabi, Y.M. Acinetobacter is the most common pathogen associated with late-onset and recurrent ventilator-associated pneumonia in an adult intensive care unit in Saudi Arabia. *Int. J. Infect. Dis.* 2013, 17, e696–e701. [CrossRef]


83. Wei, J.; Peng, N.; Liang, Y.; Li, K.; Li, Y. Phage therapy: Consider the past, embrace the future. Appl. Sci. 2020, 10, 7654. [CrossRef]


90. Gellings, P.S.; Wilkins, A.A.; Morici, L.A. Recent advances in the pursuit of an effective Acinetobacter baumannii vaccine. Pathogens 2020, 9, 1066. [CrossRef]


97. Tabrizi, C.A.; Walcher, P.; Mayr, U.B.; Pachón, J.; McConnell, M.J. A lipopolysaccharide-free outer membrane vesicle vaccine protects against Acinetobacter baumannii infection. Pathogens 2020, 9, 1066. [CrossRef] [PubMed]


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