

Supplemental figure

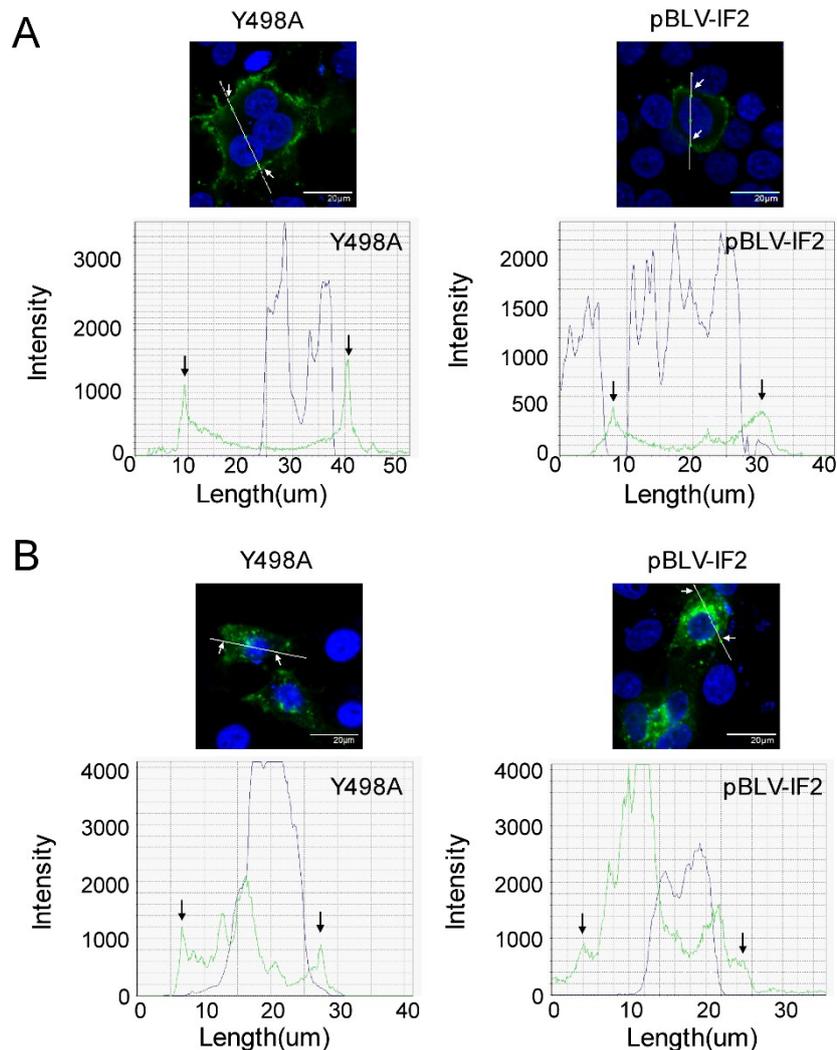


Figure S1. Quantification of intensity of Env protein on the cell membrane by line scan measurement. HeLa cells (1.0×10^5) were seeded on a coverslip in a 12 well plate the day prior to transfection and transfected with each pBLV-IF2, each pEnv, or control vector using FuGENE HD. **(A)** To detect cell surface gp51, cells were fixed and stained using an anti-gp51 MAb followed by Alexa Fluor 488-conjugated anti-Mouse IgG. **(B)** To detect intracellular gp51, cells were fixed, permeabilized with 0.5% Triton X-100, and stained with an anti-gp51 MAb followed by Alexa Fluor 488-conjugated anti-Mouse IgG. **(A and B)** Fluorescence intensity maps were plotted for linear transects drawn through the nuclei by line scan measurements using FV10-ASW 4.02 microscope software and fluorescence intensities on the cell surface were measured from these data (lower panel). Width of each line were thinner than 1 pixel. White in photo and black in graph arrows indicate that positions of the plasma membrane.

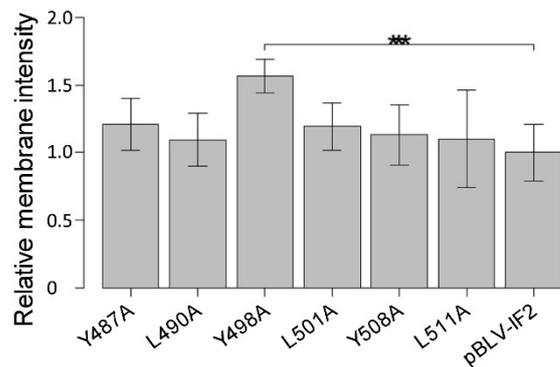
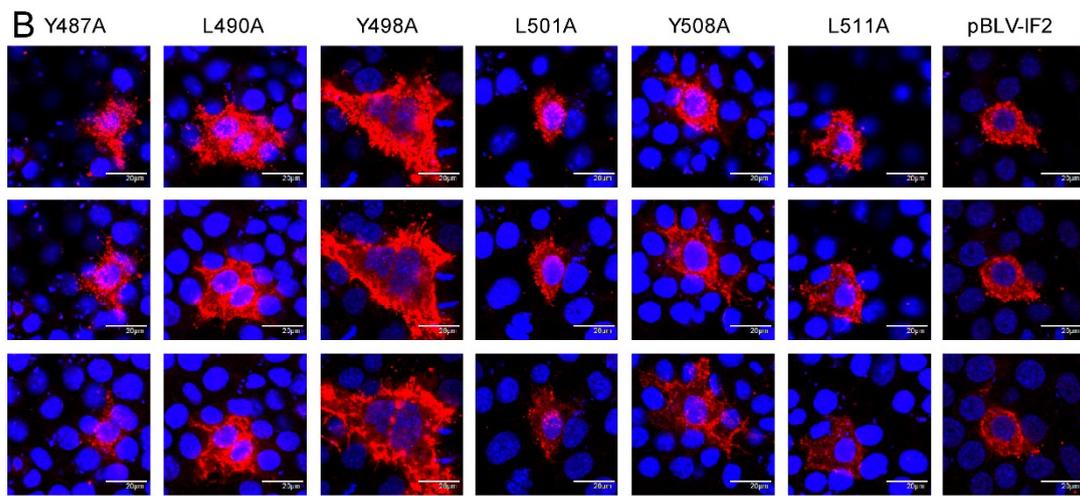
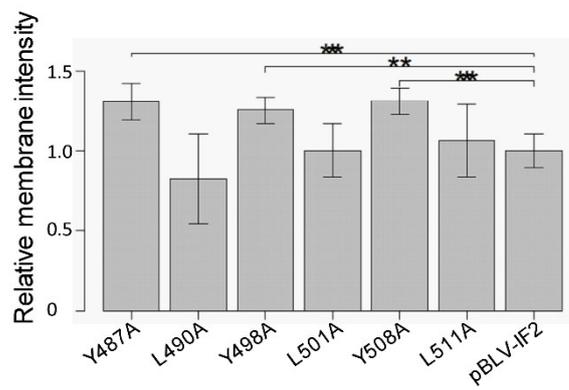
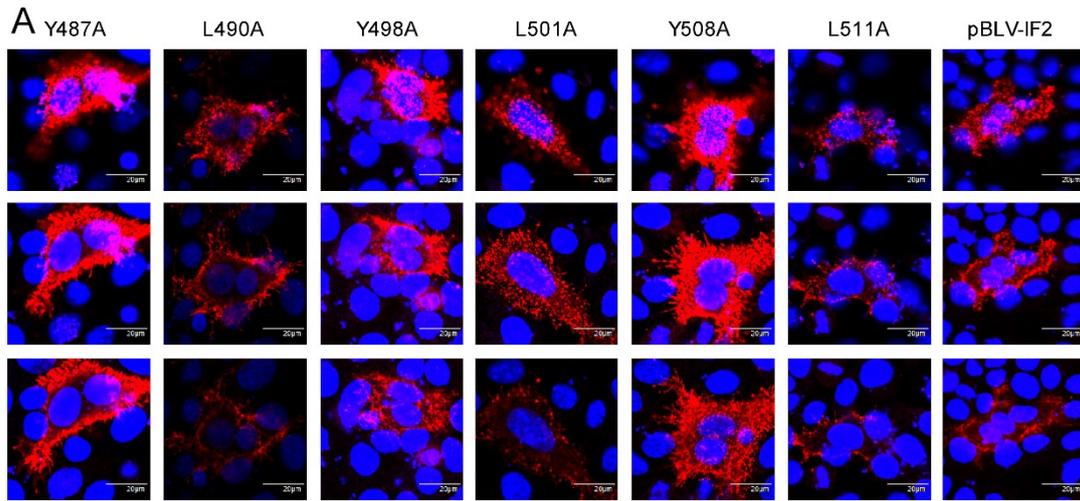


Figure S2. Z-stack analysis of localization of gp51 by mutant forms of the infectious molecular clone pBLV-IF2 in three optical section. HeLa cells (1.0×10^5) were seeded on a coverslip in a 12 well plate the day prior to transfection and transfected with 2 μ g of either wild-type pBLV-IF2, mutant pBLV-IF2, or control pBluescript II SK (-) using 8 μ L of FuGENE HD. **(A)** To detect cell surface gp51, cells were fixed and stained using an anti-gp51 MAb followed by Alexa Fluor 594-conjugated anti-Mouse IgG (upper panel). **(B)** To detect intracellular Env protein, cells were fixed, permeabilized with 0.5% Triton X-100, stained with anti-BLVgp51 MAb followed by Alexa Fluor 594-conjugated anti-Mouse IgG, and observed using an FV-1000 fluorescence microscope (upper panel). **(A and B)** For Z-stack analysis, three images were recorded at 2.0 μ m intervals. Fluorescence intensities on the cell surface were measured by line scan measurements through each cell using FV10-ASW 4.02 microscope software (under panel) and normalized by the mean intensity of pBLV-IF2. The results show the relative intensities of 10 cells expressing gp51. Each column and error bar represents the mean \pm SD of intensity for all cells. All values were analyzed by two-way analysis of variance (ANOVA) with the Dunnett test. The asterisk indicates a statistically significant difference (* $p < 0.05$; ** $p < 0.01$, *** $p < 0.001$).

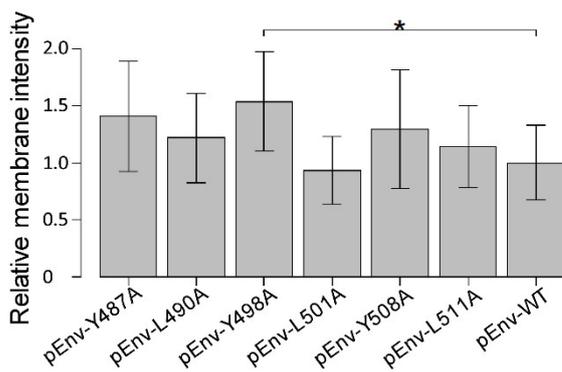
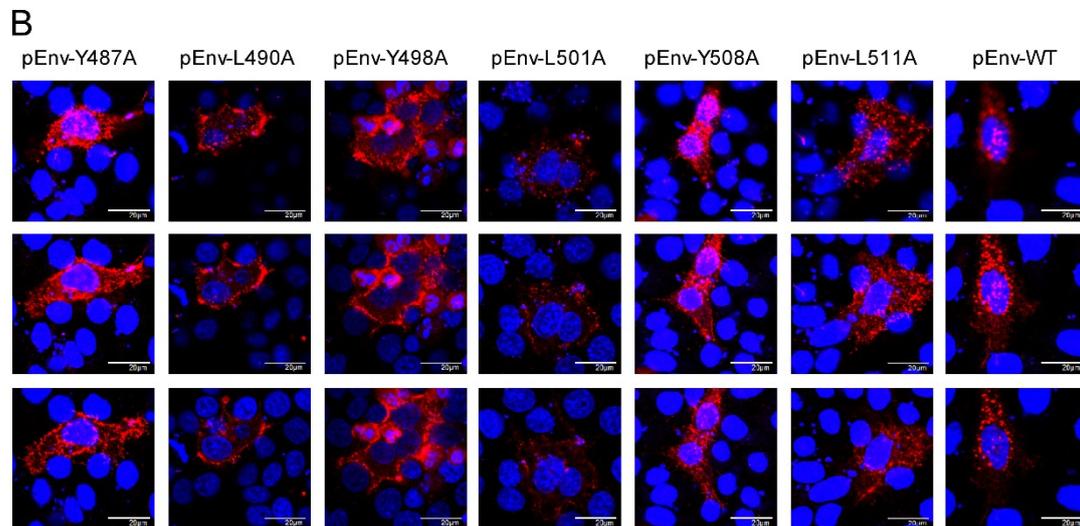
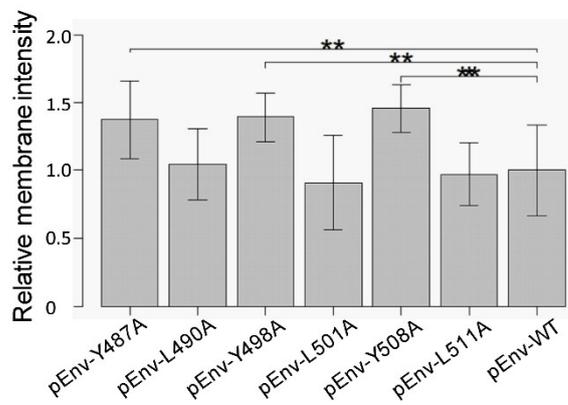
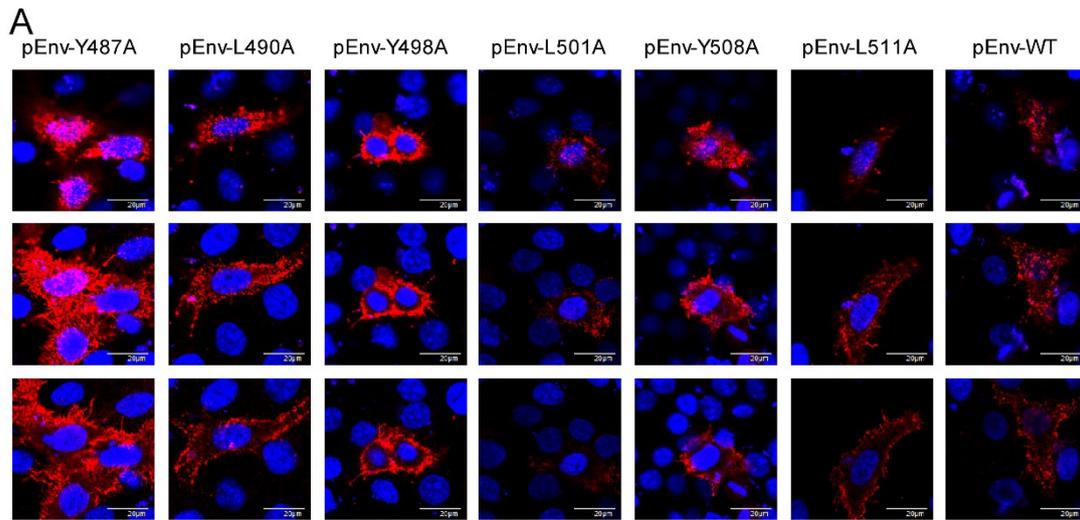


Figure S3. Z-stack analysis of localization of gp51 by mutant forms of the Env expression plasmid pEnv in three optical section. HeLa cells (1.0×10^5) were seeded on a coverslip in a 12 well plate the day prior to transfection and transfected with 2 μg of either wild-type pEnv, mutant pEnv, or the control pME-18neo using 8 μL of FuGENE HD. **(A)** To detect cell surface gp51, cells were fixed and stained using an anti-gp51 MAb followed by Alexa Fluor 594-conjugated anti-Mouse IgG (upper panel). **(B)** To detect intracellular Env protein, cells were fixed, permeabilized with 0.5% Triton X-100, stained with anti-BLVgp51 MAb followed by Alexa Fluor 594-conjugated anti-Mouse IgG, and observed using an FV-1000 fluorescence microscope (upper panel). **(A and B)** For Z-stack analysis, three images were recorded at 2.0 μm intervals. Fluorescence intensities on the cell surface were measured by line scan measurements through each cell using FV10-ASW 4.02 microscope software (under panel) and normalized by the mean intensity of pEnv-WT. The results show the relative intensities of 10 cells expressing gp51. Each column and error bar represents the mean \pm SD of intensity for all cells. All values were analyzed by two-way analysis of variance (ANOVA) with the Dunnett test. The asterisk indicates a statistically significant difference ($*p < 0.05$; $**p < 0.01$, $***p < 0.001$).