

Article

Using a Statistical Model to Examine the Effect of COD:SO₄²⁻ Ratio, HRT and LA Concentration on Sulfate Reduction in an Anaerobic Sequencing Batch Reactor

Rajesh Singh †, Chungman Moon ‡, Sathyanarayan S. Veeravalli, Saravanan R. Shanmugam, Subba Rao Chaganti and Jerald A. Lalman *

Department of Civil and Environmental Engineering, University of Windsor, 401 Sunset Ave., Windsor, ON N9B 3P4, Canada; E-Mails: rajesh.singh@cug.ac.in (R.S.); cmoon@kier.re.kr (C.M.); sevilim@uwindsor.ca (S.S.V.); ramiah@uwindsor.ca (S.R.S.); chaganti@uwindsor.ca (S.R.C.)

† Current address: School of Environment and Sustainable Development, Central University of Gujarat, Sector-30, Gandhinagar 382030, India.

‡ Current address: Korea Institute of Energy Research, 152 Gajeong-ro, Yuseong-gu 305343, Daejeon, Korea.

* Author to whom correspondence should be addressed; E-Mail: lalman@uwindsor.ca; Tel.: +1-519-253-3000 (ext. 2519); Fax: +1-519-971-3686.

External Editor: Miklas Scholz

Received: 29 August 2014; in revised form: 30 October 2014 / Accepted: 5 November 2014 / Published: 14 November 2014

Abstract: Taguchi statistical design, an orthogonal array (OA) method, was used to study the impact of the COD/SO₄²⁻ ratio, hydraulic retention time (HRT) and linoleic acid (LA) concentration on sulfate (SO₄²⁻) reduction in an anaerobic sequencing batch reactor using glucose as the electron donor. Based on the OA, optimum condition for maximum SO₄²⁻ reduction was evaluated. Increasing the COD/SO₄²⁻ ratio and HRT caused decreasing SO₄²⁻ reduction while increased SO₄²⁻ reduction was observed with increasing LA concentration (1 g L⁻¹). In control (not fed LA) cultures, higher SO₄²⁻ reduction (87% ± 3%) was observed at a low COD/SO₄²⁻ ratio of 0.8. This indicates that increasing SO₄²⁻ reduction was observed at increasing SO₄²⁻ loading rates. In general, results from this study reveal that limiting the substrate concentration with high SO₄²⁻ levels (low COD/SO₄²⁻ ratio) favors high SO₄²⁻ removal. Surface plots were used to evaluate the significant interactions between the experimental factors. Accuracy of the model was verified using an analysis of residuals.

Optimum conditions for maximum SO_4^{2-} reduction (97.61%) were observed at a COD/ SO_4^{2-} ratio of 0.8 (level 1), 12 h HRT (level 1) together with 1000 mg L^{-1} LA addition (level 3). In general, the Taguchi OA provided a useful approach for predicting the percent SO_4^{2-} reduction in inhibited mixed anaerobic cultures within the factor levels investigated.

Keywords: sulfate reduction; taguchi orthogonal array; mixed anaerobic culture; hydraulic retention time (HRT); COD/ SO_4^{2-} ratio

1. Introduction

The sulfate ion (SO_4^{2-}) is found in natural environments such as sediments, seawater and areas rich in decaying organic matter. Sulfate is also released in effluents from many industries such as pulp and paper processing, coal powered power plants, edible oil industries, tannery operations, molasses fermentation and mining [1,2]. Effluents generated from these industries also contain other sulfur species, which include thiosulfate, sulfite, sulfide and dithionite [3].

In mining operations, minerals, such as iron and zinc are converted to reduced metal sulfides. These sulfide compounds are oxidized with the release of metals ions and SO_4^{2-} (Reactions (1) and (2); Table 1). Under acidic conditions, dissolution of heavy metals from metal oxides and carbonates results in the formation of metal and SO_4^{2-} containing wastewater known as acid mine drainage (AMD) [4–6].

Discharging AMD can cause serious threats to the environment. Sulfate, an electron acceptor, is converted to hydrogen sulfide (H_2S) in the presence of electron donors such as hydrogen (H_2) or easily degradable organic chemicals [7]. Biological SO_4^{2-} reduction is a promising methodology to treat AMD due to the combined removal of acidity, SO_4^{2-} and heavy metals. Sulfate removal is accomplished by SO_4^{2-} reducing bacteria (SRB). SRBs utilize electron donors, such as volatile fatty acids (VFAs), alcohols and H_2 . SRBs often out-compete methane producing bacteria (MPB) for substrates, such as H_2 (Reactions (3) and (4); Table 1). When H_2 is utilized as an electron donor, SRBs produce H_2S and hydroxide ions (Reaction (5); Table 1).

Table 1. Standard Gibb's free energies for selected reaction stoichiometries.

Reaction No.	Stoichiometric Reaction	ΔG° (kJ mol ⁻¹)
(1)	$2\text{FeS}_2 + 7\text{O}_2 + 2\text{H}_2\text{O} \rightarrow 2\text{Fe}^{2+} + 4\text{SO}_4^{2-} + 4\text{H}^+$	-2168.0
(2)	$\text{ZnS} + 2\text{O}_2 \rightarrow \text{Zn}^{2+} + \text{SO}_4^{2-}$	-690.0
(3)	$4\text{H}_2 + \text{HCO}_3^- + \text{H}^+ \rightarrow \text{CH}_4 + 3\text{H}_2\text{O}$	-135.6
(4)	$4\text{H}_2 + \text{H}^+ + \text{SO}_4^{2-} \rightarrow 4\text{H}_2\text{O} + \text{HS}^-$	-152.2
(5)	$8\text{H}_2 + 2\text{SO}_4^{2-} \rightarrow \text{H}_2\text{S} + \text{HS}^- + 5\text{H}_2\text{O} + 3\text{OH}^-$	-146.9

Note: The Gibb's free energy values were calculated using data from Thauer *et al.* [8].

Competition between MPBs and SRBs is also dependent on the substrate concentration (Reactions (3) and (4); Table 1). Note if the substrate COD/ SO_4^{2-} ratio is large or with decreasing SO_4^{2-} levels, MPBs out-compete SRB for available electrons. Conversely, SRBs out-compete MPBs if the COD/ SO_4^{2-} ratio is low. A minimum COD/ SO_4^{2-} mol ratio of 0.67 is required for SO_4^{2-} reduction [9]. The percent SO_4^{2-} reduction is variable with different COD/ SO_4^{2-} ratios [10,11]. SRB

and MPB competition is also dependent on the operational pH with SRB growth favored at high pH [12]. Since the chemical equilibrium of different sulphide species is pH dependent [13,14], pH is a crucial factor affecting the competition between SRBs and MPBs.

Hydraulic retention time (HRT) can also affect competition between SRBs and MPBs. Lower HRT is favorable for SRB growth when compared to MPBs. The doubling time of many SRBs is less than MPBs. For example, *Desulfotomaculum acetoxidans* has a doubling time of 30 h when grown on acetate at 35 °C [15]. In comparison, the doubling time of *Methanosarcina barkeri* is 43 h when grown on acetate at 37 °C [16].

Controlling the electron flow to SRBs and MPBs is affected by factors such as pH, the COD/SO₄²⁻ ratio and HRT. Another factor controlling the activity of SRBs and MPBs is inhibitory chemicals. Successful inhibition of MPB growth will favor SRB growth and increase the quantity of SO₄²⁻ reduced. Diverting the fraction of substrate electron flow from MPBs to SRBs is achievable using different treatment methods, which selectively inhibit methanogenic growth. Among the physical methods, heat treatment is used to inhibit non-spore forming MPB [17]. However, due to the high cost associated with heat treatment, the method is unsuitable for full-scale application. Alternate methods to heat treatment include utilizing chemical inhibitors. Various chemical inhibitors have been used successfully to inhibit MPBs [18]. Inhibitors specific to inhibiting methanogens include 2-bromo ethane sulfonic acid (2-BESA). Other methanogenic and non-methanogenic inhibitors, which have been extensively studied, include saturated long chain fatty acids (SLCFAs) and unsaturated long chain fatty acids (ULCFAs). Lauric acid (C12:0), a SLCFA, and as linoleic acid (LA, C18:2), a ULCFA, are able to suppress gram positive bacteria and methanogens [19,20].

In natural habitats and engineered systems, microbial processes are affected by a combination of different factors. The effect of multiple factors on biological SO₄²⁻ reduction can be examined using statistical methods, such as the Taguchi design [21]. A significant difference between Taguchi's optimization technique and other similar methods is the ability to reduce process variability by involving factors that cause variability in the experimental design, modeling and optimization process [22]. The Taguchi method has been used in many biotechnological applications Rao *et al.* [21]. The method has been used by many researchers to optimize the operation of microbial processes [21,23,24].

The Taguchi method provides a systematic approach to understand manufacturing, microbial as well as environmental processes by assisting to identify factors which affects the process/product characteristics [25]. The method is robust and can be applied in the manufacture of automotive parts, plastics and semi-conductors [25]. The Taguchi method is used to rapidly and accurately gather data which is useful in the design and production of low-cost processes [26]. Hence, the objectives of this study are to examine the effect of the COD/SO₄²⁻ ratio, HRT and LA concentration on mesophilic biological SO₄²⁻ reduction in anaerobic sequencing batch reactors (ASBRs) using a Taguchi design.

2. Materials and Methods

2.1. Inoculum Source

The anaerobic inoculum was procured from an up-flow anaerobic sludge blanket reactor (UASBR) located at a brewery wastewater treatment facility (Guelph, ON, Canada) (designated as culture A) and at

the municipal wastewater treatment plant (Chatham, ON, Canada) (designated as culture B). Culture A and culture B were selected based on sources of MPBs and SRBs, respectively. The volatile suspended solid (VSS) of culture A and B was 50 and 20 g VSS L⁻¹, respectively. Cultures A and B were diluted with basal medium to 25 and 12 g VSS L⁻¹ in 9 L reactors, respectively (designated as reactor A and B). The bioreactors were operated in accordance with procedures reported by Ray *et al.* [27]. Reactors A and B were operated at 37 °C in a sequencing batch mode with a 14 d HRT and a feed concentration of 2000 mg glucose L⁻¹. The pH of the reactors was maintained at 7.0 ± 0.5. In addition to glucose, reactor B was acclimated incremental to increasing SO₄²⁻ levels of 250 mg L⁻¹ to 2000 mg L⁻¹ for 2 months. During the acclimation period, the quantity of gas and VFAs were monitored to establish quasi-steady state conditions. Inoculum for the experiments under consideration was combined from reactors A (80%) and B (20%) and diluted with basal medium to 8 g VSS L⁻¹. The basal medium composition used for dilution and feed was adapted from Wiegant and Lettinga [28]. All the chemicals for basal medium were procured from ACP Chemicals Inc. (Montreal, QC, Canada) and Sigma Aldrich (Oakville, ON, Canada). The feed substrate (glucose) was procured from Spectrum Chemicals, Gardena, CA, USA.

2.2. Sulfate Reduction Studies

Two 7-L (total volume) continuous stirred tank reactors (CSTRs) (New Brunswick Scientific, New Brunswick, NJ, USA) with a 5 L working volume were used to conduct the experiments. The CSTRs (R1 and R2) were operated as ASBRs at 37 ± 1 °C. Liquid samples were collected at the end of each cycle. Continuous mixing of the reactor contents during the reaction phase was conducted at 200 rpm. The pH (6.5 ± 0.1) was maintained using 1 M NaOH (base) and 0.5 M HCl (acid). The ASBRs (R1 and R2) were seeded with the inoculum from reactors A and B (8 g VSS L⁻¹) and then purged with nitrogen (N₂) (99.99% purity, Praxair, Windsor, ON, Canada) for 5 min to maintain anaerobic conditions. The experimental reactors (R1 and R2) were operated under identical conditions with a feed concentration of 2000 mg glucose L⁻¹ (2.134 g COD L⁻¹) as a carbon source. The SO₄²⁻ concentration was varied according to the COD/SO₄²⁻ ratio shown in Table 2.

Reactors R1 and R2 were operated as follows: 40 min settling; 10 min decanting; and 10 min fill. The reaction times maintained were 5 h, 11 h and 17 h for HRT values of 12 h, 24 h and 36 h, respectively. The volume decanted per cycle was constant at 2.5 L and the HRT was calculated using Equation (1) [29].

$$\text{HRT} = \frac{(\text{Working volume of the reactor})}{(\text{Volume decanted per cycle})(\text{No. of cycles per day})} \quad (1)$$

At each HRT, the reactors were operated until they attained quasi-steady state conditions (constant SO₄²⁻ reduction with ±10% variation). Different LA levels (0, 0.5 and 1.0 g L⁻¹) were fed to cultures according to experimental conditions shown in Table 2. Cultures were incubated with LA for 24 h prior to initiating the experiment (substrate addition).

2.3. Analytical Methods

Biogas production was monitored using a tipping bucket gas meter [30]. The biogas composition was determined by gas chromatography (GC) [7]. The detection limits for CH₄ and H₂ were 0.0032 kPa

(0.5 mL/bottle (160 mL)) and H₂S was 0.0315 kPa (5 mL/bottle (160 mL)), respectively. The liquid samples collected at the end of each cycle were analyzed for SO₄²⁻ using an ion chromatography (IC) [7]. The detection limits for the SO₄²⁻ was 0.5 mg L⁻¹. The total suspended solids (TSS) and VSS levels were measured according to *Standard Methods* [31].

Table 2. Design matrix for experimental factors and corresponding response function at different factor levels.

Exp. No.	COD/SO ₄ ²⁻ Ratio ¹		HRT (h)		LA conc. (mg L ⁻¹)		Experimental SO ₄ ²⁻ Reduction (%) ²	Predicted SO ₄ ²⁻ Reduction (%)
	X ₁	Level	X ₂	Level	X ₃	Level		
1	0.8	1	12	1	0	1	86.5 ± 2.6	83.5
2	0.8	1	24	2	500	2	65.8 ± 1.9	67.8
3	0.8	1	36	3	1000	3	80.6 ± 0.7	81.6
4	1.6	2	12	1	500	2	75.1 ± 1.9	76.1
5	1.6	2	24	2	1000	3	78.2 ± 3.7	75.2
6	1.6	2	36	3	0	1	58.3 ± 2.7	60.3
7	2.4	3	12	1	1000	3	89.9 ± 6.0	91.9
8	2.4	3	24	2	0	1	61.5 ± 8.6	62.5
9	2.4	3	36	3	500	2	64.5 ± 2.9	61.5

Notes: LA = linoleic acid; HRT = hydraulic retention time; COD = Chemical oxygen demand; SO₄²⁻ = sulfate;

¹ COD/SO₄²⁻ ratio of 0.8 denotes a glucose COD concentration of 2.134 g L⁻¹ and SO₄²⁻ concentration of 2.668 g L⁻¹.

Similarly in order to attain a COD/SO₄²⁻ ratio of 1.6 and 2.4, SO₄²⁻ concentration of 1.334 and 0.889 g L⁻¹ were used by keeping a constant glucose COD concentration of 2.134 g L⁻¹; ²values shown in mean ± standard deviation represents the average sulfate reduction from two reactor samples for at least three consecutive cycles;

2.4. Taguchi Design

The optimization methodology adopted in this study was divided into three phases, namely planning, conducting, and analysis. Each phase had a separate objective, interconnected sequence wise to achieve the overall optimization process using Qualitek-4 software (Nuteck Inc., Bloomfield Hills, MI, USA). Qualitek-4 software is equipped to use L4–L64 arrays with the selection of 2–63 factors and with 1, 3, and 4 levels for each factor.

2.4.1. Fractional Factorial Design of Experiments (FFDOE) (Phase 1)

The initial step in phase 1 was to identify different key parameters/factors to be optimized in the anaerobic process, which have a critical influence on the percent of SO₄²⁻ removed. The normal practice has been to experiment with a feasible range so that the variation inherent in the process does not mask the factor effect. Factors were selected and the ranges were assigned based on data from work reported by Moon *et al.* [7,32] and Kaksomen *et al.* [32]. Three factors (HRT, COD/SO₄²⁻ ratio, LA concentration) with significant influence on the SO₄²⁻ removal rate were selected for the Taguchi orthogonal array (OA) study. The levels for the three factors are shown in Table 2. The L9 OA can handle up to four factors at three levels with eight degrees of freedom. Since only three factors were examined in this study, the fourth column in the OA was left empty. Orthogonality is not lost by maintaining one or more columns of an array empty [33]. Taguchi's OA are used to estimate main

effects using only a few experimental runs. An OA (n, q, s, t) is an $n \times p$ array with entries from a set of s distinct symbols such that for any collection of t columns of the array, each of the s^t row vectors appears equally often in the matrix [34].

After selecting the levels, the OAs was created for the parameter design indicating the number and conditions for each experiment. Next, experiments were conducted as indicated in the completed array to gather data on the effect on the performance measure. The final step in phase 1 is to conduct the data analysis by assessing the effect of different parameters on the performance measure. An experimental design matrix was generated to define the data analysis procedure and a L9 OA for the control parameters to fit a specific study was selected. In the present OA, the three levels of factor variation were considered and the size of experimentation was represented by symbolic arrays L9 (the 9 experimental trials are shown in Table 2).

2.4.2. Sulfate Removal ASBR Experiments with Selected Factors and Levels (Phase 2)

Phase 2 was focused on conducting the experiments according to the Taguchi L9 OA (Table 2). The details of the inoculum source and detailed experimental methodology are outlined in Sections 2.1 and 2.2, respectively. The analytical methods used in this study to quantify the experimental response (%SO₄²⁻ reduction) are outlined in Section 2.3.

2.4.3. Analysis of Experimental Data (AED) and Prediction of Performance (POP) (Phase 3)

The SO₄²⁻ removal rate data obtained from the L-9 experiments were analyzed using the Qualitek-4 software with the “bigger-is-better” quality characteristics selected to determine the optimum conditions (higher SO₄²⁻ removal rate) and to identify individual factor influence on the SO₄²⁻ removal rate. In the Taguchi’s method, quality is measured by the deviation of a characteristic from a target value using the loss function (Equation (2)).

$$L(y) = k(y - m)^2 \quad (2)$$

where k denotes the proportionality constant, m represents the target value and y is the experimental value obtained for each trial.

The experimental data processed using the Qualitek-4 software with the bigger is better quality characteristics determined the optimum conditions for SO₄²⁻ removal. In the optimization, the bigger-is-better quality characteristic for the loss function is represented as Equation (3).

$$L(y) = k\left(\frac{1}{y^2}\right) \quad (3)$$

The expected loss function can be represented by Equation (4):

$$E[L(y)] = k E\left(\frac{1}{y^2}\right) \quad (4)$$

where $E(1/y^2)$ can be estimated from a sample of as Equation (5):

$$\sum_{i=1}^n \left[\frac{1}{y_i^2} \right] / n \quad (5)$$

Verification of the model was performed by an analysis of residuals. The residuals for the OA were calculated using the difference between the models predicted response and the experimental response at identical factor levels within the design space under consideration. The Anderson-Darling (AD) test was used to determine whether the residuals follow a normal distribution. The AD test at 5% level of significance was used to confirm the accuracy of the model based on the distribution of residuals. Three-dimensional surface plots were used to evaluate the effect of any two experimental factors on the experimental response. The ANOVA, AD plots and surface plots were generated using the MINITAB 16 statistical software (MINITAB Inc., State College, PA, USA).

3. Results and Discussion

3.1. Experimental Design Analysis

The Taguchi method is based on OAs providing a systematic, simple and efficient approach [35]. This method provides an approach which allows for a realistic arrangement of the experimental data sets with the understanding system, parameter, and tolerance designs [35]. The Taguchi OA is used to identify relationships between experimental independent variables and response dependent variable (%SO₄²⁻ reduction). The residual quantity of SO₄²⁻ measured at the end of each experimental run in the effluent was used to compute the percent of SO₄²⁻ removed (Table 2). This response variable was used to predict the optimum response using the three factors and three levels. The regression coefficients computed for the experimental response (%SO₄²⁻ reduction) were used to derive a model equation involving the three independent factors (Equation (6)).

$$\begin{aligned} \text{Sulfate reduction (\%)} &= 60.762 - 2.838 \times \left(\text{COD}/\text{SO}_4^{2-} \text{ ratio}\right) - 7.995 \times (\text{HRT}) + 7.069 \\ &\times (\text{LA concentration}) + 4.23 \times \left(\text{COD}/\text{SO}_4^{2-} \text{ ratio}\right)^2 + 7.338 \\ &\times (\text{HRT})^2 + 7.361 \times (\text{LA concentration})^2 \end{aligned} \quad (6)$$

3.2. Analysis of Variance

ANOVA was conducted to analyze the experimental response (%SO₄²⁻ reduction) at different conditions and to determine variation in contribution of each factor to the response variable (Table 3). The Fisher statistic (F-test) was used to establish whether the factors under investigation have any significant effects on the quality characteristic. In particular, the F ratio is used to determine the significance of the different experimental factor. The calculated F ratios indicate all the individual factors are statistically significant at a 95% confidence limit. The p values were used to determine the significance of each factor on SO₄²⁻ reduction. Based on the p values, the HRT contributed the maximum impact (49.99%) on the overall SO₄²⁻ reduction followed by LA with 41.42% (Figure 1 and Table 3).

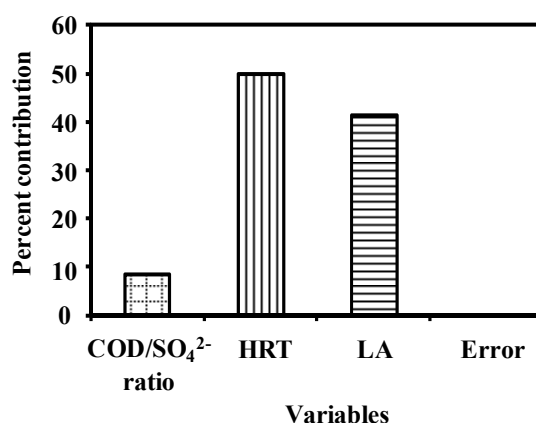
The COD/SO₄²⁻ ratio showed the least impact at the individual level (8.58%). The results from the study indicate that both HRT and LA concentration contributed more than 91% towards SO₄²⁻ reduction.

Table 3. ANOVA table.

Factor	DOF (<i>f</i>)	Sum of Squares (<i>s</i>)	Mean Squares	Variance (<i>v</i>)	F ratio (<i>F</i>) ¹	Pure Sum (<i>S'</i>)	Percent p (%) ²
COD/SO ₄ ²⁻	2	84.57	42.29	42.29	845,749.5 ³	84.57	8.58
HRT	2	492.67	246.33	246.33	4,926,686.4 ³	492.67	50.00
LA	2	408.16	204.08	204.08	4,081,612.0 ³	408.16	41.42
Error	2	0.001	0.0005	0.001			0.002
Total	8	985.40	492.70				100.00

Notes: F ratio (*F*) = Mean square error/residual square error; ¹Critical $F_{0.05, 2, 8} = 4.46$; ²Percent p (%) = (Sum of squared deviations/total sum of squared deviations) × 100; ³denotes significant at 95% confidence level.

Figure 1. Percent contribution of each variable on the sulfate removal rate. (1. HRT = hydraulic retention time; LA = linoleic acid; 2. Percent contribution of each experimental variable was estimated using ANOVA).



3.3. Effect of Factors on the Response Variables

3.3.1. Main Effects Plot

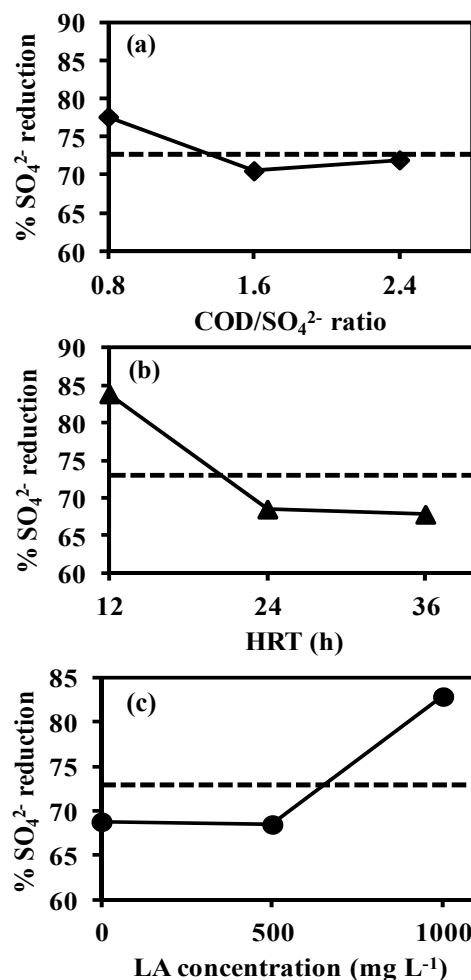
The main effects plot was used to establish the effect of each experimental factor on the response variable. The average response for each factor without considering the effects of other experimental factors is shown in the plots. Hence, interpretation of the plots must be conducted with caution. The experimental response variation between $61.5\% \pm 8.6\%$ and $89.9\% \pm 6.0\%$ (Table 2) indicates the effect of the different factors on SO₄²⁻ reduction.

Effect of COD/SO₄²⁻ Ratio

The COD/SO₄²⁻ ratio is a major factor affecting SO₄²⁻ reduction [36]. According to Velasco *et al.* [36], for a given SO₄²⁻ concentration, the feed COD/SO₄²⁻ ratio was used to control H₂S production, which in-turn was used for metal precipitation. Varying quantities of the % SO₄²⁻ removed was observed in cultures fed different COD/SO₄²⁻ ratios (Figure 2a). The main effects plot showed a maximum mean SO₄²⁻ removal of 78% at a COD/SO₄²⁻ ratio of 0.8. With increasing COD/SO₄²⁻ ratios of 1.6 and 2.4, the mean SO₄²⁻ reduction reached approximately 70% irrespective of the experimental HRT and LA concentration. These results indicate that low COD/SO₄²⁻ ratios are favorable for high SO₄²⁻ removal.

Studies conducted by Choi and Rim [9] reported reduced SRB activity at COD/SO₄²⁻ ratios exceeding 2.7 for acetate and hydrogen electron donors. They attributed the reduced SRB activity to competition by MPBs. In similar work by El Bayoumy *et al.* [37], they concluded that SRB growing on lactate and acetate with COD/SO₄²⁻ ratios between 0.75 and 2.25 was enhanced in comparison to ratios greater than 2.25. Studies by Velasco *et al.* [36] have indicated that COD/SO₄²⁻ ratios greater than 1.5 resulted in increasing sulfide levels while at lower COD/SO₄²⁻ ratios, sulfur species such as H₂S, dissolved sulfide was produced. Higher SO₄²⁻ reduction at low COD/SO₄²⁻ ratios is likely attributed to higher SRBs growth rates under these conditions. Evidence by Erdirencelebi *et al.* [10] also support the argument that at higher COD/SO₄²⁻ ratios, SRBs are unable to compete with MPBs for electrons derived from substrate oxidation.

Figure 2. Impact of selected experimental factors on percent sulfate reduction. (a) Effect of COD/SO₄²⁻ ratio; (b) Effect of HRT; (c) Effect of LA concentration.



Notes: 1. Average values are shown for the model; 2. Dashed line (-----) indicates the mean value of the percent sulfate reduction; 3. HRT = hydraulic retention time; LA = linoleic acid.

Effect of Hydraulic Retention Time

Assessing the impact of HRT on SO₄²⁻ removal was conducted after optimizing the COD/SO₄²⁻ ratio. According to Neculita *et al.* [38], increased treatment efficiency was observed with increasing HRT. Reduced treatment efficiency with decreasing HRT at a constant COD/SO₄²⁻ ratio was reported

by Zhou *et al.* [39]. These authors reported decreasing SO_4^{2-} removal efficiencies from 89% to 82% as the HRT was decreased from 24 h to 12 h at a constant COD/ SO_4^{2-} ratio of 4. In this study, a mean experimental response of 84% SO_4^{2-} reduction was observed at a 12 h HRT (Figure 2b).

Lower mean percent SO_4^{2-} removals of 69% and 68% observed in cultures operating at 24 h and 36 h HRT indicated that long HRT conditions are unfavorable for SO_4^{2-} reduction. The low SO_4^{2-} removals might be due to the ability of MPBs competing with SRB for the available substrate. MPBs are able to compete with SRBs for substrate derived electrons and subsequently produce CH_4 . Higher SO_4^{2-} removal at lower HRT (12 h) is likely associated with elevated growth rates of SRB in comparison to MPBs [37]. MPB have longer doubling times and are washed out at lower HRT thus favoring SO_4^{2-} reduction [16].

Effect of Linoleic Acid Concentration

A mean SO_4^{2-} removal (experimental response) of 69% was observed in control cultures (not fed LA). Cultures fed 0.5 g L^{-1} LA performed the same as the controls with mean SO_4^{2-} removal reaching approximately 68% (Figure 2c). However, with 1 g L^{-1} LA, the SO_4^{2-} removal was more effective with a 22% increase. This result indicates that adding 1 g L^{-1} LA was effective in selectively inhibiting MPBs and re-directing the substrate derived electrons to SRBs. In work conducted by Ray *et al.* [40] and Chowdhury *et al.* [41], they indicated that methanogenic inhibition by a threshold LA level lead to H_2 production. In comparison, Moon *et al.* [7] concluded no significant difference in SO_4^{2-} reduction was detected at low (0.5 g L^{-1}) and high (1.5 g L^{-1}) LA levels using glucose fed batch cultures maintained at pH 6.0 to 7.5 and COD/ SO_4^{2-} ratios varying from 0.5 to 2.5. This difference in comparison to the work reported herein could be due to no pH control in the batch studies. Additionally, in comparison to the work reported by Moon *et al.* [7], variation in HRT might have exerted a significant effect at varying LA levels.

3.3.2. Surface Plots

Interaction between the experimental factors is depicted using the surface plot (Figure 3). The effects of COD/ SO_4^{2-} ratio and HRT (Figure 3a) suggest that maximum SO_4^{2-} reduction (>80%) was observed at a COD/ SO_4^{2-} ratio of 0.8 and a 12 h HRT. Similar trends were observed with a low COD/ SO_4^{2-} ratio of 0.8 and an elevated LA concentration of 1 g L^{-1} (Figure 3b). The effect of HRT and LA concentration on SO_4^{2-} removal is shown in Figure 3c. Lower HRT (12 h) and higher LA concentration (1 g L^{-1}) resulted in maximum SO_4^{2-} removal. In general, from these surface plots, a combination of lower HRT and COD/ SO_4^{2-} ratio together with higher LA concentration resulted in maximum SO_4^{2-} removal.

3.4. Model Verification

The response variable computed using the model correlated reasonably well with the experimental data. The R^2 value for predicted *versus* experimental SO_4^{2-} reduction (%) was 0.9592 (data not shown). The residuals (model predicted value - experimental value) for the experimental response were used to assess the adequacy of the fit. The Anderson-Darling (AD) plot confirmed a normal

distribution of the residuals. The observed AD statistic for the model response was 0.290 (Supplementary Figure A1). This value is smaller than the critical AD value of 0.752 for a sample size of 18 at a 5% significance level. The observed p -value (Supplementary Figure A1) of 0.572 (greater than 0.05) also confirms a normal distribution of residuals. This suggests that the model-predicted response values correlated reasonably well with the experimental response values (Table 2) over the factor space under consideration. The results obtained from this study are comparable with data reported in literature (Table 4). In general, the results (% SO_4^{2-} reduction) reported in literature for various reactor systems fed with different type of substrates obtained higher SO_4^{2-} reduction at higher COD/ SO_4^{2-} ratios (>2 ; Table 4).

Figure 3. Surface plots for the experimental response (% sulfate reduction) (a) COD/ SO_4^{2-} ratio versus HRT (at constant LA = 0.5 g L⁻¹); (b) COD/ SO_4^{2-} ratio versus LA (at constant HRT = 24 h); (c) HRT versus LA (at constant COD/ SO_4^{2-} ratio = 1.6).

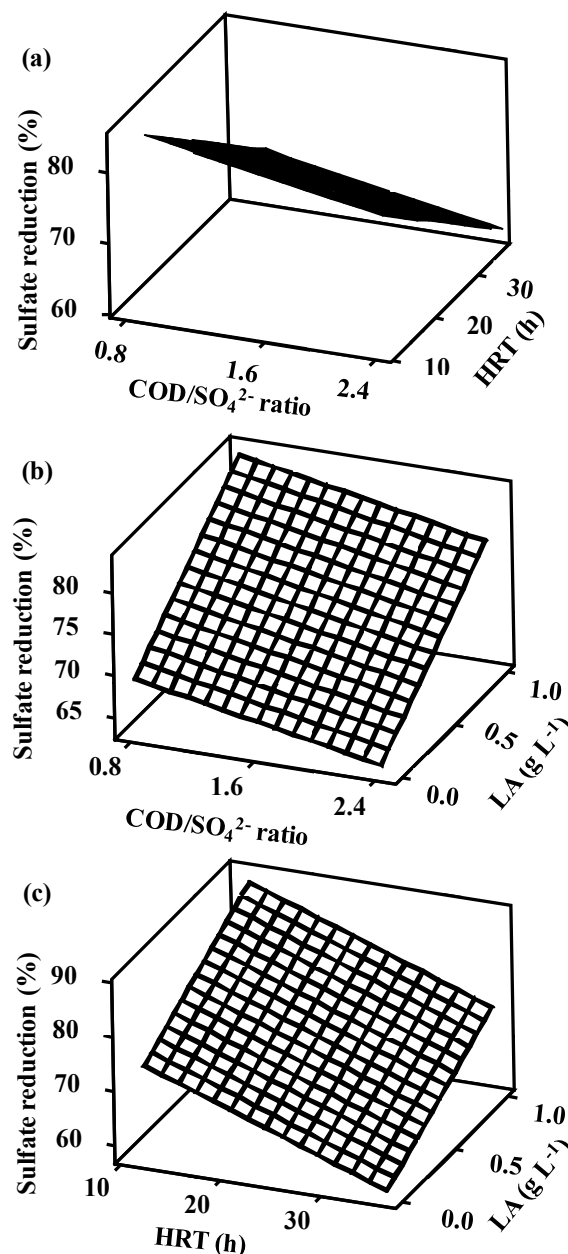


Table 4. Comparison of percent sulfate reduction under different operational conditions.

COD/SO ₄ ²⁻ Ratio	Reactor Type; Mode of Operation	Temp. (°C)	pH	SO ₄ ²⁻ Reduction (%)	Substrate	HRT	Ref.
2.5	UASBR; Continuous	30	7.0 ± 0.5	94 ± 1	Ethanol	4 d	[36]
4	FBR; Continuous	35	7.4 ± 0.2	90	Ethanol	6.5 h	[32]
3.2, 4, 5	UASBR; Continuous	30–33	7.3 ± 0.7	70, 81, 74	Glucose	24 h	[10]
3.15, 2.7	CSTR; Continuous	30	NR	29, 28	Glucose	NR	[10]
2.7, 1.23, 0.6	Serum bottle; Batch	30	NR	9, 4, 4.5	Acetate	NA	[10]
0.41, 1.03, 2.07	Serum bottle; Batch	35 ± 1	7.3 ± 0.1	26, 60, 93	Propionate	NA	[11]
1, 4	UASBR; Continuous	55	6.0	25–35, 65	Sucrose	10 h	[42]
6.67	UASBR; Continuous	35 ± 1	7.0–7.5	80–86	Sulfate rich vinasse	4.86 days	[43]
4	AFR; Continuous	37 ± 0.5	9.5	97.8 ± 1.1	Ethanol	18 h	[39]
0.8, 1.6, 2.4	ASBR; Sequencing batch	37 ± 0.1	6.5 ± 0.1	87 ± 3, 58 ± 3, 62 ± 9	Glucose	12, 36, 24 h	This study *

Notes: UASBR = Upflow anaerobic sludge blanket reactor; FBR = fluidized bed reactor; CSTR = continuous stirred tank reactor; ASBR = anaerobic sequential batch reactor, ABR = anaerobic filter reactor; NR = not reported; NA = not applicable; *: denotes % sulfate reduction in LA untreated (Control cultures).

Erdirencelebi *et al.* [10] reported maximum SO₄²⁻ reduction of 81% and 29% in UASBR and CSTR, respectively, using COD/SO₄²⁻ ratios >3 and mixed anaerobic cultures fed glucose at neutral pH 7.0. High SO₄²⁻ reduction (86.5% ± 2.6%; Table 2) in control cultures (LA unfed cultures) at a low COD/SO₄²⁻ ratio of 0.8 and a 12 h HRT indicate that higher SO₄²⁻ levels is associated with high SO₄²⁻ removal in comparison to data reported by other researchers (Table 4). In comparison, statistically the same percent SO₄²⁻ reduction (89.9% ± 6%; Table 2) was observed in the presence of 1000 mg L⁻¹ LA, a 12 h HRT and a COD/SO₄²⁻ ratio of 2.4. In the control cultures with a low COD/SO₄²⁻ ratio of 0.8 (*i.e.*, at high SO₄²⁻ concentration), low methane production (data not shown) was coupled with high SO₄²⁻ reduction. Since LA is an effective methanogenic inhibitor (at threshold levels), the LA treated cultures with high levels of SO₄²⁻ reduction at all COD/SO₄²⁻ ratios indicated that the substrate-derived electrons were utilized for SO₄²⁻ reduction rather than CH₄ formation.

3.5. Factor Interactions and Their Influence on Sulfate Reduction

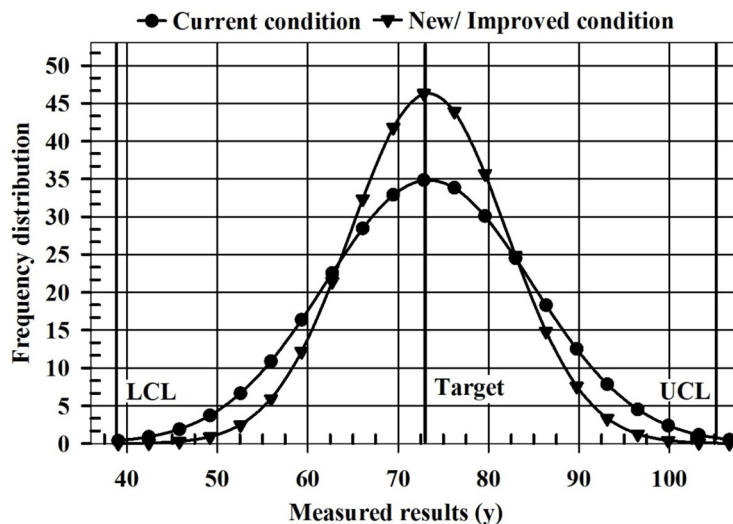
The average value of the different factors together with the interaction effects of other experimental factors at assigned factor levels on percent SO₄²⁻ reduction is shown in Supplementary Table A1. The difference among the levels (L1–L2, L1–L3, L3–L2, L2–L1, L3–L2, and L3–L1) of each factor indicates the relative influence on the response (Supplementary Table A1). A larger difference is associated with a strong influence on the response variable. The data clearly indicate that HRT showed the greatest influence (83.8%) at level 1 (12 h HRT) when compared to the other factors. The next factors were the LA concentration (level 3) and COD/SO₄²⁻ ratio (level 1; Supplementary Table A1). Notice the decreasing percent SO₄²⁻ reduction from 83.8% to 63.8% is associated with increasing HRT from 12 h to 36 h (Supplementary Table A1). Increasing the LA concentration from 0 to 1 g L⁻¹ showed an increase in SO₄²⁻ reduction from 68.8% to 82.9% (Supplementary Table A1). In comparison, minimum variation in the percent SO₄²⁻ removed was observed with varying COD/SO₄²⁻ ratios.

The Qualitek 4 software generated interaction effects were analyzed individually to examine the impact of different factors on the overall SO_4^{2-} reduction. In general, interaction effects are studied because of the possibility of one factor interacting with one or all of the other factors. The interaction severity index [SI] was calculated to determine the influence of the experimental factors at varying factor levels (Supplementary Table A2). The analysis indicates that the COD/ SO_4^{2-} ratio and LA concentration had the largest SI (59.33%) followed by the COD/ SO_4^{2-} ratio and HRT (37.65%; Supplementary Table A2). The SI value for the HRT and LA concentration was the lowest (24.84) among the factors investigated. Note that in the interactions, experimental variables with the least impact factor (COD/ SO_4^{2-} ratio (p% = 8.582); Table 3) were associated with a stronger impact factor (HRT (p% = 49.99) and LA concentration (p% = 41.42); Table 3). Data from this study indicate that the influence of selected factors on SO_4^{2-} reduction was independent of the individual influence.

3.6. Optimum Conditions for Sulfate Reduction

The Qualitek 4 software was used to examine the interaction effect of the experimental variables at various levels. The optimum process performance with major factor contributions is shown in Supplementary Table A3. Among the selected factors, HRT had the largest positive impact on SO_4^{2-} reduction. The data shows the relative interactions of the parameters on SO_4^{2-} reduction. The contribution by each individual factor is the key for enforcing control over SO_4^{2-} reduction. The expected improvement on SO_4^{2-} reduction in mixed anaerobic culture using the experimental variables is shown in Figure 4.

Figure 4. Variation reduction plot showing the performance distribution of sulfate removal under current and improved conditions. (LCL = lower control limit; UCL = upper control limit.)



The normal distribution profiles are shown for current and improved conditions assuming the optimum performance is a target. The average SO_4^{2-} removal shown is approximately 73.9% (Figure 4). The improved and current average percent SO_4^{2-} removal is the same; however, the improved condition frequency is larger when compared to the current condition frequency. A summary of the two conditions is shown in Table 5.

Table 5. Plotting parameters in the variation reduction plot.

Parameters	Current Condition	New/Improved Condition
Mean	72.49	72.49
Standard deviation	11.50	7.93
C_p	1.00	1.45
C_{pk}	1.00	1.45
Quality characteristic (QC)	Bigger is better	Bigger is better
Lower control limit (LCL)	37.99	37.99
Upper control limit (UCL)	106.99	106.99

Notes: C_p represents the capability index expressed in terms of a number (ratio) indicating the narrowness of the population distribution within the LCL and UCL; C_{pk} represents the capability index is very similar to C_p which captures the position of the mean performance as well as the variation of the data within the specification limits; LCL = Mean - (3 × standard deviation of current condition); UCL = Mean + (3 × standard deviation of current condition).

In comparison to the current condition, the standard deviation for the improved condition is smaller. The current condition is derived from the experimental response (%SO₄²⁻ reduction) while the improved condition is based on the minimization of the variation in the experimental response. The C_p and C_{pk} values are designated as capability indices [44]. C_p is a measure of the process capability with respect to the difference between the upper control limit (UCL) and lower control limit (LCL). The C_{pk} value measures the process variation with respect to the mean. A high the C_{pk} indicate the capability of the process to meet its requirements. For the improved condition case, the capability index is larger when compared to the current condition. A capability index greater than 1.33 (Table 5) indicate the percent SO₄²⁻ removals are within the tolerances (LCL and UCL). The optimum conditions for SO₄²⁻ removal was determined by the Qualitek 4 software based on the results obtained using the Taguchi OA.

The optimum conditions for maximum SO₄²⁻ removal were observed at a COD/SO₄²⁻ ratio of 0.8, a 12 h HRT together with 1 g L⁻¹ LA (Supplementary Table A3). Under the optimum conditions, the maximum SO₄²⁻ reduction attained was 97.6%. The total contribution from the experimental factors on SO₄²⁻ reduction was 24.2%. The observed 73.4% average performance of the mixed microbial cultures and 24.2% contribution from all experimental factors revealed the potential of these variables and their interaction on SO₄²⁻ reduction in the ASBRs.

4. Conclusions

The Taguchi OA was used to evaluate the percent SO₄²⁻ reduction using glucose as substrate under different experimental conditions. The factors investigated in this study included the COD/SO₄²⁻ ratio, HRT and LA concentration. In general, the percent SO₄²⁻ removed decreased with increasing COD/SO₄²⁻ ratio and HRT levels and increased with increasing LA concentration. An analysis of the residuals indicates a normal distribution. The surface plots and ANOVA indicates significant interactions between the experimental factors investigated. The Taguchi model predicted an optimum SO₄²⁻ removal of 97.6% at a COD/SO₄²⁻ ratio of 0.8 (level 1), a 12 h HRT (level 1) and 1000 mg L⁻¹ LA (level 3). The maximum SO₄²⁻ removal of 87% ± 3% was obtained at a lower feed COD/SO₄²⁻ ratio (high SO₄²⁻ loading conditions) in combination a lower HRT (12 h) in the control cultures

(without LA addition). The results obtained from this current study indicated that higher biological SO_4^{2-} reduction using anaerobic cultures could be achieved in an ASBR at high SO_4^{2-} levels.

Acknowledgments

Financial support for this work was provided by the Natural Sciences and Engineering Research Council of Canada (Grant No. 261797-2009), the Canada Research Chair program (Grant No. 950-203725) and the University of Windsor (13320).

Author Contributions

The experimental work was conducted by Rajesh Singh, Chungman Moon, Sathyanarayanan S. Veeravalli and Saravanan R. Shanmugam; the manuscript was written by Saravanan R. Shanmugam and Jerald A. Lalman; Data analysis and the model development were performed by Saravanan R. Shanmugam, Sathyanarayanan S. Veeravalli, Subba Rao Chaganti and Jerald A. Lalman.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Austin, G.T. *Shreve's Chemical Process Industries*, 1st ed.; McGraw Hill: New York, NY, USA, 1984.
2. Shin, H.S.; Sae-Eun, O.; Chae-Young, L. Influence of sulphur compounds and heavy metals on the methanisation of tannery wastewater. *Water Sci. Technol.* **1997**, *35*, 239–245.
3. Pol, L.W.H.; Lens, P.N.L.; Stams, A.J.M.; Lettinga, G. Anaerobic treatment of sulphate-rich wastewaters. *Biodegradation* **1998**, *9*, 213–224.
4. Dugan, P.R. Bacterial ecology of strip mine areas and its relationship to the production of acidic mine drainage. *Ohio J. Sci.* **1975**, *75*, 266–279.
5. Foucher, S.; Battaglia-Brunet, F.; Ignatiadis, I.; Morin, D. Treatment by sulfate-reducing bacteria of Chessy acid-mine drainage and metals recovery. *Chem. Eng. Sci.* **2001**, *56*, 1639–1645.
6. Johnson, D.B.; Hallberg, K.B. Pitfalls of passive mine water treatment. *Rev. Environ. Sci. Biotechnol.* **2002**, *1*, 335–343.
7. Moon, C.; Singh, R.; Chaganti, S.R.; Lalman, J.A. Modeling sulfate removal by inhibited mesophilic mixed anaerobic communities using a statistical approach. *Water Res.* **2013**, *47*, 2341–2351.
8. Thauer, R.K.; Jungermann, K.; Decker, K. Energy conservation in chemotrophic anaerobic bacteria. *Bacteriol. Rev.* **1977**, *41*, 100–180.
9. Choi, E.; Rim, J.M. Competition and inhibition of sulfate reducers and methane producers in anaerobic treatment. *Water Sci. Technol.* **1991**, *23*, 1259–1264.
10. Erdirencelebi, D.; Ozturk, I.; Cokgor, E.U.; Tonuk, G.U. Degree of sulfate-reducing activities on COD removal in various reactor configurations in anaerobic glucose and acetate-fed reactors. *Clean Soil Air Water* **2007**, *35*, 178–182.

11. Uberoi, V.; Bhattacharya, S.K. Interactions among sulfate reducers, acetogens, and methanogens in anaerobic propionate systems. *Water Environ. Res.* **1995**, *67*, 330–339.
12. Omil, F.; Bakker, C.D.; Pol, L.W.H.; Lettinga, G. Effect of pH and low temperature shocks on the competition between sulphate reducing bacteria and methane producing bacteria in UASB reactors. *Environ. Technol.* **1997**, *18*, 255–264.
13. Hao, O.J.; Chen, J.M.; Huang, L.; Buglass, R.L. Sulfate-reducing bacteria. *Crit. Rev. Environ. Sci. Technol.* **1996**, *26*, 155–187.
14. Okabe, S.; Nielsen, P.H.; Jones, W.L.; Characklis, W.G. Sulfide product inhibition of *Desulfovibrio desulfuricans* in batch and continuous cultures. *Water Res.* **1995**, *29*, 571–578.
15. Widdel, F.; Pfennig, N. Sporulation and further nutritional characteristics of *Desulfotomaculum acetoxidans*. *Arch. Microbiol.* **1981**, *129*, 401–402.
16. Smith, M.R.; Mah, R.A. Growth and methanogenesis by *Methanosarcina* Strain 227 on acetate and methanol. *Appl. Environ. Microbiol.* **1978**, *36*, 870–879.
17. Oh, S.E.; van Ginkel, S.; Logan, B.E. The relative effectiveness of pH control and heat treatment for enhancing biohydrogen gas production. *Environ. Sci. Technol.* **2003**, *37*, 5186–5190.
18. Shanmugam, S.R.; Chaganti, S.R.; Lalman, J.A.; Heath, D.D. Effect of inhibitors on hydrogen consumption and microbial population dynamics in mixed anaerobic cultures. *Int. J. Hydrogen Energy* **2014**, *39*, 249–257.
19. Galbraith, H.; Miller, T.B. Physicochemical effects of long-chain fatty acids on bacterial cells and their protoplasts. *J. Appl. Bacteriol.* **1973**, *36*, 647–658.
20. Lalman, J.; Bagley, D.M. Effects of C18 long chain fatty acids on glucose, butyrate and hydrogen degradation. *Water Res.* **2002**, *36*, 3307–3313.
21. Rao, R.S.; Kumar, C.G.; Prakasham, R.S.; Hobbs, P.J. The Taguchi methodology as a statistical tool for biotechnological applications: A critical appraisal. *Biotechnol. J.* **2008**, *3*, 510–523.
22. Myer, R.H.; Montgomery, D.C. *Response Surface Methodology: Process and Product Optimization Using Designed Experiment*, 2nd ed.; John Wiley and Sons: New York, NY, USA, 2002.
23. Prakasham, R.S.; Rao, C.S.; Rao, R.S.; Rajesham, S.; Sarma, P.N. Optimization of alkaline protease production by *Bacillus* sp. using Taguchi methodology. *Appl. Biochem. Biotechnol.* **2005**, *120*, 133–144.
24. Prakasham, R.S.; Rao, C.S.; Rao, R.S.; Sarma, P.N. Enhancement of acid amylase production by an isolated *Aspergillus awamori*. *J. Appl. Microbiol.* **2007**, *102*, 204–211.
25. Antony, F.; Warwood, S.; Fernandes, K.; Rowlands, H. Process optimization using Taguchi methods of experimental design. *Work Study* **2001**, *50*, 51–57.
26. Roy, R.K. *Design of Experiments Using Taguchi Approach: 16 Steps to Product and Process Improvement*; John Wiley and Sons: New York, NY, USA, 2001.
27. Ray, S.; Chowdhury, N.; Lalman, J.A.; Seth, R.; Biswas, N. Impact of initial pH and linoleic acid (C18:2) on hydrogen production by a mesophilic anaerobic mixed culture. *J. Environ. Eng. ASCE* **2008**, *134*, 110–117.
28. Wiegant, W.M.; Lettinga, G. Thermophilic anaerobic digestion of sugars in upflow anaerobic sludge blanket reactors. *Biotechnol. Bioeng.* **1985**, *27*, 1603–1607.
29. Shizas, L.; Bagley, D.M. Improving anaerobic sequencing batch reactor performance by modifying operational parameters. *Water Res.* **2002**, *36*, 363–367.

30. Speece, R.E. Gas Flow Totalizer. U.S. Patent 4064750 A, 27 December 1977.
31. American Publishers Health Association (APHA). *Standard Methods for the Examination of Water and Wastewater*, 20th ed.; APHA: Washington, DC, USA, 1999.
32. Kaksonen, A.H.; Franzmann, P.D.; Puhakka, J.A. Effects of hydraulic retention time and sulfide toxicity on ethanol and acetate oxidation in sulfate-reducing metal-precipitating fluidized-bed reactor. *Biotechnol. Bioeng.* **2004**, *86*, 332–343.
33. Wang, J.X.; Roush, M.L. *What Every Engineer Should Know about Risk Engineering and Management*; Marcel Dekker, Inc.: New York, NY, USA, 2000.
34. Evangelaras, H.; Koukouvinos, C.; Lappas, E. An efficient algorithm for the identification of isomorphic orthogonal arrays. *J. Discrete Math. Sci. Cryptogr.* **2006**, *9*, 125–132.
35. Phadke, M.S.; Dehnad, K. Optimization of product and process design for quality and cost. *Qual. Reliab. Eng. Int.* **1988**, *4*, 105–112.
36. Velasco, A.; Ramirez, M.; Volke-Sepulveda, T.; Gonzalez-Sanchez, A.; Revah, S. Evaluation of feed COD/sulfate ratio as a control criterion for the biological hydrogen sulfide production and lead precipitation. *J. Hazard. Mater.* **2008**, *151*, 407–413.
37. El Bayoumy, M.A.; Bewtra, J.K.; Ali, H.I.; Biswas, N. Sulfide production by sulfate reducing bacteria with lactate as feed in an upflow anaerobic fixed film reactor. *Water Air Soil Poll.* **1999**, *112*, 67–84.
38. Neculita, C.M.; Zagury, G.J.; Bussiere, B. Effectiveness of sulfate-reducing passive bioreactors for treating highly contaminated acid mine drainage: I. Effect of hydraulic retention time. *Appl. Geochem.* **2008**, *23*, 3442–3451.
39. Ray, S.; Saady, N.M.C.; Lalman, J.A. Diverting electron fluxes to hydrogen in mixed anaerobic communities fed with glucose and unsaturated C18 long chain fatty acids. *J. Environ. Eng. ASCE* **2010**, *136*, 568–575.
40. Chowdhury, N.; Lalman, J.A.; Seth, R.; Ndegwa, P. Biohydrogen production by mesophilic anaerobic fermentation of glucose in the presence of linoleic acid. *J. Environ. Eng.* **2007**, *133*, 1145–1152.
41. Lopes, S.I.C.; Wang, X.; Capela, M.I.; Lens, P.N.L. Effect of COD/SO₄²⁻ ratio and sulfide on thermophilic (55 °C) sulfate reduction during the acidification of sucrose at pH 6. *Water Res.* **2007**, *41*, 2379–2392.
42. Barrera, E.L.; Spanjers, H.; Romero, O.; Rosa, E.; Dewulf, J. Characterization of the sulfate reduction process in the anaerobic digestion of a very high strength and sulfate rich vinasse. *Chem. Eng. J.* **2014**, *248*, 383–393.
43. Zhou, J.M.; Song, Z.Y.; Yan, D.J.; Liu, Y.L.; Yang, M.H.; Cao, H.B.; Xing, J.M. Performance of a haloalkaliphilic bioreactor and bacterial community shifts under different COD/SO₄²⁻ ratios and hydraulic retention times. *J. Hazard. Mater.* **2014**, *274*, 53–62.
44. Van Aartsengel, A.; Kurtoglu, S. *Handbook on Continuous Improvement Transformation: The Lean Six Sigma Framework and Systematic Methodology for Implementation*; Springer: New York, NY, USA, 2013.