

Article

Uptake and Effects of *Cylindrospermopsis*: Biochemical, Physiological and Biometric Responses in The Submerged Macrophyte *Egeria densa* Planch

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Received: 21 September 2020; Accepted: 22 October 2020; Published: 26 October 2020



Abstract: *Cylindrospermopsis* (CYN) is being detected in surface waters more commonly and frequently worldwide. This stable, extracellular cyanotoxin causes protein synthesis inhibition, thus posing a risk to aquatic biota, including macrophytes, which serve as primary producers. Nevertheless, data regarding the effects caused by environmental concentrations of CYN is still limited. In the presented study, the uptake of CYN at environmental concentrations by the submerged macrophyte *Egeria densa* was investigated. Bioaccumulation, changes in the plant biomass, as well as shoot-length were assessed as responses. Variations in the cellular H₂O₂ levels, antioxidative enzyme activities, as well as concentrations and ratios of the photosynthetic pigments were also measured. *E. densa* removed 54% of CYN within 24 h and up to 68% after 336 h; however, CYN was not bioaccumulated. The antioxidative enzyme system was activated by CYN exposure. Pigment concentrations decreased with exposure but normalized after 168 h. The chlorophyll *a* to *b* ratio increased but normalized quickly thereafter. Carotenoids and the ratio of carotenoids to total chlorophylls increased after 96 h suggesting participation in the antioxidative system. Growth stimulation was observed. The ability to remove CYN and resistance to CYN toxicity within 14 days proved *E. densa* as suitable for phytoremediation; nonetheless, prolonged exposure (32 days) resulted in adverse effects related to CYN uptake, which needs to be studied further.

Keywords: *cylindrospermopsis*; *Egeria densa*; uptake; oxidative stress; pigment contents; phytoremediation

1. Introduction

The cyanobacterial toxin *cylindrospermopsis* (CYN) has been recognized as an environmental hazard and has been detected in surface waters globally [1]. To date, known species of CYN producing cyanobacteria belonging to Nostocales and Oscillatoriales, specifically *Cylindrospermopsis raciborskii*, *Raphidiopsis curvata*, *Aphanizomenon flos-aquae*, *Anabaena* spp., *Umezakia natans*, and *Lynngbya* spp. [2]. CYN concentrations of up to 92 µg/L have been reported in surface waters containing a bloom, which far exceed the guideline value of 1 µg/L for safe consumption [1]. Considering the widespread distribution of CYN producers and the ever-increasing frequency of occurrence and prolonged blooms of CYN producing cyanobacteria, the toxin poses a likely risk to both humans and the balance of aquatic ecosystems [1,3]. Risks to human health may be presented when

cyanobacterial harmful algal blooms occur in drinking and recreational water sources [1] or consuming toxin-laden tissues [3]. CYN is an alkaloid with zwitterionic properties comprising of one tricyclic guanidine attached to a hydroxymethyluracil [4], with two natural structural variants currently known: 7-epi-cylindrospermopsin (7-epi-CYN) and 7-deoxycylindrospermopsin (7-deoxy-CYN) [5,6]. De la Cruz et al. [1] showed the structure of CYN and its variants as seen below (Figure 1):

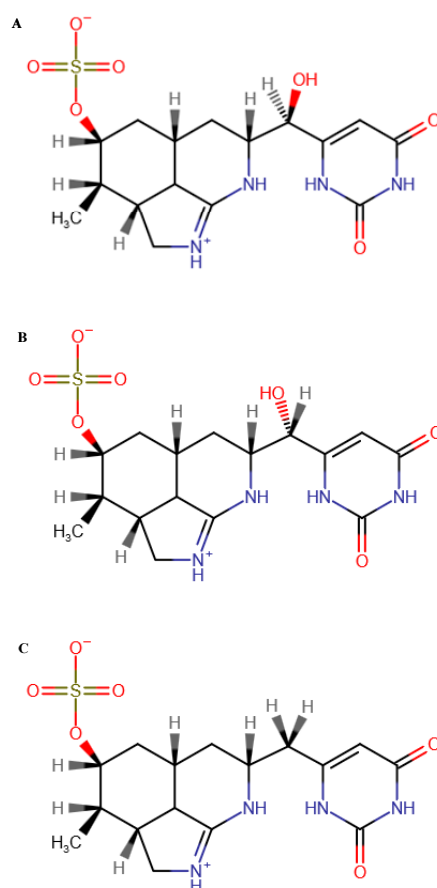


Figure 1. (A) Structure of cylindrospermopsin (CYN) and its analogues: (B) 7-epi-cylindrospermopsin (7-epi-CYN) and (C) 7-deoxycylindrospermopsin (7-deoxy-CYN). Redrawn from De la Cruz et al. [1] using Chemspace (2018).

CYN is highly water-soluble due to the hydroxymethyluracil [7]; is stable under a varied range of light, heat, and pH conditions [8]; and is often present in surface waters extracellularly [9]. Together with its capability to covalently bind to DNA/RNA [10] and hinder protein synthesis, CYN could potentially affect countless aquatic plant and animal species [3,8]. Studies focusing on CYN uptake by aquatic plants, biochemical and physiological changes, or consequences on growth linked to CYN exposure are scarce. Due to the role of macrophytes as the primary producers in oligotrophic ecosystems supporting the dependent higher trophic levels [11] and their participation in nutrient cycling control, it is pertinent to investigate the effects of CYN on macrophytes to understand the comprehensive effect on the ecosystem.

Some studies investigating the adaptation of CYN in aquatic plants have reported negligible uptake related to low bioconcentration factors (BCF) after exposures to high CYN concentrations, far exceeding those recorded in nature [12–14]. For *Hydrilla verticillata* (BFC 0.045), White et al. [12] reported 15 ng CYN/g fresh weight (FW) after seven days of exposure to 400 µg/L CYN. *Landoltia punctata* (*Spirodella oligorrhiza* previously) exposed to crude extracts of *C. raciborskii* contained 30 ng CYN/g FW plant biomass after six days of exposure to up to 500 µg/L CYN [13]. In *Azolla filiculoides*, exposed to

crude cyanobacterial extracts containing 50, 500, and 5000 µg/L CYN for seven days, 1.31 ± 0.11 µg CYN/g plant material FW was taken up resulting in a BCF of 0.40 ± 0.04 , which could be quantifiable in the macrophytes exposed to the maximum CYN concentration exclusively [14]. A recent study with *L. minor* exposed to 25 µg/L of pure CYN for 168 h [15], reported the detection of free CYN in the plant tissues ranging from 0.82 ± 0.03 ng CYN/g FW to 0.85 ± 0.01 ng CYN/g FW and highest BCF calculated was 0.095 ± 0.03 after 24 h. The same study also found that soluble CYN in the exposed plant biomass represented only up to 0.30% of CYN removed from the surroundings.

Some studies on plants exposed to CYN have reported oxidative stress promotion, i.e., the increase of glutathione peroxidase (GPx) and glutathione S-transferase (GST) activities to counteract reactive oxygen species (ROS). Rice plants (*Oryza sativa*) exposed to 2.5 µg/L CYN showed increased GST and GPx enzyme activities in the plant roots and leaves after nine days [16]. In *A. filiculoides* exposed to 5 µg/L CYN, the GST activity also increased after seven days [14]. Common lettuce *Lactuca sativa* [17] exposed to 1 µg/L, 10 µg/L, and 100 µg/L CYN mainly showed increases in the GST and GPx activities in roots at all exposure concentrations after five days and ten days. In *L. minor* exposed to CYN, the catalase (CAT) activity increased with exposure to 2.5 and 25 µg/L CYN within 168 h [18]. In the same study, the enzyme activities of GST and glutathione reductase (GR) amplified after 24 h exposure with 25 µg/L CYN. After 168 h, the activities peaked only at the highest CYN concentration [18]. Presumably, CAT acts as the initial protective response in controlling oxidative stress, followed by the activation of GST and GR during longer exposures to greater CYN concentrations.

On the subject of plant growth alterations, the results in previous studies vary and seem to depend on the CYN concentration, which could inhibit or stimulate growth. For *A. filiculoides*, an aquatic fern, growth was inhibited by 99.8% after seven days of exposure to 5000 µg/L CYN; while it grew unaltered at lower concentrations [14]. The authors also reported increases in the photosynthetic pigment concentrations as well as protein contents. For *L. minor* and *Wolffia arrhiza* exposure to both a natural cyanobacterial fraction as well as CYN as a single pure toxin (concentrations from 10 to 20,000 µg/L) caused growth alterations, comprising decreased frond numbers and biomass after five days [19]. Root growth stimulus in *H. verticillata* exposed to 400 µg/L CYN [20], increased biomass in *Landoltia punctata* exposed to 117 g/L CYN [13] and reduced fresh weight of leaves in *O. sativa* exposed to 2.5 µg/L CYN [16] have also been linked to CYN exposure regardless of purified toxin or crude extract exposure.

Research regarding the environmental implications of CYN exposure in aquatic environments has increased [1,3,7,10]; however, the uptake, subsequent effects on a physiological and morphological scale in macrophytes exposed to CYN at concentrations detected in the environment have not been studied comprehensively. Such studies will allow a better understanding of the environmental fate of CYN and the responses of plants' defense mechanisms, to further evaluate their fitness for phytoremediation [21] and to achieve a comprehensive understanding of how the ecosystem is affected by CYN.

The submerged macrophyte *Egeria densa* native from South America is widely distributed around the world [22]. It was selected for this study due to its favorable characteristics, including rapid growth and abundance in nature. The current study investigated the macrophyte's ability to assimilate CYN and the corresponding biochemical reactions in response to pure CYN uptake. The variations in the antioxidative enzyme activities, the pigment and carotenoids, in addition to the biomass and shoot length served as gauges of *E. densa*'s physiological status as a function of interaction with CYN. Thus, the study indirectly evaluated the fitness of the macrophyte for use in phytoremediation or as bio-indicator to assess the environmental impact of CYN in aquatic environments.

2. Materials and Methods

2.1. Plant Material

E. densa shoots were provided by Extraplant (Extra-group GmbH, Münster, Germany). The macrophyte *E. densa* was cultured in modified Provasoli's medium according to Flores-Rojas et al. [15]

in pre-cultivated glass tanks (60 cm × 30 cm × 30 cm) at 25 ± 1 °C under a day to night cycle of 14 h light to 10 h darkness using under cool, white fluorescent light at an irradiance of 38 E/m²/s [23]. The same cultivation parameters were applied during the treatments. The plants were pre-acclimated in the exposure containers for seven days prior to the start of the study.

2.2. Chemicals

CYN (purity > 95%) was acquired from Alexis Biochemicals (Lausen, Switzerland) as a pure toxin in powder form, and a stock solution was prepared in 70% methanol. All chemicals were analytical grade and acquired from Sigma-Aldrich, Inc. (Darmstadt, Germany) unless stated otherwise.

2.3. Experimental Design

CYN was diluted in the Provasoli's medium to four final exposure concentrations (0.025, 0.25, 2.5, and 25 µg/L). These concentrations were based on CYN concentrations previously detected in aquatic environments [1,24,25]. All experiments were conducted with non-axenic plants under non-sterile conditions. Per exposure treatment, *E. densa* shoots amounting to a total FW of 3.5 ± 0.5 g were individually subjected to each of the four specified CYN concentrations (150 mL) under the controlled culture conditions described above for a period of 336 h. The treatments were set up in parallel with their corresponding negative controls, which consisted of the exposure medium without CYN in replicates of four.

Due to the limit of quantification (10 pg on column; see Section 2.4) of the quantitative method, the uptake of CYN with time could only be studied with the highest exposure concentration (25 µg/L CYN). From the treatment and control sets, samples were collected after specific time points (4, 8, 24, 48, 96, 168, and 336 h) to quantify the CYN remaining in the media employing liquid chromatography-tandem mass spectrometry (LC-MS/MS; Section 2.4). As a means to assess the spontaneous CYN degradation, an additional positive control, which consisted of 25 µg/L CYN medium lacking plants, ($n = 4$) was run in parallel.

The set-up was duplicated in order to assess the cellular H₂O₂ concentration and the enzymatic activity after 24 h, with all four exposure concentrations and after 4, 8, 48, 96, 168, and 336 h with concentrations of 2.5 µg/L and 25 µg/L CYN ($n = 5$). The analyses of the H₂O₂ level and antioxidative stress enzyme activities were accomplished as described in Sections 2.5 and 2.6, respectively. The photosynthetic pigment contents were measured after 24 h with exposure of *E. densa* to all four CYN concentrations and after 48, 96, 168, and 336 h with 2.5 µg/L and 25 µg/L CYN ($n = 5$) as described in Section 2.7. Following exposure, the plants were rinsed with water (de-ionized) twice to eliminate any externally adhering toxin. The samples were shock-frozen and stored (−80 °C) until further analysis. Plant growth experiment was carried out with all four CYN concentrations for 32 days. Growth parameters were measured and analyzed, as described in Section 2.8.

2.4. Analysis of CYN

Soluble CYN was isolated from the plant biomass as described by Esterhuizen-Londt et al. [26] with minor modifications. The frozen plant samples were powdered while frozen and lyophilized (LIO-5P freeze-dryer Kambič Laboratorijska oprema doo, Semič, Slovenija) for 18 h (−50.3 °C; 6.1 mbar). The freeze-dried samples (500 mg) were further processed utilizing a TissueLyser LT (Qiagen, Hilden, Germany), then dissolved in 95% acetonitrile (ACN), and treated for half an hour in an ultrasonic water bath (Alpax, GmbH & Co. KG). Using an overhead Intelli-Mixer RM-2 (Neolab, Heidelberg, Germany), the samples were mixed for half an hour at ambient temperature in darkness, and after that centrifugation (20,800× *g*, 15 min, 4 °C; Eppendorf Centrifuge 5417 R, Hamburg, Germany). ACN (95%) was used to resuspend the resulting pellets, which were again centrifuged. The CYN in the pooled extracts, as well as the exposure media, were quantified via LC-MS/MS.

Matrix partition was accomplished on a Kinetex HILIC column (2.6 µm, 2.1 × 100 mm) via liquid chromatography (1200 infinity Series, Agilent, Waldbronn, Germany) coupled to triple quadrupole

mass spectrometry (model 6460 Triple Q™, Agilent, Waldbronn, Germany) with electrospray ionization (Jet-Stream, Agilent) as shown by Esterhuizen-Londt et al. [26]. The temperature of the column heater was 35 °C. A sample volume of 10 µL was injected, which was analyzed on the system at a flow rate of 0.5 mL/min. A gradient was used to separate the matrix, starting with 95% ACN (MS grade) for 3 min, which decreased to 50% for 4 min and re-equilibrated for 3 min. Using this program, CYN had a retention time of 4.1 min. The MS-MS parameters were set up using MRM mode (positive mode) with a mass transfer of 416 (Q1) to 176 and 194 (Q3) for CYN. The drying gas temperature and flow were 320 °C and 12 L/min, respectively, and the sheath gas temperature and flow were 380 °C and 12 L/min using nitrogen gas. The capillary and nozzle voltage was 4500 V and 1200 V, respectively, with a 25 psi nebulizer pressure. The plots used for calibration were linear ($R^2 = 0.998$) between 0.01 and 100 µg/L. The limit of quantification was set at 10 pg on column ($S/N \geq 5$).

CYN concentrations in exposure media and free CYN in plant tissue quantified via LC-MS/MS were used to calculate the BCFs, according to Equation (1) [27] as:

$$BCF = C_B/C_{WD}, \quad (1)$$

where C_B is the chemical concentration in the organism (ng/g) and C_{WD} is the freely dissolved chemical concentration in the water (ng/mL).

2.5. Hydrogen Peroxide (H_2O_2) Determination

Cellular H_2O_2 was determined colorimetrically following the methods of Jana and Choudhuri [28]. First, 0.3 g FW of macerated plant biomass was mixed with 3 mL of 50 mM pH 7 sodium phosphate buffer, following a centrifugation step (20,800× g, 15 min, 4 °C). The supernatant (750 µL) was combined with 250 µL of 0.1% titanium chloride in 20% H_2SO_4 (v/v). Following a short incubation at ambient temperature, the absorption was measured photometrically at 410 nm (extinction coefficient $\epsilon = 0.28$ L/mmol·cm) and expressed as µmol/g FW.

2.6. Enzyme Extraction and Activity Measurement

The enzyme preparation was completed following the procedure published by Pflugmacher [29]. The frozen plant material (1.5 g FW) was macerated in liquid nitrogen. The powder was suspended in 0.1 M sodium phosphate buffer (pH 6.5) containing 20% glycerol, 1 mM ethylenediaminetetraacetic acid (EDTA), and 1.4 mM dithioerythritol (DTE). Cellular remains were removed by centrifugation at 10,600× g for 10 min at 4 °C (Eppendorf Centrifuge 5417 R, Hamburg, Germany). Dissolved proteins were precipitated by ammonium sulfate at 80% saturation, followed by centrifugation (20,800× g, 60 min, 4 °C). The resultant pellet was liquefied in 20 mM sodium phosphate buffer (pH 7.0) and purified using Sephadex G-25 columns (NAP-columns, Amersham GE Healthcare, Uppsala, Sweden). The protein concentration was quantified at 595 nm after applying Bradford reagent [30]. The protein extracts were kept at −80 °C until further processing.

CAT (EC 1.11.1.6) enzyme activity was evaluated as described by Claiborne [31], which assesses the decrease of H_2O_2 as a decreased absorption at 240 nm ($\epsilon = 0.0361$ L/mmol·cm). Peroxidase (POD; EC 1.11.1.7) was measured as outlined by Pütter [32]. The substrate guaiacol was reduced by H_2O_2 to octahydrotetraguaiacol, which was measured at 436 nm ($\epsilon = 25.5$ L/mmol·cm). The activity of GST (EC 2.5.1.18) was assessed by measuring the conjugation of 1-chloro-2,4-dinitrobenzene (CDNB) with glutathione (GSH) at 340 nm (extinction coefficient $\epsilon = 9.6$ L/mmol·cm) as shown by Habig et al. [33]. GR (EC 1.6.4.2) activity was determined by observing the rate of nicotinamide adenine dinucleotide phosphate (NADPH) oxidation, dependent on the reduction of glutathione disulfide (GSSG) at 340 nm ($\epsilon = 6.4$ L/mmol·cm) as described by Carlberg and Mannervik [34].

2.7. Photosynthetic Pigment Contents

The chlorophyll concentration was measured according to Inskeep and Bloom [35] and the carotenoid contents according to Wellburn [36]. The frozen *E. densa* biomass was powdered in liquid nitrogen. From this powder, 0.1 g FW was dissolved in 5 mL of N,N-dimethylformamide (N,N-DMF) and kept at 4 °C in complete darkness for three days, followed by centrifugation (Eppendorf Centrifuge 5417 R, Hamburg, Germany) at 20,800× g (15 min at 4 °C). Quantification was performed spectrophotometrically at 647 nm, 664.5 nm (for chlorophyll a and b respectively), and 480 nm (for carotenoids) using cuvettes made of quartz. The chlorophyll a to b ratio (chl a/b), a bioindicator of cellular stress and effects on photosynthesis [37], was calculated by dividing the concentration of chlorophyll a by the concentration of chlorophyll b.

2.8. Plant Growth Parameters

E. densa shoots containing the apical meristems were exposed to all four concentrations of CYN in a volume of 150 mL for 32 days. Shoots were 9 ± 2 cm in length and 0.71 ± 0.1 g.

The shoots were pre-acclimated for seven days in the exposure vessels to prevent biochemical changes resulting from this preparation. Five shoots were used for each treatment and control, and both length and fresh weight of plants were measured from the second day over a period of 32 days. The increase in biomass as fresh weight and the length of the shoots were expressed as a percentage, compared to those at the start of experiments. Additionally, the number of new shoots was counted during the experiment.

2.9. Statistical Analysis

The data were statistically assessed on SPSS software (IBM SPSS Statistics, Version 20; New York, NY, USA). Normality of the data sets was tested by Shapiro–Wilk and the homogeneity by Levene's test. Statistical significance between the exposure concentrations and the corresponding controls was identified via one-way ANOVA followed by Tukey's post-hoc at a significance level (alpha) of 0.05. If the data sets failed normality and homogeneity, non-parametric Levene's test was employed. Then, significant differences between treatments and controls were established by the non-parametric Kruskal–Wallis test followed by non-parametric Post-hoc comparisons. Data sets on the concentration of CYN in exposure media and carotenoids to total chlorophyll ratio were analyzed using parametric ANOVA tests followed by Tukey's post-hoc. Data sets corresponding to the enzymatic activity of CAT and GR, contents of chlorophyll a and carotenoids were compared using both parametric (ANOVA) and non-parametric (Kruskal–Wallis) tests. For the statistical analysis of free CYN in tissue, the enzymatic activity of POD and GST, contents of chlorophyll b, total chlorophyll, and chlorophyll a to chlorophyll b ratio the non-parametric test Kruskal–Wallis were used. Growth parameters were analyzed by T-test comparing control and each treatment (toxin concentration) for each exposure time at an alpha of 0.05 (where $p < 0.05$ represents statistical significance).

3. Results

3.1. CYN Uptake

No traces of CYN contamination could be detected in the negative control plants, i.e., macrophytes, which were cultivated in Provisoli's void of CYN to test for pre-contamination. In the positive control media, 29.8% of the CYN had broken down after 24 h, which increased to 37% after 336 h (Figure 2A).

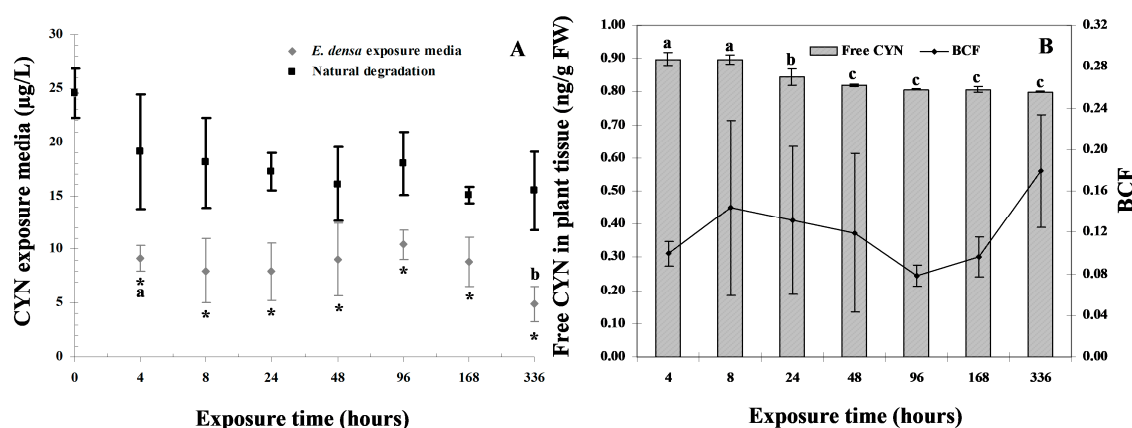


Figure 2. (A) The concentration of dissolved CYN in the exposure media in the presence (◆ *Egeria densa* in exposure media containing 25 µg/L CYN; treatment) and absence (■ Natural degradation; positive control) of *E. densa* with time. (B) CYN concentration taken up by *E. densa* with time and corresponding bioconcentration factors (BCFs). Data represent the mean value ± standard deviation ($n = 4$). Significances compared to the media control are indicated by the asterisks ($* p < 0.05$). Different letters indicate differences at the level of $p < 0.05$ of CYN in media (A) and free CYN in the plant biomass (B) by comparing the exposure times.

In the treatments, i.e., exposure media including plants, the CYN concentration was significantly reduced compared to the positive control over the entire exposure period ($p < 0.05$) (Figure 2A). The CYN concentration in the media containing plants decreased after 4 h from 24.53 ± 2.32 µg/L to 9.15 ± 1.17 µg/L equating to 62.70% CYN lost. The media CYN concentration continued to decrease within the first 24 h to 7.91 ± 2.71 µg/L ($p < 0.05$) (Figure 2A), equating to a removal percentage of 67.8%. Considering the natural degradation percentage observed, the percentage of CYN removed by *E. densa* in one day (24 h) was 54%. After 48, 96, and 168 h, the treatment media CYN concentrations remained significantly lower in relation to the control ($p < 0.05$). However, comparing 8, 24, 48, 96 and 168 h, the CYN removal percentages by *E. densa* remained unchanged ($p > 0.05$). The lowest CYN concentration in the exposure media with plants occurred after 336 h (4.94 ± 1.63 µg/L) and differed significantly with the CYN concentration at 4 h ($p < 0.05$). At the end of the experiment (after 336 h), up to 80% of the CYN was removed, of which 68% corresponded to CYN removal by *E. densa* only. The CYN removal rate after 4 h, which was 9.62 ng/min, was the highest measured during the study, followed by 5.17 ng/min after 8 h, while after 48, 96, and 168 h, further uptake was not evidenced with CYN removal rates less than 1 ng/min. Although uptake continued to occur after 336 h, the rate of CYN removal was 0.15 ng/min.

Soluble CYN was found present in the plant biomass at all time point tested ranging from 0.799 ± 0.002 ng/g FW to 0.899 ± 0.019 ng/g FW (Figure 2B). Free CYN in the plant biomass showed significant dissimilarities between the different exposure times ($p < 0.05$). After 4 and 8 h, free CYN in plant tissues were similar but differed from the plant tissues at 24 and 48 h ($p < 0.05$), which also tended to decrease. After 96, 168, and 336 h, free CYN in the plant tissues remained similar. The bioconcentration factors (BCF), which was estimated by dividing CYN in the plant tissue (ng/g FW) by CYN in the corresponding water media (ng/mL) for each exposure time, (Figure 2B) ranged from 0.079 ± 0.010 (96 h) to 0.179 ± 0.054 (336 h). However, BCF values were not found to be statistically significant ($p > 0.05$).

Considering the residual CYN in the media after the exposures with *E. densa*, the amount of free CYN detected in the whole plant biomass represented between 0.05% to 0.08% of the total amount of CYN removed from the exposure medium.

3.2. Hydrogen Peroxide (H₂O₂)

A significant increase in the intracellular H₂O₂ level after 24 h of exposure was only detected in treatments exposed to 25 µg/L CYN (Figure 3A) ($p < 0.05$); which was 1.27 times greater than the control. Prior increases of H₂O₂ were also observed after 4 h with exposure to 2.5 µg/L (1.22 times) and 25 µg/L (1.24 times) CYN, but both values did not differ between them ($p = 1$). After 96 h, only the level of H₂O₂ in the treatment with 25 µg/L CYN increased significantly (1.43 times) compared to the control ($p < 0.05$). A further significant increase of H₂O₂ occurred after 336 h only at 25 µg/L CYN, which was 1.37 times higher than the control ($p < 0.05$) (Figure 3B).

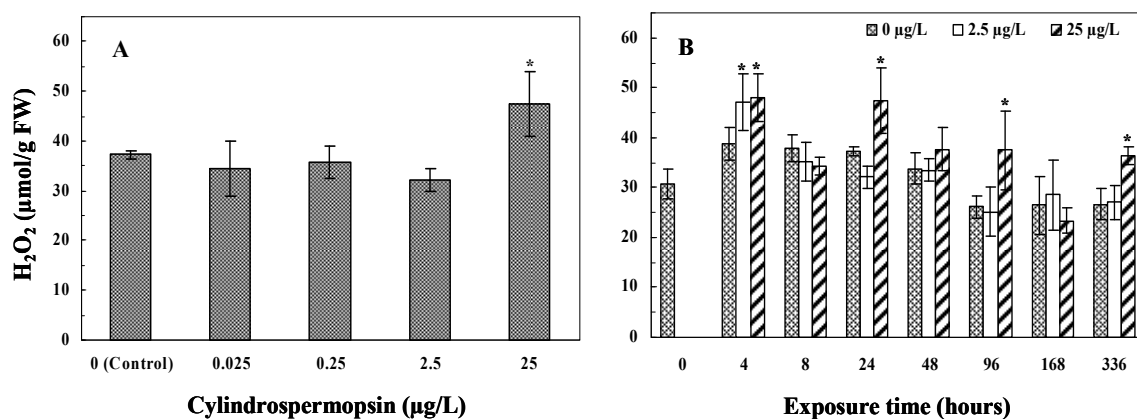


Figure 3. Levels of H₂O₂ in *E. densa* (A) with exposure to four concentrations of CYN for 24 h and (B) after exposures with time to 2.5 µg/L and 25 µg/L CYN. Data represent mean H₂O₂ concentration \pm standard deviation of five independent samples, each determined two times ($n = 5$). Significance compared to the control is shown by an asterisk (* $p < 0.05$).

3.3. Enzyme Activity

3.3.1. CAT Activity

After 24 h of exposure, the CAT activity increased significantly in the 0.25, 2.5, and 25 µg/L CYN treatments compared to control ($p < 0.05$); activities with exposure to 0.25 µg/L and 2.5 µg/L differed from 25 µg/L ($p < 0.05$). The maximum activity for CAT corresponded with the highest CYN concentration after 24 h (2.89 times) (Figure 4A). From the first hours of exposure up to the end of the experiment, CAT activity was mainly higher with exposure to the highest concentration, i.e., 25 µg/L (Figure 4B). After 4 h, CAT activity at 25 µg/L CYN was also higher than that of the corresponding control ($p < 0.05$), meanwhile after 8 h the highest activity of CAT corresponded to exposure with 2.5 µg/L CYN ($p < 0.05$). After 48 h, CAT activity at 2.5 µg/L (1.52 times) and 25 µg/L (1.58 times) diminished but remained higher than the corresponding controls ($p < 0.05$), followed by increases after 96 h ($p < 0.05$). Here CAT showed the second highest activity at 25 µg/L CYN (2.64 times). Within 168 h and 336 h, CAT activities at 2.5 µg/L (1.96 times and 1.81 times respectively) and 25 µg/L (2.45 times and 2.38 times respectively) remained higher compared to the respective control ($p < 0.05$) but tended to decrease (at 2.5 µg/L) or stayed similarly (at 25 µg/L).

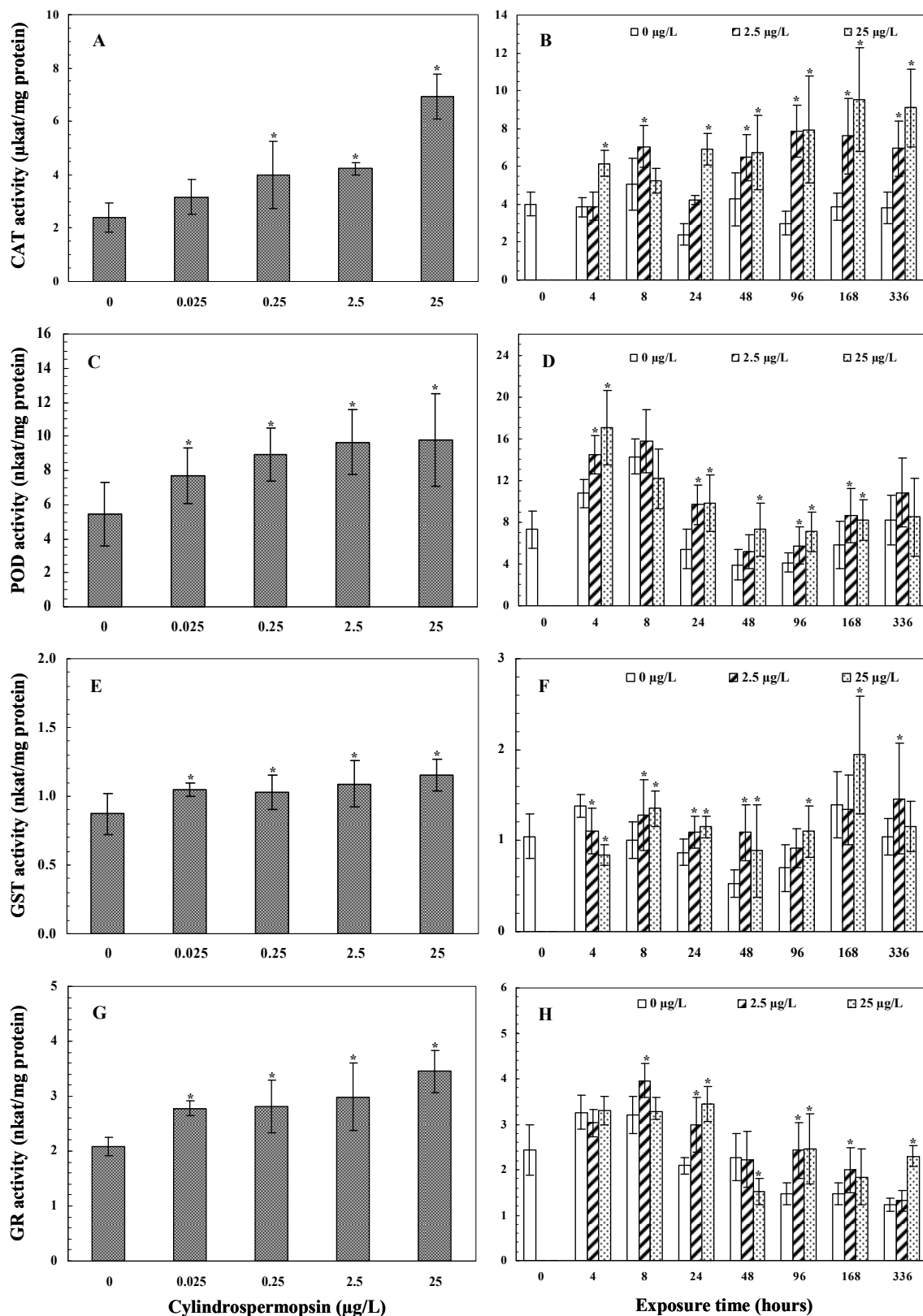


Figure 4. The (A) CAT, (C) POD, (E) GST, and (G) GR activities in *E. densa* after exposure to four concentrations of CYN for 24 h. The (B) CAT, (D) POD, (F) GST, and (H) GR activities in *E. densa* with CYN exposures at 2.5 and 25 µg/L for 336 h. Data represent the means ± standard deviation of five independent samples ($n = 5$), each determined three times. Significance compared to the control is shown by an asterisk ($* p < 0.05$).

3.3.2. POD Activity

The activity of POD showed significant increases with all four exposure concentrations after 24 h compared to the control ($p < 0.05$), but no differences were found among them ($p > 0.05$) (Figure 4C). After 4 h of exposure, POD activity increased significantly with exposure to 2.5 $\mu\text{g/L}$ (1.34 times) and 25 $\mu\text{g/L}$ (1.58 times) compared to the control ($p < 0.05$) (Figure 4D). The highest POD activity occurred after 48 h of exposure at 25 $\mu\text{g/L}$ and was 1.87-fold higher than that of the control ($p < 0.05$). Within 96 h and 168 h, POD activity showed significant differences with 2.5 $\mu\text{g/L}$ and 25 $\mu\text{g/L}$ CYN compared to the corresponding control ($p < 0.05$) but remained similar between them for both exposure times. POD activity also tended to decrease slightly in this time interval, remaining similar to the control after 336 h ($p > 0.05$).

3.3.3. GST Activity

A significant enhancement ($p < 0.05$) of GST activity was found in plants treated with all four CYN concentrations after 24 h but differences between them were not showed ($p > 0.05$) (Figure 4E). After the first 4 h of exposure, GST activity decreased significantly with 2.5 $\mu\text{g/L}$ and 25 $\mu\text{g/L}$ CYN compared to the control ($p < 0.05$) (Figure 4F). However, the GST activity increased significantly for both CYN concentrations after 8 h and until 48 h of exposure ($p < 0.05$). The maximum GST activity was reported after 48 h and corresponded to 2.5 $\mu\text{g/L}$ (2.06 times) followed by 1.68 times to 25 $\mu\text{g/L}$ at the same time exposure. After 96 and 168 h, further significant increases in the GST activity were observed only with 25 $\mu\text{g/L}$ CYN exposure, which were respectively 1.58 times and 1.39 times higher than the corresponding controls ($p < 0.05$). After 336 h, GST activity increased significantly only at 2.5 $\mu\text{g/L}$ CYN (1.40 times) compared to the corresponding control ($p < 0.05$).

3.3.4. GR Activity

The GR activity increased significantly for all four CYN concentrations after 24 h ($p < 0.05$) (Figure 4G). Only GR activities at 0.025 $\mu\text{g/L}$ and 0.25 $\mu\text{g/L}$ differed from GR activity at 25 $\mu\text{g/L}$ ($p < 0.05$). Activities of GR at 2.5 $\mu\text{g/L}$ after 8, 24, 96, and 168 h were significantly increased compared to the control ($p < 0.05$) (Figure 4H). The GR activity at 25 $\mu\text{g/L}$ showed significant enhancements ($p < 0.05$) after 24, 96, and 336 h showing the maximum GR activity (1.86 times) after 336 h. The GR activity after 48 h at 25 $\mu\text{g/L}$ CYN showed a significant decrease compared to the control ($p < 0.05$).

3.4. Photosynthetic Pigment Contents

The chlorophyll *a*, chlorophyll *b*, total chlorophyll, and carotenoid content in *E. densa* reacted similarly after 24 h with decreases observed only with 25 $\mu\text{g/L}$ CYN exposure compared to the respective control ($p < 0.05$) (Figure 5A,C,E,G). Further decreases of pigment contents with 25 $\mu\text{g/L}$ CYN concentration were seen at 48 h ($p < 0.05$) (Figure 5B,D,F,H); however, these returned to the same concentrations as the corresponding control within 96 h ($p > 0.05$). Only the carotenoids showed an increase after 96 h compared to the corresponding control ($p < 0.05$) (Figure 5H). Decreases of pigment contents were again seen after 168 h ($p < 0.05$), which then returned to the normal stated compared to the control ($p > 0.05$) until 336 h. With exposure to 2.5 $\mu\text{g/L}$ CYN, the chlorophyll *a*, chlorophyll *b*, total chlorophyll, and carotenoid contents showed significant decreases after 48 h. Chlorophyll *b* and total chlorophyll returned to the same concentrations as the control within 96 h and 336 h (Figure 5D,F). Chlorophyll *a* with 2.5 $\mu\text{g/L}$ CYN showed a further decrease after 96 h, followed by normalization within 168 h and 336 h (Figure 5B). Likewise, carotenoids decreased again ($p < 0.05$) but later (168 h), returning to the same concentration as the control after 336 h (Figure 5H).

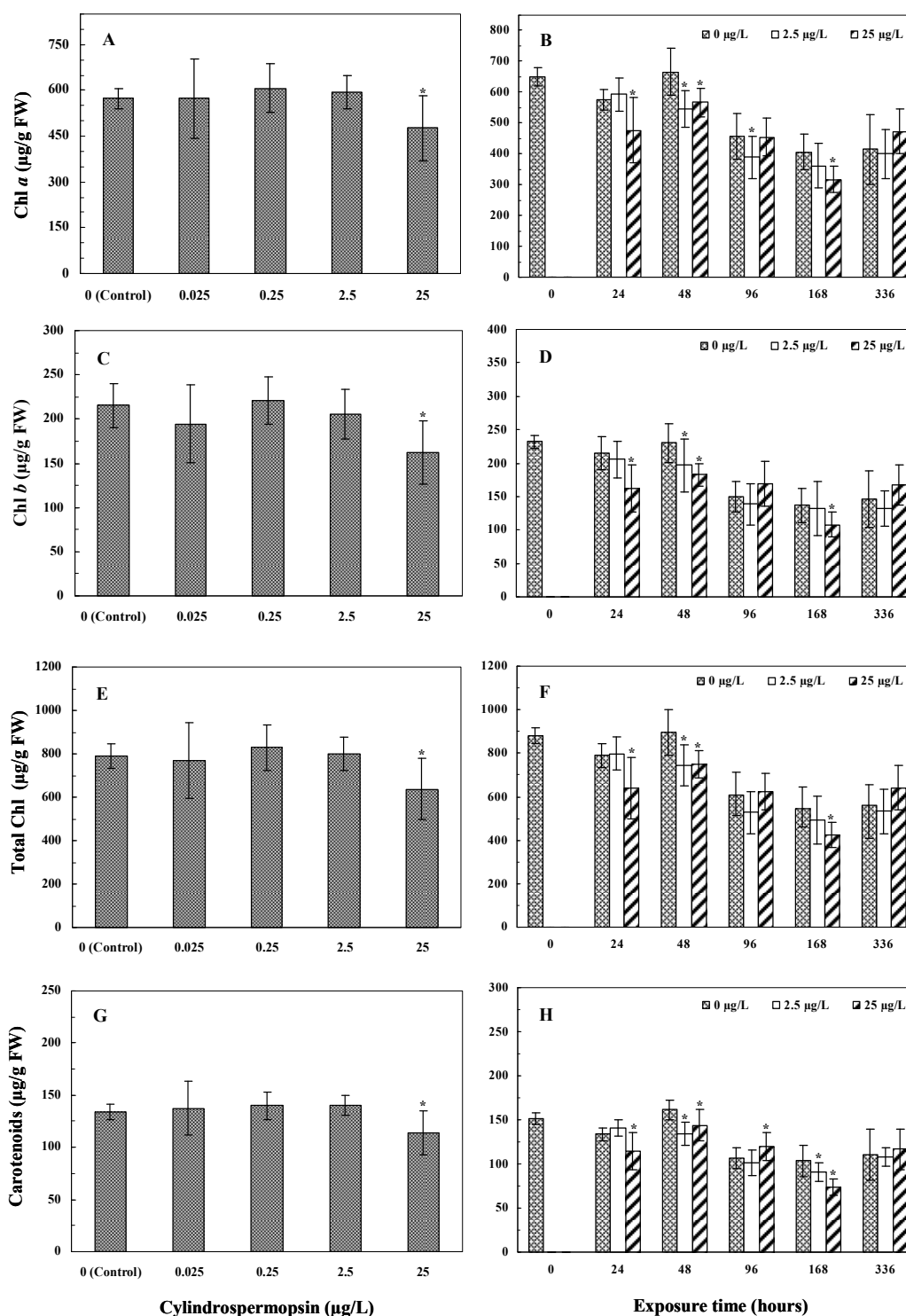


Figure 5. The (A) chlorophyll *a*, (C) chlorophyll *b*, (E) total chlorophyll, and (G) carotenoid content in *E. densa* exposed to four CYN concentrations for 24 h and (B) chlorophyll *a*, (D) chlorophyll *b*, (F) total chlorophyll, and (H) carotenoid content exposed to 2.5 µg/L and 25 µg/L CYN for 336 h. The data represent average pigment concentration ± standard deviation of five independent samples ($n = 5$), each determined three times. Significance compared to the control is shown by an asterisk ($* p < 0.05$).

The ratio of chlorophyll *a* to chlorophyll *b* increased significantly with exposure to 2.5 µg/L and 25 µg/L after 24 h ($p < 0.05$) (Figure 6A,B). After 48 h with 2.5 µg/L CYN, chlorophyll *a* to chlorophyll *b* ratio returned to similar ratios as the control, thus remaining until 336 h exposure (Figure 6A).

With 25 $\mu\text{g/L}$ CYN exposure, the chlorophyll *a* to *b* ratio continued to increase after 48 h ($p < 0.05$) and returned to the same ratios as the control within 96 h and 336 h ($p > 0.05$) (Figure 6B). The carotenoids to total chlorophyll ratio only increased significantly compared to the control after 96 h with both 2.5 $\mu\text{g/L}$ and 25 $\mu\text{g/L}$ CYN exposure ($p < 0.05$) (Figure 6C,D).

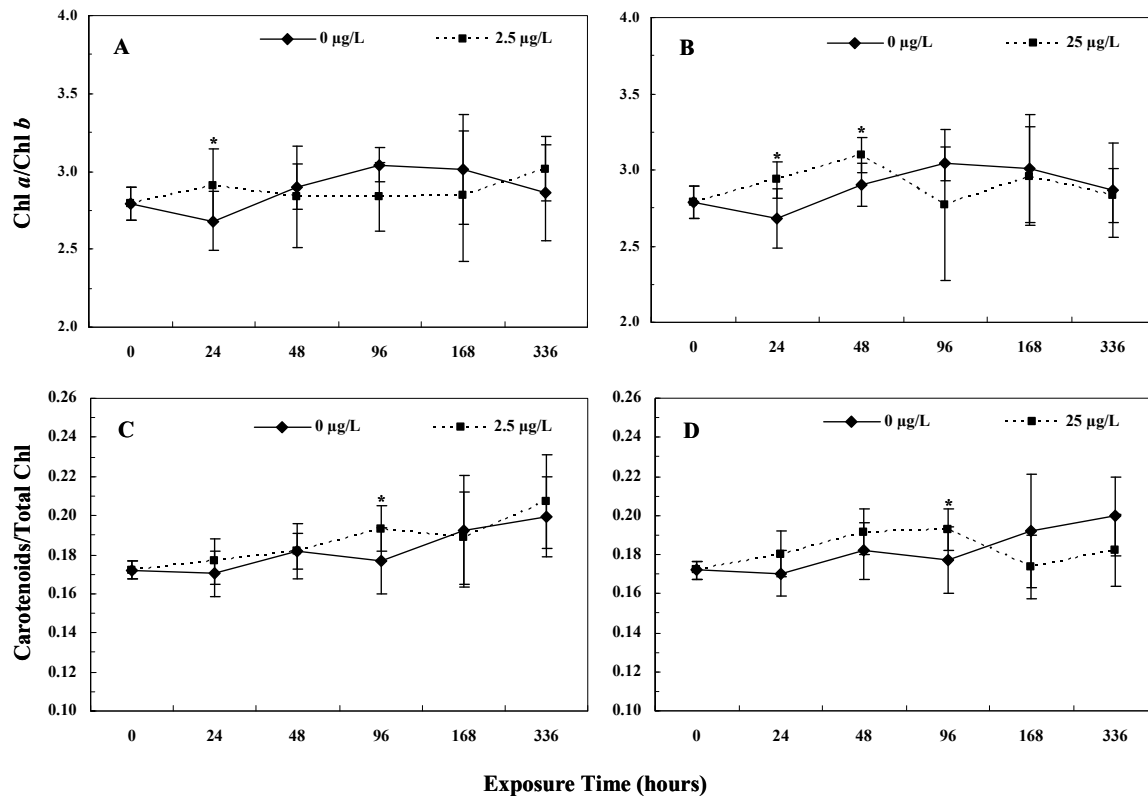


Figure 6. The ratio of chlorophyll *a* to *b* in *E. densa* exposed to (A) 2.5 $\mu\text{g/L}$ and (B) 25 $\mu\text{g/L}$ CYN as well as the ratio of carotenoids to total chlorophyll with exposure to (C) 2.5 $\mu\text{g/L}$ and (D) 25 $\mu\text{g/L}$ CYN. Significance compared to the control is shown by an asterisk ($*p < 0.05$).

3.5. Plant Growth

3.5.1. Changes in Fresh Weight

E. densa treated with CYN showed an increase in fresh weight within the first two days for all four exposure concentrations used ($p < 0.05$), of which increases with exposure to 2.5 $\mu\text{g/L}$ CYN, followed by 25 $\mu\text{g/L}$, were most prominent (Figure 7A,B,C,D). At 0.025 $\mu\text{g/L}$ CYN, plants started to show mild inhibitory effects after four days, following by 11 and 14 days ($p < 0.05$) (Figure 7A). Stagnation in the increase of fresh weight was seen after 21 days, but the decrease only was significant after 32 days ($p < 0.05$). Plants exposed to 0.25 $\mu\text{g/L}$ CYN did not show statistically significant decreases of fresh weight within 14 days ($p > 0.05$), but the decrease of fresh weight became significant within 28 and 32 days ($p < 0.05$) (Figure 7B). With exposure to 2.5 $\mu\text{g/L}$, *E. densa* showed a significant decrease in fresh weight after 32 days ($p < 0.05$) (Figure 7C). While with 25 $\mu\text{g/L}$ CYN, stagnation followed by significant decreases of fresh weight occurred after 14 days and continued until day 32 ($p < 0.05$) (Figure 7D).

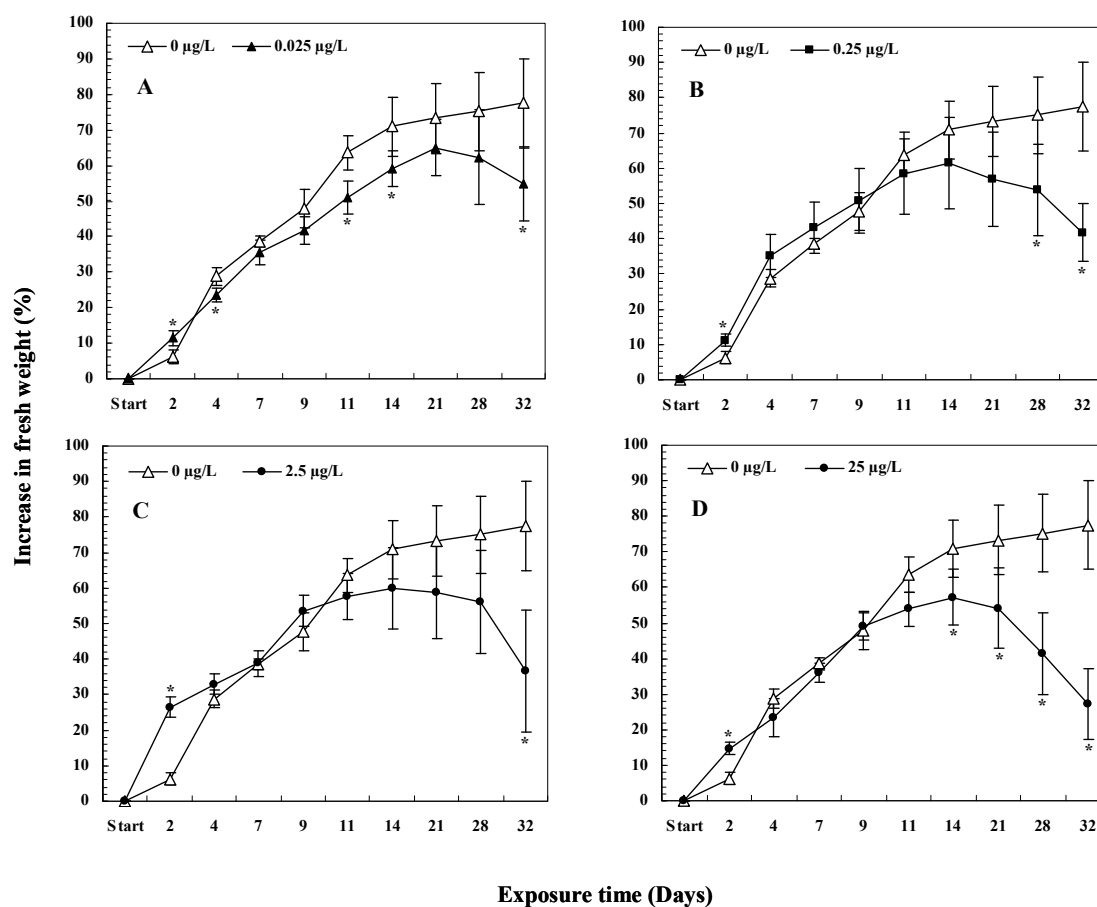


Figure 7. The growth of *E. densa* denoted as the average percentage increase of fresh weight ($n = 5$) \pm standard deviation as a function of (A) 0.025 µg/L, (B) 0.25 µg/L, (C) 2.5 µg/L, and (D) 25 µg/L CYN exposure. Significance compared to the control is shown by an asterisk ($*p < 0.05$).

3.5.2. Changes in Plant Lengths and New Shoots

E. densa shoots exposed to 0.025 µg/L and 0.25 µg/L CYN, in general, did not show changes in plant lengths (Figure 8A,B). Increases of shoot length were seen in plants exposed to 2.5 µg/L and 25 µg/L CYN (Figure 8C,D), which were significant at 2.5 µg/L within two days and seven days (Figure 8C); likewise, increases with 25 µg/L CYN exposure were seen after seven days and 11 days ($p < 0.05$) (Figure 8D). The stagnation of shoot length growth was seen after 14 days, for which the inhibitory was significant only after 14 days with 25 µg/L CYN and 32 days with both 2.5 µg/L and 25 µg/L CYN concentrations ($p < 0.05$).

The appearance of new shoots in *E. densa* differed according to the CYN concentrations. With exposure to 0.025 µg/L CYN, new shoots appeared after two days, followed by seven days, but this was less than compared to the control ($p < 0.05$) (Figure 8E). Within seven days and 14 days, the number of new shoots between days did not vary but were less compared to the control ($p < 0.05$). After 21 days, the new shoots reached a similar number compared to the control, and this remained so until day 32 ($p > 0.05$). The number of new shoots that developed albeit exposure to 0.25 µg/L was similar compared to those seen developing in the control ($p > 0.05$) with an increase only after 28 days. At 2.5 µg/L CYN, the amount of new shoots was below that observed in the controls and showed significant differences within nine days and 21 days, followed by 32 days ($p < 0.05$). The total number of shoots remained similar between day nine and day 32. At 25 µg/L, new shoots appeared later, i.e., after seven days, reaching the maximum amount after 11 days and remained unchanged until day 32. The number of new shoots during the 32-days exposure was always below that of the controls ($p < 0.05$).

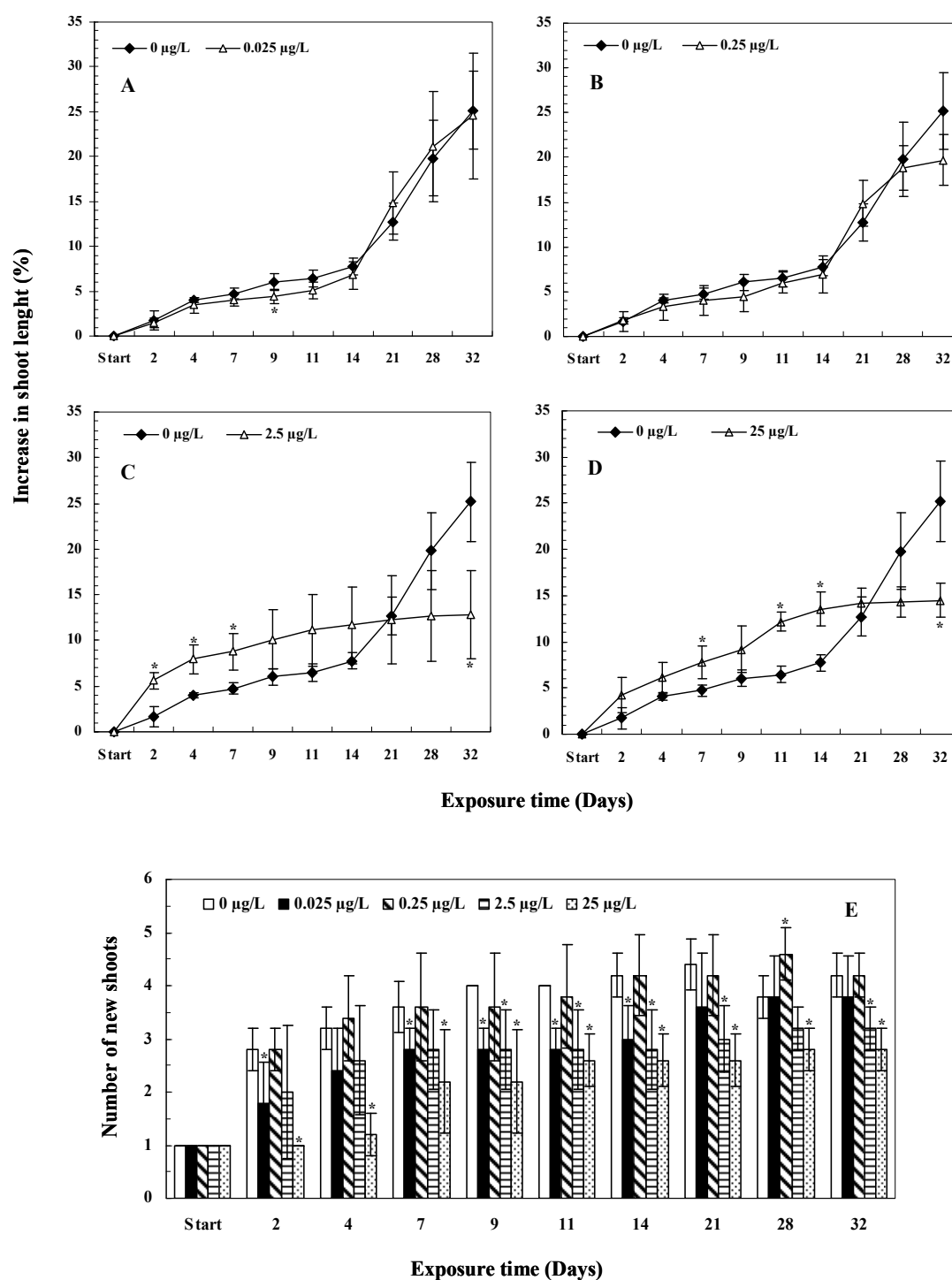


Figure 8. The growth of *E. densa* as the mean percentage increase in shoot length ($n = 5$) with exposure to (A) 0.025 µg/L, (B) 0.25 µg/L, (C) 2.5 µg/L, and (D) 25 µg/L CYN, as well as the mean number of new shoots with exposure to the four CYN concentrations (E). Significance compared to the control is shown by an asterisk (* $p < 0.05$).

4. Discussion

The lowered CYN concentrations in the exposures compared to the positive controls, which lacked plants, together with rapid removal of CYN within the first 4 h of exposure, confirmed that even though natural degradation occurred [8], *E. densa* was responsible for more than 50% of the toxin removal observed, which reached up to 65% at the end of the experiment. The results indicate that this

macrophyte is more efficient than *L. minor* [15], which reached a CYN removal percentage of 43% after 24 h of treatment with the same CYN concentration and experimental conditions [15]. The findings also suggest that *E. densa* adsorbed CYN most efficiently at the start of the exposure, concurring with previous studies on CYN uptake in macrophytes [12,15] as well as other cyanotoxins [38–41]. The rapid partial toxin removal suggests that CYN adhesion could occur on the cell wall due to its zwitterionic properties [4]. It is important to mention that after CYN uptake within the first hours, further uptake of CYN continues until 336 h. There is an interval time (48 h–168 h), in which the CYN concentrations in the exposure media remained similar, suggesting that the uptake is substantially slowed after 24 h. In this interval, changes in the metabolism of *E. densa* were promoted by CYN, tending to stabilize within 168 h as evidenced by alterations of the antioxidant enzyme activity and the concentrations of photosynthetic pigments. Whether the structure of CYN is modified or metabolized in plant cells as the plant continues to take up CYN, is an unanswered question and requires further investigations. The low amounts of free CYN detected in plant tissue, which decreased between 24 and 48 h, and then remained unchanged from 96 to 336 h, suggest that internalized CYN could be enzymatically attached, altered or metabolized within the plant cells averting detection [12]. A study which exposed different animal and plant proteins to CYN revealed that the proteins extracts and concentrations influenced the soluble CYN detectable. However, the study reported that when incubating CYN with amino acids and oxidized or reduced glutathione, all of the added toxin remained soluble proposing protein binding as a reason the CYN being undetectable previously [42]. The low BCFs indicating no bioaccumulation ($BCF < 1$) reported here agree with earlier studies related to CYN exposure at higher and similar concentrations [12–15]. Here, *E. densa* exposed to 25 $\mu\text{g/L}$ CYN took up $0.065\% \pm 0.015\%$ per exposure organism, which is similar to uptake reported for the same species exposed to 50 $\mu\text{g/L}$ [26]. These values are lower compared to the percentage of CYN per exposure organism (0.30%) reported in *L. minor* [15]. The low concentration of CYN remaining in the media and the amounts of free CYN in *E. densa* also support the possibility that the cyanotoxin could be adhering to plant cells components. However, the former studies, together with the data currently presented, do not confirm the association of CYN to proteins and need further investigation.

In general, CAT, POD, GST, and GR activity did not increase dose-dependently after 24 h of exposure to CYN. Increased H_2O_2 levels with time coincided with the increased CAT and POD activities at 2.5 and 25 $\mu\text{g/L}$ CYN. While CAT and POD activities correlated with CYN removal within 4 hours and 24 h, further increases of them can be linked with changes in the content of photosynthetic pigments and growth parameters as plant response to CYN exposure. The maintenance of high activities mainly of CAT during the experiment can be explained by the further CYN uptake at 336 h by *E. densa*.

Although H_2O_2 is involved in a range of biochemical reactions and signaling cascades necessary plant growth and is a constant byproduct of normal metabolism [43], excessive H_2O_2 needs to be eliminated from the cells. H_2O_2 is mainly formed in chloroplasts and mitochondria during electron transfer [43]. This study shows that concentrations of 2.5 $\mu\text{g/L}$ and 25 $\mu\text{g/L}$ CYN promote high CAT activities within 336 h, which is a link to the high levels of H_2O_2 within 336 h mainly at the highest CYN concentration. From the results, it can be inferred that this defense mechanism remains active against oxidative stress in which CAT plays a major role. Equivalent observations were obtained in a previous study with *L. minor*, in which CAT activity increased in treatments with 2.5 and 25 $\mu\text{g/L}$ CYN within 168 h [18]. CAT, as a member of the superoxide dismutase (SOD) system, acts as the first defense against oxygen toxicity [44]. SOD dismutates O_2^- to molecular oxygen and H_2O_2 , which is reclaimed by CAT [45,46]. Studies about CYN effects on SOD system are scarce, but it has been demonstrated that the activity of CAT and SOD are induced in aquatic plants by exposure to heavy metals [45,47]. In addition, in *E. densa* exposed to MCs, the activity of CAT and SOD increased after two days of exposure [22]. The levels of H_2O_2 remaining unchanged compared to the control after 48 and 168 h are explained by the enhancements of POD activity at the same time. POD is involved in phenolic metabolism expending H_2O_2 as substrate and plays an important role in the biosynthesis of lignin and how plants respond to pathogens [43]. This study shows high POD activities within 168 h, which together with

CAT can maintain H₂O₂ levels similar to control after 48 and 168 h. Flores-Rojas et al. [18] found that in *L. minor* exposed to CYN (0.025–25 µg/L), the activity of POD was not significantly promoted. In contrast to *Lemna* sp. [48], *E. densa* has higher percentages of lignin in stems and forms roots along its stems [49], which can transport nutrients and control homeostasis. Since *E. densa* is a submerged macrophyte, it is more exposed to CYN than the floating macrophyte *L. minor*, inducing more ROS production, which leads the antioxidative activation of POD and CAT.

GST and GR in *E. densa* tended to remain active after 24 and 168 h. An interesting result after 4 h of exposure to 2.5 and 25 µg/L CYN is lowered GST activity, accompanied by GR activity similar to control. This scenario suggests that GST is activated before 4 h as a response against acute stress, followed by an imbalance between GSH/GSSG [50]. The remaining activity of GR in the first hours can prove the recovery of GSH [50]. Both enzymes tend to maintain high activities between 24 and 96 h at the highest CYN concentration. Notably is the lower activity of GR together with higher GST activity after 48 h with 25 µg/L CYN. This indicates other imbalance between GSH/GSSG, in which enhancement of GSH can occur because of changes in the growth and concentration of photosynthetic pigments as strategies to stress tolerance [51]. Further high activities of GST and GR at 96 h tend to diminish the generation of ROS.

The outcomes of CYN exposure on the chlorophyll *a* and *b* contents presented in previous studies have been contradictory [14,20]. In *L. minor*, increased chlorophyll *a* and carotenoids concentrations resulted within 24 h at CYN concentrations under 25 µg/L [15]. *A. filiculoides* exposed to CYN experienced increased carotenoid levels [14]. In the current study, decreased chlorophyll *a*, chlorophyll *b*, total chlorophyll, and carotenoids contents were observed after 24 h mainly at 25 g/L CYN, which can be linked with higher H₂O₂ concentrations and higher activities of antioxidative enzymes. The slower metabolism of photosynthetic pigments, which also involves the production of H₂O₂ [43], could be a strategy to counteract the high levels of H₂O₂. This scenario remains after 48 h with a tendency to stabilize the chlorophyll *a* to chlorophyll *b* ratios. Subsequent stabilization of chlorophyll *a* and chlorophyll *b* levels, accompanied by enhancements of carotenoids and ratios of carotenoids to total chlorophyll at 96 h with 25 µg/L CYN may be vital for maintaining the complex chlorophyll-carotenoid binding proteins, which are needed to absorb and convert light energy during photosynthesis. The increased carotenoids levels after 96 h also suggest their activity in the antioxidative system response to combat excessive ROS formation promoted by CYN [52]. Subsequent decreases of the photosynthetic pigment levels but accompanied by balanced ratios of chlorophyll *a* to chlorophyll *b* and carotenoids to total chlorophyll after 168 h showed the stabilization of photosynthetic system in *E. densa* toward the end of the experiment. Changes at 2.5 µg/L CYN also show stabilization of photosynthetic system but in a shorter time.

Regarding the growth response, in two studies pertaining to *L. punctata* and *H. verticillata* [13,20], the relative growth was stimulated with exposure to CYN. In *A. filiculoides*, the increased growth in the presence of CYN was ascribed as being a defense strategy [14]. For *L. minor* exposed to 25 µg/L CYN, Flores-Rojas et al. [15] reported an increase in the number of fronds, below that of the control, which normalized after 14 days. Increased fresh weight in *E. densa* within the first two days, even at lower CYN concentrations, accompanied by a tendency to increase the shoot length in plants exposed to 2.5 and 25 µg/L CYN within the first 11 days, can be a strategy to combat toxicity. Growth also produces enhancements of ROS and the subsequent increased antioxidative enzymatic activity. Fresh weight gain within the first two days coincided with increased antioxidative enzymatic activities even to lower CYN concentrations (after 24 h) and changes in photosynthetic pigments levels mainly noted in plants exposed to 2.5 and 25 µg/L CYN. The subsequent fresh weight stabilization between 4 and 14 days, accompanied by the stimulation of shoot length within 11 days could be a reaction to maintain the balance of homeostatic processes in the plant. Additionally, decreases of the photosynthetic pigment levels maintaining their ratios after 336 h show a strategy to balance ROS, which is also generating by shoot length stimulation. After 336 h (14 days), *E. densa* reacted against CYN uptake again, as evident by the changes observed in growth parameters. Although decreases in fresh weight

were more evident within 28 and 32 days at 2.5 and 25 µg/L CYN, the stagnation after 14 days and decreases of shoot length until the end of the 32-days exposure seem to be a better indicator of changes due to the further CYN uptake after 336 h. The delayed appearance of new shoots with exposure to 25 µg/L CYN can indicate inhibition caused by CYN but at the same time a response toward ROS imbalance. Remarkably, chlorosis was observed during the experiment, which occurred after 14 days mainly in plants exposed to 25 µg/L; however, they remained alive during the 32-days exposure and forming small new leaves. Reduced plant vitality and biomass production can result from damage or inhibited biosynthesis of the photosynthetic pigments [53]. Although the photosynthetic system in *E. densa* tended to balance within 336 h, appearance of chlorosis in some shoots and not so robust plants after 14 days and until 32-days exposure can confirm the alteration of this. Changes in the morphological pattern of development in plants are one of the acclimation strategies in response to environmental stresses [51,53], which together with the biochemical and physiological responses can be mechanisms of resistance to CYN in *E. densa* within the fourteen first days. However, this study cannot confirm how long this aquatic plant maintains the regulation of its metabolism after a subsequent CYN uptake because this depends on the plant condition, CYN remaining concentration and exposure time. In aquatic ecosystems, high concentrations of CYN can be found even after months of an algal bloom [3], thus increasing the risk of affecting the macrophyte communities. The monitoring of biomarkers as CAT, carotenoids, and the ratios of chlorophyll *a* to chlorophyll *b* and carotenoids to total chlorophyll can elucidate the metabolism regulation and health condition of macrophytes.

5. Conclusions

In the presented study, CYN exposure at environmental concentrations did not cause bioaccumulation of the toxin in *E. densa*. The antioxidative enzymatic system remained active during the whole exposure period of 336 h with exposure to both 2.5 and 25 µg/L CYN exposure. In response to CYN exposure, the pigment concentrations decreased, but the ratios of chl *a/b* and carotenoids/total chl elevated for a short period tending to balance, probably to combat the adverse effects of the xenobiotic. The increase in the carotenoid content after 96 h suggests the intervention of these in the antioxidant system. These results indicate that *E. densa* can tolerate CYN toxicity. Moreover, stimulation of the fresh weight at the short time and shoot length within the first 14 days with 2.5 and 25 µg/L CYN support this evidence. However, this study also showed stagnation and inhibition after 14 days, and this could respond to the CYN uptake by *E. densa* after 336 h exposure. Although *E. densa* seems to be unaffected and even capable of combat adverse effects linked to CYN exposure for 336 h, it is necessary to investigate the responses of aquatic macrophytes to long exposures in order to elucidate the stagnation and inhibition growth and the possibility of plant health recovery. While the bioaccumulation of CYN was not found, the remotion of this toxin by *E. densa* over 65% and its moderate tolerance within 14 days apparently without lethal effects, open the possibility to consider *E. densa* as a candidate for sustainable phytoremediation, but CYN uptake needs further investigations. Even though the removal percentage exhibited by *E. densa* would not likely result to remediation of CYN in the environment to under the accepted guideline value of 1 µg/L, the macrophyte could contribute in a phytoremediation system comprising of several species, such as in the case of the Green Liver System, to form a comprehensive CYN remediation solution. According to the results, *E. densa* also showed changes in appearance and biomass due to exposure to CYN. In aquatic environments, these morphological changes can affect the communities of macrophytes depending on CYN concentrations and time of exposure. This study confirms that exposures to concentrations of 2.5 µg/L CYN and greater can be harmful to biota. The biochemical and physiological responses contribute to the understanding of the morphological changes and uptake abilities. In addition, measures of antioxidative enzymes and photosynthetic pigments can be used as biomarkers of plant health. Further studies with more macrophytes can make it possible to develop additional tools to control CYN in aquatic environments. *E. densa* proved to be sensitive to CYN exposure with apparent biochemical and morphological responses, but its use as bioindicator needs additional investigation,

particularly in mesocosm experiments. These experiments must be carried out taking into account the physical and chemical parameter changes in water and sediment, the light conditions, and the relationship between macrophyte species, e.g., using floating emergent and submerged macrophytes, simulating a natural environment under controlled conditions.

Author Contributions: Conceptualization, N.C.F.-R. and M.E.; methodology, N.C.F.-R.; validation, M.E.; formal analysis, N.C.F.-R.; investigation, N.C.F.-R.; resources, M.E.; data curation, N.C.F.-R.; writing—original draft preparation, N.C.F.-R.; writing—review and editing, M.E.; supervision, M.E.; project administration, M.E.; funding acquisition, M.E. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The authors thank Stephan Pflugmacher for the support during the conceptualization and Sandra Kühn for technical assistance in the laboratory. Administrative and technical support provided by Technische Universität Berlin. Open access funding provided by the University of Helsinki.

Conflicts of Interest: The authors declare no conflict of interest.

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